

Immersed membranes configuration for the microfiltration of fruit-based suspensions

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1	Immersed membranes configuration for the microfiltration of fruit-based
2	suspensions
3	
4	Keywords: Fruit-based suspensions; Microfiltration; Immersed membranes; Productivity; Selectivity.
5	
6	Highlights:
7	• Interest of using immersed membranes microfiltration for fruit-based suspensions
8	• Productivity in line with what is reported in other domains
9	Selectivity goals towards compounds of interest reached
10	• Interesting alternative to conventional cross-flow filtration for small production units
11	
12	
13	Abstract
14	Microfiltration is widely used to ensure the athermal stabilization, clarification and concentration of various
15	fruit-based suspensions (e.g. fruit juices, food by-products, wine). However, the performances of membrane
16	filtration remain highly challenged by membrane fouling. To prevent membrane fouling, cross-flow
17	filtration is generally performed. Nevertheless, this intensive working mode is considered as highly energy
18	consuming due to the intensive pumping required to circulate the suspension at high velocities. In the light
19	of this, immersed membranes configurations have been developed in many fields, as they allow working in
20	energy-friendly operating conditions. Thus, this work investigated for the first time the performances of an
21	immersed membranes configuration for fruit-based suspensions microfiltration, in terms of productivity
22	(membrane fouling, permeate flux) and selectivity (clarification, concentration of bioactive compounds).
23	This study focused on three fruit-based suspensions: a grapefruit juice and two winery by-products.
24	Concerning the process selectivity, pilot-scale experiments showed that immersed membranes filtration
25	allowed producing retentate and permeate of quality as least as good as the one related to conventional
26	cross-flow filtration. Concerning the process productivity, cross-flow filtration allowed reaching higher
27	fluxes compared to immersed membranes filtration, in accordance with the conventional order of magnitude
28	specific to each configuration. Immersed membranes configuration could find interesting applications
29	within small production units of fruit juices and/or industries dealing with the valorization of low added-
30	value byproducts thanks to its various advantages (high compactness, easy handling and mobility, low
31	investment and operational costs).

33 List of abbreviations and variables:

32

A	Absorbance (/)
С	Total monomeric anthocyanins content, malvidin-3-glucoside equivalent (mg.L ⁻¹)
C_i	Concentration of targeted compounds in the initial feed suspension
C_{p}	Concentration of targeted compounds in the permeate
ĎF	Dilution factor (/)
DM	Dry Matter $(g.L^{-1})$
I	Permeate flux (m.s ⁻¹)
Ĩ	Path length (1.0 cm)
MW	Molecular weight of malvidin-3-glucoside (463.3 g.mol ⁻¹)
NTU _i	Turbidity of the initial feed suspension (NTU)
NTU_p	Turbidity of the permeate (NTU)
PES	Polyethersulfone
C _{clarif}	Clarification ratio %)
Cret	Retention ratio (%)
R_h	Total resistance to permeation (m ⁻¹)
Red Extract	Red grape pomace extract
Seed	Grape seeds extract
Extract	
SIS	Insoluble suspended solids
ТМР	Transmembrane pressure (Pa)
VRR	Volume reduction ratio
ε	Molar extinction coefficient of malvidin-3-glucoside (28 000 L.mol ⁻¹ .cm ⁻¹)
μ_p	Permeate dynamic viscosity (Pa.s)
μ_s	Suspension dynamic viscosity (Pa.s)

34

35 **1.** Introduction

36 Microfiltration is widely used to ensure the clarification and the concentration of various fruit-based 37 suspensions such as fruit juices [1–4], agro-food by-products [5–7] or wine [8]. This solid-liquid separation technic allows producing high quality products thanks to its high selectivity and low operating temperatures. 38 However, the performances of membrane filtration remain highly challenged by membrane fouling. Fruit-39 40 based suspensions, well known to be heterogeneous suspensions containing colloids and larger suspended 41 insoluble solids (SIS) dispersed in a continuous aqueous phase, are considered as highly-fouling 42 suspensions. During the microfiltration of such complex suspensions, particles deposition is considered as 43 one of the main causes of membrane fouling [9,10]. This type of fouling is mainly governed by the balance 44 between convective forces (permeate flow), leading particles to the membrane, and back-transport forces, 45 removing particles away from the membrane surface [11].

So far, a wide range of filtration configurations have been studied for fruit juices microfiltration, such as 46 47 cross-flow filtration using organic plane [3,12] or hollow-fiber [13,14] membranes, organic or inorganic tubular membranes [2,4,15–20]. In spite of the diversity of membrane shape and material and operating 48 49 conditions, the use of high shear stress at the membrane surface is always a common feature to enhance the back-transport mechanisms and thus increase the permeate fluxes [21]. However, this intensive working 50 51 mode is well known to be highly energy consuming due to the intensive pumping required to circulate the 52 suspension at high velocities. Moreover, the use of important shear forces, leading to high turbulences, has 53 been reported to induce particles size modifications [22,23] that could impact the suspension characteristics (fouling propensity, nutritional and sensorial properties). 54

55 In the light of this, immersed membranes configuration (out-to-in filtration) could be an interesting 56 alternative for the microfiltration of fruit-based suspensions. In this configuration, the membrane (plane or hollow fiber) is immersed in the suspension and the filtration is generally ensured by permeate suction at 57 58 constant flux. Thus, the filtration operation is conducted in conditions close to that of dead-end filtration, 59 associated with limited back-transport forces and low permeation fluxes. Despite the low permeation fluxes 60 commonly applied in such conditions, immersed membranes filtration have been widely and successfully 61 used in many fields, notably for water treatment (e.g. drinking water production) and for wastewater treatment for the filtration of more heterogeneous suspensions (e.g. microalgae suspensions [24-26], 62 activated sludge [27–30]). For these applications, the relatively low productivity is generally offset by the 63 great packing density of the membrane, by the low cost of organic membranes and the low energy 64 65 consumption of the process [31,32].

However, as far as the authors are aware, the performances of this filtration configuration remain little 66 67 studied for fruit-based suspensions microfiltration. At present time, no studies have yet characterized the 68 productivity and the selectivity of this filtration configuration for such applications. In the light of this, the 69 aim of this work was to investigate the performances of an immersed membranes configuration for fruit-70 based suspensions microfiltration, in terms of membrane fouling and selectivity. This study focused on three 71 different agro-food suspensions: a grape fruit juice, a grape pomace extract and a grape seeds extract. Firstly, 72 a specific experimental strategy was conducted in order to define the optimal operating conditions of this 73 system. Secondly, based on the previously identified operating conditions, filtration performances were 74 analyzed in terms of membrane fouling and selectivity (clarification and/or concentration of targeted compounds). Finally, a comparison of immersed membranes configuration performances with conventional 75 76 side-stream membranes configuration ones (cross-flow filtration) was proposed and discussed.

78 2. Material and methods

79 2.1. Fruit-based suspensions

80 2.1.1. Selection

81 Three agro-food suspensions were studied in this work: a grapefruit juice, a red grape pomace extract and a82 grape seeds extract.

Grapefruit juice was chosen as it is among the most popular citrus fruits worldwide [33,34]. Moreover, its
microfiltration offers several applications in fruit-juices industries as it allows producing a high quality
clarified and stabilized juice rich in phenolic compounds (mainly naringin and narirutin [35]) and a
concentrated pulpy fraction rich in carotenoids (mainly lycopene and beta-carotene [36]). These products
find useful applications in industries (pharmaceutical, cosmetic, food) thanks to their therapeutic, nutritional
and sensorial properties.
Red grape pomace and grape seeds extracts were chosen for being among the main by-products (produced

by considerable tonnage) by winery industries [37]. Their valorization is a major economic and ecological challenge, for which clarification is a key pre-treatment step. The microfiltration of red grape pomace and grape seed extracts allows producing a clarified permeate rich in phenolic compounds, notably flavonoids like tannins and anthocyanins in the case of red grape pomace extract [38,39]. This permeate can be easily valorized through additional extraction steps.

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96 **2.1.2.** Procurement, characterization and conservation

In this study, grapefruit juice was produced by squeezing Star Ruby grapefruit (*Citrus grandis (L.) Osbeck*) 97 (Spain) in a semi-industrial extractor (Automatic orange juicer, model 32, SANTOS, Vaulx-en-Velin, 98 99 France) and pre-filtered through a stainless steel sieve (1 mm mesh size). Red grape pomace extract (named 100 Red Extract) and grape seeds extract (Seed Extract) were purchased from a local distillery located in the South of France. The extracts were obtained by industrial solid-liquid extraction in sulphited water, after 101 102 grinding of the raw residues generated during winemaking (i.e. red grape pomaces and grape seeds). The 103 three suspensions were stored at - 20 °C and thawed before use. Their main physicochemical and chemical 104 characteristics (dry matter (DM), turbidity (NTU), pH, Brix degree (°Brix), dynamic viscosity (μ_s) and 105 suspended insoluble solids (SIS)) were determined according to the protocols and methods described by 106 [37,40].

108 2.2. Immersed membranes filtration experiments

109 2.2.1. Experimental set-up

A schematic illustration of the experimental equipment is presented in figure 1. It consisted of a 6.3 L stirred 110 stainless steel feed tank, linked to a 1.8 L external filtration unit containing immersed organic hollow-fiber 111 membranes (main characteristics given in table 1). A low flow-rate pump (item 3 on figure 1, 520S IP31 112 peristaltic pump, Watson-Marlow, Massachusetts, USA) allowed the juice flowing through the filtration 113 unit (flow velocity of 3.5×10⁻² m.s⁻¹, corresponding to Reynolds number of around 500 in the external 114 filtration unit with hollow-fiber membranes and to a 3 s⁻¹ shear rate at the membrane surface). A cryostat 115 116 connected to a water jacket on the recycling loop, maintained the system at a constant temperature of $25 \pm$ 117 2 °C.





Table 1: Main characteristics of the immersed membranes.

Configuration	Hollow-fiber
Material	PES
Average pore size (µm)	0.1
Intrinsic membrane resistance R_m^* (m ⁻¹)	2.9×10 ¹¹
Water permeability * (L.h ⁻¹ .m ⁻² .bar ⁻¹)	1240
Manufacturer	Polymem (France)
Filtration area (m ²)	1.8×10 ⁻¹

128

* Experimental measurement at 25 °C

129

The out-to-in permeation flow was ensured by pump suction (item 7 on figure 1, 621F/RE IP55 peristaltic pump, Watson-Marlow, Massachusetts, USA) at constant flux (*J*). The evolution of membrane fouling during the filtration runs was estimated though the monitoring of the transmembrane pressure evolution (TMP) by an Almemo 2690-8 computer-controlled device (Ahlborn GmbH, Germany) connected to a pressure sensor. Indeed, according to Darcy's law (equation 1), in constant flux experiments, an increase of TMP is directly related to an increase of the resistance to permeation:

$$J = \frac{TMP}{\mu_p R_h} \tag{1}$$

137 With *J* the permeate flux $(m.s^{-1})$

138 *TMP* the transmembrane pressure (Pa)

139 μ_p the permeate dynamic viscosity (Pa.s)

140 R_h the total resistance to permeation (m⁻¹).

141

142 **2.2.2.** Definition of the optimal operating filtration conditions: flux-stepping experiments

Pre-filtration tests were performed to define the optimal operating flux of the immersed membranes filtration 143 system for each suspension. As stated before, membrane fouling is mainly governed by the equilibrium 144 between convective and back-transport forces. Even if important permeate fluxes are generally needed to 145 146 ensure the process sustainability, excessive fluxes can be counterproductive as they lead to important 147 convective transports of foulant particles toward the membrane surface. Choosing an optimal permeate flux 148 is therefore of crucial interest to control membrane fouling. Among all flux concepts that have been studied 149 to guide permeate flux selection, critical and threshold fluxes concepts are particularly adopted when dealing 150 with immersed membranes filtration [21].

Critical flux is defined as being the flux below which TMP remains strictly constant. According to many 151 152 studies, critical flux is generally really low and its determination is time consuming [21,41,42], which limits its industrial application. In the light of this, the authors focused on the concept of threshold flux, defined 153 154 as being the flux at which the rate of fouling increases significantly. This flux concept is more applicable for industrial applications, as less time is required for its determination and it generally matches with 155 156 acceptable value of fluxes.

- Threshold flux identification was based on progressive increasing flux-steps under total recycle mode 157 158 (retentate and permeate were systematically returned to the feed tank) and on simultaneous TMP monitoring. In this work, the filtration was initially operated at a constant permeate flux of 2 L.h⁻¹.m⁻² for 10 minutes. 159 After 10 minutes, the flux was increased and the filtration was operated for another 10 minutes. Like so, 160 161 flux was gradually increased at 10-minutes intervals. The values of flux and flux-steps duration were chosen
- 162 according to values reported in the literature [43-45].
- 163 Based on the experimental results, the fouling rate increase was evaluated through the determination of
- 164 dTMP/dt values for each constant flux-step, representing the TMP increase during the last 5 minutes of each
- flux-step. A threshold value of $dTMP/dt = 1.0 \times 10^{-5}$ bar.s⁻¹ was chosen in accordance with values used in the 165
- literature [43–46]. Thus, when dTMP/dt remained lower than 1.0×10^{-5} bar.s⁻¹, fouling was considered as 166
- being low. On the opposite, fouling increase was considered significant when dTMP/dt values were higher 167 than 1.0×10^{-5} bar.s⁻¹. Therefore, the threshold flux value was defined as being the highest flux at which 168 dTMP/dt remained lower than 1.0×10^{-5} bar.s⁻¹.
- 169
- 170

171 2.2.3. Immersed membrane filtration performances

Once the threshold flux was identified for each suspension, filtration experiments under batch concentration 172 173 mode (until Volume Reduction Ratio of 2) were performed at constant flux below or equal to the identified 174 threshold flux (sub-critical conditions).

175 The performances of immersed membranes configuration in terms of *membrane fouling* were estimated through the monitoring of the TMP during the filtration running. 176

The performances of immersed membranes configuration in terms of *selectivity* were evaluated by 177 178 performing sampling on the initial feed suspensions and on the permeates resulting from their filtration 179 under batch concentration mode. Depending on the suspension, various separation objectives were expected 180 from microfiltration experiments (table 2).

 Table 2: Separation objectives related to the microfiltration of grapefruit juice, Red and Seed Extracts

105 _					
	Suspension	Grapefruit juice	Red Extract	Seed Extract	
_	Separation objectives	Clarified permeate rich in total phenolic compounds Retentate concentrated in	Clarified permeate rich in total phenolic compounds and anthocyanins	Clarified permeate rich in total phenolic compounds	
		carotenoids (lycopene and β-carotene)			
184		· · · · · · · · · · · · · · · · · · ·			
185	The ability of imme	ersed membranes filtration to r	produce a clarified permeate	was measured through the	
186	analysis of turbidity	loss, evaluated though the clari	fication ratio C_{clarif} (%), calc	ulated as follows (equation	
187	2):				
188		$C_{clarif} = 1$	$-\frac{NTU_p}{NTU_i} \times 100$	(2)	
189					
190	The selectivity of the filtration towards valuable compounds was characterized through the retention ratio				
191	C_{ret} (%) representing the percentage of targeted compound retained by the membrane (equation 3):				
192	$C_{ret} = 1 - \frac{c_p}{c_i} \times 100 \tag{3}$				
193	With NTU_p and C	C_p the turbidity and the concent	ration of targeted compounds	in the permeate	
194	NTU_i and C_i	$_i$ the turbidity and the concentra	tion of targeted compounds in	the initial feed suspension.	
195					
196	In the light of the	above, various physicochemica	al and biochemical analyses	were carried out for each	
197	suspension and related permeate.				
198					
199	Turbidity measuren	nents were performed on wate	er diluted extracts to fall in	the turbidimeter precision	
200 201	domain ranging betw	ween 0 and 50 NTU (Hanna LP	2000, Hanna instruments, Sz	zeged, Hungary).	
202	Total phenolic con	tent was measured by spectro	ophotometry, according to a	modified Folin Ciocalteu	
203	method. Firstly, the	suspensions were prepared in a	n ethanol/distilled water (25:	75, v/v) solution. After the	
204	addition of 184 μ L	of distilled water, 24 µL of san	nple, 12 µL of Folin Ciocalte	eu reagent and 30 μ L of 20	

% (w/v) Na₂CO₃ solution in a 96-well microplate (MultiSkan Spectrum, Thermo Scientific), the resulting
mixture was incubated at 25 °C in the darkness for 1h. The absorbance was then measured at 765 nm. Gallic
acid was used as a standard for calibration. Results were expressed as milligrams gallic acid equivalent
(GAE) per liter of sample.

209

Total monomeric anthocyanins content was determined using the pH differential method [47]. The sample
 absorbance was measured at pH 1.0 and 4.5 at 510 nm (the wavelength of maximum absorbance) and at 700
 nm to correct haze. Measurements were performed using a spectrophotometer (UV 2450, Shimadzu, Kyoto,
 Japan). Total monomeric anthocyanins were expressed as follows, as malvidin-3-glucoside equivalent
 (mg.L⁻¹) [48] (equation 4):

215 $C = \frac{A \times MW \times DF}{\varepsilon \times l} \times 10^3$ (4)

216 With *MW* the molecular weight (463.3 g.mol⁻¹) and ε the molar extinction coefficient

217 (28 000 L.mol⁻¹.cm⁻¹) of malvidin-3-glucoside, respectively

218 *DF* the dilution factor

219 l the path length (1.0 cm)

220 *A* the sample absorbance, calculated as follows (equation 5):

221
$$A = (A_{510} - A_{700})_{pH1.0} - (A_{510} - A_{700})_{pH4.5}$$
(5)

222

223 *Carotenoids (lycopene and \beta-carotene) content* was evaluated thanks to an extraction step followed by 224 HPLC analysis. Carotenoids were firstly extracted from the samples though two successive extraction steps 225 using ethanol/hexane (4/3 (v/v) containing 0.1% of BHT as antioxidant) as extraction solvent, under stirring 226 [49,50]. At each step, residue was separated from the liquid phase by filtration using an $n^{\circ}2$ porosity filter 227 funnel. Ethanol and hexane were successively used to wash the residue. Organic phases were transferred to 228 a separating funnel and successively washed with 10% sodium chloride and distilled water. The aqueous 229 layer was removed and the hexanic extract was collected and dried with a rotary evaporator at 30 °C. The 230 dried carotenoids extracts were then dissolved in 1 mL of dichloromethane/methyl tert-butyl ether/methanol solution (50/40/10 (v/v/v)). Secondly, HPLC analysis of carotenoids were conducted as described by 231 232 Polidori et al. (2018) using an Agilent 1100 liquid chromatograph (Massy, France) equipped with a 233 photodiode array detector and a C_{30} separation column (250 × 4.6 mm i.d., 5 µm YMC, EUROP Gmbh, 234 Germany) [50].

236 **3. Results**

237 **3.1.** Feed suspensions characterization

The main physicochemical and chemical characteristics (dry matter (DM), turbidity (NTU), pH, Brix degree (°Brix), dynamic viscosity (μ_s) and suspended insoluble solids (SIS)) of the three fruit-based suspensions are given in table 3.

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243

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Table 3: Physicochemical characteristics of the studied suspensions.

	Grapefruit juice	Red Extract	Seed Extract
DM (g.L⁻¹)	105.2 ± 0.2	34.5 ± 0.2	22.6 ± 0.3
Turbidity (NTU)	3720 ± 230	1000 ± 70	1560 ± 250
рН (/)	3.1 ± 0.1	3.6 ± 0.1	4.3 ± 0.1
°Brix (g/100 g)	9.9 ± 0.1	3.6 ± 0.1	3.1 ± 0.1
μ_s (mPa.s)	1.7 ± 0.1	1.4 ± 0.1	1.5 ± 0.1
SIS (g.L⁻¹)	1.5 ± 0.1	1.6 ± 0.1	1.5 ± 0.1

245 246

All assays were performed at 25 ± 2 °C and values provided are the average of three replicates.

Among all studied products, grapefruit juice appeared to be the most different suspension with high turbidity, DM and Brix values. The two winery byproducts were quite similar, apart from a slightly lower DM value for Seed Extract and a higher turbidity value for Red Extract. These observations are consistent with the different raw materials and manufacturing processes related to each suspension. For the three suspensions, the SIS represented a very small part of the total dry matter with low SIS/DM ratios. It is a coherent observation, since a great part of their dry matter consists in sugar, estimated through the °Brix.

255

254 **3.2.** Definition of the optimal operating domain: flux-stepping experiments

Figure 2 presents the TMP evolution as function of time during flux-stepping experiments performed ongrapefruit juice (a), Red (b) and Seed Extracts (c).



Figure 2: TMP versus time during flux-stepping experiments of grapefruit juice (a), Red Extract (b) and Seed Extract (c) using 0.1
 μm PES hollow-fiber membranes.

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For the three suspensions, it can be noticed that the TMP progressively increased in greater or lesser degree for each flux-step, showing that fouling evolved differently depending on the imposed operating flux. Based on these experimental results, the increase of the fouling rate was evaluated through the determination of dTMP/dt values for each constant flux-step and each suspension (figure 3).



Figure 3: dTMP/dt versus flux during flux-stepping experiments of grapefruit juice, Red Extract and Seed Extract using 0.1 μm PES hollow-fiber membranes. Lines drawn to guide the eye.

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272

For low operating fluxes, a similar fouling behavior was observed for the three suspensions with values of 273 274 dTMP/dt lower than the threshold value of 1.0×10^{-5} bar.s⁻¹ (low fouling rate). Contrariwise, once specific 275 values of fluxes were exceeded, different fouling behaviors were identified. Concerning winery byproducts, 276 a significant increase of the fouling rate was observed between 7 and 13 L.h⁻¹.m⁻², with dTMP/dt values reaching 2.1×10^{-5} and 8.5×10^{-5} bar.s⁻¹ for Red and Seed Extracts, respectively. Therefore, the threshold flux 277 was defined as being around 7 $L.h^{-1}.m^{-2}$ for both winery byproducts in the studied operating conditions. 278 Concerning grapefruit juice, the significant increase of the fouling rate was observed between 5 and 10 L.h⁻ 279 1 .m⁻² fluxes, with a dTMP/dt increase reaching 1.3×10^{-4} bar.s⁻¹. Therefore, a threshold flux of around 5 L.h⁻¹ 280 ¹.m⁻² was identified for this suspension. 281

282

283 It can be noticed that the threshold flux related to grapefruit juice seemed slightly lower than the ones 284 identified for Red and Seed Extracts. Moreover, the fouling rate increased differently depending on the suspension. Above threshold flux, dTMP/dt values were substantially higher for grapefruit juice than for 285 Seed Extract or Red Extract. These differences are not surprising since each suspension is characterized by 286 specific biochemical and physicochemical properties. This result highlights the important impact of the 287 suspension characteristics on membrane fouling and fouling mechanisms. However, despite these 288 289 differences, the threshold fluxes of the three studied suspensions were in line with the threshold values reported for immersed membranes filtration in other fields. Indeed, threshold fluxes ranging between 5 and 290

10 L.h⁻¹.m⁻² were reported during immersed membranes filtration of milk protein concentrate solutions [43],
activated sludge [44], alginate, yeast and bentonite solutions [45].

293

294 3.3. Immersed membrane filtration performances

295 3.3.1. Membrane fouling

For all suspensions, the membrane fouling evolution in immersed membranes configuration was evaluated in sub-critical conditions, sustainable conditions for long-time filtration running. With a view to comparing the fouling behavior of the three suspensions, a same constant flux of 5 L.h⁻¹.m⁻² was chosen as operating flux. Figure 4 presents the evolution of TMP as function of VRR during filtration of grapefruit juice, Red and Seed Extracts.



301

302 *Figure 4:* TMP as function of VRR during filtration of grapefruit juice, Red extract and Seed Extract (batch concentration mode).

303

For the three suspensions, a continuous increase of TMP was observed between VRR 1 and VRR 2,reflecting an increase in membrane fouling.

306 Different fouling behaviors can be distinguished for the three suspensions. At the beginning of the filtration (between VRR 1 and 1.2), TMP remained almost constant for Red Extract with TMP surrounding 8.0×10^{-3} 307 bar and dTMP/dt close to zero. Contrariwise, an important and immediate fouling was observed for Seed 308 Extract and grapefruit juice, with TMP increasing from 1.0×10^{-2} to 3.7×10^{-2} bar and from 1.5×10^{-2} to 2.5×10^{-2} 309 ² bar, respectively. Starting VRR 1.2, fouling increased for Red Extract while it stabilized for grapefruit 310 juice leading to a final TMP of around 4.0×10⁻² bar at VRR 2 for both suspensions. Concerning Seed Extract 311 312 filtration, a continuous fouling was observed throughout the remainder of the filtration operation, with TMP reaching around 1.0×10^{-1} bar at VRR 2. 313

These differences of fouling behavior might be related to the physicochemical characteristics of each 314 suspension (table 3). Indeed, the three suspensions presented different dynamic viscosities, pH, DM, Brix 315 and turbidity and these parameters are known for having a significant impact on membrane fouling [40]. 316 317 However, considering the complexity of these biological suspensions, these physicochemical properties are 318 not sufficient to explain the different fouling behaviors of the studied suspensions. Indeed, membrane fouling results from complex interactions between the suspension compounds and the membrane and 319 320 between the suspension compounds themselves [51,52]. These phenomena are governed not only by the 321 operating conditions of the filtration operation and the physicochemical characteristics of the suspension, 322 but also by the biochemical nature of the suspension compounds. Hence, the differences of fouling behavior 323 between the studied suspensions might also be strongly related to the specific biochemical properties of 324 each suspension.

325

In order to estimate the importance of the fouling rate increase during a running operation, dTMP/dt were calculated for each suspension. Between VRR 1 and VRR 2, dTMP/dt values remained lower than the threshold value for the three suspensions, with dTMP/dt of 5.3×10^{-6} , 5.5×10^{-6} and 5.9×10^{-6} bar.s⁻¹ for grapefruit juice, Red and Seed Extracts, respectively. In the light of this, it seems that the fouling rates remained sustainable throughout the filtration operation, in batch concentration mode. This observation supports the interest of using the critical or threshold flux concept as pre-filtration test in order to define the optimal operating conditions prior to effective filtration experiments.

333

334 3.3.2. Membrane selectivity

The selectivity of immersed membranes was evaluated trough the characterization of turbidity loss (clarification), the retention of total phenolic compounds for the three suspensions, and the retention of carotenoids for grapefruit juice.

338 Concerning clarification efficiency, C_{clarif} of 99.9 %, 99.5% and 99.3 % were obtained for grapefruit juice,

Red and Seed Extracts, respectively, with permeates turbidities lower than 5 NTU for the three suspensions.
These results are in accordance with values reported in the literature during microfiltration of melon and

341 pomegranate juices [2,12]. For the three suspensions, immersed membranes filtration allowed decreasing

342 significantly the initial turbidity of the suspension leading to a clarified permeate.

344 Concerning phenolic compounds, initial feed suspensions presented very different concentrations of total phenolic compounds, with 145, 4076 and 5517 mg GAE per liter of grapefruit, Red Extract and Seed Extract, 345 respectively. These quantitative differences between grapefruit juice and winery byproducts are consistent 346 347 since winery byproduct are known for their particular richness in phenolic compounds [38,39]. As mentioned before, these phenolic compounds include, among others, naringin and narirutin for grapefruit 348 349 juice, tannins for Red and Seed Extracts and anthocyanins for Red Extract. Anthocyanins were also 350 quantified in Red Extract and there concentration was around 560 mg malvidin-3-glucoside.L⁻¹ which is in 351 the range of what is generally reported in the literature (from 300 to 900 mg malvidin-3-glucoside. L^{-1}) 352 [48,53]. These differences concerning at the same time the type and the concentration of bioactive 353 compounds in each suspension could have an impact on the membrane selectivity during the filtration operation. Table 4 presents the C_{ret} regarding bioactive compounds specific to each studied suspension. 354

355

Table 4: Retention ratios (%) regarding total phenolic compounds, anthocyanins, β- carotene, lycopene during grapefruit juice,
 Red and Seed Extracts microfiltration.

358

	Grapefruit juice	Red Extract	Seed Extract
Total phenolic compounds	8	14	3
Anthocyanins	/	17	/
β-carotene	96	/	/
Lycopene	96	/	/

359 Regardless of the suspension, phenolic compounds (including anthocyanins) were mainly recovered in the permeate with low Cret ranging between 3 and 17 %. Similar results have been reported during 360 microfiltration of pineapple juice (retention of 7 % of total phenolic content [14]), pomegranate juice 361 (retention of 16.5 % of total polyphenols and 11.7 % of anthocyanins [54]), red raspberry juice (retention of 362 363 16% of anthocyanins [17]). This is a consistent observation since most of phenolic compounds are smaller 364 than the membrane pore size $(0.1 \,\mu\text{m})$ with molecular weight generally ranging between 300 and 3000 Da 365 [55], even though some highly polymerized tannins (condensed tannins) have been reported to reach 20,000 Da [56]. However, membrane selectivity to phenolic compounds cannot be discussed only in terms of their 366 nominal molecular weight. Indeed, in the case of winery byproducts, low molecular weight phenolic 367 368 compounds can interact with each other or with other compounds, leading to the formation of complexes 369 with higher average size (e.g. tannin-anthocyanin, anthocyanin–anthocyanin, tannin-proteins complexes) 370 [8]. Thus, the higher C_{ret} observed for Red Extract phenolic compounds (14 %) and anthocyanins (17 %) 371 could be explained by complexation phenomena, leading to the formation of compounds larger than the

membrane pore size. The partial retention of phenolic compounds and anthocyanins could also be relatedtheir adsorption on/in the membrane layer [51].

374

375 Concerning carotenoids (β -carotene and lycopene), the concentration of lycopene was higher than that of 376 beta-carotene in grapefruit, with respective concentrations of 1.75 et 0.18 mg.L⁻¹. These differences of 377 concentrations did not have an influence on C_{ret} ratios, which were greater than 96 % for both compounds. 378 Despite their relatively low molecular weight (536.87 g.mol⁻¹), almost all carotenoids were retained by the 379 membrane. This phenomenon has been reported in many studied and is explained by the strong association 380 of carotenoids with the cell fragments membranes (i.e. pulp) due to their hydrophobicity [2]. Therefore, 381 carotenoids are mainly concentrated in the retentate during fruit juice microfiltration [2,50].

382

In the light of the above, immersed membranes filtration allowed producing (i) a permeate rich in phenolic compounds and clarified for the three suspensions and (ii) a retentate concentrated in carotenoids for grapefruit juice.

386

387 3.4. Comparison with conventional cross-flow filtration using side-stream membranes

388 The previous analysis allowed characterizing the performances of immersed membranes configuration in terms of productivity and selectivity. In order to validate the efficiency of immersed membranes 389 390 configuration for fruit-based suspensions microfiltration, previous productivity and selectivity results were compared with the performances of cross-flow filtration using side-stream tubular membranes. Indeed, this 391 392 filtration configuration remains the most used for fruit juices microfiltration [2,16–20]. Cross-flow filtration 393 experiments were performed under batch mode concentration (until VRR 2), using operating conditions 394 generally used when dealing with fruit-based suspensions microfiltration (0.1 µm inorganic tubular membranes, constant pressure of 1.5 bar [2,16,18,57]). The cross-flow velocity in each membrane and in 395 396 the loop tubes (with diameter close to the membrane one) was around 5 m.s⁻¹, corresponding to a Reynolds number and a membrane shear rate of 7 300 and 22 000 s⁻¹, respectively (turbulent flow) ([57]). 397

Steady-state fluxes (flux decay was observed for the three suspensions, followed by a pseudo-equilibrium of the flux) of 80 L.h⁻¹.m⁻², 30 L.h⁻¹.m⁻² and 24 L.h⁻¹.m⁻² were obtained for grapefruit juice, Red and Seed Extracts, respectively. These different flux values are in line with permeate flux values reported during microfiltration of citrus fruit juices (20 - 80 L.h⁻¹.m⁻² [57,58]) and winery byproducts (35 L.h⁻¹.m⁻² [38,59]). However, it is interesting to note that the ranking of the fouling potential obtained during cross-flow filtration experiments was slightly different than the one observed during immersed membranes filtration experiments. While Red Extract presented a low fouling potential close to that of grapefruit juice during 405 immersed membranes filtration experiments, it was one of the most foulant suspension during cross-flow 406 filtration experiments, with low flux $(30 \text{ L.h}^{-1}.\text{m}^{-2})$ compared to grapefruit juice. These differences of fouling

407 behavior might be related to the different hydrodynamic conditions of each studied filtration configuration.

408 However, an extensive characterization of the fouling behaviors of these suspensions (fouling mechanisms,

409 involved particles, etc.) while using immersed membranes filtration or cross-flow filtration is necessary to

410 go further on this path.

Concerning productivity, fluxes obtained for the three suspensions during cross-flow filtration were much 411 higher (5 to 16 times greater) than the threshold flux of 5 L.h⁻¹.m⁻² identified during immersed membranes 412 filtration. These differences of order of magnitude are in line with the fluxes generally reported for these 413 414 two configurations. Xue et al. (2015) compared the performances of an immersed membranes system with 415 a side-stream one (cross-flow) during waste leachate treatment and reported fluxes almost 20 times higher 416 when using cross-flow filtration [60]. Indeed, cross-flow filtration is an intensive process that works with high shear rates to ensure a high productivity but requires high energy consumption and investment cost. 417 418 On the opposite, immersed membranes system can be considered as an extensive process (gentle operating 419 conditions, low fluxes) for which productivity is generally improved by increasing the membrane surface 420 thanks to low investment costs and limited energy consumption [61,62]. Despite its relatively low productivity, immersed membranes configuration could offer interesting applications in fruit juices and 421 422 agro-food byproducts processing. Indeed, this simple processing system might be a convenient filtration 423 configuration for small agro-food producers with limited investment capacities or for the valorization byproducts with low added-value. However, further analysis are needed to confirm these suggestions, 424 425 notably concerning the process selectivity.

426

In order to compare the selectivity of both configurations, C_{clarif} and C_{ret} were calculated for cross-flow filtration experiments, based on analysis performed on the permeates collected at the end of the previous filtration experiments (batch concentration mode) and on the feed suspensions. C_{clarif} similar to the ones obtained for immersed membranes filtration were observed at the end of cross-flow filtration experiments (99.8 %, 96.4 % and 97.3 % for grapefruit juice, Red and Seed Extracts, respectively). Concerning C_{ret} , slight differences were observed between immersed membranes filtration and cross-flow filtration (figure 5).

434





Figure 5: Comparison of immersed membranes filtration (a) and cross-flow filtration (b) selectivity.

439 Concerning grapefruit juice carotenoids, both configurations presented very close C_{ret} (93 – 99 %). As 440 during immersed membranes filtration, most of carotenoids were concentrated in the retentate during cross-441 flow filtration experiments.

442 When comparing phenolic compounds for the three suspensions and anthocyanins for Red Extract, cross-443 flow filtration presented higher C_{ret} (10 – 33 %) than immersed membranes filtration (3 – 17 %).

444 These differences could be explained by adsorption mechanisms occurring on the mineral tubular membrane 445 used for cross-flow filtration experiments, as reported by Vladisavljević et al. (2013) [17]. Moreover, the differences of C_{ret} could also be related to the different hydrodynamic conditions specific to each 446 447 configuration. Indeed, the high cross-flow velocities and high TMP specific to cross-flow filtration might 448 enhance the interactions between the phenolic compounds and between phenolic compounds and other Red 449 Extract compounds, leading to the formation of complexes larger than the membrane average pore diameter [63,64]. Considering the many differences between both configurations, it is difficult to state which 450 phenomenon is responsible of C_{ret} differences. 451

To sum up, immersed membranes filtration allowed producing retentate and permeate of quality as least as good as the one obtained with conventional cross-flow filtration. Concerning the process productivity, crossflow filtration was the most interesting filtration configuration in terms of flux values for the three studied suspensions. However, the productivity of immersed membranes configuration could be easily increased by increasing the membrane surface (low cost of the membranes and great packing density) and thanks to the low energy consumption of this system. Moreover, immersed membranes system could find interesting application for small production units thanks to its compactness, easy handling and mobility.

459 **4.** Conclusion

The aim of this work was to investigate the possibility of using an immersed membranes configuration for the microfiltration of fruit-based suspensions. The study focused on three different agro-food suspensions, for which microfiltration is widely used for clarification and concentration purpose, a grapefruit juice and two winery byproducts. The performances of a pilot-scale immersed membranes system were characterized in terms of membrane permeability (membrane fouling, permeate flux) and selectivity (clarification, concentration of targeted compounds).

466 Flux-stepping experiments performed under total recycle mode followed by filtration experiments under 467 batch concentration mode allowed defining and validating the optimal operating domains of the studied 468 system. The optimal permeate flux of immersed membranes configuration was found to be around 5 - 7L.h⁻¹.m⁻² for the three studied suspensions, which is in line with permeate fluxes reported in other fields 469 where immersed configurations have been widely and successfully used. Selectivity analysis showed that 470 471 immersed membranes filtration allowed reaching the selectivity goals specific to each suspension by 472 producing (i) a clarified permeate rich in phenolic compounds and (ii) a retentate concentrated in carotenoids 473 for grapefruit juice. Finally, immersed membranes filtration productivity and selectivity results were 474 compared with performances of conventionally used cross-flow filtration with tubular membranes. The 475 main outcomes were that immersed membranes filtration allowed producing retentate and permeate with 476 quality as least as good as the one related to conventional cross-flow filtration. Concerning the process 477 productivity, cross-flow filtration allowed reaching high fluxes compared to immersed membranes 478 filtration. These differences were in line with the respective order of magnitude of fluxes reported for both 479 configurations. However, thanks to its high compactness, easy handling and mobility, low investment and 480 operational costs, immersed membranes configuration could find interesting applications in small 481 production units of fruit juices and industries dealing with the valorization of byproducts with low added-482 value. Immersed membranes configuration could be an affordable and simple process to perform the *in situ* 483 clarification/concentration of fruit-based suspensions, limiting loss and wastage due to processing delay.

All these findings offer new prospects for immersed membranes configuration applied to fruit-based
 suspensions microfiltration. Further investigations would be of great interest to get more insight into the
 involved fouling mechanisms occurring during immersed membranes filtration of fruit-based suspensions.

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