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Title: Trace element contamination in fish impacted by bauxite red mud disposal in the Cassidaigne canyon (NW French Mediterranean)

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Abstract:

From 1966 to 2015, the Gardanne alumina refinery discharged some 20 million tons of bauxite residue (called red mud) into the Cassidaigne Canyon (northwest French Mediterranean) with impacts on local ecosystem functioning. Although these red muds contained high levels of trace elements (TE), in particular titanium (Ti), vanadium (V), aluminium (Al) and arsenic (As), surprisingly, their impacts on fish contamination levels and the risk related to fish consumption have been little studied until now. Here, 11 trace elements (Al, As, Cd, Cr, Co, Cr, Mn, Ni, Pb, Ti and V) were analyzed in muscle and, when possible, liver, from 1 308 fish of 26 species from an impacted zone in the vicinity of the Cassidaigne Canyon and a reference zone, unaffected by red mud disposals. Moreover, 66 arsenic speciation analyses were performed. Although the impact of human activities on the levels of fish contamination by trace elements is generally not easy to assess *in situ* because it is blurred by interaction with biological effects, we highlighted significant contamination of the fish species collected from the Cassidaigne Canyon, especially by the main trace elements attributable to the discharges of the Gardanne alumina refinery, namely Al, V and Ti. Moreover, inorganic toxic As concentrations were higher in the impacted zone. The results of this baseline research also confirmed the concern previously raised regarding Hg in Mediterranean organisms and that trace element contamination levels in fish are generally negatively related to fish length for all TE except Hg.

Keywords: Trace element, bioaccumulation, arsenic speciation, ICP-MS, fish, industrial discharge

1 Introduction

2 Hazards to human health stemming from exposure to individual contaminants or groups of

3 contaminants have emerged as a major concern in recent years (e.g. Copat et al., 2013; Damiano et
4 al., 2011; Miniero et al., 2014; Percin et al., 2011; Suarez-Serrano et al., 2010; Turkmen et al., 2009).

5 Among the wide range of toxic substances contaminating the marine environment, specific focus has
6 been given to trace elements (Castro-Gonzalez and Mendez-Armenta, 2008). Naturally present in the

7 Earth's crust, trace elements (TE) are also found in the marine environment (Bryan, 1971; Mason,

8 2013). However, human activities have considerably increased their spread into aquatic ecosystems

9 (Asante et al., 2010; Nriagu and Pacyna, 1988; Squadrone et al., 2016), especially those in marine

10 coastal areas (Ourgaud et al., 2018). TE are generally classified into two categories on the basis of

11 their metabolic role and their regulation by organisms. Essential TE (e.g. iron, copper and zinc) are

12 involved in metabolic processes, including functional and structural ones (Amiard et al., 1987). They

13 are generally efficiently regulated by marine animals, and present a narrow range of variation in their

14 tissues, except when ambient concentrations in seawater or food reach very high concentrations

15 (Amiard et al., 1987; Chapman, 1996; Vallee and Auld, 1990), or when they are involved in a specific

16 metabolic activity at a specific life stage (e.g. juvenile growth; Endo et al., 2008). On the contrary,

17 some TE (e.g. cadmium, lead and mercury) are not essential for marine life and their concentrations

18 in organisms depend mainly on environmental levels (Amiard et al., 1987). Whether essential or non-

19 essential, TE become toxic above a certain threshold (Amiard et al., 1987).

20 From 1966 to 2015, the Gardanne alumina refinery discharged bauxite residue (called red mud)

21 through a pipe at 320 meters depth in the Cassidaigne Canyon in the northwest French

22 Mediterranean. This mud then spread along the canyon axis and on its lateral flanks to great depth

23 (>2000 m). A centimetric red layer of bauxite residue can now be observed along the canyon axis,

24 more than 50 km away from the pipe. The red mud contained high levels of TE, in particular titanium

25 (Ti), vanadium (V), aluminium (Al), and to a lesser extent arsenic (As) (Dauvin, 2010; Fontanier et al.,

26 2012). These elements are present in several mineral oxo-hydroxide phases (hematite, goethite,
27 ilmenite, rutile, anatase, brookite and kaolinite) that are not dissolved by the industrial process used
28 to extract alumina from bauxite ore at the Gardanne alumina refinery (Fontanier et al., 2012). Red
29 mud disposal into the Cassidaigne Canyon was expected to lower the biodiversity and abundance of
30 marine communities and to increase the potential uptake of bioavailable TE into the tissues of
31 marine organisms, their bioaccumulation through food webs and ultimately into human fish-
32 consuming communities (Ellis et al., 1995; Gray et al., 2003; Metian et al., 2013; Swales et al., 1998;
33 Williams et al., 1999).

34 Since the 1960s, several environmental impact studies of the Gardanne alumina production activity
35 have been performed to document the effects of these disposals on biodiversity (Dauvin, 2010) but,
36 surprisingly, up to now the impact of red mud residues in the Cassidaigne Canyon on TE
37 contamination of fish and the risk related to fish consumption have been studied only twice. In 2004,
38 9 TE were analyzed in 28 samples from 12 fish species and in 2013, 27 TE were analyzed in 11
39 samples from 8 fish species (ANSES, 2016; Dauvin, 2010). These studies did not identify any
40 cumulated risk linked with fish consumption (Dauvin, 2010). However, their conclusions were limited:
41 (i) because the number of fish analyzed per species was very low; and (ii) because thresholds were
42 not compared with baseline values measured in a control zone, precluding any clear conclusion on
43 actual contamination. Indeed, for elements without regulatory thresholds such as Ti and V, a robust
44 impact assessment should include a comparison of trace metal concentrations in fish from the
45 impacted area and from a non-impacted area chosen as control. Moreover, As is one of the
46 elements present in the red muds discharged in the Cassidaigne Canyon. Arsenic can biomagnify in
47 food webs (Francesconi, 2010; Rahman et al., 2012), but this element exists in a number of chemical
48 forms in nature, all with very different toxicity characteristics, with inorganic species being the most
49 hazardous (Saha et al., 1999). Therefore, the determination of the total As content is certainly not
50 sufficient for estimating its potential toxic impact and its speciation must be studied. Indeed, very
51 few data are available on As speciation in the Mediterranean context (see Fattorini et al., 2004;

52 Storelli et al., 2005; Storelli and Marcotrigiano, 2000). To our knowledge, As speciation in fish
53 collected close to the Cassidaigne Canyon has never been studied.
54 In this context, the specific aim of this study was to assess, for the first time, the direct impact of red
55 mud disposal in the Cassidaigne Canyon on the TE concentrations, including As species, of several
56 common fish species of commercial interest. The hypothesis tested here was that the burden for fish
57 captured close to the Cassidaigne Canyon would be higher for red mud-related trace elements, in
58 particular in Al, Ti and V, than those of fish from other non-impacted areas.

59

60 Materials and methods

61 Study area

62 The Cassidaigne Canyon is a short canyon (<50 km) located in the northern part of the western
63 Mediterranean Sea, east of the Gulf of Lions (Fig. 1). Its head borders Cassis Bay, only 8 km from the
64 coast, and presents in its deeper part two narrow passages located 3 km and 17 km, respectively,
65 from the shelf break (170 m depth). This canyon has been considerably affected by the massive
66 disposal of bauxite residues (Dauvin, 2010) that lasted almost 50 years and is estimated at 20 million
67 tons (ANSES, 2016). Below 350 m, the entire seabed along the canyon axis was covered by the red
68 mud flow which spread more than 60 km from the pipe, with a significant proportion of the mud
69 being propagated at depths >2000 m (Dauvin, 2010). Today, this red mud also covers steep inclined
70 rock and is found underneath overhangs (Fabri et al., 2014). During strong north-eastern Mistral
71 wind events, especially during winter, quantities of this red mud could be transported northward,
72 from the canyon to the continental shelf (Dauvin, 2010).

73 To assess the impacts of red mud disposal on fish contamination, two zones were considered (Fig. 1).
74 The first was adjacent to the Cassidaigne canyon head (called 'impacted zone', IZ), where detectable
75 impact was most likely to occur (see Dauvin, 2010; Fontanier et al., 2012). The second zone (called

76 'reference zone', RZ) was in the vicinity of the Stoechades Canyon. As water masses in this part of the
77 Mediterranean flow westward along the continental margin (Gatti et al., 2006), the Stoechades
78 Canyon is far from the influence of industrial discharge and thus not impacted by bauxite residues
79 (MC. Fabri, pers. com.). The head of the Stoechades Canyon is located in the Gulf of Hyères, less than
80 5 km from the coast, providing similar bathymetry and physico-chemical conditions that allow
81 comparing the two canyons. (Fig. 1).

82

83 Fish sampling

84 Twenty-six commercial species were selected in this study with regard to their importance in the
85 communities of both systems and in the diet of the local population. In addition, these species
86 encompass a wide range of feeding habits and living environments. The sampling can thus be
87 considered representative of all fish functional groups occurring in the area. The aim of the sampling
88 program was to obtain similar species from both the impacted and the reference zone. Fish were
89 collected during 40 fishing operations (16 in the impacted zone, 24 in the reference zone) conducted
90 between June 15th 2015 and July 27th 2015 by professional fishermen using different fishing gears
91 such as bottom and pelagic trawls, trammel nets, hooks, etc. The geographical coordinates of the
92 fishing operations were systematically collected by an on-board scientific observer.

93 Individual fish were stored on board and transferred to the laboratory in ice boxes where all fish
94 were measured (total length to the nearest mm) and weighed (total mass to the nearest 0.1 g)
95 before dissection. Specific measurements were performed in the muscle, the main edible tissue in
96 fish). White dorsal muscle was sampled, individually placed in polyethylene bags and kept frozen at -
97 20°C until chemical analysis. For larger species, namely *Conger conger*, *Merluccius merluccius*,
98 *Scyliorhinus canicula*, *Galeus melastomus* and *Raja clavata*, livers were also collected when possible,
99 as liver is the organ mostly involved in detoxication processes.

100

101 Determination of trace element concentrations

102 For each sample, concentrations in 11 TE (Al, As, Cd, Co, Cr, Hg, Mn, Ni, Pb, Ti and V) were analyzed.

103 The analyses of total TE were carried out using a 7700x ICP-MS (Agilent Technologies, Courtaboeuf,

104 France), equipped with a third-generation octopole reaction system (ORS3) using helium as the

105 collision gas. Further details of the instrument settings and performance criteria of this method,

106 validated in-house and accredited by the French Committee of Accreditation (COFRAC), were

107 described in Chevallier et al., (2015). Briefly, 0.2 to 2 g of sample was weighed precisely in a quartz

108 digestion vessel and then wet-oxidised with a mixture of 3 mL of ultra-pure water and 3 mL of ultra-

109 pure HNO₃ (67% v/v) in a Multiwave 3000 closed microwave digestion system (Anton-Paar,

110 Courtaboeuf, France). After cooling to room temperature, the digested samples were transferred

111 into 50 mL polyethylene tubes into which a solution of a mixture of internal standards (yttrium,

112 scandium and rhenium) at 2 µg L⁻¹ and ultrapure water was added to the final volume before analysis

113 using ICP-MS. One randomly selected vessel was filled with reagents only and taken through the

114 entire procedure as a blank. The limits of detection (LOD) were estimated at 0.042, 0.001, 0.0003,

115 0.0005, 0.005, 0.004, 0.0025, 0.025, 0.0012, 0.025, 0.0005 mg.kg⁻¹ wet mass (mg.kg⁻¹ wm) whereas

116 the limits of quantification were estimated at 0.083, 0.002, 0.0005, 0.001, 0.010, 0.008, 0.005, 0.050,

117 0.0025, 0.050, 0.0001 mg.kg⁻¹ wm for Al, As, Cd, Cr, Hg, Mn, Ni, Pb, Ti and V respectively.

118 Chemical speciation measurements were carried out on the seafood species in which total arsenic

119 exposure was most prevalent: sea breams (gilthead sea bream *Sparus aurata*, black spot sea bream

120 *Pagellus bogaraveo*, common two-banded sea bream *Diplodus vulgaris*, white sea bream *Diplodus*

121 *sargus* and red sea bream *Pagellus erythrinus*), redfish and rays. These species contribute more than

122 65% of total arsenic exposure for human consumers. For each species, between 2 and 8 composite

123 samples per zone were analyzed. Each composite sample consisted of 3 to 5 sub-samples. Arsenic

124 speciation was performed by HPLC/ICP-MS (HPLC Ultimate 300, Dionex, Voisins le bretonneux,

125 France; ICP-MS X-Series^{II}, Thermo Scientific, Courtaboeuf, France) coupling in standard mode after

126 microwave-assisted extraction (Multiwave 3000, Anton-Paar). This method was validated in-house,
127 accredited by COFRAC, and described by Leufroy et al. (2011). The following arsenic species were
128 analyzed: As(III), As(V), monomethylarsonic acid (MA), dimethylarsinic acid (DMA), and
129 arsenobetaine (AsB). For all these species, an LOD/LOQ of 0.002/0.005 mg As kg⁻¹ wm was assessed
130 under robust conditions. These speciation analyses were used to determine the levels of inorganic
131 arsenic (sum of As(III) and As(V)). In parallel, the total arsenic concentration in the composite
132 samples was also measured by the ICP-MS method described above for total TE analysis.

133 Several internal quality controls (IQCs) and associated criteria were used to ensure the reliability of
134 the results. Each run included a standard calibration, blanks, several certified reference materials
135 (CRMs): TORT-2 lobster hepatopancreas, ERM-CE278k mussel tissue and SRM 1548a typical diet,
136 chosen to cover the 11 elements of interest (for total TE analysis) and BCR-627 tuna fish tissue (for As
137 speciation analysis), two different spiked sample solutions, several fish samples including two
138 samples analyzed in duplicate and a mid-range standard analyzed for every eight samples and at the
139 end of the sequence. The set criteria were as follows: calibration ($r^2 > 0.995$), blanks (values < limit of
140 quantification (LOQ)), internal standards (values within 70 and 130% of the target value), mid-range
141 standards (values within 80 and 120% of the target value), spiked standard solutions (spike recovery
142 within 70 and 130% of the theoretical spiked standard value), CRMs (Z-score < ± 2) and duplicates
143 (acceptable if the relative standard deviation (RSD) $\leq 20\%$ when the mean value $\geq 5 \times \text{LOQ}$ or RSD \leq
144 40% when the mean value $\geq \text{LOQ}$ and $< 5 \times \text{LOQ}$). When the acceptance criteria were not met, the
145 results were discarded and the samples were re-analyzed (Millour et al., 2010).

146 All concentrations are given in mg.kg⁻¹ wm. When data were available in both muscle and liver, the
147 liver to muscle concentration ratios were calculated.

148

149 Statistical analyses

150 The statistical analysis of the results obtained on both matrices (muscle and liver) of fish was
151 performed only for species presenting a minimum sample size of 15 individuals per zone (IZ and RZ).
152 For these species, Pearson correlation tests were used to investigate the relationship between the TE
153 levels and fish length. Moreover, spatial variations in TE concentrations were analyzed according to
154 the zone for each matrix separately. To do this, PERMANOVAs were performed. They allow handling
155 complex, unbalanced multiple-factor designs and do not assume normal error distributions
156 (Anderson, 2001). Euclidean distance similarity matrices were generated and the factor 'sampling
157 area' was treated as fixed. Since it can affect contamination levels, fish length was considered as a
158 covariate. Data were $\log(X+1)$ transformed prior to statistical analyses. In each case, p-values were
159 calculated by 999 random permutations of residuals (Anderson, 2001). Concentrations below LOD
160 were set to 0 whereas concentrations below LOQ were set to LOD. Due to the large number of
161 species and TE analyzed, a large number of statistical analyses were performed simultaneously in this
162 study, thereby increasing the proportion of false positives. Therefore, a multiple testing correction
163 procedure was conducted to control the risk of reporting too many false positives and we used a
164 false discovery rate (FDR) method to adjust for the inflation of p-values (Benjamini and Hochberg,
165 1995). All data processing and statistical analyses were performed using R software (R Core Team,
166 2014) and PRIMER 6 software with the PERMANOVA add-on (Clarke and Warwick, 2001). The
167 significance level for the tests was consistently set at $\alpha = 0.05$.

168

169 Results

170 A total of 1 308 fish of 26 species were collected during this study (769 individuals and 23 species at
171 IZ; 539 individuals and 21 species at RZ, with 18 species collected in both zones) (Table 1, 2).

172 Considering the two matrices (muscle and liver), 1 597 analyses of 11 trace elements were
173 performed (928 for IZ and 669 for RZ). Among the 26 species sampled, only 9 (*Conger conger*, *Coris*
174 *julis*, *Diplodus vulgaris*, *Helicolenus dactylopterus*, *Merluccius merluccius*, *Mullus surmuletus*, *Pagellus*

175 *bogaraveo*, *Scorpaena porcus* and *Scyliorhinus canicula*) reached the minimum sampling size required
176 to investigate spatial variations and the relationship between TE and fish length. These species
177 represented more than 67% of the total chemical analyses performed. With regards to As speciation,
178 only 66 analyses were performed due to financial constraints (Table 3). This number was too small to
179 correctly explore differences between the two sampling areas and test the effect of fish size on
180 contamination but these results nonetheless provide preliminary information on the burdens of this
181 little investigated TE.

182 The trace element concentrations in the muscle and the liver of different fish species collected from
183 the two sampling areas are given in Table 2. In muscle, the distribution patterns in the TE
184 concentrations were as follows: As > Al > Hg > Mn > Ti > Pb > Cr > V > Ni > Cd > Co for IZ and As > Al >
185 Hg > Mn > Ti > Pb > Cr > V > Ni > Co > Cd for RZ. In the liver, the sequence was: As > Al > Mn > Hg > Cd
186 > Ti > Pb > Co > V > Cr > Ni for IZ and As > Al > Mn > Hg > Cd > V > Pb > Co > Ti > Cr > Ni for RZ.

187 Liver to muscle concentration ratios were consistently higher than 1 for Al, Cd, Co, Cr, Mn, Pb and V
188 and reached up to 360 for Cd in *M. merluccius* from RZ. Ratios were lower than 1 (*i.e.* higher
189 concentrations in muscle than in liver) for As and Hg. Due to the high number of values under LOD, it
190 was generally not possible to calculate those ratios for Ni. Finally, the liver to muscle concentration
191 ratios for Ti were below 1 for *C. conger* and *S. canicula* but over 1 for *M. merluccius* and *Raja clavata*.

192 Contamination levels in muscle differed significantly between species for all TE except Ni (Table 1),
193 and no general specific trend could be observed. Nevertheless, the highest concentrations of As were
194 found in the shark species *S. canicula* ($p < 0.001$) and the highest concentrations of Hg were found in
195 *H. dactylopterus* and *S. canicula* ($p < 0.001$). Considering livers, species-dependent patterns in TE
196 concentrations were less blurred and quite similar between sampling zones, with *S. canicula*
197 presenting the highest Al, As, Cd, Co and Hg concentrations, *M. merluccius* the highest Mn
198 concentrations and *C. conger* the highest Cr, Pb and V concentrations (Table 2).

199 The only compound of inorganic As (As_i) found during this study was AS(III) while As(V)
200 concentrations were systematically below LOD. Inorganic arsenic concentrations in fish muscle were
201 not correlated with total As (As_T) concentrations (Table 3), and the average percentage of As_i
202 (relative to As_T) ranged between 0.01 and 2.69% for all the species considered and in both sampling
203 zones. This percentage of As_i was systematically higher in individuals from IZ than those from RZ.

204 The relationships between fish length and concentrations differed between trace elements, species
205 and tissues (Table 4). The vast majority of relationships, whether significant or not, were nonetheless
206 negative, even for high concentrations of TE in the disposal. On the contrary, Hg was the only TE with
207 a positive relationship for all species and tissues. Finally, As exhibited an intermediate pattern as
208 positive relationships were observed for 8 species × tissue cases studied, 5 of which were significant
209 (muscle of *C. conger*, *S. canicula* and *P. bogaraveo*, liver of *S. canicula* and *C. conger*).

210 Regarding the spatial trends, TE concentrations varied according to the sampling zone but the nature
211 of the variation differed according to the species and the tissue considered (Table1, 2). Indeed, 36
212 out of the 132 comparisons of TE concentrations between sampling areas remained significant after
213 adjusting for FDR (muscle and liver grouped), with 23 and 13 comparisons displaying significantly
214 higher concentrations in IZ and RZ, respectively. For the main TE from the Gardanne alumina refinery
215 discharges, namely Al, V and Ti, when spatial variations existed, the highest concentrations in fish
216 muscle were systematically observed in IZ. By contrast, for As, Co, Hg, Mn and Pb, no clear pattern
217 was shown with concentrations higher in IZ for some species and higher in RZ for others. Finally, for
218 Cd, Cr and Ni, no difference in TE concentrations were observed between the two zones whatever
219 couple of species × tissue was considered.

220

221 Discussion

222 Very few studies have assessed the impact of the red mud disposal in the Cassidaigne Canyon on
223 contamination levels in fish. To our knowledge, this is the first time that this has been investigated so

224 thoroughly, with the analysis of 11 TE for a large number of individuals (n=1308) from 26 different
225 fish species. In addition, two tissues (muscle and liver) were taken into account, comparing, when
226 possible, contamination levels between the impacted zone and a reference one. Moreover, although
227 several studies have already been conducted on TE in Mediterranean fish (e.g. Canli and Atli, 2003;
228 Iamiceli et al., 2015; Mille et al., 2018), some elements such as Al, Ti and V have rarely been
229 investigated (Eisler, 2010). Lastly, the present study provides a baseline dataset for various
230 Mediterranean fish species and allows a preliminary investigation of certain factors affecting
231 contamination.

232 A large number of the fish analyzed in this study presented concentration levels higher than the
233 limits defined by the European Commission in fish muscle (EC, 2006). A total of 444 individuals from
234 15 different species (*C. conger*, *C. julis*, *D. sargus*, *D. vulgaris*, *G. melastomus*, *H. dactylopterus*,
235 *M. merluccius*, *M. surmuletus*, *M. barbatus*, *P. bogaraveo*, *R. clavata*, *S. canicula*, *T. mediterraneus*,
236 *T. trachurus* and *Z. faber*) had Hg concentration levels above the European regulatory limit (0.5
237 mg.kg⁻¹ wm in teleosts, 1 mg.kg⁻¹ wm in sharks), confirming the concern raised previously regarding
238 Hg in Mediterranean organisms (Cossa et al., 2012; Harmelin-Vivien et al., 2009; Koenig et al., 2013).
239 The straightforward pattern observed for Hg in the present study may be largely driven by biological
240 and biogeochemical mechanisms at play at the scale of the whole NE Mediterranean (Cossa and
241 Coquery, 2005) rather than from the local influence of red mud disposals. For Pb and Cd, the number
242 of individuals with concentrations above the EU regulatory limits in fish muscle (0.3 mg.kg⁻¹ wm and
243 0.05 mg.kg⁻¹ wm for Pb and Cd, respectively), were lower, 8 and 19, respectively, which represent
244 between 0.6 and 1.5% of the total number of fish analyzed. Between 75 and 90% of these threshold
245 overruns concerned IZ, confirming the conclusions of Ourgaud et al. (2018), who found that these TE
246 were more concentrated in *S. porcus* collected in the bay of Marseille (close to the Cassidaigne
247 Canyon) than in those from the Gulf of Hyères. There is no regulatory limit for total As in fish muscle,
248 but arsenic exposure is a significant worldwide environmental health concern. Overall, the data we
249 obtained here appear of great concern with regard to total As concentrations. However, the risk of

250 potential As poisoning through fish consumption should be further assessed through the specific
251 determination of the inorganic As content in fish tissues. Most of the As in the tissues of marine
252 animals is bound in organoarsenic forms (Neff, 1997) and considered of low toxicity (Hindmarsh,
253 2000). In this study, the main form of As found in fish was arsenobetaine which accounts for up to
254 100% of As in certain species (Table 3). This form is not toxic to humans (Neff, 1997).

255 In both sampling zones, our results clearly showed significant differences in the bioaccumulation of a
256 range of TE between the 26 selected species, suggesting a diversity response among fish (Table 1, 2).
257 Indeed, the accumulation of TE in fish does not directly reflect their simple concentration in the
258 ambient environment as it is affected by environmental factors, species biology and the physico-
259 chemical properties of the contaminant (Cossa et al., 2012; Cresson et al., 2015). Thus, assessing a
260 site-specific contamination pattern requires good understanding of all these parameters although
261 the interpretation of the results remains complex because of the existence of interactions between
262 them. Notwithstanding some general trends could still be discerned. Some TE such as Hg are well-
263 known for their biomagnification property, namely the trend for increasing concentrations along
264 food webs (Bryan et al., 1979). This process in particular is responsible for the high Hg concentrations
265 generally observed in high trophic level species such as marine apex predators (Endo et al., 2008) like
266 *S. canicula*, *M. merluccius*, *G. melastomus*, *Z. faber*, etc. In this study, Hg concentrations in fish muscle
267 from IZ varied between 0.014 mg.kg⁻¹ in *L. aurata* and 1.613 m. kg⁻¹ in *S. canicula* (Table 1). The inter-
268 specific variation in mean trophic levels, ranging from 2.8 for *L. aurata* to 3.8 *S. canicula* (Cresson et
269 al., 2014b; Froese and Pauly, 2014; Stergiou and Karpouzi, 2001), could partially explain this
270 differences in the TE concentrations between these species. Moreover, an increasing number of
271 studies have documented relatively high levels of TE contamination in shelf-edge and deep species
272 (Cresson et al., 2014a; Hornung et al., 1993), especially when compared to coastal species
273 (Chouvelon et al., 2012). The 26 species selected here live at drastically different depths, from the
274 surface down to a few meters depth for *C. julis*, *L. ramada* and *L. aurata*, to several hundreds of
275 meters for *G. melastomus*, *H. dactylopterus* and *P. bogaraveo* (Froese and Pauly, 2014). In this study,

276 *C. julis*, *L. ramada* and *L. aurata* were captured between 5 and 35 m depth whereas *G. melastomus*,
277 *H. dactylopterus* and *P. bogaraveo* were caught between 141 and 391m depth. This may explain a
278 part of the inter-specific variations in TE contamination and the lower TE concentrations generally
279 observed in *L. ramada*, *L. aurata* and *C. julis* (Table 1), Finally, inter-specific differences in TE
280 concentrations may also be driven by contrasting life-history and behavior. Factors such as preferred
281 habitat (e.g. benthic and pelagic), longevity, diet and foraging behavior (e. g. vertical migrations and
282 type of prey consumed), metabolic rates and biochemical response (Burger, 2007; Zhao et al., 2012)
283 have been demonstrated to affect rates of TE uptake and excretion from both food and water
284 pathways. Previous works showed that species that are continuously in contact with contaminants in
285 sediment have developed higher detoxication abilities, resulting in lower body burdens while being
286 exposed to the same contaminant load as pelagic species (Sole et al., 2006). This may explain why TE
287 concentrations in the benthic ray *R. clavata* are generally lower than those of the demersal *S. canicula*
288 (Table1, 2), despite similar trophic levels (Froese and Pauly, 2014; Stergiou and Karpouzi, 2001) and
289 living depth (both species were captured between 100 and 380m depth).

290 Fish length is often recognized to be of importance in determining the rate of physiological processes
291 that influence uptake, distribution and elimination of TE (Canli and Atli, 2003). However, except for
292 Hg, our analysis revealed no conclusive evidence that mean TE concentrations were systematically
293 linked to fish length, which is in good agreement with the results reported in the literature. Indeed,
294 although it is generally accepted that Hg load in fish body is strongly affected by size it is still not
295 clear if the other TE accumulations are size-dependent. This is supported by the fact that most TE
296 (with the exception of As and Hg) do not biomagnify along aquatic food webs (Fey et al., 2019; Xu
297 and Wang, 2002). The available literature on the correlation between TE concentrations in muscle
298 tissue and fish length and weight have shown that a dependency exists for some species (e.g. Gobert
299 et al., 2017), while for other species, no dependency is found (e.g. Mille et al., 2018). The absence of
300 relationship likely indicates that fish regulate some TE at a certain concentration required for
301 efficient metabolic activities (Canli and Atli, 2003; Hornung et al., 1993). Moreover, when significant,

302 the relationships between TE (other than Hg) and fish lengths are generally negative (Endo et al.,
303 2008). This may result from a higher metabolic activity in fast-growing young individuals, requiring
304 higher amounts of these elements than older individuals or through a higher excretion rate and/or
305 dilution of metal burden with growth, and lower metabolism (Eisler, 2010). In addition, length is
306 commonly used as a proxy of age and chronic exposure to contaminants, as length is quickly
307 measured, while accurate age determination requires complex and time-consuming procedures (*e.g.*
308 otolith or vertebrae reading for teleost and chondrichthyans, respectively). This pattern is nonetheless
309 blurred by the fact that growth rate affects the age - length relationship. As a result, individuals of
310 the same length but from zones with different nutritional inputs (and thus different growth rates)
311 may have different age and may have been thus exposed to contaminant during different periods of
312 time.

313 Many studies have shown that TE accumulate in various organs of fish at different levels (Canli and
314 Atli, 2003; Dural et al., 2006; Fernandes et al., 2007; Long and Wang, 2005; Mormede and Davies,
315 2001; Uluturhan and Kucuksezgin, 2007). With the exception of Hg, As and Ti, the liver to muscle
316 concentration ratios calculated in this study were generally over 1 and reached up to 360. This is not
317 surprising because, in general, TE are preferentially accumulated in the liver of aquatic organisms
318 (Henry et al., 2004), which likely results from detoxification processes (Eisler, 2010). For Hg, higher
319 concentrations are commonly found in fish muscle. Indeed, methylmercury is the predominant form
320 of Hg in fish, ranging from 80% to 100% (Bloom, 1992; Chauvelon et al., 2018; Magalhaes et al.,
321 2007) and is mainly accumulated in fish muscle due to its affinity to protein (Amlund et al., 2007;
322 Harris et al., 2003). This difference in chemical affinity explains why the liver to muscle concentration
323 ratios for Hg ranged here from 0.47 to 0.97. Considering As, Eisler (2010) stated that hepatic
324 concentrations are usually higher than those found in muscle tissues, a pattern not observed in the
325 present study as the liver to muscle concentration ratios ranged between 0.34 and 0.90. Such ratios
326 below 1 have already been reported in benthic (*Pteromylaeus bovinus* and *Myliobatis aquila*) and
327 pelagic (*Pteroplatytrygon violacea*) rays from the northern Adriatic Sea (Slejkovec et al., 2014), in *S.*

328 *canicula* from the North Sea (De Gieter et al., 2002), in 22 fish species from the New Caledonia
329 lagoon (Metian et al., 2013), and in 32 teleost species from the island of Guam (Pacific Ocean)
330 (Denton et al., 2006). Therefore, the As partition between muscle and liver seems to be species-
331 dependent (e.g. Hellou et al., 1996; Raimundo et al., 2013; Raimundo et al., 2015) and possibly
332 related to differences in the bioavailability of forms of this element (Francesconi, 2010). Finally, very
333 few data exist on Ti concentrations in fish and, to our knowledge, the accumulation of this element in
334 different organs has never been studied. The distinct accumulation patterns observed here may
335 result from differences in the physiological response of the species, as well as differences in life-
336 history and behavior of fish.

337 As confirmed by sediment analysis (Fontanier et al., 2012), the Cassidaigne Canyon has been
338 subjected to considerable TE inputs, mainly in Al, Ti, V from the alumina refinery's by-products. In
339 general, these TE are considered non-essential for marine life and their concentrations in organisms
340 mainly depend on their environmental levels (Amiard et al., 1987). Therefore, it was expected that
341 fish from IZ would contain consistently higher concentrations of red mud-related TE than those from
342 RZ. Among all the species, Al was consistently more accumulated in fish from IZ, likely resulting from
343 industrial releases. Aluminum is also the TE most concentrated in red mud. Laboratory experiments
344 showed that the bauxite from the Gardanne plant releases substantial quantities of Al in seawater
345 (Pagano et al., 2002). In the marine environment (pH 8.0-8.3, salinity 35), this Al is theoretically
346 mostly present in two forms: the aluminate anion $\text{Al}(\text{OH})_4^-$ and to a lesser extent neutral aluminum
347 hydroxide $\text{Al}(\text{OH})_3$ (Millero et al., 2009). There are very few studies on the transfer of aluminate
348 anions and aluminum hydroxide to aquatic organisms and even fewer focusing on fish species. Our
349 results suggested that fish are able to at least partially accumulate these Al forms. This observation
350 can be extended to other living organisms as Al bioaccumulation has also been reported in marine
351 plants collected nearby a coastal bauxite sludge disposal site (Malea and Haritonidis, 1989). For the
352 other TE attributable to the Gardanne alumina refinery discharges (Ti and V), we found that when
353 spatial variations existed, concentrations in fish muscle were systematically higher in IZ than in RZ,

354 but this difference concerned a limited number of species. It could be hypothesized that they are not
355 readily bioavailable for species and/or they are poorly transferred through the food chain up to apex
356 predators. This result is similar to those of Bourcier (1969), Bourcier & Zibrowius (1973) and
357 Fontanier et al. (2012), who observed that there was no chemical impact of red muds on
358 macrobenthic fauna and the structure of deep-sea foraminifera communities in the Cassidaigne
359 Canyon. They suggested that most of the toxic TE in the red mud impacting the sites are not
360 bioavailable but simply locked in solid phases.

361

362 Conclusion

363 The impact of the human activities on the levels of fish contamination by TE is not easy to assess *in*
364 *situ* because it is blurred by the interaction of biological effects (e.g. Bouchoucha et al., 2018; Cresson
365 et al., 2014a; Metian et al., 2013). A thorough sampling campaign and numerous analyses allowed
366 highlighting the significant contamination of fish species collected from the Cassidaigne Canyon area,
367 especially for Al and to a lesser extent for Ti and V. Although not statistically proved, it was also
368 shown that fish collected close to the alumina refinery pipe had higher Asi concentrations than those
369 from the reference area. In the next step, it would be interesting to assess human dietary exposure
370 to these metals through the consumption of fish from the Cassidaigne Canyon zone. This would allow
371 better evaluation of the health risk for humans consuming these fish. Moreover, the solubility of
372 many TE is highly redox-sensitive, thus the elemental concentrations in seawater change accordingly
373 (e.g. Morford and Emerson, 1999). Consequently, if the redox conditions change in the sites
374 contaminated by red mud, this would influence the solubility of TE, making them bioavailable to
375 marine organisms. Therefore, it seems essential to continue regular and long-term monitoring of
376 marine organisms from this area to prevent any further environmental deterioration and to assess
377 human exposure.

378

379

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388 References

389 Amiard JC, Amiardtriquet C, Berthet B, Metayer C. Comparative study of the patterns of
390 bioaccumulation of essential (Cu, Zn) and non-essential (Cd, Pb) trace metals in various
391 estuarine and coastal organisms. *Journal of Experimental Marine Biology and Ecology* 1987;
392 106: 73-89.

393 Amlund H, Lundebye AK, Berntssen MHG. Accumulation and elimination of methylmercury in Atlantic
394 cod (*Gadus morhua* L.) following dietary exposure. *Aquatic Toxicology* 2007; 83: 323-330.

395 Anderson JM. Permutation tests for univariate or multivariate analysis of variance and regression.
396 *Canadian Journal of Fisheries and Aquatic Sciences* 2001; 58: 626-639.

397 ANSES. Note d'appui scientifique et technique de l'Agence nationale de sécurité sanitaire de
398 l'alimentation , de l'environnement et du travail relatif à l'impact potentiel sur la santé
399 humaine du rejet en Méditerranée d'effluents issus des activités de transformation de
400 minerai de bauxite, 2016, pp. 74.

401 <https://www.anses.fr/en/system/files/ERCA2014SA0223.pdf>

402 Asante KA, Agusa T, Kubota R, Mochizuki H, Ramu K, Nishida S, et al. Trace elements and stable
403 isotope ratios ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) in fish from deep-waters of the Sulu Sea and the
404 Celebes Sea. *Marine Pollution Bulletin* 2010; 60: 1560-1570.

405 Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to
406 multiple testing. *Journal of the Royal statistical society: series B (Methodological)* 1995; 57:
407 289-300.

408 Bloom NS. On the chemical form of mercury in edible fish and marine invertebrate tissue. *Canadian*
409 *Journal of Fisheries and Aquatic Sciences* 1992; 49: 1010-1017.

410 Bouchoucha M, Brach-Papa C, Gonzalez J-L, Lenfant P, Darnaude A. Growth, condition and metal
411 concentration in juveniles of two *Diplodus* species in ports. *Marine Pollution Bulletin* 2018;
412 126: 31-42.

413 Bourcier M. Ecoulement des boues rouges dans le canyon de la Cassidaigne (décembre 1968).
414 *TeHthys, France* 1969; 1: 779.

415 Bourcier M, Zibrowius H. Les «boues rouges» déversées dans le canyon de la Cassidaigne (région de
416 Marseille). *Observations en soucoupe plongeante SP* 1973; 350: 811-842.

417 Bryan G. The effects of heavy metals (other than mercury) on marine and estuarine organisms.
418 *Proceedings of the Royal Society of London B: Biological Sciences* 1971; 177: 389-410.

419 Bryan G, Waldichuk M, Pentreath R, Darracott A. Bioaccumulation of marine pollutants. *Philosophical*
420 *Transactions of the Royal Society of London. Series B, Biological Sciences* 1979: 483-505.

421 Burger J. A framework and methods for incorporating gender-related issues in wildlife risk
422 assessment: Gender-related differences in metal levels and other contaminants as a case
423 study. *Environmental Research* 2007; 104: 153-162.

424 Canli M, Atli G. The relationships between heavy metal (Cd, Cr, Cu, Fe, Pb, Zn) levels and the size of
425 six Mediterranean fish species. *Environmental Pollution* 2003; 121: 129-136.

426 Castro-Gonzalez MI, Mendez-Armenta M. Heavy metals: Implications associated to fish consumption.
427 *Environmental Toxicology and Pharmacology* 2008; 26: 263-271.

428 Chapman DV. *Water quality assessments: a guide to the use of biota, sediments, and water in*
429 *environmental monitoring*: E & Fn Spon London, 1996.

430 Chevallier E, Chekri R, Zinck J, Guérin T, Noël L. Simultaneous determination of 31 elements in
431 foodstuffs by ICP-MS after closed-vessel microwave digestion: Method validation based on
432 the accuracy profile. *Journal of Food Composition and Analysis* 2015; 41: 35-41.

433 Chouvelon T, Cresson P, Bouchoucha M, Brach-Papa C, Bustamante P, Crochet S, et al. Oligotrophy as
434 a major driver of mercury bioaccumulation in medium-to high-trophic level consumers: A
435 marine ecosystem-comparative study. *Environmental Pollution* 2018; 233: 844-854.

436 Chouvelon T, Spitz J, Caurant F, Mendez-Fernandez P, Autier J, Lassus-Debat A, et al. Enhanced
437 bioaccumulation of mercury in deep-sea fauna from the Bay of Biscay (north-east Atlantic) in
438 relation to trophic positions identified by analysis of carbon and nitrogen stable isotopes.
439 *Deep-Sea Research Part I-Oceanographic Research Papers* 2012; 65: 113-124.

440 Clarke KR, Warwick RM. *Change in marine communities: an approach to statistical analysis and*
441 *interpretation*. PRIMER-E Limited Plymouth, UK, 2001.

442 Copat C, Arena G, Fiore M, Ledda C, Fallico R, Sciacca S, et al. Heavy metals concentrations in fish and
443 shellfish from eastern Mediterranean Sea: Consumption advisories. *Food and Chemical*
444 *Toxicology* 2013; 53: 33-37.

445 Cossa D, Coquery M. The Mediterranean mercury anomaly, a geochemical or a biological issue. In:
446 Saliot A, editor. *The Mediterranean Sea*. Springer Berlin Heidelberg, Berlin, Heidelberg, 2005,
447 pp. 177-208.

448 Cossa D, Harmelin-Vivien M, Mellon-Duval C, Loizeau V, Averty B, Crochet S, et al. Influences of
449 bioavailability, trophic position, and growth on methylmercury in hakes (*Merluccius*
450 *merluccius*) from northwestern Mediterranean and northeastern Atlantic. *Environmental*
451 *Science & Technology* 2012; 46: 4885-4893.

452 Cresson P, Bouchoucha M, Miralles F, Elleboode R, Mahe K, Maruszczak N, et al. Are red mullet
453 efficient as bio-indicators of mercury contamination? A case study from the French
454 Mediterranean. *Marine Pollution Bulletin* 2015; 91: 191-199.

455 Cresson P, Fabri MC, Bouchoucha M, Papa CB, Chavanon F, Jadaud A, et al. Mercury in organisms
456 from the Northwestern Mediterranean slope: Importance of food sources. *Science of The*
457 *Total Environment* 2014a; 497: 229-238.

458 Cresson P, Ruitton S, Ourgaud M, Harmelin-Vivien M. Contrasting perception of fish trophic level
459 from stomach content and stable isotope analyses: A Mediterranean artificial reef
460 experience. *Journal of Experimental Marine Biology and Ecology* 2014b; 452: 54-62.

461 Damiano S, Papetti P, Menesatti P. Accumulation of heavy metals to assess the health status of
462 swordfish in a comparative analysis of Mediterranean and Atlantic areas. *Marine Pollution*
463 *Bulletin* 2011; 62: 1920-1925.

464 Dauvin JC. Towards an impact assessment of bauxite red mud waste on the knowledge of the
465 structure and functions of bathyal ecosystems: The example of the Cassidaigne canyon
466 (north-western Mediterranean Sea). *Marine Pollution Bulletin* 2010; 60: 197-206.

467 De Gieter M, Leermakers M, Van Ryssen R, Noyen J, Goeyens L, Baeyens W. Total and toxic arsenic
468 levels in North Sea fish. *Archives of Environmental Contamination and Toxicology* 2002; 43:
469 406-417.

470 Denton GRW, Concepcion LP, Wood HR, Morrison RJ. Trace metals in marine organisms from four
471 harbours in Guam. *Marine Pollution Bulletin* 2006; 52: 1784-1804.

472 Dural M, Goksu MZL, Ozak AA, Derici B. Bioaccumulation of some heavy metals in different tissues of
473 *Dicentrarchus labrax* L, 1758, *Sparus aurata* L, 1758 and *Mugil cephalus* L, 1758 from the
474 Camlik lagoon of the eastern coast of Mediterranean (Turkey). *Environmental Monitoring and*
475 *Assessment* 2006; 118: 65-74.

476 EC. Commission Regulation (EC) No 1881/2006 of 19 December 2006 setting maximum levels for
477 certain contaminants in foodstuffs. Off J Eur Union 2006; 364: 5-24.

478 Eisler R. Chapter 3 - Fishes. In: Eisler R, editor. Compendium of Trace Metals and Marine Biota.
479 Elsevier, Amsterdam, 2010, pp. 39-220.

480 Ellis DV, Poling GW, Baer RL. Submarine tailings disposal (STD) for mines: An introduction. Marine
481 Georesources & Geotechnology 1995; 13: 3-18.

482 Endo T, Hisamichi Y, Haraguchi K, Kato Y, Ohta C, Koga N. Hg, Zn and Cu levels in the muscle and liver
483 of tiger sharks (*Galeocerdo cuvier*) from the coast of Ishigaki Island, Japan: Relationship
484 between metal concentrations and body length. Marine Pollution Bulletin 2008; 56: 1774-
485 1780.

486 Fabri MC, Pedel L, Beuck L, Galgani F, Hebbeln D, Freiwald A. Megafauna of vulnerable marine
487 ecosystems in French mediterranean submarine canyons: Spatial distribution and
488 anthropogenic impacts. Deep-Sea Research Part II-Topical Studies in Oceanography 2014;
489 104: 184-207.

490 Fattorini D, Alonso-Hernandez CM, Diaz-Asencio M, Munoz-Caravaca A, Pannacciulli FG, Tangherlini
491 M, et al. Chemical speciation of arsenic in different marine organisms: Importance in
492 monitoring studies. Marine Environmental Research 2004; 58: 845-850.

493 Fernandes C, Fontainhas-Fernandes A, Peixoto F, Salgado MA. Bioaccumulation of heavy metals in
494 *Liza saliens* from the Esmoriz-Paramos coastal lagoon, Portugal. Ecotoxicology and
495 Environmental Safety 2007; 66: 426-431.

496 Fey P, Bustamante P, Bosserelle P, Espiau B, Malau A, Mercader M, et al. Does trophic level drive
497 organic and metallic contamination in coral reef organisms? Science of the Total
498 Environment 2019; 667: 208-221.

499 Fontanier C, Fabri MC, Buscail R, Biscara L, Koho K, Reichart GJ, et al. Deep-sea foraminifera from the
500 Cassidaigne Canyon (NW Mediterranean): Assessing the environmental impact of bauxite red
501 mud disposal. Marine Pollution Bulletin 2012; 64: 1895-1910.

502 Francesconi KA. Arsenic species in seafood: Origin and human health implications. Pure and Applied
503 Chemistry 2010; 82: 373-381.

504 Froese R, Pauly D. Fishbase. Available: www.fishbase.org, 2014.

505 Gatti J, Petrenko A, Devenon JL, Leredde Y, Ulses C. The Rhone river dilution zone present in the
506 northeastern shelf of the Gulf of Lion in December 2003. Continental Shelf Research 2006;
507 26: 1794-1805.

508 Gobert S, Pasqualini V, Dijoux J, Lejeune P, Durieux EDH, Marengo M. Trace element concentrations
509 in the apex predator swordfish (*Xiphias gladius*) from a Mediterranean fishery and risk
510 assessment for consumers. Marine Pollution Bulletin 2017; 120: 364-369.

511 Gray JE, Greaves IA, Bustos DM, Krabbenhoft DP. Mercury and methylmercury contents in mine-
512 waste calcine, water, and sediment collected from the Palawan Quicksilver Mine, Philippines.
513 Environmental Geology 2003; 43: 298-307.

514 Harmelin-Vivien M, Cossa D, Crochet S, Banaru D, Letourneur Y, Mellon-Duval C. Difference of
515 mercury bioaccumulation in red mullets from the north-western Mediterranean and Black
516 seas. Marine Pollution Bulletin 2009; 58: 679-685.

517 Harris HH, Pickering IJ, George GN. The chemical form of mercury in fish. Science 2003; 301: 1203-
518 1203.

519 Hellou J, Zitko V, Friel J, Alkanani T. Distribution of elements in tissues of yellowtail flounder
520 *Pleuronectes ferruginea*. Science of The Total Environment 1996; 181: 137-146.

521 Henry F, Amara R, Courcot L, Lacouture D, Bertho ML. Heavy metals in four fish species from the
522 French coast of the eastern English Channel and Southern Bight of the North Sea.
523 Environment International 2004; 30: 675-683.

524 Hindmarsh JT. Arsenic, its clinical and environmental significance. The Journal of Trace Elements in
525 Experimental Medicine: The Official Publication of the International Society for Trace
526 Element Research in Humans 2000; 13: 165-172.

527 Hornung H, Krom MD, Cohen Y, Bernhard M. Trace-metam content in deep-water sharks from the
528 eastern Mediterranean-Sea. *Marine Biology* 1993; 115: 331-338.

529 Iamiceli A, Ubaldi A, Lucchetti D, Brambilla G, Abate V, De Felip E, et al. Metals in Mediterranean
530 aquatic species. *Marine Pollution Bulletin* 2015; 94: 278-283.

531 Koenig S, Solé M, Fernández-Gómez C, Díez S. New insights into mercury bioaccumulation in deep-
532 sea organisms from the NW Mediterranean and their human health implications. *Science of*
533 *The Total Environment* 2013; 442: 329-335.

534 Leufroy A, Noël L, Dufailly V, Beauchemin D, Guérin T. Determination of seven arsenic species in
535 seafood by ion exchange chromatography coupled to inductively coupled plasma-mass
536 spectrometry following microwave assisted extraction: method validation and occurrence
537 data. *Talanta* 2011; 83: 770-779.

538 Long AM, Wang WX. Metallothionein induction and bioaccumulation kinetics of Cd and Ag in the
539 marine fish *Terapon jarbua* challenged with dietary or waterborne Ag and Cu. *Marine Ecology*
540 *Progress Series* 2005; 291: 215-226.

541 Magalhaes MC, Costa V, Menezes GM, Pinho MR, Santos RS, Monteiro LR. Intra- and inter-specific
542 variability in total and methylmercury bioaccumulation by eight marine fish species from the
543 Azores. *Marine Pollution Bulletin* 2007; 54: 1654-1662.

544 Malea P, Haritonidis S. Concentration of Aluminium in *Halophila stipulacea* (Forsk.) archers. and the
545 substrate of the Antikyra Gulf, Greece. *Toxicological & Environmental Chemistry* 1989; 20-21:
546 241-248.

547 Mason RP. Trace metals in aquatic systems: John Wiley & Sons, 2013.

548 Metian M, Warnau M, Chouvelon T, Pedraza F, Baena A, Bustamante P. Trace element
549 bioaccumulation in reef fish from New Caledonia: Influence of trophic groups and risk
550 assessment for consumers. *Marine Environmental Research* 2013; 87-88: 26-36.

551 Mille T, Cresson P, Chouvelon T, Bustamante P, Brach-Papa C, Bruzac S, et al. Trace metal
552 concentrations in the muscle of seven marine species: Comparison between the Gulf of Lions
553 (North-West Mediterranean Sea) and the Bay of Biscay (North-East Atlantic Ocean). *Marine*
554 *Pollution Bulletin* 2018; 135: 9-16.

555 Millero FJ, Woosley R, Ditrolio B, Waters J. Effect of Ocean Acidification on the Speciation of Metals
556 in Seawater. *Oceanography* 2009; 22: 72-85.

557 Millour S, Noël L, Chekri R, Vastel C, Kadar A, Guérin T. Internal quality controls applied in inductively
558 coupled plasma mass spectrometry multi-elemental analysis in the second French Total Diet
559 Study. *Accreditation and quality assurance* 2010; 15: 503-513.

560 Miniero R, Abate V, Brambilla G, Davoli E, De Felip E, De Filippis SP, et al. Persistent toxic substances
561 in Mediterranean aquatic species. *Science of The Total Environment* 2014; 494: 18-27.

562 Morford JL, Emerson S. The geochemistry of redox sensitive trace metals in sediments. *Geochimica Et*
563 *Cosmochimica Acta* 1999; 63: 1735-1750.

564 Mormede S, Davies IM. Heavy metal concentrations in commercial deep-sea fish from the Rockall
565 Trough. *Continental Shelf Research* 2001; 21: 899-916.

566 Neff JM. Ecotoxicology of arsenic in the marine environment. *Environmental Toxicology and*
567 *Chemistry* 1997; 16: 917-927.

568 Nriagu JO, Pacyna JM. Quantitative assessment of worldwide contamination of air, water and soils by
569 trace-metals. *Nature* 1988; 333: 134-139.

570 Ourgaud M, Ruitton S, Bourgogne H, Bustamante P, Churlaud C, Guillou G, et al. Trace elements in a
571 Mediterranean scorpaenid fish: Bioaccumulation processes and spatial variations. *Progress in*
572 *Oceanography* 2018; 163: 184-195.

573 Pagano G, Méric S, De Biase A, Iaccarino M, Petruzzelli D, Tunay O, et al. Toxicity of bauxite
574 manufacturing by-products in sea urchin embryos. *Ecotoxicology and Environmental Safety*
575 2002; 51: 28-34.

576 Percin F, Sogut O, Altinelataman C, Soylak M. Some trace elements in front and rear dorsal ordinary
577 muscles of wild and farmed bluefin tuna (*Thunnus thynnus* L. 1758) in the Turkish part of the
578 eastern Mediterranean Sea. *Food and Chemical Toxicology* 2011; 49: 1006-1010.

579 R Core Team. R: A Language and Environment for Statistical Computing. R Foundation for Statistical
580 Computing, Vienna, Austria, 2014.

581 Rahman MA, Hasegawa H, Lim RP. Bioaccumulation, biotransformation and trophic transfer of
582 arsenic in the aquatic food chain. *Environmental Research* 2012; 116: 118-135.

583 Raimundo J, Anes B, Caetano M, Giacomello E, Menezes G, Vale C. Trace-element concentrations in
584 muscle and liver of 11 commercial fish species from Condor Seamount, Azores Archipelago
585 (Portugal). *Deep-Sea Res* 2013; 98: 137-147.

586 Raimundo J, Vale C, Martins I, Fontes J, Graca G, Caetano M. Elemental composition of two
587 ecologically contrasting seamount fishes, the bluemouth (*Helicolenus dactylopterus*) and
588 blackspot seabream (*Pagellus bogaraveo*). *Marine Pollution Bulletin* 2015; 100: 112-121.

589 Saha JC, Dikshit AK, Bandyopadhyay M, Saha KC. A review of arsenic poisoning and its effects on
590 human health. *Critical Reviews in Environmental Science and Technology* 1999; 29: 281-313.

591 Slejkovec Z, Stajnko A, Falnoga I, Lipej L, Mazej D, Horvat M, et al. Bioaccumulation of Arsenic Species
592 in Rays from the Northern Adriatic Sea. *International Journal of Molecular Sciences* 2014; 15:
593 22073-22091.

594 Sole M, de la Parra LMG, Alexandre-Grimaldo S, Sarda F. Esterase activities and lipid peroxidation
595 levels in offshore commercial species of the NW Mediterranean Sea. *Marine Pollution*
596 *Bulletin* 2006; 52: 1708-1716.

597 Squadrone S, Burioli E, Monaco G, Koya MK, Prearo M, Gennero S, et al. Human exposure to metals
598 due to consumption of fish from an artificial lake basin close to an active mining area in
599 Katanga (DR Congo). *Science of The Total Environment* 2016; 568: 679-684.

600 Stergiou KI, Karpouzi VS. Feeding habits and trophic levels of Mediterranean fish. *Reviews in Fish*
601 *Biology and Fisheries* 2001; 11: 217-254.

602 Storelli MM, Busco VP, Marcotrigiano GO. Mercury and arsenic speciation in the muscle tissue of
603 *Scylliorhinus canicula* from the Mediterranean sea. *Bulletin of Environmental Contamination*
604 *and Toxicology* 2005; 75: 81-88.

605 Storelli MM, Marcotrigiano GO. Organic and inorganic arsenic and lead in fish from the South Adriatic
606 Sea, Italy. *Food Additives and Contaminants* 2000; 17: 763-768.

607 Suarez-Serrano A, Alcaraz C, Ibanez C, Trobajo R, Barata C. *Procambarus clarkii* as a bioindicator of
608 heavy metal pollution sources in the lower Ebro River and Delta. *Ecotoxicology and*
609 *Environmental Safety* 2010; 73: 280-286.

610 Swales S, Storey AW, Roderick ID, Figa BS, Bakowa KA, Tenakanai CD. Biological monitoring of the
611 impacts of the Ok Tedi copper mine on fish populations in the Fly River system, Papua New
612 Guinea. *Science of The Total Environment* 1998; 214: 99-111.

613 Turkmen M, Turkmen A, Tepe Y, Tore Y, Ates A. Determination of metals in fish species from Aegean
614 and Mediterranean seas. *Food Chemistry* 2009; 113: 233-237.

615 Uluturhan E, Kucuksezgin F. Heavy metal contaminants in Red Pandora (*Pagellus erythrinus*) tissues
616 from the Eastern Aegean Sea, Turkey. *water research* 2007; 41: 1185-1192.

617 Vallee BL, Auld DS. Zinc coordination, function, and structure of zinc enzymes and other proteins.
618 *Biochemistry* 1990; 29: 5647-5659.

619 Williams TM, Weeks JM, Apostol AN, Miranda CR. Assessment of mercury contamination and human
620 exposure associated with coastal disposal of waste from a cinnabar mining operation,
621 Palawan, Philippines. *Environmental Geology* 1999; 39: 51-60.

622 Xu Y, Wang WX. Exposure and potential food chain transfer factor of Cd, Se and Zn in marine fish
623 *Lutjanus argentimaculatus*. *Marine Ecology Progress Series* 2002; 238: 173-186.

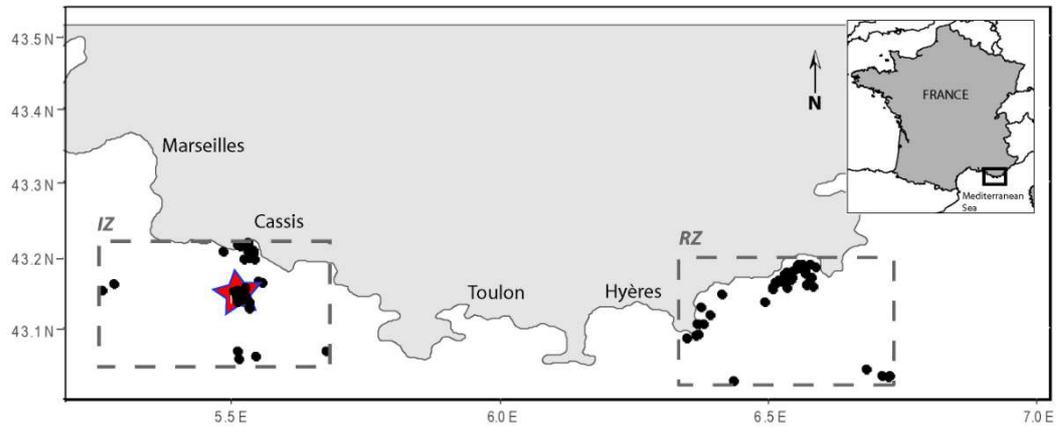
624 Zhao S, Feng CH, Quan WM, Chen XF, Niu JF, Shen ZY. Role of living environments in the
625 accumulation characteristics of heavy metals in fishes and crabs in the Yangtze River Estuary,
626 China. *Marine Pollution Bulletin* 2012; 64: 1163-1171.

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631 Figures
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633 **Figure 1:** Location of the sampling station and sampling zones. The star represents the Gardanne
634 alumina refinery pipeline outlet.

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Table 1: Mean TE concentrations \pm standard deviation (in mg.kg⁻¹ wm) measured in fish muscle from the red mud impacted (IZ) and the reference (RZ) zones. Minimum and maximum values are given in brackets.

Species	zone	Tissue	N	Total length (mm)	Total mass (g)	Al	As	Cd	Co	Cr	Hg
<i>Chelon labrosus</i>	IZ	muscle	1	406	860.3	0.320	1.659	0.000	0.002	0.021	0.128
<i>Conger conger</i>	IZ	muscle	31	710 \pm 171 (416 - 1119)	799.5 \pm 643.2 (94.5 - 2894.4)	0.062 \pm 0.159 (0.000 - 0.858)	22.115 \pm 11.886 (5.308 - 53.193)	0.003 \pm 0.007 (0.000 - 0.038)	0.002 \pm 0.003 (0.000 - 0.015)	0.021 \pm 0.029 (0.000 - 0.109)	0.38 \pm 0.339 (0.07 - 1.872)
<i>Conger conger</i>	RZ	muscle	23	716 \pm 203 (334 - 1035)	817.5 \pm 672.6 (50.8 - 2341.7)	0.032 \pm 0.102 (0.000 - 0.465)	26.867 \pm 19.009 (8.613 - 86.156)	0.003 \pm 0.008 (0.000 - 0.041)	0.001 \pm 0.001 (0.000 - 0.003)	0.017 \pm 0.023 (0.000 - 0.085)	0.345 \pm 0.218 (0.066 - 1.003)
<i>Coris julis</i>	IZ	muscle	32	128 \pm 1.9 (103 - 212)	20.9 \pm 12.5 (8.6 - 83.5)	0.106 \pm 0.112 (0.000 - 0.389)	4.213 \pm 2.266 (1.08 - 10.100)	0.001 \pm 0.001 (0.000 - 0.002)	0.007 \pm 0.003 (0.000 - 0.017)	0.031 \pm 0.035 (0.000 - 0.145)	0.148 \pm 0.105 (0.095 - 0.700)
<i>Coris julis</i>	RZ	muscle	31	125 \pm 14 (86 - 147)	17.0 \pm 5.3 (5.1 - 29.4)	0.149 \pm 0.237 (0.000 - 1.11)	7.633 \pm 7.786 (0.586 - 33.938)	0.004 \pm 0.015 (0.000 - 0.085)	0.009 \pm 0.004 (0.000 - 0.019)	0.014 \pm 0.019 (0.000 - 0.065)	0.174 \pm 0.245 (0.035 - 1.369)
<i>Dicentrarchus labrax</i>	RZ	muscle	1	349	346.6	0.000	0.675	0.000	0.002	0.027	0.222
<i>Diplodus sargus</i>	IZ	muscle	4	269 \pm 21 (235 - 289)	357.2 \pm 86.9 (214.6 - 433.3)	0.026 \pm 0.051 (0.000 - 0.102)	5.28 \pm 2.908 (2.203 - 9.221)	0.001 \pm 0.000 (0.001 - 0.001)	0.004 \pm 0.001 (0.003 - 0.004)	0.016 \pm 0.012 (0.000 - 0.028)	0.756 \pm 0.364 (0.429 - 1.264)
<i>Diplodus sargus</i>	RZ	muscle	39	245 \pm 45 (173 - 375)	308.7 \pm 189.1 (101 - 969.4)	0.385 \pm 1.784 (0.000 - 11.076)	11.911 \pm 6.165 (1.218 - 24.651)	0.001 \pm 0.001 (0.000 - 0.005)	0.007 \pm 0.008 (0.000 - 0.047)	0.009 \pm 0.016 (0.000 - 0.073)	0.429 \pm 0.296 (0.084 - 1.369)
<i>Diplodus vulgaris</i>	IZ	muscle	28	226 \pm 20 (189-279)	203.5 \pm 59.0 (83.6 - 359.4)	0.262 \pm 0.154 (0.000 - 0.683)	4.346 \pm 1.188 (1.585 - 7.514)	0.001 \pm 0.005 (0.000 - 0.024)	0.003 \pm 0.002 (0.000 - 0.010)	0.016 \pm 0.045 (0.000 - 0.228)	0.442 \pm 0.176 (0.124 - 0.786)
<i>Diplodus vulgaris</i>	RZ	muscle	53	213 \pm 24 (149 - 249)	164.0 \pm 56.8 (46.9 - 283.9)	0.052 \pm 0.083 (0.000 - 0.304)	6.919 \pm 3.132 (3.209 - 15.573)	0.001 \pm 0.001 (0.000 - 0.003)	0.005 \pm 0.002 (0.000 - 0.011)	0.016 \pm 0.019 (0.000 - 0.096)	0.389 \pm 0.159 (0.082 - 0.798)
<i>Engraulis encrasicolus</i>	IZ	muscle	60	139 \pm 8 (121-157)	16.8 \pm 2.9 (10.2 - 25.5)	0.030 \pm 0.074 (0.000 - 0.363)	3.804 \pm 0.759 (1.688 - 5.900)	0.005 \pm 0.002 (0.002 - 0.009)	0.01 \pm 0.002 (0.006 - 0.017)	0.019 \pm 0.018 (0.000 - 0.059)	0.119 \pm 0.046 (0.035 - 0.247)
<i>Engraulis encrasicolus</i>	RZ	muscle	6	118 \pm 11 (100 - 129)	10.6 \pm 2.7 (5.5 - 12.5)	1.769 \pm 2.503 (0.160 - 5.711)	4.664 \pm 0.846 (3.830 - 6.252)	0.005 \pm 0.002 (0.000 - 0.007)	0.016 \pm 0.004 (0.011 - 0.022)	0.035 \pm 0.019 (0.017 - 0.070)	0.084 \pm 0.02 (0.065 - 0.118)
<i>Galeus melastomus</i>	IZ	muscle	32	459 \pm 81 (323 - 578)	283.4 \pm 147.3 (81.8 - 545.7)	0.574 \pm 0.583 (0.000 - 3.090)	33.122 \pm 7.735 (22.517 - 55.173)	0.001 \pm 0.002 (0.000 - 0.006)	0.001 \pm 0.002 (0.000 - 0.007)	0.019 \pm 0.017 (0.000 - 0.074)	1.47 \pm 0.842 (0.553 - 3.833)
<i>Galeus melastomus</i>	RZ	muscle	4	327 \pm 31 (277 - 354)	89.8 \pm 23.1 (51.2 - 110.0)	1.923 \pm 1.016 (0.812 - 2.829)	28.503 \pm 9.637 (17.587 - 41.050)	0.001 \pm 0.000 (0.001 - 0.001)	0.003 \pm 0.001 (0.002 - 0.005)	0.038 \pm 0.005 (0.031 - 0.042)	0.908 \pm 0.139 (0.799 - 1.089)
<i>Helicolenus dactylopterus</i>	IZ	muscle	37	187 \pm 31 (114 - 248)	109.6 \pm 51.2 (22.2 - 264.9)	0.694 \pm 0.683 (0.137-3.364)	5.281 \pm 2.442 (2.111 - 12.100)	0.001 \pm 0.001 (0.000 - 0.006)	0.001 \pm 0.002 (0.000 - 0.005)	0.029 \pm 0.086 (0.000 - 0.525)	1.045 \pm 0.456 (0.273 - 2.730)
<i>Helicolenus dactylopterus</i>	RZ	muscle	46	194 \pm 33 (139-291)	130.7 \pm 72.2 (37.6 - 402)	0.217 \pm 0.301 (0.000 - 1.839)	10.166 \pm 5.484 (2.690 - 25.008)	0.001 \pm 0.001 (0.000 - 0.003)	0.003 \pm 0.002 (0.000 - 0.010)	0.013 \pm 0.024 (0.000 - 0.137)	1.099 \pm 0.368 (0.382 - 2.059)
<i>Liza aurata</i>	IZ	muscle	12	281 \pm 24 (241-330)	193.3 \pm 71.1 (111.7 - 388.8)	0.064 \pm 0.084 (0.000 - 0.217)	0.561 \pm 0.253 (0.285 - 0.960)	0.000 \pm 0.000 (0.000 - 0.000)	0.01 \pm 0.002 (0.006 - 0.013)	0.027 \pm 0.025 (0.000 - 0.076)	0.014 \pm 0.01 (0.000 - 0.028)
<i>Liza aurata</i>	RZ	muscle	7	328 \pm 25 (282 - 359)	307.3 \pm 65.6 (212.6 - 385.9)	0.050 \pm 0.089 (0.000 - 0.218)	1.302 \pm 0.477 (0.832 - 2.230)	0.000 \pm 0.000 (0.000 - 0.000)	0.011 \pm 0.006 (0.000 - 0.018)	0.024 \pm 0.036 (0.000 - 0.088)	0.021 \pm 0.012 (0.013 - 0.047)
<i>Liza ramada</i>	IZ	muscle	13	319 \pm 50 (239 - 389)	288.7 \pm 144.8 (105.7 - 506.2)	0.591 \pm 0.447 (0.101 - 1.770)	0.992 \pm 0.589 (0.243 - 2.091)	0.000 \pm 0.000 (0.000 - 0.000)	0.006 \pm 0.003 (0.002 - 0.010)	0.019 \pm 0.019 (0.000 - 0.06)	0.094 \pm 0.095 (0.007 - 0.321)
<i>Merluccius merluccius</i>	IZ	muscle	53	354 \pm 132 (233 - 795)	439.1 \pm 518.4 (89.1 - 2305.2)	0.460 \pm 0.474 (0.000 - 2.139)	6.512 \pm 2.707 (3.353 - 18.323)	0.002 \pm 0.007 (0.000 - 0.043)	0.001 \pm 0.005 (0.000 - 0.021)	0.005 \pm 0.011 (0.000 - 0.041)	0.454 \pm 0.524 (0.08 - 3.073)

<i>Merluccius merluccius</i>	RZ	muscle	24	384 ± 54 (280 - 476)	456.9 ± 185.6 (147.3 - 858.4)	0.131 ± 0.16 (0.000 - 0.487)	5.070 ± 2.194 (2.405 - 11.150)	0.000 ± 0.001 (0.000 - 0.004)	0.000 ± 0.000 (0.000 - 0.002)	0.006 ± 0.009 (0.000 - 0.026)	0.497 ± 0.39 (0.104 - 1.593)
<i>Mugil cephalus</i>	IZ	muscle	1	425	969.5	0.105	3.597	0.000	0.003	0.061	0.327
<i>Mullus barbatus</i>	IZ	muscle	77	171 ± 25 (121 - 230)	57.7 ± 27.7 (11.7 - 140.7)	1.209 ± 1.434 (0.000 - 9.152)	24.775 ± 7.771 (1.43 - 50.586)	0.001 ± 0.002 (0.000 - 0.013)	0.008 ± 0.004 (0.003 - 0.022)	0.017 ± 0.02 (0.000 - 0.091)	0.46 ± 0.338 (0.06 - 1.743)
<i>Mullus barbatus</i>	RZ	muscle	2	212 ± 28 (185 - 239)	115.0 ± 45.6 (70.4 - 159.5)	0.203 ± 0.163 (0.088 - 0.318)	13.222 ± 10.906 (5.510 - 20.933)	0.000 ± 0.000 (0.000 - 0.000)	0.007 ± 0.002 (0.005 - 0.008)	0.044 ± 0.037 (0.018 - 0.07)	0.217 ± 0.247 (0.042 - 0.391)
<i>Mullus surmuletus</i>	IZ	muscle	68	181 ± 29 (135 - 267)	75.2 ± 41.1 (27.2 - 236.1)	1.294 ± 3.004 (0.116 - 24.632)	15.112 ± 5.732 (5.647 - 34.549)	0.000 ± 0.001 (0.000 - 0.004)	0.005 ± 0.002 (0.002 - 0.016)	0.014 ± 0.015 (0.000 - 0.065)	0.426 ± 0.51 (0.102 - 2.585)
<i>Mullus surmuletus</i>	RZ	muscle	72	188 ± 30 (152 - 315)	88.4 ± 57.9 (39.1 - 418.8)	0.268 ± 0.453 (0.000 - 2.32)	11.828 ± 5.166 (3.090 - 28.257)	0.000 ± 0.000 (0.000 - 0.002)	0.004 ± 0.003 (0.000 - 0.012)	0.014 ± 0.022 (0.000 - 0.112)	0.132 ± 0.098 (0.022 - 0.403)
<i>Pagellus bogaraveo</i>	IZ	muscle	27	268 ± 19 (236 - 307)	267.9 ± 63.6 (149.6 - 420.6)	0.328 ± 0.436 (0.000 - 2.083)	5.87 ± 2.535 (2.250 - 11.900)	0.001 ± 0.002 (0.000 - 0.010)	0.004 ± 0.003 (0.000 - 0.012)	0.009 ± 0.015 (0.000 - 0.066)	0.318 ± 0.174 (0.092 - 1.041)
<i>Pagellus bogaraveo</i>	RZ	muscle	30	240 ± 10 (221 - 265)	204.5 ± 24.4 (167.5 - 273.8)	0.203 ± 0.157 (0.000 - 0.671)	5.417 ± 1.138 (3.212 - 7.746)	0.001 ± 0.001 (0.000 - 0.002)	0.006 ± 0.002 (0.003 - 0.014)	0.017 ± 0.041 (0.000 - 0.220)	0.329 ± 0.194 (0.183 - 0.995)
<i>Pagellus erythrinus</i>	RZ	muscle	6	172 ± 11 (154 - 188)	68.3 ± 10.7 (55.1 - 83.1)	0.948 ± 0.191 (0.743 - 1.27)	5.06 ± 1.275 (3.640 - 6.350)	0.000 ± 0.000 (0.000 - 0.000)	0.002 ± 0.003 (0.000 - 0.006)	0.003 ± 0.007 (0.000 - 0.016)	0.171 ± 0.049 (0.091 - 0.234)
<i>Raja clavata</i>	IZ	muscle	21	605 ± 116 (440 - 826)	1493.4 ± 940.0 (431.4 - 3820.1)	0.559 ± 1.143 (0.000 - 4.987)	76.051 ± 30.462 (1.404 - 119.085)	0.099 ± 0.300 (0.001 - 1.325)	0.013 ± 0.025 (0.000 - 0.088)	0.005 ± 0.009 (0.000 - 0.029)	1.185 ± 0.723 (0.156 - 3.765)
<i>Raja clavata</i>	RZ	muscle	5	749 ± 68 (669 - 849)	2293.3 ± 650.1 (1567.7 - 3181.4)	0.080 ± 0.129 (0.000 - 0.296)	89.171 ± 22.568 (68.400 - 119.268)	0.001 ± 0.001 (0.001 - 0.002)	0.002 ± 0.001 (0.000 - 0.003)	0.015 ± 0.014 (0.000 - 0.028)	1.429 ± 0.74 (0.739 - 2.323)
<i>Sardina pilchardus</i>	IZ	muscle	68	141 ± 11 (124 - 193)	21.3 ± 5.9 (12.5 - 54.7)	0.393 ± 0.968 (0.000 - 7.817)	5.805 ± 1.341 (4.028 - 12.264)	0.001 ± 0.002 (0.000 - 0.007)	0.011 ± 0.002 (0.007 - 0.016)	0.009 ± 0.014 (0.000 - 0.073)	0.1 ± 0.042 (0.046 - 0.334)
<i>Sardinella aurita</i>	RZ	muscle	2	240 ± 6 (234 - 246)	101.3 ± 6.9 (94.5 - 108.0)	0.068 ± 0.096 (0.000 - 0.136)	6.230 ± 1.061 (5.480 - 6.980)	0.001 ± 0.000 (0.001 - 0.001)	0.003 ± 0.004 (0.000 - 0.006)	0.007 ± 0.009 (0.000 - 0.013)	0.103 ± 0.000 (0.103 - 0.103)
<i>Scorpaena porcus</i>	IZ	muscle	39	187 ± 39 (129 - 262)	155.2 ± 99.7 (45.1 - 392.6)	0.808 ± 0.861 (0.000 - 3.689)	4.190 ± 2.439 (0.735 - 11.700)	0.001 ± 0.001 (0.000 - 0.006)	0.001 ± 0.001 (0.000 - 0.005)	0.013 ± 0.02 (0.000 - 0.096)	0.151 ± 0.049 (0.072 - 0.307)
<i>Scorpaena porcus</i>	RZ	muscle	71	160 ± 21 (119 - 218)	83.6 ± 37.4 (33.7 - 201.7)	0.778 ± 0.745 (0.108 - 3.74)	4.826 ± 3.0590 (0.577 - 12.600)	0.001 ± 0.001 (0.000 - 0.006)	0.003 ± 0.004 (0.000 - 0.015)	0.012 ± 0.026 (0.000 - 0.153)	0.171 ± 0.061 (0.072 - 0.361)
<i>Scyliorhinus canicula</i>	IZ	muscle	63	417 ± 59 (244 - 509)	245.1 ± 90.8 (39.4 - 400.4)	1.259 ± 1.727 (0.196 - 11.497)	37.735 ± 17.946 (15.030 - 112.802)	0.065 ± 0.221 (0.000 - 1.541)	0.012 ± 0.026 (0.000 - 0.163)	0.032 ± 0.026 (0.000 - 0.144)	1.613 ± 1.335 (0.233 - 8.955)
<i>Scyliorhinus canicula</i>	RZ	muscle	82	370 ± 62 (244 - 500)	183.6 ± 103.9 (41.3 - 485.4)	1.299 ± 0.995 (0.000 - 4.815)	41.341 ± 19.253 (0.020 - 118.615)	0.002 ± 0.003 (0.000 - 0.026)	0.007 ± 0.005 (0.000 - 0.034)	0.042 ± 0.034 (0.000 - 0.254)	0.845 ± 0.309 (0.000 - 1.862)
<i>Sparus aurata</i>	IZ	muscle	24	252 ± 15 (232 - 306)	233.9 ± 46.1 (183.6 - 419.6)	0.031 ± 0.075 (0.000 - 0.254)	4.816 ± 3.0760 (1.263 - 10.910)	0.000 ± 0.000 (0.000 - 0.002)	0.004 ± 0.002 (0.002 - 0.009)	0.009 ± 0.016 (0.000 - 0.069)	0.15 ± 0.087 (0.077 - 0.411)
<i>Sparus aurata</i>	RZ	muscle	4	261 ± 45 (223 - 336)	264.7 ± 150.2 (150.2 - 519.6)	0.147 ± 0.033 (0.117 - 0.189)	11.948 ± 8.455 (4.940 - 23.710)	0.000 ± 0.000 (0.000 - 0.000)	0.003 ± 0.001 (0.002 - 0.005)	0.009 ± 0.011 (0.000 - 0.024)	0.193 ± 0.166 (0.056 - 0.421)
<i>Trachurus mediterraneus</i>	IZ	muscle	8	331 ± 20 (307 - 366)	283.4 ± 49.6 (238 - 393.6)	0.441 ± 0.420 (0.000 - 1.081)	1.593 ± 1.122 (0.549 - 3.889)	0.002 ± 0.004 (0.000 - 0.011)	0.005 ± 0.001 (0.003 - 0.008)	0.023 ± 0.024 (0.000 - 0.074)	0.511 ± 0.133 (0.342 - 0.741)
<i>Trachurus mediterraneus</i>	RZ	muscle	30	272 ± 22 (247 - 318)	165.2 ± 41.8 (115.3 - 256.8)	0.298 ± 0.260 (0.000 - 0.921)	1.815 ± 1.113 (0.735 - 4.63)	0.001 ± 0.002 (0.000 - 0.009)	0.007 ± 0.005 (0.000 - 0.026)	0.011 ± 0.019 (0.000 - 0.071)	0.378 ± 0.135 (0.194 - 0.697)
<i>Trachurus trachurus</i>	IZ	muscle	34	337 ± 37 (281 - 425)	352.1 ± 130.8 (180 - 659.1)	0.176 ± 0.110 (0.000 - 0.427)	5.429 ± 2.0460 (1.478 - 10.51)	0.002 ± 0.005 (0.000 - 0.029)	0.008 ± 0.004 (0.000 - 0.022)	0.015 ± 0.03 (0.000 - 0.173)	0.947 ± 0.334 (0.399 - 1.88)
<i>Zeus faber</i>	IZ	muscle	36	322 ± 96 (131 - 511)	581.6 ± 514.5 (38.1 - 2278.2)	0.142 ± 0.340 (0.000 - 2.030)	0.619 ± 0.308 (0.302 - 1.406)	0.001 ± 0.004 (0.000 - 0.019)	0.001 ± 0.001 (0.000 - 0.004)	0.013 ± 0.03 (0.000 - 0.133)	0.569 ± 0.496 (0.046 - 2.251)

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<i>Zeus faber</i>	RZ	muscle	1	355	612.2	0.130	0.469	0.000	0.002	0.101	0.25
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663 **Table 1 (continued):** Mean TE concentrations \pm standard deviation (in mg.kg⁻¹ wm) measured in fish muscle from the zone (IZ) impacted by red mud and the
664 reference (RZ) zones. Minimum and maximum values are given in brackets.
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Species	zone	tissue	N	Total length (mm)	Total mass (g)	Mn	Ni	Pb	Ti	V
<i>Chelon labrosus</i>	IZ	muscle	1	406	860.3	0.069	0.000	0.005	0.001	0.001
<i>Conger conger</i>	IZ	muscle	31	710 \pm 171	799.5 \pm 643.2	0.346 \pm 0.260	0.000 \pm 0.000	0.007 \pm 0.007	0.067 \pm 0.114	0.006 \pm 0.0140
				(416 - 1119)	(94.5 - 2894.4)	(0.07 - 1.035)	(0.000 - 0.000)	(0.000 - 0.031)	(0.000 - 0.54)	(0.000 - 0.077)
<i>Conger conger</i>	RZ	muscle	23	716 \pm 203	817.5 \pm 672.6	0.515 \pm 0.383	0.000 \pm 0.000	0.006 \pm 0.008	0.070 \pm 0.115	0.004 \pm 0.004
				(334 - 1035)	(50.8 - 2341.7)	(0.058 - 1.273)	(0.000 - 0.000)	(0.000 - 0.031)	(0.000 - 0.466)	(0.000 - 0.017)
<i>Coris julis</i>	IZ	muscle	32	128 \pm 1.9	20.9 \pm 12.5	0.182 \pm 0.058	0.007 \pm 0.024	0.025 \pm 0.019	0.059 \pm 0.06	0.030 \pm 0.017
				(103 - 212)	(8.6 - 83.5)	(0.097 - 0.304)	(0.000 - 0.109)	(0.000 - 0.076)	(0.000 - 0.254)	(0.000 - 0.090)
<i>Coris julis</i>	RZ	muscle	31	125 \pm 14	17.0 \pm 5.3	0.196 \pm 0.074	0.003 \pm 0.011	0.014 \pm 0.012	0.114 \pm 0.127	0.048 \pm 0.043
				(86 - 147)	(5.1 - 29.4)	(0.107 - 0.415)	(0.000 - 0.051)	(0.000 - 0.043)	(0.000 - 0.513)	(0.007 - 0.191)
<i>Dicentrarchus labrax</i>	RZ	muscle	1	349	346.6	0.151	0.000	0.000	0.000	0.000
<i>Diplodus sargus</i>	IZ	muscle	4	269 \pm 21	357.2 \pm 86.9	0.064 \pm 0.01	0.000 \pm 0.000	0.005 \pm 0.007	0.000 \pm 0.000	0.007 \pm 0.003
				(235 - 289)	(214.6 - 433.3)	(0.056 - 0.076)	(0.000 - 0.000)	(0.000 - 0.015)	(0.000 - 0.000)	(0.003 - 0.011)
<i>Diplodus sargus</i>	RZ	muscle	39	245 \pm 45	308.7 \pm 189.1	0.061 \pm 0.032	0.015 \pm 0.065	0.023 \pm 0.121	0.044 \pm 0.086	0.004 \pm 0.004
				(173 - 375)	(101 - 969.4)	(0.035 - 0.177)	(0.000 - 0.382)	(0.000 - 0.757)	(0.000 - 0.509)	(0.000 - 0.020)
<i>Diplodus vulgaris</i>	IZ	muscle	28	226 \pm 20	203.5 \pm 59.0	0.115 \pm 0.051	0.017 \pm 0.068	0.024 \pm 0.070	0.019 \pm 0.038	0.016 \pm 0.023
				(189-279)	(83.6 - 359.4)	(0.067 - 0.266)	(0.000 - 0.334)	(0.000 - 0.368)	(0.000 - 0.131)	(0.000 - 0.13)
<i>Diplodus vulgaris</i>	RZ	muscle	53	213 \pm 24	164.0 \pm 56.8	0.102 \pm 0.082	0.000 \pm 0.000	0.025 \pm 0.029	0.019 \pm 0.036	0.019 \pm 0.0260
				(149 - 249)	(46.9 - 283.9)	(0.032 - 0.475)	(0.000 - 0.000)	(0.000 - 0.125)	(0.000 - 0.197)	(0.000 - 0.154)
<i>Engraulis encrasicolus</i>	IZ	muscle	60	139 \pm 8	16.8 \pm 2.9	0.345 \pm 0.091	0.006 \pm 0.025	0.004 \pm 0.002	0.045 \pm 0.051	0.021 \pm 0.012
				(121-157)	(10.2 - 25.5)	(0.039 - 0.533)	(0.000 - 0.157)	(0.000 - 0.012)	(0.000 - 0.287)	(0.002 - 0.073)
<i>Engraulis encrasicolus</i>	RZ	muscle	6	118 \pm 11	10.6 \pm 2.7	0.478 \pm 0.210	0.028 \pm 0.023	0.072 \pm 0.100	0.115 \pm 0.105	0.032 \pm 0.021
				(100 - 129)	(5.5 - 12.5)	(0.262 - 0.879)	(0.000 - 0.051)	(0.000 - 0.216)	(0.039 - 0.257)	(0.018 - 0.072)
<i>Galeus melastomus</i>	IZ	muscle	32	459 \pm 81	283.4 \pm 147.3	0.118 \pm 0.024	0.007 \pm 0.036	0.053 \pm 0.068	0.191 \pm 0.147	0.003 \pm 0.004
				(323 - 578)	(81.8 - 545.7)	(0.081 - 0.198)	(0.000 - 0.201)	(0.005 - 0.305)	(0.054 - 0.754)	(0.000 - 0.019)
<i>Galeus melastomus</i>	RZ	muscle	4	327 \pm 31	89.8 \pm 23.1	0.210 \pm 0.098	0.000 \pm 0.000	0.141 \pm 0.104	0.218 \pm 0.03	0.005 \pm 0.002
				(277 - 354)	(51.2 - 110.0)	(0.140 - 0.354)	(0.000 - 0.000)	(0.035 - 0.280)	(0.182 - 0.247)	(0.002 - 0.006)
<i>Helicolenus dactylopterus</i>	IZ	muscle	37	187 \pm 31	109.6 \pm 51.2	0.096 \pm 0.037	0.006 \pm 0.036	0.086 \pm 0.100	0.127 \pm 0.111	0.004 \pm 0.005
				(114 - 248)	(22.2 - 264.9)	(0.055 - 0.276)	(0.000 - 0.221)	(0.006 - 0.435)	(0.000 - 0.647)	(0.000 - 0.026)
<i>Helicolenus dactylopterus</i>	RZ	muscle	46	194 \pm 33	130.7 \pm 72.2	0.086 \pm 0.025	0.004 \pm 0.019	0.003 \pm 0.003	0.038 \pm 0.031	0.002 \pm 0.002
				(139-291)	(37.6 - 402)	(0.051 - 0.167)	(0.000 - 0.103)	(0.000 - 0.012)	(0.000 - 0.105)	(0.000 - 0.009)
<i>Liza aurata</i>	IZ	muscle	12	281 \pm 24	193.3 \pm 71.1	0.094 \pm 0.027	0.010 \pm 0.023	0.001 \pm 0.003	0.021 \pm 0.036	0.003 \pm 0.003
				(241-330)	(111.7 - 388.8)	(0.064 - 0.155)	(0.000 - 0.060)	(0.000 - 0.008)	(0.000 - 0.114)	(0.000 - 0.011)
<i>Liza aurata</i>	RZ	muscle	7	328 \pm 25	307.3 \pm 65.6	0.093 \pm 0.054	0.018 \pm 0.025	0.008 \pm 0.01	0.010 \pm 0.017	0.007 \pm 0.005
				(282 - 359)	(212.6 - 385.9)	(0.057 - 0.196)	(0.000 - 0.053)	(0.000 - 0.029)	(0.000 - 0.036)	(0.000 - 0.013)
<i>Merluccius merluccius</i>	IZ	liver	48	364 \pm 135	471.6 \pm 534.3	1.69 \pm 0.545	0.008 \pm 0.027	0.015 \pm 0.019	0.130 \pm 0.229	0.034 \pm 0.030
				(233 - 795)	(97.7 - 2305.2)	(0.081 - 2.748)	(0.000 - 0.147)	(0.000 - 0.107)	(0.000 - 1.27)	(0.000 - 0.135)
<i>Merluccius merluccius</i>	IZ	muscle	53	354 \pm 132	439.1 \pm 518.4	0.145 \pm 0.194	0.031 \pm 0.186	0.01 \pm 0.044	0.018 \pm 0.042	0.002 \pm 0.003
				(233 - 795)	(89.1 - 2305.2)	(0.067 - 1.151)	(0.000 - 1.314)	(0.000 - 0.308)	(0.000 - 0.229)	(0.000 - 0.022)

<i>Merluccius merluccius</i>	RZ	muscle	24	384 ± 54 (280 - 476)	456.9 ± 185.6 (147.3 - 858.4)	0.082 ± 0.018 (0.054 - 0.121)	0.023 ± 0.111 (0.000 - 0.544)	0.026 ± 0.063 (0.000 - 0.293)	0.011 ± 0.021 (0.000 - 0.057)	0.000 ± 0.001 (0.000 - 0.002)
<i>Mugil cephalus</i>	IZ	muscle	1	425	969.5	0.055	0.000	0.009	0.001	0.001
<i>Mullus barbatus</i>	IZ	muscle	77	171 ± 25 (121 - 230)	57.7 ± 27.7 (11.7 - 140.7)	0.159 ± 0.048 (0.05 - 0.332)	0.005 ± 0.02 (0.000 - 0.101)	0.014 ± 0.023 (0.000 - 0.192)	0.076 ± 0.069 (0.000 - 0.35)	0.006 ± 0.006 (0.002 - 0.046)
<i>Mullus barbatus</i>	RZ	muscle	2	212 ± 28 (185 - 239)	115.0 ± 45.6 (70.4 - 159.5)	0.119 ± 0.025 (0.101 - 0.136)	0.000 ± 0.000 (0.000 - 0.000)	0.048 ± 0.006 (0.043 - 0.052)	0.050 ± 0.071 (0.000 - 0.100)	0.003 ± 0.001 (0.002 - 0.004)
<i>Mullus surmuletus</i>	IZ	muscle	68	181 ± 29 (135 - 267)	75.2 ± 41.1 (27.2 - 236.1)	0.17 ± 0.058 (0.098 - 0.442)	0.001 ± 0.01 (0.000 - 0.084)	0.01 ± 0.018 (0.000 - 0.082)	0.058 ± 0.051 (0.000 - 0.212)	0.005 ± 0.003 (0.000 - 0.020)
<i>Mullus surmuletus</i>	RZ	muscle	72	188 ± 30 (152 - 315)	88.4 ± 57.9 (39.1 - 418.8)	0.136 ± 0.041 (0.076 - 0.266)	0.002 ± 0.010 (0.000 - 0.069)	0.016 ± 0.011 (0.000 - 0.048)	0.045 ± 0.047 (0.000 - 0.228)	0.002 ± 0.003 (0.000 - 0.020)
<i>Pagellus bogaraveo</i>	IZ	muscle	27	268 ± 19 (236 - 307)	267.9 ± 63.6 (149.6 - 420.6)	0.098 ± 0.032 (0.067 - 0.21)	0.012 ± 0.060 (0.000 - 0.312)	0.023 ± 0.037 (0.000 - 0.190)	0.092 ± 0.109 (0.000 - 0.547)	0.002 ± 0.003 (0.000 - 0.013)
<i>Pagellus bogaraveo</i>	RZ	muscle	30	240 ± 10 (221 - 265)	240.5 ± 24.4 (167.5 - 273.8)	0.101 ± 0.061 (0.059 - 0.375)	0.000 ± 0.000 (0.000 - 0.000)	0.029 ± 0.035 (0.004 - 0.130)	0.012 ± 0.035 (0.000 - 0.166)	0.001 ± 0.001 (0.000 - 0.004)
<i>Pagellus erythrinus</i>	RZ	muscle	6	172 ± 11 (154 - 188)	68.3 ± 10.7 (55.1 - 83.1)	0.112 ± 0.028 (0.076 - 0.148)	0.000 ± 0.000 (0.000 - 0.000)	0.082 ± 0.046 (0.042 - 0.162)	0.116 ± 0.064 (0.071 - 0.208)	0.004 ± 0.006 (0.000 - 0.012)
<i>Raja clavata</i>	IZ	muscle	21	605 ± 116 (440 - 826)	1493.4 ± 940.0 (431.4 - 3820.1)	0.309 ± 0.279 (0.116 - 1.047)	0.000 ± 0.000 (0.000 - 0.000)	0.004 ± 0.009 (0.000 - 0.037)	0.030 ± 0.044 (0.000 - 0.166)	0.014 ± 0.045 (0.000 - 0.202)
<i>Raja clavata</i>	RZ	muscle	5	749 ± 68 (669 - 849)	2293.3 ± 650.1 (1567.7 - 3181.4)	0.160 ± 0.023 (0.145 - 0.200)	0.000 ± 0.000 (0.000 - 0.000)	0.006 ± 0.011 (0.000 - 0.025)	0.000 ± 0.000 (0.000 - 0.000)	0.000 ± 0.001 (0.000 - 0.002)
<i>Sardina pilchardus</i>	IZ	muscle	68	141 ± 11 (124 - 193)	21.3 ± 5.9 (12.5 - 54.7)	0.414 ± 0.098 (0.118 - 0.744)	0.023 ± 0.024 (0.000 - 0.115)	0.01 ± 0.004 (0.000 - 0.024)	0.066 ± 0.039 (0.000 - 0.246)	0.038 ± 0.023 (0.000 - 0.092)
<i>Sardinella aurita</i>	RZ	muscle	2	240 ± 6 (234 - 246)	101.3 ± 6.9 (94.5 - 108.0)	0.829 ± 0.021 (0.814 - 0.844)	0.000 ± 0.000 (0.000 - 0.000)	0.021 ± 0.012 (0.012 - 0.029)	0.053 ± 0.028 (0.033 - 0.073)	0.02 ± 0.028 (0.000 - 0.04)
<i>Scorpaena porcus</i>	IZ	muscle	39	187 ± 39 (129 - 262)	155.2 ± 99.7 (45.1 - 392.6)	0.117 ± 0.069 (0.059 - 0.468)	0.003 ± 0.010 (0.000 - 0.050)	0.04 ± 0.047 (0.000 - 0.188)	0.096 ± 0.066 (0.000 - 0.377)	0.003 ± 0.004 (0.000 - 0.014)
<i>Scorpaena porcus</i>	RZ	muscle	71	160 ± 21 (119 - 218)	83.6 ± 37.4 (33.7 - 201.7)	0.119 ± 0.045 (0.063 - 0.350)	0.006 ± 0.016 (0.000 - 0.089)	0.041 ± 0.054 (0.000 - 0.325)	0.107 ± 0.057 (0.035 - 0.252)	0.004 ± 0.006 (0.000 - 0.025)
<i>Scyliorhinus canicula</i>	IZ	muscle	63	417 ± 59 (244 - 509)	245.1 ± 90.8 (39.4 - 400.4)	0.272 ± 0.301 (0.121 - 1.794)	0.002 ± 0.013 (0.000 - 0.084)	0.024 ± 0.045 (0.000 - 0.321)	0.112 ± 0.113 (0.000 - 0.435)	0.009 ± 0.014 (0.000 - 0.087)
<i>Scyliorhinus canicula</i>	RZ	muscle	82	370 ± 62 (244 - 500)	183.6 ± 103.9 (41.3 - 485.4)	0.226 ± 0.082 (0.000 - 0.726)	0.013 ± 0.062 (0.000 - 0.521)	0.039 ± 0.037 (0.000 - 0.248)	0.117 ± 0.108 (0.000 - 0.678)	0.009 ± 0.01 (0.000 - 0.057)
<i>Sparus aurata</i>	IZ	muscle	24	252 ± 15 (232 - 306)	233.9 ± 46.1 (183.6 - 419.6)	0.089 ± 0.020 (0.058 - 0.127)	0.022 ± 0.108 (0.000 - 0.527)	0.003 ± 0.004 (0.000 - 0.014)	0.014 ± 0.028 (0.000 - 0.093)	0.001 ± 0.001 (0.000 - 0.004)
<i>Sparus aurata</i>	RZ	muscle	4	261 ± 45 (223 - 336)	264.7 ± 150.2 (150.2 - 519.6)	0.070 ± 0.023 (0.036 - 0.090)	0.000 ± 0.000 (0.000 - 0.000)	0.007 ± 0.005 (0.000 - 0.012)	0.059 ± 0.040 (0.000 - 0.088)	0.002 ± 0.001 (0.000 - 0.002)
<i>Trachurus mediterraneus</i>	IZ	muscle	8	331 ± 20 (307 - 366)	283.4 ± 49.6 (238 - 393.6)	0.085 ± 0.022 (0.049 - 0.111)	0.000 ± 0.000 (0.000 - 0.000)	0.007 ± 0.008 (0.000 - 0.021)	0.045 ± 0.062 (0.000 - 0.177)	0.002 ± 0.001 (0.001 - 0.003)
<i>Trachurus mediterraneus</i>	RZ	muscle	30	272 ± 22 (247 - 318)	165.2 ± 41.8 (115.3 - 256.8)	0.102 ± 0.019 (0.057 - 0.135)	0.024 ± 0.046 (0.000 - 0.170)	0.02 ± 0.017 (0.002 - 0.083)	0.061 ± 0.069 (0.000 - 0.35)	0.002 ± 0.002 (0.000 - 0.006)
<i>Trachurus trachurus</i>	IZ	muscle	34	337 ± 37 (281 - 425)	352.1 ± 130.8 (180 - 659.1)	0.096 ± 0.025 (0.071 - 0.197)	0.016 ± 0.062 (0.000 - 0.326)	0.008 ± 0.013 (0.000 - 0.056)	0.037 ± 0.050 (0.000 - 0.195)	0.001 ± 0.001 (0.000 - 0.004)
<i>Zeus faber</i>	IZ	muscle	36	322 ± 96 (131 - 511)	581.6 ± 514.5 (38.1 - 2278.2)	0.086 ± 0.040 (0.035 - 0.164)	0.010 ± 0.040 (0.000 - 0.223)	0.005 ± 0.016 (0.000 - 0.071)	0.021 ± 0.050 (0.000 - 0.271)	0.000 ± 0.000 (0.000 - 0.002)

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<i>Zeus faber</i>	RZ	muscle	1	355	612.2	0.059	0.000	0.004	0.000	0.000
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668 **Table 2:** Mean TE concentrations \pm standard deviation (in $\text{mg}\cdot\text{kg}^{-1}\text{wm}$) measured in fish liver from the zone (IZ) impacted by red mud and the reference (RZ)
 669 zones. Minimum and maximum values are given in brackets.
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Species	zone	Tissue	N	Total length (mm)	Total mass (g)	Al	As	Cd	Co	Cr	Hg
<i>Conger conger</i>	IZ	liver	30	696 \pm 157	729.7 \pm 525.3	0.173 \pm 0.131	19.809 \pm 9.831	0.117 \pm 0.082	0.021 \pm 0.007	0.07 \pm 0.084	0.331 \pm 0.203
				(416 - 1020)	(94.5 - 2161.8)	(0.000 - 0.535)	(5.278 - 48.487)	(0.000 - 0.263)	(0.000 - 0.377)	(0.042 - 1.033)	
<i>Conger conger</i>	RZ	liver	21	691 \pm 195	817.5 \pm 672.6	0.334 \pm 0.458	24.035 \pm 14.403	0.117 \pm 0.127	0.019 \pm 0.009	0.073 \pm 0.208	0.286 \pm 0.211
				(334 - 1010)	(50.8 - 2341.7)	(0.000 - 1.931)	(6.931 - 59.475)	(0.001 - 0.503)	(0.000 - 0.038)	(0.000 - 0.950)	(0.055 - 0.867)
<i>Merluccius merluccius</i>	IZ	liver	48	364 \pm 135	471.6 \pm 534.3	1.963 \pm 2.563	5.194 \pm 1.859	0.071 \pm 0.101	0.033 \pm 0.016	0.028 \pm 0.044	0.442 \pm 0.729
				(233 - 795)	(97.7 - 2305.2)	(0.000 - 14.479)	(2.803 - 12.569)	(0.000 - 0.514)	(0.000 - 0.072)	(0.000 - 0.256)	(0.06 - 4.644)
<i>Merluccius merluccius</i>	RZ	liver	23	388 \pm 51	469.7 \pm 178.9	0.922 \pm 0.699	3.961 \pm 1.217	0.06 \pm 0.049	0.025 \pm 0.012	0.026 \pm 0.054	0.261 \pm 0.219
				(284 - 476)	(147.3 - 858.4)	(0.000 - 3.13)	(2.413 - 8.010)	(0.012 - 0.208)	(0.006 - 0.053)	(0.000 - 0.165)	(0.064 - 0.972)
<i>Raja clavata</i>	IZ	liver	21	605 \pm 116	1493.4 \pm 940.0	0.819 \pm 0.712	33.368 \pm 20.832	0.291 \pm 0.278	0.04 \pm 0.023	0.005 \pm 0.008	0.666 \pm 0.613
				(440 - 826)	(431.4 - 3820.1)	(0.000 - 2.885)	(13.88 - 81.629)	(0.001 - 1.255)	(0.000 - 0.090)	(0.000 - 0.029)	(0.261 - 2.476)
<i>Raja clavata</i>	RZ	liver	5	749 \pm 68	2293.3 \pm 650.1	1.539 \pm 1.146	30.499 \pm 4.233	0.346 \pm 0.230	0.078 \pm 0.025	0.006 \pm 0.013	0.669 \pm 0.521
				(669 - 849)	(1567.7 - 3181.4)	(0.463 - 3.349)	(26.315 - 36.691)	(0.130 - 0.653)	(0.046 - 0.104)	(0.000 - 0.028)	(0.237 - 1.425)
<i>Scyliorhinus canicula</i>	IZ	liver	60	417 \pm 58	244.1 \pm 91.2	1.873 \pm 1.727	29.800 \pm 16.703	0.202 \pm 0.220	0.059 \pm 0.042	0.036 \pm 0.064	1.129 \pm 1.191
				(244 - 509)	(39.4 - 400.4)	(0.238 - 7.033)	(3.827 - 81.501)	(0.000 - 0.971)	(0.003 - 0.186)	(0.000 - 0.393)	(0.121 - 6.146)
<i>Scyliorhinus canicula</i>	RZ	liver	81	370 \pm 62	182.7 \pm 104.2	2.594 \pm 2.362	27.772 \pm 12.341	0.167 \pm 0.149	0.068 \pm 0.034	0.036 \pm 0.037	0.496 \pm 0.383
				(244 - 500)	(41.3 - 485.4)	(0.466 - 14.108)	(2.900 - 63.125)	(0.001 - 0.857)	(0.026 - 0.168)	(0.000 - 0.169)	(0.077 - 2.182)

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680 **Table 2 (continued):** Mean TE concentrations \pm standard deviation (in mg.kg⁻¹ wm) measured in fish liver from the zone (IZ) impacted by red mud and the
 681 reference (RZ) zones. Minimum and maximum values are given in brackets.
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Species	zone	tissue	N	Total length (mm)	Total mass (g)	Mn	Ni	Pb	Ti	V
<i>Conger conger</i>	IZ	liver	30	696 \pm 157 (416 - 1020)	729.7 \pm 525.3 (94.5 - 2161.8)	0.934 \pm 0.214 (0.280 - 1.361)	0.022 \pm 0.045 (0.000 - 0.186)	0.069 \pm 0.044 (0.000 - 0.169)	0.021 \pm 0.043 (0.000 - 0.151)	0.096 \pm 0.064 (0.003 - 0.331)
<i>Conger conger</i>	RZ	liver	21	691 \pm 195 (334 - 1010)	817.5 \pm 672.6 (50.8 - 2341.7)	1.119 \pm 0.213 (0.768 - 1.553)	0.000 \pm 0.000 (0.000 - 0.000)	0.102 \pm 0.150 (0.008 - 0.657)	0.055 \pm 0.079 (0.000 - 0.208)	0.131 \pm 0.096 (0.006 - 0.368)
<i>Merluccius merluccius</i>	IZ	liver	48	364 \pm 135 (233 - 795)	471.6 \pm 534.3 (97.7 - 2305.2)	1.69 \pm 0.545 (0.081 - 2.748)	0.008 \pm 0.027 (0.000 - 0.147)	0.015 \pm 0.019 (0.000 - 0.107)	0.130 \pm 0.229 (0.000 - 1.27)	0.034 \pm 0.030 (0.000 - 0.135)
<i>Merluccius merluccius</i>	RZ	liver	23	388 \pm 51 (284 - 476)	469.7 \pm 178.9 (147.3 - 858.4)	1.444 \pm 0.383 (0.924 - 2.629)	0.014 \pm 0.037 (0.000 - 0.113)	0.015 \pm 0.025 (0.000 - 0.117)	0.022 \pm 0.035 (0.000 - 0.095)	0.017 \pm 0.012 (0.002 - 0.042)
<i>Raja clavata</i>	IZ	liver	21	605 \pm 116 (440 - 826)	1493.4 \pm 940.0 (431.4 - 3820.1)	0.798 \pm 0.336 (0.129 - 1.506)	0.003 \pm 0.014 (0.000 - 0.066)	0.015 \pm 0.012 (0.000 - 0.049)	0.043 \pm 0.062 (0.000 - 0.255)	0.051 \pm 0.049 (0.000 - 0.231)
<i>Raja clavata</i>	RZ	liver	5	749 \pm 68 (669 - 849)	2293.3 \pm 650.1 (1567.7 - 3181.4)	0.859 \pm 0.185 (0.571 - 1.023)	0.013 \pm 0.028 (0.000 - 0.063)	0.019 \pm 0.008 (0.008 - 0.027)	0.036 \pm 0.056 (0.000 - 0.128)	0.081 \pm 0.044 (0.04 - 0.151)
<i>Scyliorhinus canicula</i>	IZ	liver	60	417 \pm 58 (244 - 509)	244.1 \pm 91.2 (39.4 - 400.4)	0.873 \pm 0.435 (0.208 - 2.165)	0.012 \pm 0.042 (0.000 - 0.211)	0.026 \pm 0.017 (0.003 - 0.079)	0.078 \pm 0.084 (0.000 - 0.343)	0.038 \pm 0.034 (0.002 - 0.168)
<i>Scyliorhinus canicula</i>	RZ	liver	81	370 \pm 62 (244 - 500)	182.7 \pm 104.2 (41.3 - 485.4)	1.147 \pm 0.437 (0.488 - 3.143)	0.011 \pm 0.026 (0.000 - 0.112)	0.029 \pm 0.02 (0.007 - 0.151)	0.074 \pm 0.084 (0.000 - 0.442)	0.129 \pm 0.332 (0.007 - 2.421)

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692 **Table 3:** Speciation of arsenic in muscle of fish collected from the zone (IZ) impacted by red mud and the reference zone (RZ) (in mg.kg⁻¹ wm). Asi = inorganic
 693 arsenic (= As(III) because As(V) was not found in this study), MMA = monomethylarsonic acid, DMA = dimethylarsinic acid, AsB = arsenobetaine. Minimum
 694 and maximum values are given between brackets.
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Species	Zone	N	Length (mm)	mass (g)	Total As	Asi	MMA	DMA	AsB
<i>Diplodus sargus</i>	IZ	3	264 ± 26 (235 - 285)	335 ± 108.2 (214.6 - 424)	6.23 ± 4.40 (2.34 - 11)	0.033 ± 0.019 (0.013 - 0.048)	0.001 ± 0.001 (0.000 - 0.002)	0.004 ± 0.006 (0.000 - 0.011)	5.69 ± 4.39 (1.81 - 10.5)
<i>Diplodus sargus</i>	RZ	7	245 ± 32 (191 - 282)	311.6 ± 121.5 (151 - 481.1)	11.73 ± 6.60 (2.8 - 22.5)	0.037 ± 0.015 (0.022 - 0.059)	0.000 ± 0.000 (0.000 - 0.000)	0.001 ± 0.002 (0.000 - 0.005)	10.58 ± 6.71 (2.01 - 21.7)
<i>Diplodus vulgaris</i>	IZ	5	227 ± 10 (213 - 239)	204.4 ± 33.6 (165.8 - 240)	4.44 ± 1.42 (2.64 - 6.41)	0.06 ± 0.015 (0.037 - 0.073)	0.000 ± 0.000 (0.000 - 0.000)	0.005 ± 0.012 (0.000 - 0.026)	4.35 ± 1.56 (2.43 - 6.52)
<i>Diplodus vulgaris</i>	RZ	8	213 ± 9 (201 - 225)	164.6 ± 19.3 (145.3 - 195.6)	8.93 ± 4.08 (4.42 - 15.9)	0.081 ± 0.009 (0.069 - 0.099)	0.000 ± 0.001 (0.000 - 0.002)	0.007 ± 0.002 (0.002 - 0.009)	8.43 ± 4.23 (3.80 - 15.5)
<i>Helicolenus dactylopterus</i>	IZ	5	186 ± 14 (168 - 201)	108.7 ± 24.6 (76.8 - 135)	5.52 ± 2.71 (2.46 - 9.18)	0.021 ± 0.009 (0.014 - 0.033)	0.000 ± 0.000 (0.000 - 0.000)	0.001 ± 0.001 (0.000 - 0.002)	4.33 ± 2.45 (1.79 - 7.61)
<i>Helicolenus dactylopterus</i>	RZ	6	191 ± 21 (163 - 219)	127 ± 36.3 (86.3 - 177.4)	11.38 ± 7.30 (3.7 - 23.6)	0.04 ± 0.029 (0.017 - 0.095)	0.006 ± 0.01 (0.000 - 0.019)	0.001 ± 0.001 (0.000 - 0.002)	7.82 ± 5.74 (2.48 - 18.0)
<i>Pagellus bogaraveo</i>	IZ	7	269 ± 11 (259 - 286)	267.4 ± 38.8 (240.5 - 352.5)	6.03 ± 2.31 (2.99 - 9.9)	0.057 ± 0.009 (0.044 - 0.072)	0.000 ± 0.001 (0.000 - 0.002)	0.006 ± 0.003 (0.000 - 0.01)	2.39 ± 0.71 (1.44 - 3.66)
<i>Pagellus bogaraveo</i>	RZ	7	240 ± 5 (229 - 246)	203.9 ± 12.4 (180.8 - 219.3)	5.67 ± 1.55 (3.51 - 8.31)	0.049 ± 0.02 (0.029 - 0.09)	0.003 ± 0.007 (0.000 - 0.018)	0.009 ± 0.007 (0.000 - 0.022)	2.79 ± 1.31 (2.02 - 5.67)
<i>Pagellus erythrinus</i>	RZ	2	172 ± 3 (170 - 174)	68.3 ± 7.5 (63 - 73.6)	4.73 ± 1.51 (3.67 - 5.8)	0.041 ± 0.001 (0.04 - 0.042)	0.006 ± 0.001 (0.005 - 0.007)	0.013 ± 0.007 (0.008 - 0.018)	4.58 ± 1.61 (3.44 - 5.71)
<i>Raja clavata</i>	IZ	5	604 ± 72 (497 - 699)	1474 ± 628.7 (677.6 - 2349.3)	98.50 ± 19.94 (86 - 132)	0.033 ± 0.015 (0.016 - 0.057)	0.053 ± 0.021 (0.021 - 0.072)	0.000 ± 0.000 (0.000 - 0.000)	95.7 ± 19.9 (80.1 - 125)
<i>Raja clavata</i>	RZ	2	744 ± 38 (717 - 771)	2293.3 ± 286 (2091 - 2495.5)	100.5 ± 27.58 (81 - 120)	0.018 ± 0.003 (0.016 - 0.021)	0.084 ± 0.045 (0.052 - 0.116)	0.000 ± 0.000 (0.000 - 0.000)	110 ± 26.8 (91.4 - 129)
<i>Sparus aurata</i>	IZ	7	253 ± 10 (242 - 272)	233.9 ± 27.1 (204.9 - 289.9)	5.82 ± 3.29 (1.59 - 10.2)	0.033 ± 0.026 (0.011 - 0.088)	0.000 ± 0.000 (0.000 - 0.000)	0.007 ± 0.003 (0.002 - 0.013)	5.91 ± 3.92 (0.971 - 11.6)
<i>Sparus aurata</i>	RZ	2	261 ± 26 (243 - 280)	264.7 ± 99.5 (194.4 - 335.1)	12.62 ± 10.02 (5.53 - 19.7)	0.032 ± 0.002 (0.031 - 0.033)	0.000 ± 0.000 (0.000 - 0.000)	0.004 ± 0.005 (0.000 - 0.007)	13.1 ± 10.1 (5.99 - 20.3)

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Table 4: Pearson correlation coefficients between TE concentrations and length in the muscle and the liver of fish collected in this study. Significant differences are highlighted by bold characters (*p < 0.05, **p < 0.01, ***p < 0.001)

Species	Tissue	N IZ	N RZ	Al	As	Cd	Co	Cr	Hg	Mn	Ni	Pb	Ti	V
<i>Conger conger</i>	muscle	31	23	-0.057	0.56***	-0.036	-0.058	-0.337*	0.535***	-0.293*		-0.42**	-0.469***	-0.290*
<i>Coris julis</i>	muscle	32	31	-0.072	-0.258*	-0.044	-0.347**	-0.067	0.07	-0.252*	-0.080	-0.163	-0.042	-0.458***
<i>Diplodus vulgaris</i>	muscle	28	53	0.096	0.075	0.116	-0.162	-0.041	0.738***	-0.116	0.308**	-0.342**	-0.344**	-0.125
<i>Helicolenus dactylopterus</i>	muscle	37	46	-0.251*	-0.049	-0.101	-0.193	0.007	0.529***	-0.577***	0.021	-0.289**	0.133	-0.241*
<i>Merluccius merluccius</i>	muscle	53	24	-0.320**	-0.084	0.027	0.041	-0.016	0.851***	-0.043	-0.038	0.272*	0.147	-0.077
<i>Mullus surmuletus</i>	muscle	68	72	-0.167*	0.136	0.077	0.181*	0.13	0.470***	-0.004	0.052	-0.034	-0.043	0.122
<i>Pagellus bogaraveo</i>	muscle	27	30	0.004	0.270*	0.091	-0.274*	-0.043	0.226	0.045	0.081	0.087	0.152	-0.033
<i>Scorpaena porcus</i>	muscle	39	71	-0.363***	0.151	-0.171	-0.167	-0.046	0.338***	-0.163	-0.112	-0.311**	-0.357***	-0.382***
<i>Scyliorhinus canicula</i>	muscle	63	82	-0.120	0.250**	0.206	0.119	-0.352***	0.467***	0.166*	-0.167*	-0.004	-0.343***	0.092
<i>Conger conger</i>	liver	30	21	-0.197	0.472***	0.158	0.525***	-0.496***	0.444**	0.151	-0.214	-0.270	-0.248	0.323*
<i>Merluccius merluccius</i>	liver	48	23	0.402**	-0.186	0.583***	0.254*	-0.173	0.748***	-0.449***	-0.170	0.276*	0.472***	0.555***
<i>Scyliorhinus canicula</i>	liver	60	81	-0.182*	0.370***	0.337***	0.299***	-0.309***	0.434***	-0.260**	-0.211*	0.1	-0.314***	0.177*

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Graphical abstract

