

Serum IgG Concentrations in Adult Patients Experiencing Virus-Induced Severe Asthma Exacerbations

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1 Serum IgG concentrations in adult patients experiencing virus-induced severe asthma

2 exacerbations

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30 Abstract:

Background: Patients experiencing severe asthma exacerbations have a poorer quality of life
and an increase in morbidity and mortality. Viruses are frequently involved in asthma
exacerbations.

34 Objective: Determining the value of measuring serum immunoglobulin G (IgG) 35 concentrations in asthma exacerbations and their link with viral infections in patients 36 hospitalised for asthma has been assessed.

Methods: Patients hospitalised for asthma exacerbation were included in an observational
study from the 1 January to 31 December 2015. Serum IgG concentrations on admission were
compared between patients with a positive upper airway viral sample to those with a negative
viral sample.

41 Results: Amongst the 82 patients included, those with positive viral nasopharyngeal samples 42 (n=40) presented with lower serum IgG concentrations during exacerbation than those with a negative viral sample (n=42) (10.1 \pm 2.3 g/l vs. 11.5 \pm 3.6 g/l, p <0.05). The median 43 44 concentration of serum IgG was lower in patients hospitalised for more than three days 45 compared to those hospitalised for less than three days (10.0 g/l [8.2-12.4] vs. 11.4 g/l [10.1-12.8], p < 0.05) and in patients who received oral corticosteroid therapy for more than five 46 47 days compared to those treated with oral steroids for less than five days (10.1 [8.3-12.2] vs. 48 11.6 g/l [10.0-13.8], p <0.05).

49 Conclusion: Serum IgG was significantly lower when asthma exacerbations were associated
50 with positive viral samples. The patients with lower serum IgG concentrations required longer
51 hospitalisations and longer courses of steroids.

52

53 Highlights

54 What is already known about this topic? Many patients suffering from asthma experience 55 virus-induced asthma exacerbations but the pathophysiology is not fully understood and no 56 biomarkers have been found to predict severity.

57 What does this article add to our knowledge? Patients hospitalised for asthma exacerbations 58 associated with a positive virus sample have lower serum IgG than their negative 59 counterparts. Longer hospital stays and a longer duration of oral steroids were linked to lower

- 60 serum IgG concentrations.
- 61 How does this study impact current management guidelines? Serum IgG quantification during
- 62 asthma exacerbations may be lower in patients with positive virus samples but more data are
- 63 needed.
- 64
- 65 Key words: asthma, immunoglobulin G, virus, exacerbation
- 66
- 67 Abbreviations:
- 68 ACT: Asthma control test
- 69 BMI: Body mass index
- 70 CVID: Common variable immune deficiency
- 71 COPD: Chronic obstructive pulmonary disease
- 72 FEV1: Forced expiratory volume in one second
- 73 FVC: Forced vital capacity
- 74 GINA: Global Initiative for Asthma
- 75 Ig: Immunoglobulin
- 76 IgG: Immunoglobulin G
- 77 IgE: Immunoglobulin E
- 78 PCR: Polymerase Chain Reaction
- 79 ROC curves: Receiver operating characteristics curves
- 80 RSV: Respiratory syncytial virus
- 81

82 Introduction

Asthma is a chronic respiratory disease associated with airway inflammation (1). This disease
affects approximately 334 million people worldwide and its prevalence is increasing (2).
According to the latest World Health Organization (WHO) estimates released in December
2016, there were 383,000 asthma-related deaths in 2015 (3). The majority of these deaths
occurs in low- and middle-income countries (3).

88 Exacerbations play an important role in asthma morbidity and mortality (4). An exacerbation 89 is defined as acute or sub-acute episodes of progressive worsening respiratory symptoms (5). A severe exacerbation corresponds to an emergency consultation or hospitalisation (6). 90 91 Asthma is an increasingly common cause of emergency department consultations (7). Twenty 92 percent of asthma patients have experienced exacerbations requiring treatment within the 93 emergency department or hospitalisation and these patients account for more than 80% of the 94 total direct costs of asthma (8). Mortality amongst patients hospitalised for asthma 95 exacerbations accounts for one-third of all asthma-related deaths (9).

96 Infections with pulmonary tropism are very frequently involved in triggering asthma 97 exacerbations (40 to 80% of cases) (10) and are more often associated with failure of 98 conventional treatments in asthmatic patients (11). Rhinovirus, human metapneumovirus, 99 enterovirus, coronavirus and respiratory syncytial virus (RSV) are the most common viruses 100 found in asthma exacerbations (12). Rhinovirus is particularly related to asthma exacerbations 101 and is found in 60% of children experiencing asthma exacerbations (12). In adults, rhinovirus 102 also seems to be frequently involved in asthma exacerbations (13).

103 Regarding virus-induced asthma exacerbations, the antiviral response is complex and involves
104 immune cells leading to the release of inflammatory mediators (14, 15). Immunoglobulins
105 (Ig) play a key role in the anti-viral response but knowledge remains limited about their

106 involvement in virus-induced asthma exacerbations. Interestingly, it has recently been shown 107 that patients with primary hypogammaglobulinemia are more likely to suffer from viral 108 infections, including rhinovirus, compared to healthy subjects, in spite of intravenous 109 immunoglobulin replacement (16). The prevalence of asthma appears to be higher in patients 110 with various immune deficiencies including hypogammaglobulinemia, with an estimated 111 prevalence of 15% in the studies (17). The mechanism causing the increased prevalence of 112 viral infections in patients with hypogammaglobulinemia is unknown. Secretory IgA 113 deficiency in the bronchial mucosa has been suggested (16). To our knowledge, the link 114 between viral infections and serum immunoglobulin in asthmatic patients is poorly understood. 115

116 The purpose of this project is to compare the serum immunoglobulin G (IgG) concentrations 117 between patients with an upper airway specimen that tested positive for a virus to those with a 118 negative viral specimen at the time of hospitalisation for asthma exacerbations.

119

120 **Population and Methods:**

121

122 Population

An observational study was conducted from 1 January to 31 December 2015. Patients with asthma were recruited from the Department of Respiratory Medicine of the University Hospital of Tours, located in the centre of France, as well as from the Department of Respiratory Medicine of the University Centre of Reunion, a French island located in the Indian Ocean. The socioeconomic statuses of the patients are very close because both locations are managed in the same way by the French government. Inclusion criteria were patients aged 18 or older and hospitalised for an asthma exacerbation. An exacerbation was

130 defined as the increase in respiratory symptoms for more than 24 hours or the use of oral 131 corticosteroid therapy. Asthma diagnoses were established on a history of respiratory 132 symptoms such as wheezing, shortness of breath, chest tightness and coughing associated 133 with variable expiratory airflow limitation prior to hospitalisation according to the Global 134 Initiative for Asthma (GINA) (1). Exclusion criteria were smoking more than ten packs of 135 cigarettes a year, chronic obstructive pulmonary disease and pregnancy. The study was 136 approved by the Institutional Review Board of the Société de pneumologie de langue 137 Française. Each patient gave an informed written consent to take part in the study.

138

139 <u>Clinical and biological data during asthma exacerbation</u>

140 Demographic data such as age, sex, body mass index, smoking status quantified in packs per 141 year (P/A), inhaled treatment prescribed prior to exacerbation, as well as the use and duration 142 of oral steroids during the exacerbation were collected. The total serum immunoglobulin E (IgE) concentration (immunoassay technique, ImmunoCAP IgE total, Thermo Fisher 143 144 Scientific, Phadia AB, Uppsala, Sweden), blood eosinophilia (Beckman Coulter LH 780 145 controlled analyser), and C-reactive protein (CRP) (Beckman Coulter LH 780 Controlled 146 Analyser) were also collected. The severity of exacerbations was characterised by the length 147 of the hospital stay including the time spent in the emergency room and if the patient needed 148 to be treated in the intensive care unit. Controlled asthma was defined as an asthma control 149 test (ACT) score greater than or equal to 20 points in the month preceding the consultation.

150

151 <u>Upper airway viral sampling</u>

152 Viral carriage was assessed in the nasopharyngeal fluid obtained by nasopharyngeal aspiration 153 performed as soon as possible by qualified nurses accustomed to the technique. The viral 154 identification was performed by multiplex real time Polymerase Chain Reaction (PCR) 155 (Tours: Anyplex II RV16, V1.1 detection, Eurobio, France and Thermal Cycler, C1000, BioRad, US Meeting; Reunion: FTD Respiratory pathogens 21, Fast track diagnostics, 156 157 Luxembourg and Rotor-Gene Q instrument, Qiagen, France). At least 16 viruses (RSV A and 158 B, Rhinovirus, metapneumovirus, myxovirus influenzae A, H1N1, B, parainfluenzae from 1 159 to 4, coronavirus 229E, NL63 and OC43, adenovirus, enterovirus) were analysed in each 160 centre. In Reunion, the panel of viruses also included coronavirus HKU1, bocavirus and 161 parechovirus.

162

163 Immunoglobulin G (IgG) serum concentration

Serum IgG concentrations and IgG subclasses were measured by the nephelometric technique on BN Prospect® (Siemens Healthcare Diagnostics, Marburg, Germany) at the time of admission to the hospital section.

167

168 Follow-up visit

Patients received a follow-up visit three months after exacerbation. Functional respiratory tests were performed via flow-volume curves using a plethysmograph (SensorMedics Vmax Encore, Carefusion, San Diego, CA). The values of both forced vital capacity (FVC) and forced expiratory volume in one second (FEV 1) were expressed as a percentage of predicted values according to age, sex, weight and height. Airway obstruction was defined by a postbronchodilator FEV1/FVC ratio of less than 70%. Measurements were taken according to ATS/ERS 2006 recommendations using the predicted values published by the ERS (18, 19). Prick tests to aeroallergens (house dust mites (Dermatophagoides pteronyssinus and Dermatophagoides farinae at both sites and Blomia tropicalis in Reunion)), mould (Alternaria alternata), animal dander (cat and dog), cockroaches (Blattella germanica), tree pollens (olive tree, birch) and herbaceous plants (ragweed, ambrosia, wormwood and grasses) were also carried out.

181

182 <u>Statistical analyses</u>

183 Quantitative data, with a normal distribution, were expressed as mean and standard deviation 184 (mean \pm SD) and quantitative data, with no normal distribution, were expressed as median 185 deviation [interquartile range]. Qualitative data were expressed as frequencies (%). A 186 Student's t-test (parametric) was used to compare continuous variables with normal 187 distribution. The Mann-Whitney test was used for quantitative data and had no normal 188 distribution. Statistical significance was defined as a p value of <0.05. Receiver operator 189 characteristics (ROC) curves were used to assess serum IgG concentrations as a potential 190 marker of a positive virus specimen. The best IgG cut-off was determined using the Youden's index. Sensitivity and specificity of IgG cut-off for a positive virus sample were also defined. 191 192 A Spearman test was used to assess the correlation between serum IgG concentrations and 193 other continuous variables. The linearity of these relations was then tested using linear 194 regression. The statistical analysis was carried out using the Graph Pad Prism 5 software (license reference: GPW6-222441-RILS-2C3D1). 195

196

197 **Results**

198 Characteristics of patients and virus distribution:

Eighty-two patients were included in this study. The patients' characteristics are summarised in Table 1. The median age was 49.5 [28.8-63.3] years old. Patients were predominantly female (76.8%) and overweight with a median body mass index (BMI) of 26.3 [22.1-31.2] kg/m². Inhaled corticosteroid treatment was used in 61% of patients before asthma exacerbation and 14.6% out of all the patients required intensive care at the time of exacerbation. The median duration of oral corticosteroid therapy and the median length of stay for the exacerbation were 7.0 days and 4.0 days, respectively.

Nasopharyngeal aspirations were positive for viruses in 48.8% of patients. The most common
viruses were rhinovirus (38%), myxovirus influenza (18%), metapneumovirus (12%) and
enterovirus (10%) (Figure 1).

209

210 Characteristics of patients according to the viral status of nasopharyngeal aspiration:

Patients with a positive or negative viral sample showed no statistically significant difference in terms of demographic and biological data (Table 2). Patients with positive nasopharyngeal aspiration for viruses were slightly younger than patients with negative nasopharyngeal aspiration for viruses (43.5 vs. 50.5 years, respectively) with no statistical significance (p >0.05). Patients with positive viral specimens also seemed to be hospitalised in intensive care units more often compared to patients harbouring negative viral specimens (20% vs. 9.5%, respectively), however the difference was not significant.

218

219 <u>Immunoglobulin blood concentration:</u>

Patients with a positive viral sample had significantly lower serum IgG concentrations than patients hosting a negative viral specimen $(10.1 \pm 2.3 \text{ g/l vs. } 11.5 \pm 3.6 \text{ g/l, p} < 0.05)$ (Table 3). No difference was found for serum IgA and IgM concentrations in both groups. IgG subclasses were similar between groups. An inverse correlation was observed between age and serum IgG concentrations (p=0.016). No other correlation was shown for the other demographic characteristics.

226

227 Serum IgG concentrations and diagnosis of a positive virus sample.

ROC curve analysing the serum IgG concentrations as a marker for predicting a specimen positive for a virus is presented in Figure 2. According to Youden's index, the best cut-off value of serum IgG to predict a positive virus sample is 12.30 g/l, the optimal sensitivity and specificity were 80.0% and 40.5%, respectively.

232

233 IgG concentration and characteristics of asthma exacerbation:

234 The median serum concentration of IgG was significantly lower in patients hospitalised for 235 more than three days than in those whose hospital stay was less than or equal to three days 236 (10.0 g/l [8.2-12.4] g/l vs. 11.4 g/l [10.1-12.8] g/l, p <0.05) (Figure 3B). Patients receiving 237 oral corticosteroids for more than five days had lower median serum IgG concentrations than 238 patients who were given corticosteroid therapy for five days or less (10.1 [8.3-12.2] g/l vs. 239 11.6 [10-13.8] g/l respectively, p <0.05) (Figure 3C). The median serum concentration of IgG 240 was not significantly different if patients were hospitalised in intensive care units or not 241 (Figure 3A). An inverse correlation was observed between the length of stay and the serum 242 IgG concentration (p=0.022 with the Spearman test) (Figure 4). No correlation was found 243 between oral steroids and serum IgG concentrations. ROC curves have been created according 244 to the length of stay and the steroid duration in order to know the reliability of serum IgG

245 concentrations as a predictive biomarker of exacerbation severity. For the length of stay 246 prediction (≤ 3 days or >3 days), the best sensitivity and specificity of IgG concentrations are 247 80.00% and 50.00%, respectively for a cut-off value of 11.05 g/l (Figure E1a). Regarding the 248 steroid duration prediction (≤ 5 days or > 5 days), for a cut-off value of 9.99 g/l, the best 249 sensitivity and specificity of IgG concentrations are 62.07% and 66.67%, respectively (Figure 250 E1b). If a positive virus sample is combined with the IgG serum concentrations, for a cut-off 251 of 10.75 g/l, the best sensitivity and specificity of the length of stay prediction are 79.17% and 252 61.11%. Regarding a cut-off of 11.05 g/l, the best sensitivity and specificity of the steroid 253 duration prediction are 82.76% and 55.00% (Figure 5).

254

255 **Discussion:**

256

Asthma patients with a positive nasopharyngeal viral sample had a lower serum IgG concentration, at the time of exacerbation, than those with a negative viral sample. Moreover, the median serum IgG concentration was lower in patients hospitalised for more than three days as well as in those who were given oral steroids for more than five days compared to their counterparts.

262 In our study, the epidemiology of viruses found in patients hospitalised for asthma 263 exacerbations was similar to data observed in the literature. Indeed, herein, rhinovirus was the 264 most frequently observed pathogen (38%). This virus is known to be more often associated with asthma exacerbations in children (20-22), and some works suggest that it is also the most 265 266 incriminated virus in severe asthma exacerbations in adults (13, 23, 24). Our patients also 267 presented with other viruses such as myxovirus influenzae. Myxovirus influenza, particularly influenza A H1N1, is responsible for a significant morbidity and mortality in asthmatic 268 269 patients (25-27). Enterovirus appears to be less frequently found in the literature (28). In our

cohort, enterovirus was always found in co-infection with rhinovirus and this represented five
out of ten patients who had co-infections. The five remaining patients were co-infected with
rhinovirus/myxovirus influenza A (n=3), metapneumovirus A and B/myxovirus influenza A
H1N1 (n=1), adenovirus/myxovirus influenza A (n=1) and Coronavirus 229E/myxovirus
influenza A (n=1). No relationship between co-infection and serum IgG concentrations or the
severity of exacerbations was found.

276 No demographic differences were found between the patients with positive virus samples and 277 those with negative virus samples. However, although the difference was not statistically 278 significant, patients with a positive virus sample were slightly younger compared to their 279 negative counterparts (43.5 vs. 50.5 years, respectively). The lack of significance is probably 280 due to the small sample size. It is well known that asthma exacerbation in young people is 281 associated with virus infections (29). BMI appears to be a bit higher in patients with positive 282 virus samples compared to patients with negative virus samples (29 vs. 25 kg/m², 283 respectively) but the difference is also not significant. The risk of viral infections in asthma 284 and obese patients has been identified elsewhere in the literature. Indeed asthma and obesity 285 have been found as independent risk factors of pandemic influenza hospitalisation (30). In our 286 study, more patients in the positive virus group required intensive care although there was no 287 significant difference. Data from the literature suggest that respiratory viruses may trigger 288 severe asthma exacerbations, particularly in young people (31).

289

The link between viral infections and asthma exacerbations is widely recognised and the impairment of viral immunity may partly explain the development of viral infections in asthma patients (32, 33). In influenza infections, IgG has been identified as being particularly involved in protecting the respiratory tract against viruses and its role seems to be superior to IgA (34). Rhinovirus-specific IgG appears to prevent and control re-infection and a high 295 concentration of specific IgG antibodies is correlated with the attenuation of respiratory 296 symptoms and a reduction in viral shedding (35). A decrease in specific viral antibodies in 297 patients with asthma has been suggested (36). However, in another study, higher antibody 298 responses to rhinovirus have been shown in asthmatic patients compared to non-asthmatic 299 controls (37). Given the study conducted by Johnston et al. on rhinovirus involvement in 300 asthma exacerbations in children, we may also speculate that the admitted patients had less 301 exposure to school-aged children in the past and therefore had not generated a sufficient 302 antibody response to the current virus(es) (38). Other mechanisms beyond adaptive immunity 303 dysfunction are involved in virus-induced asthma exacerbations. Indeed, an altered innate 304 immune anti-viral response has been observed in asthmatic patients compared to non-305 asthmatic ones (39-41). Our study does not add a mechanistic explanation for the low level of 306 serum IgG in patients with a positive virus sample.

307 Our results may suggest that serum IgG concentrations at the time of asthma exacerbation 308 would be a marker of severity. Indeed, patients with a hospitalisation length of more than 309 three days or a steroid course of more than five days show lower IgG concentrations at the 310 time of exacerbation. Amongst patients with stable COPD, reduced total IgG levels have 311 been linked to an increased risk of exacerbations and hospitalisations (42). In that study, 312 patients with serum IgG <7 g/l had a higher risk of COPD exacerbations compared to patients 313 with the highest quartile of serum IgG.. We assessed the predictive ability of serum IgG 314 concentrations as a biomarker of exacerbation severity. However, it does not seem to be a 315 useful predictor based on modest sensitivity and poor specificity. If we combine serum IgG 316 concentrations and a positive virus sample, the prediction of exacerbation severity was not 317 improved, and further studies are needed to specifically assess IgG level as a marker of 318 severity in asthma exacerbations. The prediction of next hospitalisations with IgG levels 319 would also be interesting but we cannot answer this question with our study as it was not 320 designed to answer this question. However, we may speculate that a low IgG level 321 particularly at baseline would predispose patients to virus infection and consequently increase 322 their risk of hospitalisation.

323 Our study does have several limitations. The difference in IgG concentrations between 324 patients with positive and negative viral samples is statistically significant, but it remains 325 uncertain whether this difference is clinically relevant or not. More robust studies on the role 326 of IgG are needed. In addition, the change of IgG concentrations or other immunoglobulins 327 over time was not assessed in our study. We did not measure IgG in patients in a stable 328 condition. It would have been interesting to know if the IgG decline is transient or longer-329 lasting. Indeed, in patients with chronic obstructive pulmonary disease (COPD), low IgG 330 serum concentrations at baseline seem to be associated with more severe exacerbations per 331 year (42). However, in this study, no virus analysis was performed. Finally, our study is based 332 on multiplex PCR for virus identification. Nevertheless, the list of viruses, though very 333 extensive, is not exhaustive, therefore emerging viruses cannot be ruled out.

334

335 Conclusion:

336 The results of our study show that serum IgG was significantly lower when severe asthma 337 exacerbations were associated with a positive viral sample. Serum IgG levels were lower in 338 patients with a longer length of stay and when a course of oral steroids was prescribed for a longer duration. Further studies are requested to know the exact usefulness of IgG 339 340 concentrations as a severity marker of severe asthma exacerbations. The understanding of 341 immunological mechanisms of viral infection in severe asthma exacerbations may be an interesting lead for the development of new therapies in patients for whom severe 342 343 exacerbations are a source of significant morbidity and mortality.

344

Figure 1: Details of viruses found in nasopharyngeal aspirations of hospitalised patients forasthma exacerbation.

Figure 2: Receiver operating characteristics (ROC) curve analysis of serum IgG
concentrations for the prediction of a positive virus sample. At the cut-off value of 12.30 g/l,
sensitivity and specificity are 80.0% and 40.5%, respectively.

Figure 3: Comparison of blood IgG concentrations a) between hospitalised patients in intensive care and those without ICU, b) between patients hospitalised for three days or less and those hospitalised for more than three days and c) between patients treated during five days or less with oral corticosteroids and those treated during more than five days, *p <0.05. ICU: Intensive care unit.

Figure 4: Scatter plots with regression line showing the relationship between the serum IgG concentrations and the hospital length of stay. *p=0.0058 using linear regression.

Figure 5: Receiver operating characteristics (ROC) curve analysis of serum IgG concentrations combined with a positive virus sample for a) the prediction of a length of stay duration of more than three days (for a cut-off value of 10.75 g/l, sensitivity and specificity are respectively 80.0% and 40.5%) and b) the prediction of a steroid duration of more than five days (for a cut-off value of 11.05, the best sensitivity and specificity are 82.76% and 55.00%).

Table 1: Characteristics of patients hospitalised due to asthma exacerbations.

| | n=82 |
|--|------------------|
| Age (years) | 49.5 [28.8-63.3] |
| Female | 63 (76.8) |
| Active or former smokers | 21 (25.6) |
| BMI (kg/m^2) | 26.3 [22.1-31.2] |
| Allergen sensitisation | 35 (42.7) |
| Use of an inhaled corticosteroid before hospitalisation | 50 (61.0) |
| Inhaled corticosteroid dose (beclomethasone equivalent doses) (μ g/d) | 1600 [800-2000] |
| Exacerbation ≥ 2 12 months prior to exacerbation | 15 (18.3) |
| Oral corticotherapy duration for exacerbation | 7.0 [5.0-8.5] |
| Length of hospitalisation | 4.0 [2.0-5.0] |
| Intensive care unit stay at the time of exacerbation | 12 (14.6) |
| Positive viral sampling | 40 (48.8) |
| FEV1 after hospitalisation (ml) (% predicted values) | 76 [63-88] |
| FEV1/FVC after hospitalisation (%) | 74 ± 15 |
| Blood Eosinophil (G / 1) | 0.20 [0.03-0.68] |
| Total IgE (kIU / l) | 339 [148-786] |

Quantitative data, with normal distribution, are expressed as means and standard deviation (mean \pm SD) and quantitative data, without normal distribution, are expressed in median [interquartile range]. The qualitative data are expressed as frequencies (%).

Table 2: Characteristics of patients hospitalised for asthma exacerbations with positive or
negative upper airway viral samples. FEV1: Forced expiratory volume in 1 second, FVC:
Forced vital capacity, IgE: Immunoglobulin E

| | Virus + n=40 | Virus – n=42 | р |
|--|---------------------------------|--------------------------------|--------------|
| Age (years) | 43.5 [24.3-60.3] | 50.5 [36.0-68.3] | 0.14 |
| Female | 33 (82.5) | 30 (71.4) | 0.30 |
| Active or former smokers | 11 (27.5) | 10 (23.8) | 0.80 |
| BMI (kg/m^2) | 29.0 [22.0-33.2] | 25.0 [22.3-30.4] | 0.42 |
| Allergen sensitisation | 18 (45.0) | 17 (40.5) | 0.82 |
| Use of an inhaled corticosteroid prior to hospitalisation | 24 (60.0) | 26 (61.9) | 1.00 |
| Inhaled corticosteroid dose (beclomethasone equivalent doses) (µg/d) | 1600 [800-2000] | 1600 [800-1600] | 0.91 |
| Exacerbation ≥2 12 months prior to exacerbation | 5 (12.5) | 10 (23.8) | 0.39 |
| Oral corticotherapy duration while exacerbation (days) | 7.0 [5.0-9.5] | 6.0 [5.0-8.5] | 0.15 |
| Length of hospitalisation | 4.0 [2.0-5.0] | 3.5 [2.0-5.0] | 0.31 |
| Intensive care unit stay at the time of exacerbation | 8 (20.0) | 4 (9.5) | 0.22 |
| FEV1 after hospitalisation (% predicted values) | 80.5 [61.0-90.5] | 75.5 [63.5-86.00] | 0.50 |
| FEV1/FVC after hospitalisation (%) | 75 ± 12 | 66 ± 12 | 0.052 |
| Blood Eosinophil (G/l) Total IgE (kIU/l) | 0.2 [0.0-0.4] 349 [185-1003] | 0.2 [0.0-0.8] 310 [127-516] | 0.39 0.29 |

372 Values are expressed as median and [interquartile range] or mean ± standard deviation for

373 quantitative data and raw values (%) for categorical data. A Student's t-test (parametric) is

374 used to compare continuous variables to a normal distribution. The Mann-Whitney test is used 375 for quantitative data that do not have a normal distribution

375 for quantitative data that do not have a normal distribution.

- Table 3: Comparison of serum Immunoglobulin G (IgG), Immunoglobulin A (IgA),
 Immunoglobulin M (IgM) and IgG subclass levels between patients with positive viral
 samples (Virus +) and negative (Virus -).
- 379

| | Virus + n=40 | Virus – n=42 | р |
|-------------------------|--------------------|--------------------|-------|
| $I_{\alpha}G(\alpha/l)$ | 10.1 ± 2.3 | 11.5 ± 3.6 | <0.05 |
| | 10.10 [8.15-11.68] | 11.25 [9.09-13.43] | <0.05 |
| IgA (g/l) | 2.30 [1.77-3.06] | 2.34 [1.98-2.86] | 0.70 |
| IgM (g/l) | 0.88 [0.55-1.15] | 0.75 [0.56-1.36] | 0.91 |
| IgG1 (g/l) | 4.9 [3.5-5.8] | 5.5 [4.5-7.0] | 0.059 |
| IgG2 (g/l) | 3.7 ± 1.0 | 4.1 ± 1.4 | 0.12 |
| IgG3 (g/l) | 0.58 [0.30-0.80] | 0.57 [0.43-0.86] | 0.38 |
| IgG4 (g/l) | 0.27 [0.15-0.43] | 0.27 [0.15-0.60] | 0.85 |

Values are expressed as median and [interquartile range] or mean \pm standard deviation for quantitative data and raw values (%) for categorical data. Both median [interquartile range] and mean \pm standard deviation are given for IgG. A Student test (parametric) is used to compare continuous variables to a normal distribution. The Mann-Whitney test is used for quantitative data that do not have a normal distribution.

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