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Iron removal from raw asbestos by siderophores-producing *Pseudomonas*

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Abstract

Asbestos, mineral present in soil, are highly toxic due to the presence of iron. Microbes-mineral interactions occur naturally through various processes leading to their alteration. We examined the effect of siderophore-producing *Pseudomonas* with a particular focus on the role of pyoverdine and pyochelin on raw asbestos fibers such as amosite, crocidolite and chrysotile. We compared the efficiency of pyoverdine to the iron chelating agent EDTA in the release of iron from raw asbestos fibers. Pyoverdine was able to extract iron from all the tested raw asbestos with the higher efficiency observed for chrysotile and crocidolite. When asbestos were grinded, the iron removal was more important for all types. We monitored the effect of bacterial growth and siderophores containing bacterial supernatant on raw asbestos dissolution by solution chemistry analysis and transmission electron microscopy. The siderophore-containing supernatant allowed a higher iron solubilisation than the one obtained after bacterial growth. Moreover, the iron dissolution was faster with pyoverdine-containing supernatant than pyochelin-containing supernatant, with approximately the same iron level for the maximum extraction with a delay of 48 h. Our study clearly showed the involvement of bacterial siderophores, pyoverdine and pyochelin on chrysotile, crocidolite and amosite fibers weathering.

Keywords

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Iron-Asbestos-Weathering- Siderophore-*Pseudomonas*

1. Introduction

Asbestos has been extensively used for building constructions during the industrial exploitation in 1900's before it was banned in 1997 in France. The management of huge amount of wastes is a major concern today since asbestos-containing wastes represent an environmental and health problem. Asbestos fibers inhalation leads to various types of toxic effects in human such as inflammation and fibrogenesis of lung, mesothelioma or bronchogenic carcinoma. Asbestos are natural fibrous silicates belonging to serpentine and amphibole families. Chrysotile, the only member of the asbestiform serpentine is widely present among wastes (90-95%), while amphibole can be found in a minor extent compared to chrysotile in wastes. Among these, crocidolite and amosite are the two members of the amphibole, which have been found among the greatest commercial applications. The crystalline structures of fibers of amphiboles and serpentine are well described [1,2] and it explains the opposite chemical susceptibility towards acids or bases, e.g. chrysotile fibers are easily attacked by acidic condition while amphibole are in basic solution. The chrysotile structure is composed of a double layer consisting of a tetrahedral (silicate) sheet and a magnesium hydroxide octahedral sheet (brucite). This theoretical structure can vary if Fe^{2+} substitutes with Mg^{2+} in the brucite sheet. Up to 5% iron can be encountered in fibers since it is a relatively abundant element in many soils where asbestos natural rocks are originated. A double-tetrahedral silicate chain characterized the amphibole crystalline structure combined with a layer composed of ions mostly Mg^{2+}, Ca^{2+}, Na^{+}, Fe^{2+}, or Fe^{3+} (Table 1). Asbestos toxicity is correlated to the presence of iron as a catalyst of the Fenton reaction leading to the generation of free radicals and reactive oxygen species (ROS) [3,4].

Iron is an essential nutrient for most living organism to sustain their growth, although it is mostly unavailable in the environment. Iron is naturally abundant in soil, where it can be found in iron-bearing minerals such as goethite or ferrihydrite as iron oxides or clays for example. Therefore, many strategies have been developed by organisms such as fungi, plants or bacteria to have access to this element essential for their growth. Siderophore production, small molecules with a high affinity for Fe(III) produced in iron limited conditions, has been

described as one the common weathering process for fungi, plants and bacteria in various interactions with minerals [5]. For example, the dissolution of goethite mediated by the fungus *Suillus granulatus* [6] or various iron oxides solubilized by the phytosiderophore mugineic acid has been reported [7]. Kalinowski *et al*. [8] and Rosenberg and Maurice [9] reported hornblende and kaolinite dissolution rates depending on desferrioxamine-B, a bacterial siderophore that enhanced iron solubilisation. Asbestos, as a fibrous silicate mineral, is no exception to natural weathering by biological processes. Few studies investigated the biological alteration of raw asbestos. Indeed, telluric bacteria such as *Bacillus mucilaginosus* induced a mineral dissolution with an interestingly loss of crystallinity in the serpentine fibers along with a pH decrease, organic acids and ligand secretion [10]. Other bacteria isolated from asbestos rocks or soil from several indian mines, without species identification, decreased the iron content of asbestos rocks [11]. Further studies on these mine-isolated bacteria showed the production of siderophore which could be involved in the weathering process, however with no evidence on the structure of the produced siderophores [12]. Moreover, Rajkumar *et al*. [13] described the plant growth-promoting mechanisms involving siderophore-producing-bacteria in a serpentine soil where siderophore is one of the various strategies to remove nutrient elements essential for plant growth. Few siderophores demonstrated their ability to release iron from asbestos fibers. Indeed, iron removal by desferrioxamine siderophore evidenced from raw asbestos, on chrysotile fibers [14] and amphibole fibers such as crocidolite and amosite [15]. Siderophore-producing *Pseudomonas* are widespread bacteria in soil and known for mineral-weathering capacity. The genus *Pseudomonas* produces a wide variety of siderophores and more than 98 % of different fluorescent *Pseudomonas* isolated from soil, produce detectable siderophore under iron starvation [16]. The range of siderophore concentrations found in soil is wide, from tens of micromoles to a few millimoles per litre [17]. *Pseudomonas aeruginosa* is an ubiquitous fluorescent *Pseudomonas* found in various environment [18], whose siderophore production and iron acquisition are well known [19]. *P. aeruginosa* produces two endogenous siderophores, a more energy demanding high affinity molecule, pyoverdine (PVD) [20] and a low affinity siderophore, pyochelin (PCH) [21]. Pyoverdine is composed of three distinct structural parts i) a dihydroxyquinoline chromophore which confers the color and the fluorescence of the molecule ii) a peptide chain comprising 6 to 12 amino acids bound to its

carboxyl group iii) a small dicarboxylic acid (or its monoamide) connected amidically to the NH_2-group [22–26]. Pyoverdine chelates iron with a Ka of $10^{32}M^{-1}$ and in 1:1 stoichiometric ratio, via the catechol group of the chromophore and the two hydroxyornithines of the peptide moiety [27]. The structure of pyochelin is a derivative of a salicylic acid, which chelates iron with an affinity of $10^{28.8}M^{-2}$ [28] and a 2:1 (PCH : Fe^{3+}) stoichiometry : a tetra-dentate is provided by one molecule of PCH and a bi-dentate by the second PCH to complete the hexacoordinate octahedral geometry necessary for iron chelation [29]. Some studies were shown the ability of siderophore-producing *Pseudomonas* to interact with minerals. For example, *Pseudomonas mendocina* dissolved hematite or goethite, iron oxides, by iron mobilization from these minerals by bacterial metabolism [30]. Moreover, Ferret *et al*. [31] showed the ability of *P. aeruginosa* to solubilize structural iron from smectite, involving the pyoverdine and pyochelin in release process. Smectite is, a type of clay with silicate sheet structure, which structure share a common feature with asbestos minerals.

The aim of the present study was to investigate the bioweathering process of chrysotile or amphibole type by siderophore-producing *Pseudomonas*. Therefore, we compared the efficiency of pyoverdine to the chemically iron chelator, Ethylenediaminetetraacetic acid (EDTA), in the release of iron from raw asbestos. We examined the role of the *P. aeruginosa* siderophores, the pyoverdine and pyochelin in the asbestos alteration. This work provide a better understanding of the bacterial interaction with asbestos to further develop in the future a bioremediation process based on bacterial and/or siderophore alteration.

2. Experimental

2.1. Raw asbestos preparation

Three types of raw asbestos were used (chrysotile (CHR), crocidolite (CRO) and amosite (AMO), table 1) with or without manual grinding. All asbestos samples (0.125 g) were immersed in distilled water and sterilized by autoclave (121°C for 20 min). Samples were then incubated at 70°C for 14 days before experiments for a complete sterilization of materials. The heat removed any water allowing to weight samples with precision.

2.2. Pyoverdine purification

129 *P. aeruginosa* ATCC 15692 was grown in Luria Bertani medium (LB) for 15 h at 30°C under
130 shaking condition (220 rpm). 1 mL of LB culture was centrifuged (3 min at 9871 g). To
131 obtain iron starved cells, the iron-deficient succinate medium (composition in g L^{-1}: K_2HPO_4,
132 6.0; KH_2PO_4, 3.0; $(NH_4)_2SO_4$, 1.0; $MgSO_4.7H_2O$, 0.2; sodium succinate, 4.0; pH adjusted to
133 7.0 by addition of NaOH), sterilized by 0.2 µm filtration, was used [20]. The pellet, washed
134 twice with 1 ml of succinate medium, was suspended in 1 ml of succinate medium and
135 inoculated in 15 mL of sterile succinate medium, incubated at 30°C under shaking condition
136 (220 rpm). After 24 h of growth, 7.5 mL of this culture was transferred in an Erlenmeyer flask
137 containing 1 L of succinate medium. After 24 h of incubation, the culture was centrifuged
138 (2664 g for 40 min). Then, the supernatant was filtered twice with Whatman filter and once
139 on nitrocellulose filter (0.45 µm porosity). Siderophore containing supernatant was loaded,
140 after acidification (pH 6.0), on XAD-4 column, washed with two volume of purified water
141 and eluted with one volume of 50% ethanol. The green fraction was concentrated under
142 vacuum on a rotary evaporator and lyophilized.

143 2.3. Bacterial strains and growth conditions

144 We used the wild-type *P. aeruginosa* ATCC 15692 strain and two siderophore mutants,
145 PAO6297, a pyochelin-deficient strain (ΔPCH) [32] and PAO6382, a pyoverdine-deficient
146 strain (ΔPVD) [33] (Table 2). Bacteria were grown at 30°C overnight in LB broth medium at
147 200 rpm. For iron-depleted culture, cells were harvested from a LB pre-culture (24 h
148 incubation), washed twice in succinate medium and used to inoculate fresh succinate medium.
149 The cultures were incubated for 24 h under shaking (200 rpm). A second transfer in succinate
150 medium was done in the same condition. After 24 h, the culture was centrifuged (10 min at
151 9871 g) and the pyoverdine-containing supernatants were sterilized by filtration (0.22 µm
152 porosity). To prepare the bacterial suspension, cells were washed twice with sterile succinate
153 medium. Bacterial cell density was adjusted to OD_{600} at 0.05 corresponding to 1-4 x 10^7
154 CFU/ml.
155

156 2.4. Dissolution of iron from asbestos fibers

157 Dissolution of iron from asbestos fibers by EDTA or purified pyoverdine produced by
158 *Pseudomonas aeruginosa* ATCC 15692 was compared. Both chelators were prepared in
159 succinate media at a final concentration of 200 µM. Asbestos samples of CHR, CRO and
160 AMO, grounded manually or not, were spiked into 20 mL of each chelator. Assays were
161 incubated at 30°C under shaking condition (220 rpm) during 48 h. After this first cycle

162 contact, the assays were centrifuged 30 min at 9871 g to recover supernatants. In order to
163 continue alteration, two others cycles were repeated during 48 h and 96 h in the same
164 conditions. Each assay was done in triplicate and the results are the mean of triplicate.

165 Dissolution of iron from asbestos fibers by bacteria or bacterial supernatants was done at
166 30°C under shaking conditions (220 rpm) with the 3 types of grinded asbestos. Pyoverdine-
167 containing supernatant and bacterial cell suspension were prepared as described on paragraph
168 2.3. After a first cycle of 48 h alteration, assays were centrifuged 30 min at 9871 g. For each
169 renewal, 20 ml of siderophore-containing supernatant or a bacterial suspension of OD_{600} at
170 0.05 were added to respective samples. Renewal cycles of 48 h were performed twice for both
171 conditions. Every 48 h cycle, bacterial number was determined by serially diluted and plated
172 on LB plates incubated at 30°C for 24 h. All supernatants were sterilized by filtration (0.22
173 µm porosity). Each assay was done in triplicate and the results are the mean of triplicate.
174 Samples of raw crocidolite incubated with bacteria and its abiotic control were observed by
175 transmission electron microscopy at the end of the experimental cycles.

176 In order to determine iron content in supernatant, solutions were acidified to 1% with nitric
177 acid and iron was determined by atomic absorption spectroscopy (Perkin-Elmer AAnalyst
178 400).

179

180 2.7. STEM-EDX of asbestos fibers

181 TEM (Transmission Electron Microscopy) images were recorded with a JEOL 2100
182 microscope, with a 200 kV potential applied on a LaB6 filament as an electron source.
183 Resolution of the TEM is 0.21 nm. TEM mapping was performed in STEM (Scanning
184 transmission electron microscopy) mode (resolution 2 nm) and using an SSD-EDX (Silicon
185 Drift Detector-Energy Dispersive X-ray) spectrometer to determine the chemical composition.

186

187 2.8. Statistical analysis

188 Differences of iron concentrations after pyoverdine and EDTA treatments solubilized from
189 asbestos fibers were determined using Kruskal-Wallis test (RStudio v1.0.153) followed by a
190 Conover post-hoc analysis.

191

192 **Results and discussion**

193

194 **1.1. Comparison of pyoverdine and EDTA treatments**

195 For the three asbestos, chrysotile, crocidolite and amosite, iron was released in the presence of
196 the iron-complexing agents, pyoverdine or EDTA at each renewal cycle. Whichever the
197 asbestos family, serpentine (chrysotile) or amphiboles (Amosite, Crocidolite), the iron release
198 was possible even if the crystalline structure was very different (Fig. 1). Our result showed
199 that the bacterial siderophore pyoverdine produced by fluorescent *Pseudomonas* was able to
200 remove iron from asbestos fibers. During the three incubation period, the bacterial
201 siderophore pyoverdine extracted more iron than the chemical chelator EDTA for chrysotile
202 (0.38 to 1.04 mg/L and 0.08 to 0.19 mg/L respectively) (Fig. 1A-B) and crocidolite (0.59 to
203 3.84 mg/L and 0.12 to 1.92 mg/L respectively) (Fig. 1C-D). Unlike the last two asbestos,
204 pyoverdine and EDTA revealed the same efficiency towards amosite fibers (1.07 to 3.58 and
205 0.82 to 3.64 mg/L respectively) (Fig. 1E-F). Differences between pyoverdine and EDTA
206 efficiency may be explain by their affinity and specificity for iron. Indeed, pyoverdine
207 chelates iron with an affinity of 10^{32} M^{-1} while EDTA has a lower Ka of 10^{20} M^{-1} [34]. When
208 asbestos were grinded, iron weathering increased in the presence of EDTA or pyoverdine with
209 an efficiency ranging from 1.3 times and 2 times for chrysotile and amosite respectively and 2
210 to 5 times for crocidolite (Fig. 1). Grinding lead to the amorphization of chrysotile with
211 probably the direct OH bonds between the silica and brucite sheets destroyed or opened [35].
212 Moreover, grinding may also increase the exchange surface capacity between pyoverdines
213 and asbestos, improving the extraction and accessibility of iron release. Similar results were
214 obtained by Chao *et al*. [15] where EDTA or citrate were also able to extract iron from
215 crocidolite or amosite but to a lesser extent that the bacterial siderophore desferrioxamine.
216 Werner *et al.* [36] observed the same findings on crocidolite under 30 days of contact with the
217 ranking desferrioxamine > EDTA > citrate. Ferrozine, an iron(II) chelator, was able to extract
218 iron from crocidolite and amosite, while the iron extraction from chrysotile was only possible
219 when ascorbic acid was supplemented [14]. Recently, Desferrioxamine and ferrichrome
220 extracted iron for raw chrysotile at the same level but with no synergy when organic acids
221 were supplemented or tested alone [37].

222

223 **1.2. Dissolution of iron by siderophore-producing bacteria**

We have shown that pyoverdine was able to withdraw iron from the three commonly found raw asbestos fibers. Since asbestos were extracted from soils, like numerous telluric microorganisms such as fluorescent *Pseudomonas*, we wanted to evaluate the role of *P. aeruginosa* compared to potential metabolites produced during an iron-starved culture in the release of iron. Therefore, asbestos samples were incubated with cells or bacterial supernatant of *Pseudomonas* producing only pyoverdine, only pyochelin or producing both endogenous siderophores. Removal of iron was measured after three successive contact cycles of 48 h. The amount of iron released in the presence of bacterial supernatants at any given cycle was higher than in the presence of bacterial cells for chrysotile and crocidolite (Fig. 2A, B, C, D). During the first 48 h contact cycle (T48) with chrysotile (Fig 2A), a rapid release of iron was observed for the wild-type and the ΔPch pyoverdine-containing supernatant, which dissolve iron 2 to 3 times more efficiently than the ΔPvd supernatant, producing only pyochelin. For the second cycle (T48-2), results are reversed since the ΔPvd supernatant extracted twice as much iron than the WT and ΔPch supernatant for chrysotile, results are also valid for crocidolite. Interestingly, for chrysotile and crocidolite, pyochelin containing supernatant at T48-2 cycle extracted the same iron amount than the pyoverdine containing supernatant at T48 but required twice as much time. For the last cycle (T48-3), the same tendency is noticed for chrysotile between the 3 supernatants but to a lesser extent than the T48-2. For crocidolite in the T48-3 cycle, the maximum iron content (0.78 mg/L) was observed for the WT supernatant compared to previous cycles (0.41 and 0.34 mg/L for T48 and T48-2). For amosite, a very low efficiency for all the supernatants was noticed between 0.04 and 0.17 mg/L. In the presence of bacteria, only the ΔPch strain (0.28 mg/L) seemed to be more effective in iron removal at the T48-3 cycle. No changes were observed in culture medium alone or culture medium containing either fibers or bacteria (Fig. 2.). Our results showed that iron dissolution is more rapid and important for chrysotile compared to amphibole. Such differences can be explained by crystal structure of asbestos and the iron-binding sites [38]. Indeed, in the chrysotile structure the octahedral layer (brucite) magnesium can be substituted by iron while in the tetrahedral layer, silicon may be rarely replaced by iron. In amphibole structure, iron is bound in cationic sites present in two ribbons of silicate tetrahedra. Therefore, iron is more easily available in chrysotile (brucite outer layer) compared to amphibole whose iron is trapped in two silicate layers.

Together with these high affinity iron chelators produced in the supernatant, physiological organic acids could be also encountered during bacterial growth, which may have a

258 synergistic effect in iron release [39]. Another described parameter is the grinding which
259 enhanced iron dissolution [40,41]. In our study, considering that all asbestos fibers were
260 manually grinded, this parameter could have influenced the iron solubilisation, since it was
261 difficult to obtain homogeneous grinding samples between experiments. As a consequence,
262 lower iron concentrations were observed for supernatant-treated crocidolite and amosite
263 compared to the purified pyoverdine treatment. It is well known that siderophore-mediated
264 iron removal involve siderophore adsorption on the iron-bearing mineral surface [5].
265 Therefore, the grinding may increase the specific surface area of asbestos and improve
266 accessibility of siderophores to iron on the surface. Moreover, the presence of organic acids in
267 the supernatant could have influenced the chrysotile dissolution since the amount of dissolved
268 iron at 48 h (2.54 mg/L) in the supernatant is higher compared to the purified pyoverdine
269 dissolution test (0.83 mg/L, Fig. 1B). At the second renewal, the same influence was noticed
270 with values three times higher for the supernatant (Fig. 2A) compared to the purified
271 pyoverdine (Fig. 1B). Differences could not be due to (i) pyoverdine degradation since this
272 molecule has been shown to be stable in those conditions (ii) pyoverdine concentration since
273 it was tested at the same level between the purified and the one synthesized by cells after 48
274 h.
275 Our results confirm that *Pseudomonas* was able to promote mineral dissolution concomitantly
276 with siderophore production. Compared with data related to asbestos-fungi alteration, few
277 researches were focused on asbestos-bacterial interactions. As an example of Gram-positive
278 bacteria, *Bacillus mucilaginosus* was able to accelerate serpentine powder thanks to organic
279 acids and ligands [10]. Bhattacharya *et al.* [11] isolated Gram positive and Gram negative
280 bacteria from Indian asbestos contaminated soil or asbestos rocks which induced a decrease in
281 the iron content of asbestos after bacterial weathering. Further research, focused on the Gram
282 positive previously isolated strains, revealed the production of siderophores from these
283 bacteria, with no precise identification of the produced compounds [12]. Bacterial
284 siderophores produced by *Staphylococcus* mines isolates [11,12] were also implicated in iron
285 removal from asbestos rocks, together with desferrioxamine from crocidolite fibers [42]. This
286 strategy to overcome iron limitation is also well documented for soil organisms since several
287 fungi have been reported to be involved in iron sequestration [43]. Bioweathering abilities of
288 chrysotile fibers revealed different efficiency depending on the fungi genus. Fungi such as
289 *Fusarium oxysporum* released also in the extracellular medium chelators able to extract iron
290 from both serpentine (chrysotile) and amphibole (amosite and crocidolite) asbestos fibers

[42]. Asbestos seems to be an iron source for various organisms described so far through the siderophore strategy.

1.3. Evidence of biofilm formation on asbestos fibers

In the experiment of iron dissolution by siderophore-producing bacteria, we observed that all raw asbestos incubated with bacteria were agglomerated, leading to a compact cluster, whereas the fibers resulting from deterioration in the abiotic medium remained free. In order to determine the impact of biotic condition on the material, fibers were observed by TEM (Fig. 3). As an example, crocidolite fibers incubated with *Pseudomonas aeruginosa* wild type or in the succinate medium after 48 h were analyzed. The analysis of the alterated fibers in abiotic conditions showed no detectable alteration layer. In biotic condition, fibers are covered with a biofilm explaining the agglomeration of the material. *Pseudomonas* are well known to form biofilm, which is a surface-attached bacteria embedded in a self-produced matrix of extracellular polymeric substances (EPS) that are adherent to each other and/or a surface exopolysaccharides (Fig. 3). Adhesion to surfaces has been seen as a survival strategy under energy limitation [44]. Weathering of rocks is a complex process where biofilm are able to accelerate or inhibit the alteration [45]. For example, *Pseudomonas aeruginosa* has already been described to form a biofilm on municipal waste incinerator bottom ash cementing grains together [46] or on smectite, a type of clay [31]. In leachates experiments of bottom ashes, analysis of iron concentration in solution increase in the presence of bacteria and bottom ash, while the concentration of others elements such as Ni and Zn was higher in abiotic condition compared to the biotic medium. This could be a consequence of chemical modification of the solution by bacterial development since reducing conditions together with the production of organic acids and siderophores may occur. Siderophore-promoted dissolution of iron-bearing minerals has been clearly documented and the study of Ferret *et al.* [31] showed an increase in the dissolved silicon, iron and aluminum concentrations following smectite supplementation in the presence of *Pseudomonas*. In an asbestos cement pipe in drinking water distribution system, a biofilm was evidenced containing metallic cations leading to decrease of the thickness of the pipe wall linked to the loss of hydrated cement matrix. A consequence of this bacterial activity may cause a pipe failure [47]. In our experiments, the analysis of crocidolite fibers by STEM-EDX showed a slight decrease of iron content after 48 h with bacterial contact. According to results in figure 2, the bacterial supernatant of the wild type *P. aeruginosa* allowed to deplete iron

more efficiently from fibers compared to bacterial cells. These results confirmed the active dissolution driven by the siderophore-mediated and/or bacterial process (Fig. 4).

4. Conclusions

We examined the potential bioweathering ability of siderophore-producing *Pseudomonas* and their produced siderophores on raw asbestos fibers. We clearly showed that biological chelating agents are more efficient in iron extraction than the best known chemical chelating compound EDTA. In addition, iron dissolution was more effective when fibers were manually grinded. Our results demonstrated that *Pseudomonas* and their produced siderophores, pyoverdine and pyochelin, were able to dissolve iron from asbestos fibers but not at the same efficiency. Siderophores containing supernatants extracted more iron than the bacterial cells after asbestos contact. A biofilm covering fibers was evidenced during bacterial growth and the iron content in fibers decreased after bacterial contact. Moreover, pyoverdine containing supernatant dissolved more rapidly iron from chrysotile and crocidolite fibers than pyochelin containing supernatant. Our study clearly demonstrated that either siderophores or *Pseudomonas* cells were able to weather the three types of asbestos fibers. Our research has shown that asbestos can serve as a source of nutrients for soil bacteria. Further investigations are required to extend these experiments on asbestos wastes in order to develop a bioremediation process based on bacterial and/or siderophore alteration.

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504 FIGURE LEGENDS:

505

506 **Figure 1:** Iron concentration dissolved from raw asbestos grinded or not in the presence of
507 EDTA (200 µM) (grey bars) or purified pyoverdine (200 µM) from *Pseudomonas aeruginosa*
508 ATCC 15692 (black bars) after two 48 hours-contact time and a 96 hours contact time under
509 shaking (200 rpm) at 30°C. Iron was monitored by atomic absorption spectroscopy in the
510 absence or in the presence of asbestos (0.125 g). Raw asbestos were designed as chrysotile
511 (CHR) or grinded chrysotile (CHRb), crocidolite (CRO) or grinded crocidolite (CROb),
512 amosite (AMO) or grinded amosite (AMOb). Error bars indicates standard deviation over
513 mean of triplicate experiments. Groups without significant different iron amounts were
514 indicated by a, b, and c.

515

516 **Figure 2:** Iron concentration measured in the supernatant after bacterial growth (B,D, F) or in
517 the presence of each bacterial strain's supernatant (A, C, E) after 3 renewal cycles of 48 H
518 with raw chrysotile (A, B), raw crocidolite (C, D) and raw amosite (E, F) at 30°C. The tested
519 strains were the wild-type *Pseudomonas aeruginosa* ATCC 15691 (PAO1, red bars),
520 pyocheline-deficient mutant (□PCH, green bars) and pyoverdine-deficient mutant (□PVD,
521 blue bars). Error bars indicates standard deviation over mean of triplicate experiments.

522

523 **Figure 3:** STEM images of bacterial biofilm associated to crocidolite fibers (B) compared to
524 control fibers (A). Crocidolite fibers were incubated with the wild-type *Pseudomonas*
525 *aeruginosa* in succinate medium or succinate medium without bacteria during 48 h under
526 shaking condition (220 rpm). Scale bar corresponds to 500 nm.

527

528 **Figure 4:** STEM images and STEM mappings of crocidolite fibers before contact (a and b),
529 after bacterial growth (c and d) and after incubation with bacterial supernatant (e and f) during
530 48 h under shaking conditions (220 rpm). Large images obtained from the combination of the
531 three distributions of Mg, Si, and Fe with analysis areas (a, c and e). Atomic ratios of Mg/Si
532 and Fe/Si (b, d and f). Mass percentage of iron, magnesium and silicium before and after
533 contact of raw crocidolite with *P. aeruginosa* PAO1 or bacterial supernatant (g).

534

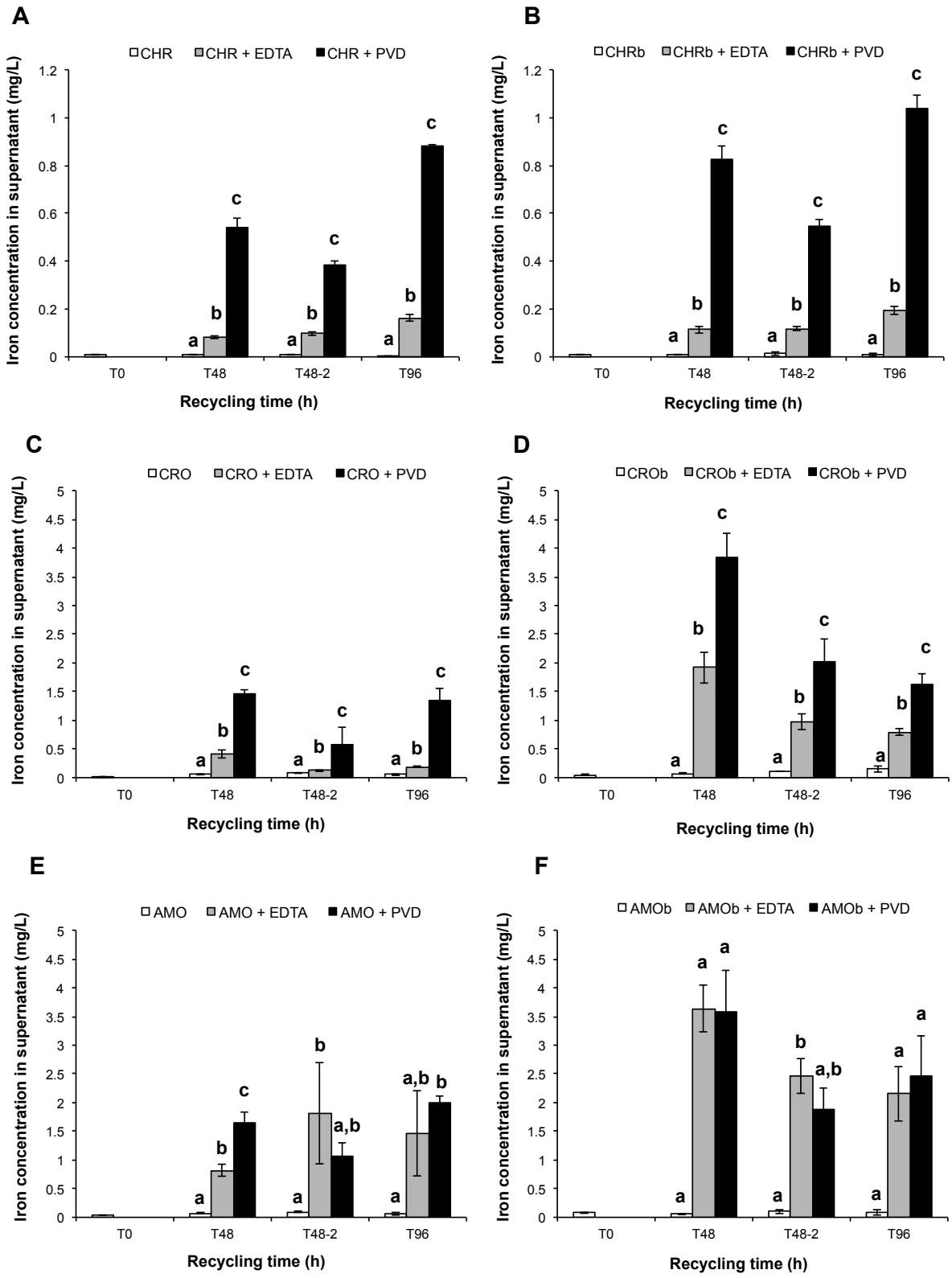
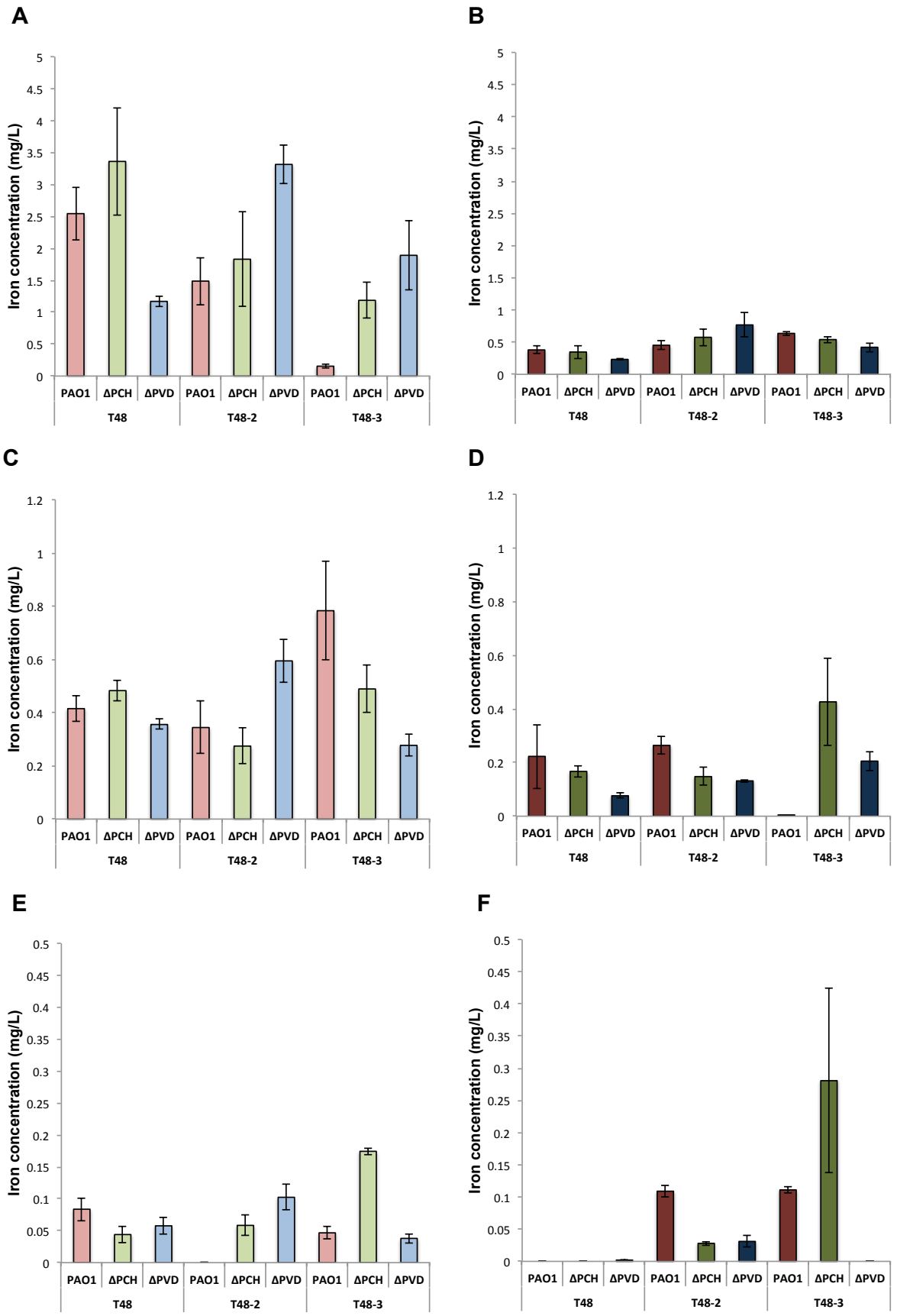


Figure 1: Iron concentration dissolved from raw asbestos grinded or not in the presence of EDTA (200 µM) (grey bars) or purified pyoverdine (200 µM) from *Pseudomonas*

aeruginosa ATCC 15692 (black bars) after two 48 hours-contact time and a 96 hours contact time under shaking (200 rpm) at 30°C. Iron was monitored by atomic absorption spectroscopy in the absence or in the presence of asbestos (0.125 g). Raw asbestos were designed as chrysotile (CHR) or grinded chrysotile (CHRb), crocidolite (CRO) or grinded crocidolite (CROb), amosite (AMO) or grinded amosite (AMOb). Error bars indicates standard deviation over mean of triplicate experiments. Groups without significant different iron amounts were indicated by a, b, and c.


A
B
C
D
E
F
Iron concentration (mg/L)
PAO1
ΔPCH
ΔPVD
T48
T48-2
T48-3

5
6 **Figure 2:** Iron concentration measured in the supernatant after bacterial growth (B,D,
7 F) or in the presence of each bacterial strain's supernatant (A, C, E) after 3 renewal
8 cycles of 48 H with raw chrysotile (A, B), raw crocidolite (C, D) and raw amosite (E, F) at
9 30°C. The tested strains were the wild-type *Pseudomonas aeruginosa* ATCC 15691
10 (PAO1, red bars), pyochelin-deficient mutant (ΔPCH, green bars) and pyoverdine-
11 deficient mutant (ΔPVD, blue bars). Error bars indicates standard deviation over mean
12 of triplicate experiments.
13
14 **In color print**

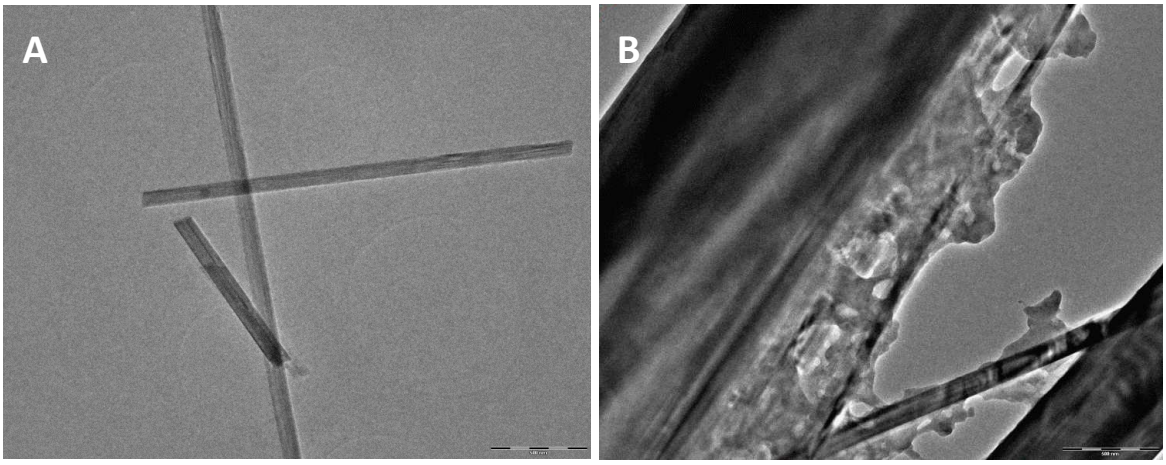
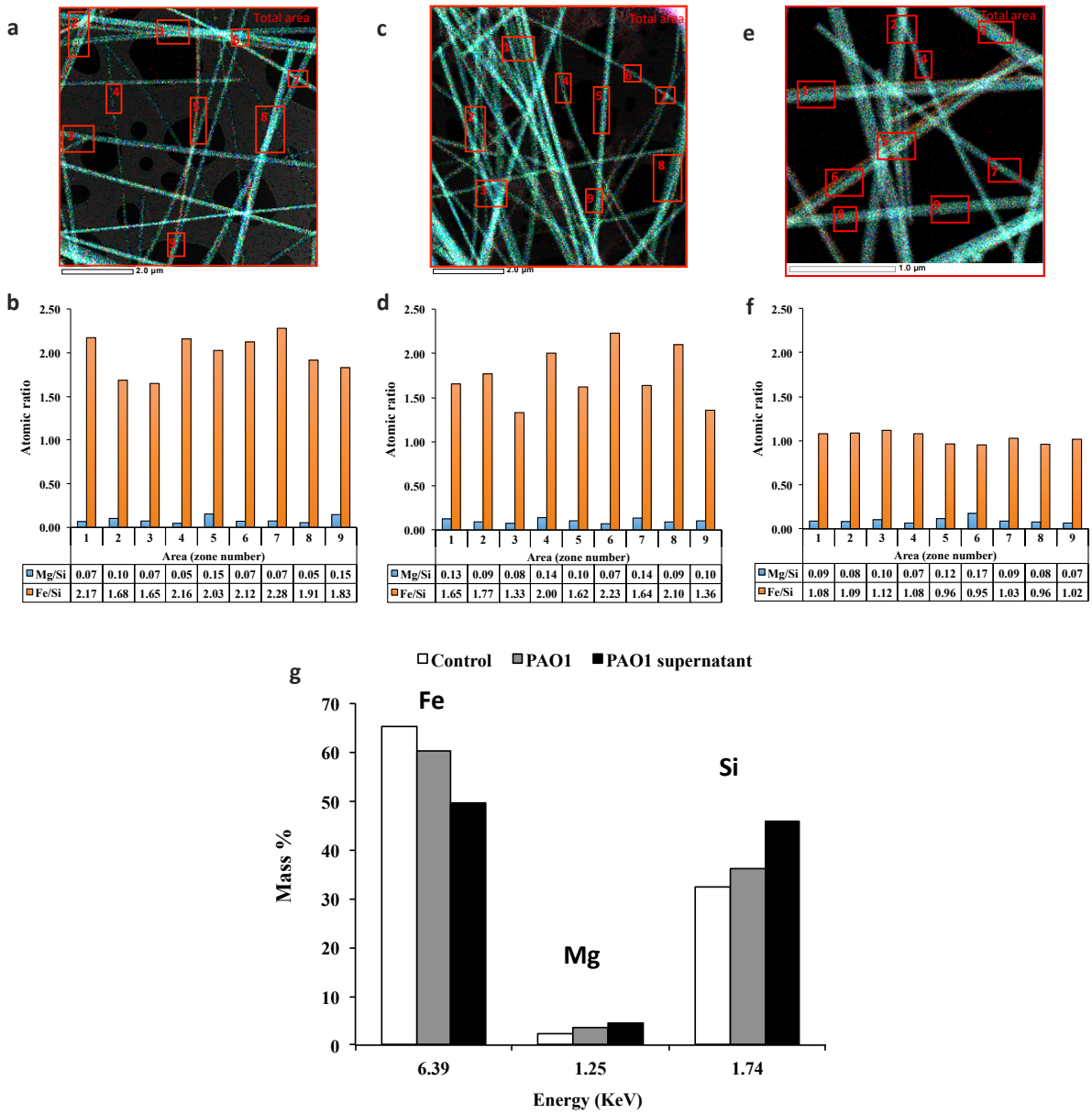


Figure 3: STEM images of bacterial biofilm associated to crocidolite fibers (B) compared to control fibers (A). Crocidolite fibers were incubated with the wild-type *Pseudomonas aeruginosa* in succinate medium or succinate medium without bacteria during 48 h under shaking condition (220 rpm). Scale bar corresponds to 500 nm.



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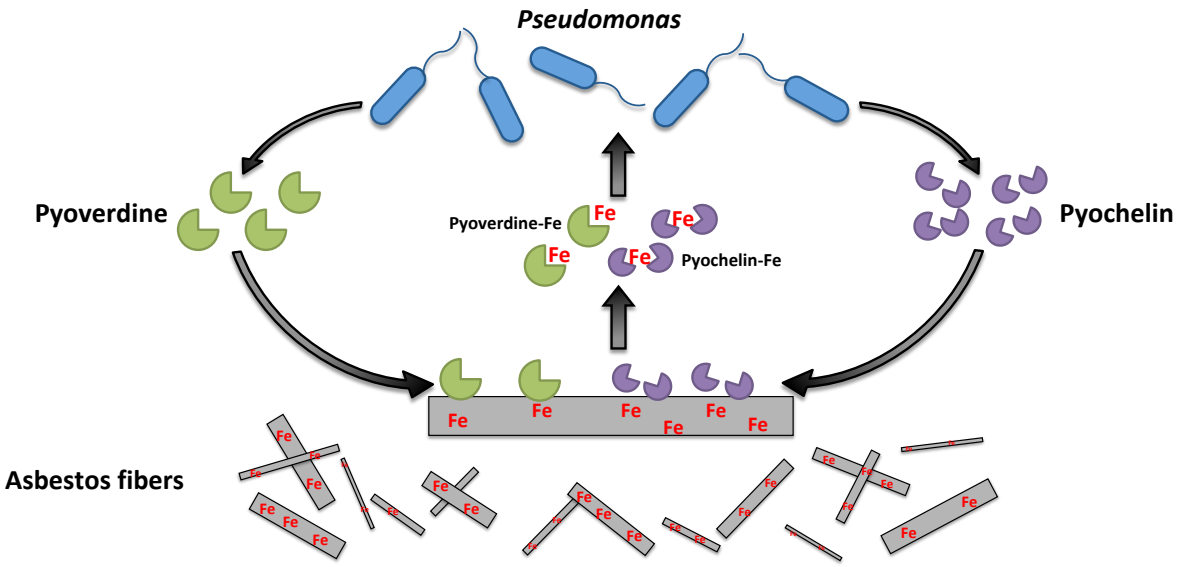
In color print

Table 1. List and composition of asbestos used.

Name	Composition	Source
Chrysotile	$Mg_6Si_4O_{10}(OH)_8$	Natural History Museum of Paris
Crocidolite	$Na_2(Fe^{3+})_2(Fe^{2+})_3Si_8O_{22}(OH)_2$	mine South Africa « Wangu Hill, Kwagulu, province of Natal Woumo
Amosite	$(Fe, Mg)_7Si_8O_{22}(OH)_2$	Penga mine- South Africa, province of Impopo

1 **Table 2.** List of the strains used.
2

Name	Relevant characteristics	Reference or source
***P. aeruginosa* strains**		
PAO1	Wild-type	ATCC15692
PAO6297	Δ*pchBA*	[32]
PAO6382	ΔpvdF	[33]

3
4

Graphical Abstract