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Partial glucocorticoid resistance in the pathophysiology of adrenal

2 cortex hyperplasia.

- 3 Géraldine VITELLIUS^{1,2} and Marc LOMBES^{1,3}
- 4 ¹INSERM UMR_S U1185, Fac Med Paris-Sud, Univ. Paris-Sud, Université Paris-Saclay, Le
- 5 Kremlin Bicêtre, F-94276, France;
- ²Service d'Endocrinologie-Diabète-Nutrition, Hôpital Robert Debré, CHU Reims, F-51100,
 France;
- ⁸ ³Service d'Endocrinologie et des Maladies de la Reproduction, Hôpitaux Universitaires Paris
- 9 Sud, AH-HP, CHU Bicêtre, F-94275, Le Kremlin Bicêtre, France.
- 10 Address all correspondence and requests for reprints to:
- 11 Dr Marc Lombès, MD, PhD, Inserm U1185, Faculté de Médecine Paris-Sud,
- 12 63, Rue Gabriel Péri, F-94276 Le Kremlin Bicêtre Cedex France.
- 13 E-mail: marc.lombes@u-psud.fr.
- 14
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19 Abstract

The pathophysiology of Bilateral Adrenal Hyperplasia (BAH) remains obscure. Even though 20 several genetic causes have been associated with this disease, often related to 21 overactivation of the cAMP signaling pathway, cortisol regulation by autocrine adrenal ACTH 22 production has also been proposed to enhance corticosteroidogenesis, cortisol secretion and 23 24 adrenal cortex growth. Recently, heterozygous glucocorticoid receptor (GR) loss-of-function mutations have been discovered in patients with BAH, highlighting a potential new genetic 25 cause of adrenal hyperplasia, as previously reported in a mouse model of GR 26 27 haploinsufficiency. However, the molecular mechanisms linking adrenal hyperplasia and GR haploinsufficiency need to be further elucidated. 28

29 1 Introduction

Morphological criteria to define adrenal hyperplasia remain scarce. Even though the 30 prevalence of adrenal mass incidentally discovered during imaging or adrenal incidentaloma 31 (AI), varies between 1-8% following autopsy studies, it seems to increase with age (1). A 32 large number of AI are nowadays discovered and frequently referred to a clinical 33 endocrinologist. Recent guidelines (2) define benign AI with strict radiological criteria (3). 34 35 Among AI, 15% are bilateral but mainly non-functional (1). Diagnosis of Bilateral Adrenal Hyperplasia (BAH) may be considered easy. However, the definition of normal adrenal 36 glands is still controversial. Initially, normal adrenal glands were defined as a right lateral and 37 medial limb measurement at 2.8 cm, with the left medial limb at 3.3 cm and lateral at 3.0 cm 38 (4). More recently, adrenal volume assessed by computerized tomography (CT) was used to 39 define normality (5). Left adrenal gland volume was estimated at 4.5 \pm 1.6 cm³ while the 40 volume of the right adrenal gland was 3.8 ± 1.3 cm³. Volume measurements appear to be 41 more reproducible among investigators compared with linear measurements. The 42 43 association between BAH and steroid excess facilitates diagnosis and is important to recognize given the potential increased morbidity and mortality. Hypercortisolism occurs in 44 12% of AI, while hyperaldosteronism or catecholamine excess is observed in 2.5% and in 7% 45 46 of cases, respectively (2). Metabolic disorders (glucose intolerance, diabetes, hypertension, 47 dyslipidemia and osteoporosis) can be associated with AI and subclinical hypercortisolism (2). Malignancy was described in 3% of cases (1) and congenital adrenal hyperplasia in 21% 48 49 (6). A new nomenclature of BAH associated with Cushing' syndrome was recently published (7). When the nodule size is <1 cm, thus referring to micronodular adrenals, the main cause 50 51 is primary pigmented nodular adrenal disease (PPNAD). However, if their size exceeds 1 cm, 52 primary bilateral macronodular adrenal hyperplasia (PBMAH) should be considered (8).

In this review, we will discuss the pathogenesis of BAH in the context of subclinical 53 hypercortisolism, highlighting the recently discovered NR3C1 mutations as recently 54 Nicolaides 55 discussed by & Chrousos (9). Adrenal pheochromocytomas or hyperaldosteronism, carcinoma, metastasis or etiologies of unilateral AI were excluded. 56

58 2. Adrenal hyperplasia.

59 **2.1. Physiology of adrenal cortex**

Cortisol release is under the control of hypothalamic corticotropin releasing hormone (CRH). 60 As illustrated in Fig. 1, CRH stimulates corticotrope cells of the anterior pituitary, which 61 secrete adrenocorticotropic hormone (ACTH). ACTH is one of the most potent physiological 62 modulators of steroidogenesis and binds to its 7-transmembrane G protein-coupled receptor 63 64 (GPCR) MC2R, expressed in all zones of the adrenal cortex. ACTH binding to MC2R, in association with melanocortin-2 receptors accessory proteins (MRAP), activated Gs protein, 65 followed by activation of adenylyl cyclase, enhancing intracellular cAMP concentration (Fig. 66 2), thus stimulating Protein Kinase A (PKA) signaling. PKA is a hetero-tetramer composed of 67 68 2 catalytic subunits, encoded by three different catalytic subunits of which C α predominates in the adrenal gland plus 2 regulatory subunits, encoded by four genes (R1 α , R1 β , R2 α and 69 70 R2B) (10). Binding of 4 cAMP molecules to PKA regulatory subunit dimers allows release and activation of the 2 PKA catalytic subunits, which phosphorylate several targets, such as 71 cAMP response element (CRE) binding protein (CREB), or steroidogenic factor 1 (SF1), 72 73 regulating StAR function. Phosphodiesterase, involved in cAMP degradation, act as negative regulators of this pathway. 74

StAR is the first step of steroidogenesis, involved in cholesterol flux into the inner
mitochondrial membrane (11). Cortisol, the final steroidogenic compound, exerts a negative
feedback loop on hypothalamic CRH and pituitary ACTH (Fig.1).

ACTH also plays a major role in the vasculature development *via* local synthesis of VEGF
and in adrenal tissue growth. ACTH may be locally produced by medullary chromaffin cells,
thus playing a major autocrine/paracrine role in the adrenal gland (12). Activation of
cAMP/PKA signaling pathway may lead to hypercortisolism (13). A functional link between
PKA and Wnt pathways was reported, as demonstrated by a cell-specific PKA-activated
downregulation of β-catenin phosphorylation in H295R cells (14).

85 2.2. Genetic abnormalities

Every enzyme or protein involved in these pathways could be mutated and responsible for
overactivation of signaling cascades resulting in hypercortisolism and adrenal hyperplasia.
(summarized in Table 1).

89 2.2.1 PPNAD

PPNAD, an autosomal dominant manner disorder, is often observed in patients with 90 91 Carney's complex. Seventy percent of index cases are familial, while the remaining cases 92 are *de novo* germline mutations (15). One hundred and thirty-five pathogenic mutations have been reported to date. PPNAD is mainly related to germline inactivating mutations of the 93 PRKAR1A gene, associated with constitutive activation of PKA (16), or phosphodiesterases 94 PDE11A and PDE8B mutations (17). PPNAD can be associated with hypercortisolism 95 leading to ACTH-independent Cushing syndrome. Of note, the glucocorticoid receptor (GR) 96 was shown to be overexpressed in PPNAD nodules, underlying paradoxical dexamethasone 97 98 (DXM) induction of cortisol secretion (18).

99 **2.2.2 PBMAH**

Expression of illicit receptors in the adrenal glands (GIP, LH/HCG, vasopressin, 100 101 catecholamines, serotonin 5 HT, angiotensin II or glucagon) was first identified as a leading 102 cause of regulatory mechanism of cortisol production in PBMAH. The prevalence of these illegitimate membrane receptors is rather high (up to 77-87% of cases), leading to 103 cAMP/PKA pathway activation and enhanced corticosteroidogenesis (19). Intra-adrenal 104 ACTH secretion responsible for an autocrine/paracrine regulation in cortisol production, was 105 106 recently reported in PBMAH (20). Activating MC2R mutations (19), or point mutations of the 107 α subunit types of G proteins responsible for altered GTPase activity (21) leading to constitutive activation of the cAMP/PKA pathway, have been also described less frequently 108 109 in PMBAH. Recently, a new tumor suppressor gene, inactivation of Armadillo Repeat Containing Family (ARMC5), was discovered after whole genome sequencing of 26 patients 110 with corticotropin-independent macronodular hyperplasia (22). ARMC5 was described in 111

familial and sporadic forms of PBMAH, but the mechanism by which ARMC5 was 112 responsible for hypercortisolism and adrenal hyperplasia remains unknown (23). Recently, 113 114 Lecog et al (24), identified somatic duplications in the 19g13.32 chromosome region in three 115 patients. These genetic rearrangements lead to a new genomic environment in which cisregulatory regions, containing glucocorticoid response elements of the ZMYND8 gene, drive 116 ectopic expression of glucose-dependent insulinotropic polypeptide receptor (GIPR) in two 117 GIP-dependent cortisol secreting adenomas and one adrenal tumor with PBMAH. PBMAH 118 119 was also described in genetic syndromes, as multiple endocrine neoplasia type 1, or APC syndrome (1). 120

121 2.2.3 Mutations of steroidogenesis players

Mutations of important players, either transporters (StAR) or enzymes (CYP11A1, HSD3B2, CYP17A1, POR, CYP11B1 and CYP21A2) involved in steroidogenesis, are responsible for adrenal hyperplasia associated with a cortisol secretion defect and high levels of ACTH.

125

126 3. Glucocorticoid resistance

127 **3.1. Definition**

Glucocorticoid hormones (including Cortisol) regulate various biological functions involved in 128 129 development, metabolism, inflammatory processes and stress. Their actions are mediated 130 mainly by GR, a member of the nuclear receptor superfamily, which is an intracellular receptor protein acting as a ligand-activated transcription factor. Human GR (hGR) is 131 132 encoded by the NR3C1 gene (MIM#138040). GR may exist in multiple isoforms generated by alternative splicing: hGRa, hGRβ, hGRγ, hGR-A and hGR-P. Eight alternative initiation sites 133 into hGRα have been previously described (11, 12), leading to distinct GR protein variants 134 135 (GRa-A to D3). GRa-A results from translation of GRa mRNA from the first AUG codon (Met¹) while the shorter GR α -B results from translation beginning from the second AUG 136 codon (Met²⁷). Even though numerous GR variants have been discovered by next generation 137 sequencing (NGS) methods (27), only twenty-eight NR3C1 loss-of-function mutations have 138 been clearly associated with a glucocorticoid resistance syndrome (MIM#615962) (28, 29, 139

140 30) while 30 index cases were studied (28, 29, 30). Eight patients out of 12 in whom an 141 adrenal CT examination was performed, presented with BAH, bringing into question the role 142 played by GR mutations and on the involvement of glucocorticoid resistance in the 143 pathophysiology of adrenal hyperplasia.

144

145 **3.2. Role of GR in adrenal cortex**

146 The presence of GR in the normal adrenal gland remains controversial. Several authors 147 reported a significant contribution of GR in the pathogenesis of hyperplastic glands (18, 31). Bourdeau et al (18) demonstrated, by RT-gPCR and immunohistochemistry, that PPNAD 148 cells and some PBMAH cells overexpress endogenous GR. Later on, Louiset et al (31) 149 showed, using PPNAD cells, the involvement of GR in a paradoxical response to cortisol by 150 measuring cortisol release by RIA in culture supernatants (31). They demonstrated that DXM 151 decreased cortisol production using a PKA inhibitor, indicating that DXM stimulates PKA 152 catalytic subunits. The GR antagonist RU486 was found to decrease basal glucocorticoid 153 154 production, suggesting that cortisol exerts an ultrashort positive feedback on its own production in PPNAD, via GR and supporting GR dependent up-regulation of PKA catalytic 155 subunit genes (18). 156

157 More recently, our group demonstrated that GR directly impacts human adrenocortical 158 H295R cell function by increasing key factors of steroidogenesis, including MC2R expression, as shown by pharmacological GR inhibition or RNA interference strategy (32). 159 Finally, the presence of GR was confirmed in human normal adrenal tissue by RT-PCR and 160 WB (33), while the GRa-D isoform, that had a reduced transcriptional activity compared to 161 162 GRa-B, was reported in normal and adrenal cortex tumors (34). Later, it was also 163 demonstrated that an overexpression of GR variant: A3669G and Bcll polymorphism, can be associated with AI. The Bcll polymorphism was also overrepresented in patients with BAH 164 (35). To conclude, the involvement of GR in adrenal cortex and in the pathogenesis of 165 different diseases with adrenal hyperplasia remains unclear, but its link with different 166 signaling pathways in the adrenal cortex needs to be further elucidated. To address this 167

issue, we recently used CRISPR/Cas 9 technology to introduce a GR loss-of-function
 mutation, previously functionally characterized (36), in human H295R cells.

170

171 **4. BAH in patients with** *NR3C1* **mutations**

The French National Hospital Clinical Research Program (PHRC), Muta-GR, was launched 172 to determine the prevalence of NR3C1 mutations in a cohort of one hundred patients 173 174 presenting with BAH, associated with high blood pressure (HBP) and/or biological 175 hypercortisolism without Cushing features. Forty patients had abnormal DXM suppression tests (plasma cortisol concentration remains >50 nmol/L) and high urinary free cortisol levels 176 sometimes associated with unsuppressed ACTH levels. Five novel heterozygous NR3C1 177 mutations were discovered, among which three have been already functionally characterized 178 (36, 37), leading to the 5% prevalence of GR mutations. The phenotype of glucocorticoid 179 resistant patients was much milder than the initial description (Table 2) (38). We recently 180 unraveled the molecular mechanisms of defective GR-dependent regulation of 11β-HSD2 in 181 182 patients carrying GR loss-of-function mutations, associated with pseudohypermineralocorticism (39). These findings emphasize the importance of GR genetic 183 screening in selected patients enabling their appropriate management and optimized follow-184 185 up. Primary fibroblast cultures were obtained after skin biopsy in four mutated patients (R469X, R477S, R491X, Q501H GR mutations) and GR haploinsufficiency was further 186 demonstrated by a reduced DXM induction of FKBP5 mRNA levels, consistent with 187 glucocorticoid resistance observed in the mutated patients. 188

The study of the first heterozygous GR stop mutation, R469X, was very informative (37). Indeed, among 8 mutated family members spanning three generations, four had BAH, among them a 9-y-old child, suggesting the involvement of GR haploinsufficiency in the pathogenesis of BAH.

However, the exact mechanisms by which GR haploinsufficiency could modulate adrenal hyperplasia remain unknown. Up to now, it was proposed that ACTH overstimulation of the adrenal gland accounts for BAH in the context of GR loss-of-function mutations, supported

196 by several human studies, including three heterozygous GR mutations associated with BAH and high ACTH levels (29, 35, 40). This was also reminiscent of the homozygous Nr3c1 197 198 gene inactivation in mouse models (41) in which 90% of these mice died 1-2 h postnatally, while an aberrant GR binding site was recovered in different tissues of the 10% of mouse 199 survivors despite overt glucocorticoid resistance (42). An enlarged adrenal gland size and 200 disorganized adrenocortical cells were reported in homozygous GRKO mice that presented 201 202 with high corticosterone and ACTH levels, consistent with altered HPA regulation (41). 203 However, a heterozygous GR mouse model was recently published in which one functional GR allele, responsible for GR haploinsufficiency, also had enlarged adrenal glands (29) but 204 normal ACTH levels, precluding an exclusive role of high circulating ACTH levels in the 205 206 pathogenesis of adrenal hyperplasia.

207

208 5. Conclusion

Adrenal hyperplasia is often associated with overactivation of cAMP signaling pathways, related to well-characterized GPCR or other genetic alterations. Given that GR is expressed in the adrenal cortex and owing to the recent discovery of heterozygous GR mutations in BAH patients, it is likely that GR haploinsufficiency might affect adrenal function and proliferation. The molecular mechanisms involved remain to be elucidated. Genome editing technology may help unravel these issues.

215 Disclosure

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this study.

218

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223

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234

235 **Abbreviations:**

cAMP, cyclic adenosine monophosphate; ACTH, adrenocorticotropic hormone; AI, adrenal
incidentaloma; BAH, bilateral adrenal hyperplasia; CAH, congenital adrenal hyperplasia;
CRH, corticotropin releasing hormone; DXM, dexamethasone; DST, DXM suppression test;
GIP, glucose-dependent insulinotropic polypeptide; GR, Glucocorticoid receptor; PBMAH,
primary bilateral macronodular adrenal hyperplasia; PHRC, Hospital Clinical Research
Program; PPNAD, primary pigmented nodular adrenal disease; PKA, protein kinase A; UFC,
Urinary-free Cortisol.

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474 Figure Legends

475 Figure 1: Hypothalamus-Pituitary-Adrenal gland axis

476 Cortisol release is under the control of hypothalamic corticotropin releasing hormone (CRH). 477 CRH stimulates secretion of pituitary adrenocorticotropic hormone (ACTH) which is 478 responsible the production of cortisol by adrenal cortex. Paracrine/autocrine regulation of 479 cortisol and ACTH secretion in adrenal glands has also been described (20, 33). Cortisol 480 exerts a negative feedback loop on hypothalamic and pituitary cells.

481

482 Figure 2: Steroidogenesis in adrenal cells

483 ACTH binds its MC2R receptor and activates a cAMP pathway leading to PKA activation

through release and activation of the 2 PKA catalytic subunits. Subsequent phosphorylation

485 of several targets allows StAR activation.

486

487 Figure 3: Bilateral adrenal hyperplasia in mutated GR patients

Examples of CT images in BAH patients; (A) left and right, lateral and medial limb >5mm without nodules;(B) nodules and right and left lateral and medial limb >5 mm; (C) Adrenal

490 macronodules >10 mm.

492 **Table 1:Genetic alteration involved in adrenal hyperplasia**

	Gene	Bilateral adrenal hyperplasia	
ACTH pathways	PRKAR1A	PPNAD	
	PDE11A	PPNAD	
	PDE8B	PPNAD and BAH	
	MC2R	PBMAH	
	GNAS	PBMAH	
	ARMC5	PBMAH	
	NEM 1	PBMAH	
	FH	PBMAH	
Steroidogenesis enzyme	StAR	lipoid CAH	
	CYP11A1 (P450scc)	lipoid CAH	
	HSD3B2	САН	
	CYP17A1	САН	
	POR	САН	
	CYP11B1	САН	
	CYP21A2	САН	
GR pathways	NR3C1	ВАН	
	chromosome 19q13 microduplication	ВАН	
Wnt/β catenin pathway	CTNNB1	PBMAH	
	АРС	PBMAH	

493

494 Genes involved in adrenal hyperplasia and corresponding altered signaling pathways.

495 ACTH, adrenocorticotropic hormone; BAH, bilateral adrenal hyperplasia; CAH, congenital

496 adrenal hyperplasia; GR, Glucocorticoid receptor; PBMAH, primary bilateral macronodular

497 adrenal hyperplasia; PPNAD, primary pigmented nodular adrenal disease.

Table 2: Phenotype comparison between mutated GR patients described in Muta-GRstudy (35) and the first mutated GR patient (38)

	Mutated patients Muta-GR study	First described patient	Normal range
Type of Mutation	Heterozygous	Homozygous	
Age (Years)	53.2 ± 7.7	58	
BMI (kg/m ²)	30.7 ± 9.5	24.01	<25
SBP (mmHg)	123 ± 4.5	190	<140
DBP (mmHg)	72 ± 4.5	130	<90
Kalemia (mmol/L)	3.6±0.2	R	3.5-5
Aldosterone (pg/mL)	17.3 ± 9.9	R	19-117
Renin (pg/mL)	47.2 ± 92.6	Normal	3-16
Adrenal Hyperplasia	Yes	No	
8-AM ACTH (pg/mL)	Ν	2.1N	<1
UFC (ULN)	1.7 ± 0.65	30-40	<1
DST	_	_	

BMI: Body mass index; SBP / Systolic Blood Pressure; DBP: Diastolic Blood Pressure; UFC:

Urinary Free Cortisol, DST: 1 mg DXM suppresion test , -: negative.



Fig. 1



Fig. 2



nodular and hyperplasia limb

Hyperplasia limb

- 510
- 511 **Fig. 3**



nodular adrenal