



HAL
open science

Circulating soluble intercellular adhesion molecule-1 (sICAM-1) after exercise-induced muscular damage: Does the use of whole-body cryostimulation influence its concentration in blood?

François Bieuzen, Christophe Hausswirth, Benoit Dugué

► To cite this version:

François Bieuzen, Christophe Hausswirth, Benoit Dugué. Circulating soluble intercellular adhesion molecule-1 (sICAM-1) after exercise-induced muscular damage: Does the use of whole-body cryostimulation influence its concentration in blood?. *Cryobiology*, 2019, 87, pp.120 - 122. 10.1016/j.cryobiol.2019.01.013 . hal-03485716

HAL Id: hal-03485716

<https://hal.science/hal-03485716>

Submitted on 20 Dec 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial 4.0 International License

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

Circulating soluble intercellular adhesion molecule-1 (sICAM-1) after exercise-induced muscular damage: does the use of whole-body cryostimulation influence its concentration in blood?

Bieuzen F^{a,b}, Hauswirth C^{a,c,d}, Dugué B^e

^aFrench National Institute of Sport (INSEP), Laboratory of Sport, Expertise and Performance - EA 7370, Research Department, Paris, France

^b Québec National Institute of Sport, Montréal, QC, Canada.

^c Laboratory of Human Motricity, Education Sport and Health - EA 6312, University of Nice Sophia Antipolis, Nice, France.

^d Mouratoglou Tennis Academy, Medical Center, Biot, France

^e University of Poitiers, Laboratoire Mobilité Vieillessement Exercice (MOVE)-EA6314, Faculty of Sport Sciences, Poitiers, France

Corresponding author:

Benoit Dugué, PhD ; Université de Poitiers et Laboratoire « Mobilité, Vieillessement, Exercice » ; 8 allée Jean Monnet, 86000, Poitiers, France.

E-mail: benoit.dugue@univ-poitiers.fr

33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56

Abstract

As soluble intercellular adhesion molecule-1 (sICAM-1) was recently hypothesized to be a key player in the mechanisms involved in exercise-induced muscular damage (EIMD), we investigated its circulating concentration changes in athletes before and after EIMD with and without the use of whole-body cryostimulation (WBC; 3 min at -110°C) at the exercise end **and repeated once a day during 4 days.**

We previously characterized plasma specimens from 11 endurance athletes who performed twice (randomized crossover design) strenuous running leading to EIMD, followed by passive recovery or WBC. Muscle soreness and inflammatory response were observed in both cases but the use of WBC induced a significant reduction in these responses (PlosOne 2011;6:e22748). We now found that sICAM-1 concentration slightly increased in both circumstances and remained elevated for 24 hours ($p<0.01$). However, no significant WBC effect was observed concerning sICAM-1 changes indicating that this compound is not a major player both in EIMD and WBC physiological impacts.

Key words: Exercise-induced muscular damage (EIMD); Intercellular adhesion molecule-1 (ICAM-1); Soluble intercellular adhesion molecule-1 (sICAM-1); Whole-body cryostimulation; Whole-body cryotherapy; Recovery

57 Intercellular adhesion molecule-1 (ICAM-1) and its soluble form (sICAM-1) have
58 recently been suspected to be key components in the mechanisms involved in
59 exercise-induced muscular damage (EIMD) [4; 3]. It has been speculated that whole-
60 body cryostimulation (WBC; 2 to 4 min at -110°C or less after exercise) which lowers
61 EIMD also impacts the expression of ICAM-1 and the amount of circulating soluble
62 ICAM-1 (sICAM-1) [4; 3].

63
64 ICAM-1 is an adhesion protein present on endothelial cells and leukocyte surfaces.
65 This factor mediates adhesion and transmigration of leukocytes through the
66 endothelium. In addition, ICAM-1 directly contributes to inflammatory responses
67 within the blood vessel wall by increasing endothelial cell activation. Surface
68 expressed ICAM-1 can be shed from the cells and then circulates as soluble ICAM-1
69 (sICAM-1) [3; 10; 7]. Initially thought to be a relevant marker of cardiovascular risk
70 factor, sICAM-1 is nowadays more frequently considered as a biomarker of
71 endothelial activation/dysfunction mimicking the amount of expressed ICAM on
72 endothelial cells and leukocytes [7].

73
74 We have recently presented that in the context of EIMD sarcomeres disrupt and then
75 leukocytes (neutrophils, monocytes, and lymphocytes) transmigrate to the injured
76 cells/tissue via ICAM-1 [4; 3]. Following this, pro-inflammatory cytokines and reactive
77 oxygen species are produced in the muscle cells by leukocytes and amplify the initial
78 muscle damage.

79
80 Ferreira et al have suggested that a drop in core temperature induced by WBC would
81 likely cause vasoconstriction of blood vessels and lower the amount of leukocytes
82 arriving to the exercise-induced muscular inflammation site [4]. These authors
83 propose a mechanism in which a decrease in the expression of ICAM-1 at the
84 endothelium level would limit diapedesis through a reduction a: in the possibilities of
85 activated leukocytes arrest, b: in a firm adherence on the endothelial cells, and c: in
86 the initial steps of the trans-endothelial cell migration process. The level of this
87 reduction in the expression of ICAM-1 could be followed by measuring the
88 concentration of circulating sICAM-1.

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

117

118

119

120

This view may be correct if the rate of shedding (splicing) remains constant. It seems accepted that sICAM-1 is an accurate marker of inflammation reflecting the amount of ICAM-1 expressed on membranes. During inflammation and infection, membrane ICAM-1 expression on leukocytes and endothelial cells tremendously increases as well as the concentration of circulating sICAM-1. In this context sICAM-1 is a relevant biomarker which correctly reflects the expression of ICAM-1 on cell membranes. However, this might not always be the case. If the shedding of ICAM-1 is regulated, one can expect changes in the concentration of sICAM-1 in blood without changes in the expression of ICAM-1. The regulation of sICAM-1 in blood is definitely not clear [3]. It has previously been shown that an increase in the shedding of the ICAM-1 can be due to increased secretases activity, stimulated by catecholamines and other factors. One study showed that exercise may induce an increase in circulating sICAM-1 and such change could be blocked by the infusion of propranolol [9]. Moreover, ~~we have shown that~~ noradrenaline is produced and released in circulation after cold stimulation [7; 1] and can theoretically induce an extra release of sICAM-1 with or without changes in the expression of ICAM-1 [2]. In this case, the release of the soluble part of the ICAM-1 will leave a truncated form of the ICAM-1 at the membrane site which will not be able to play its role as an adhesion molecule. Therefore, the possibilities of leukocyte transmigration and the subsequent muscle inflammation will be reduced. This can be seen as a paradox, as in this case, an extra release of sICAM-1 may induce a decrease in muscle inflammation.

Nevertheless, there is for the time being no information available on the changes of sICAM-1 due to the combine effects of EIMD and whole-body cryostimulation. ~~A few years ago, we~~ We have previously studied the effects of EIMD and whole-body cryostimulation in high level athletes on perceived muscle soreness, inflammation markers and pro and anti-inflammatory cytokines. We were able to show that the heavy exercise our athletes had to perform led to muscle soreness accompanied by pro-inflammatory response. The use of whole-body cryostimulation (-110°C, 3min of exposure) after the exercise ~~and repeated once a day during 4 days~~ induced in our athletes both a reduction in muscle soreness and in the concentration of circulating

121 markers of inflammation (lower blood concentration in C-reactive protein and
122 interleukin-1beta) and an increase in the concentration of anti-inflammatory
123 circulating markers (increase in the concentration in blood interleukin-1 receptor
124 antagonist) [8].

125
126 The aim of the present study was to investigate ~~whether sICAM-1 is an important~~
127 ~~regulating substance after EIMD and cryostimulation.~~ whether sICAM-1 is an
128 important regulating substance in formation of EIMD and whether cryostimulation
129 can modulate its concentration in blood (vs. passive recovery). To conduct this
130 analysis, we used well preserved plasma specimens [5] collected from a previous
131 experiment [8] where we investigated the effect of two different recovery modalities
132 (passive recovery and WBC) after a simulated trail running race that created a EIMD
133 in our athletes.

134
135 The subjects and the methods have previously been described [8]. In summary, 11
136 well-trained male runners (age 31.8 ± 2.0 years; VO_{2max} 62.0 ± 1.2 ml.min⁻¹.kg⁻¹)
137 participated in the study. All of them gave their informed consent and the study was
138 approved by the local Ethics Committee (Île-de-France XI, France; Ref. 200978). The
139 study had a crossover design. The subjects completed two simulated runs at a one-
140 month interval on a treadmill followed in a random order by a passive recovery or
141 with a recovery where WBC was used. The runs were organized on the same
142 treadmill and were designed to generate fatigue. Blood specimens were collected
143 before and after the simulated run, after the first recovery session, and before the
144 recovery sessions after 24 h, 48 h, 72 h, 96 h following the end of the run.
145 Hydration/nutrition before and during each session was standardized. Between trials,
146 low intensity training was ensured.

147
148 One week before the first simulated run, subjects were familiarized with the test
149 scheme and preliminary tests were performed to determine maximal oxygen uptake,
150 the first and the second ventilatory thresholds, and the maximal aerobic speed as
151 previously described [8]. Concerning the proper test, the race lasted 48 minutes and
152 was divided into 5 stages. The first stage consisted of a 6 minute run on treadmill on

153 a flat surface (intensity comprised between the subject first and the second
154 ventilatory thresholds), followed by 3 minute uphill (+10% gradient; ≈80% of the
155 subject maximal aerobic velocity) and 3 minute downhill (-15% gradient; first
156 ventilatory threshold). Stages 2–5 consisted of 3 min at 0°, followed by 3 min uphill
157 and 3 min downhill at the gradients and velocities as previously described [8]. After
158 the run, subjects were randomly assigned to WBC or passive recovery. WBC sessions
159 were administered in a cryogenic chamber (Zimmer MedizinSysteme GmbH, Ulm,
160 Germany) and the subjects were exposed to -110°C for 3 min. **The exposures**
161 **occurred at the end of the exercise and repeated once a day during 4 days at the**
162 **same time of the day.** Before the exposure, subjects were instructed to dry any
163 sweat, and clothe themselves in swimming trunks, a surgical mask, ear bands, gloves,
164 dry socks and sabots. After **each** WBC session, subjects spent 10 min comfortably
165 seated in an environment at 24°C wearing a bath robe. The control recovery was a
166 passive recovery during which each subject was comfortably seated in an armchair
167 for 30 min at 24°C. The subjects were not allowed to speak to anyone during the
168 experiment.

169
170

171 Blood specimens were collected from a superficial forearm vein using standard
172 venipuncture techniques and EDTA tubes (Greiner Bio-one; Frickenhausen,
173 Germany). Plasma aliquots were prepared and stored at -80°C. ELISA kits for the
174 determination of sICAM-1 were purchased from R&D Systems Europe. The intra-
175 assay imprecision (CVA) was 4.6% and 4.8% for sICAM-1 concentrations of 126 and
176 476 mg/L, respectively. All specimens were analyzed in duplicate using Dynex MRXe
177 spectrophotometer (Magellan Biosciences, Chelmsford, MA, USA).

178

179 Results are expressed in Figure 1 as means ± SD. Two-way analysis of variance for
180 repeated measurements was used to analyze the changes between cold and control
181 experiments with time and treatments (WBC versus control) as factors. We used
182 logarithm transformation as the data distribution was not Gaussian. Post hoc
183 multiple comparisons were made by PLSD test when appropriate to single out
184 statistical significance. Statistical significance was set at $p < 0.05$.

185

186 We observed a significant increase (data not shown) in the blood concentration of
187 sICAM-1 and leukocytes after exhausting exercise in athletes, who experienced a
188 significant post exercise inflammatory response, with or without the use of a 3-min
189 cryostimulation exposure as a recovery procedure at the end of the exercise. The
190 concentrations of sICAM-1 in blood obtained 1 hour and 24 hours after the end of the
191 exercise were significantly more elevated than those obtained at rest ($p < 0.01$),
192 whereas the sICAM-1 concentration obtained at the time point of 48, 72 and 96h
193 were not significantly different than those obtained at rest (Fig 1). However, no
194 significant interaction concerning the effect of cryostimulation was observed
195 concerning circulating sICAM-1.

196

197 This work confirmed that the concentration of sICAM-1 in blood increases after a
198 high intensity aerobic exercise [6]. In our previous analysis with the same subjects
199 and blood specimens, we reported a reduction both in muscle soreness and in the
200 concentration of circulating markers of inflammation (lower blood concentration in
201 C-reactive protein and interleukin-1beta) and an increase in the concentration of
202 anti-inflammatory circulating markers (increase in the concentration in blood
203 interleukin-1 receptor antagonist) [8] when athletes experienced cryostimulation
204 after exhausting exercise. Such significant physiological changes did not seem to alter
205 the concentration of sICAM-1 in blood, though a minor reduction was observed
206 compared with control group.

207

208 One limit of our study is the rather low number of participants ($n=11$) which lowers
209 the statistical power. Therefore, such trend might favor the hypothesis of a certain
210 reduction in the expression of ICAM-1 on cell membranes (endothelial cells and
211 leukocytes), a reduction in the secretase activity, or a combined effect at the
212 expression level of the protein and in the activity of secretases.

213

214 Trans-endothelial migration is divided into at least four different steps: the
215 attachment of circulating cells on to the endothelium; the activation of integrins on
216 the leukocyte cell surface; the firm adhesion of the leukocyte and transmigration of

217 the cell through the endothelial cell barrier into the sub-endothelial space [7]. The
218 role of ICAM-1 is to increase endothelial cell activation, to facilitate the firm arrest of
219 leukocytes and to initiate the leukocytes' transmigratory process [7]. However, there
220 are many other proteins (e.g. selectins, chemokines, integrins, IgSF) involved in
221 diapedesis, and cryostimulation may influence the expression of those proteins
222 involved in the transmigration process. If each of these proteins as well as ICAM-1
223 were slightly less expressed after cryostimulation, a global significant impact might
224 appear and reduces the exercise induced inflammation. Further studies dealing with
225 the impact of cryostimulation and the changes of proteins involved in trans-
226 endothelial migration would still be necessary. Also, it would be of interest to
227 investigate downstream signaling using muscle and endothelial cells in order to get a
228 deeper insight into exercise-induced muscle damage mechanisms and to rule out
229 sICAM-1 and ICAM-1's roles.

230

231

232 **References**

- 233 [1] R. Bouzigon, F. Grappe, G. Ravier, and B. Dugué: Whole- and partial-body
234 cryostimulation/cryotherapy: Current technologies and practical applications. J
235 Therm Biol 61 (2016) 67-81.
- 236 [2] B. Dugué, E. Leppänen, and R. Gräsbeck: Preanalytical factors (biological variation) and
237 the measurement of serum soluble intercellular adhesion molecule-1 in humans:
238 influence of the time of day, food intake, and physical and psychological stress. Clin
239 Chem 45 (1999) 1543-7.
- 240 [3] B. Dugué: An attempt to improve Ferreira-Junior model concerning the anti-inflammatory
241 action of whole-body cryotherapy after exercise induced muscular damage (EIMD).
242 Front Physiol 6 (2015) 35.
- 243 [4] J. Ferreira-Junior, M. Bottaro, J. Loenneke, A. Vieira, C. Vieira, and M. Bembem: Could
244 whole-body cryotherapy (below -100°C) improve muscle recovery from muscle
245 damage? Front Physiol 5 (2014) 247.
- 246 [5] J. Hartweg, M. Gunter, R. Perera, A. Farmer, C. Cull, C. Schalkwijk, A. Kok, H. Twaalfhoven,
247 R. Holman, and A. Neil: Stability of soluble adhesion molecules, selectins, and C-

248 reactive protein at various temperatures: implications for epidemiological and large-
249 scale clinical studies. *Clin Chem* 53 (2007) 1858-60.

250 ~~[5]~~ [6] Y. Koh, and J. Park: Cell adhesion molecules and exercise. *J Inflamm Res* 11 (2018)
251 297-306.

252 ~~[6]~~ [7] C. Lawson, and S. Wolf: ICAM-1 signaling in endothelial cells. *Pharmacological Reports*
253 61 (2009) 22-32.

254 ~~[7]~~ J. Leppäluoto, T. Westerlund, P. Huttunen, J. Oksa, J. Smolander, B. Dugué, and M.
255 Mikkelsen: Effects of long-term whole-body cold exposures on plasma
256 concentrations of ACTH, beta-endorphin, cortisol, catecholamines and cytokines in
257 female subjects. ~~*Scand J Clin Lab Invest* 68 (2008) 145-53.~~

258 [8] H. Pournot, F. Bieuzen, J. Louis, R. Mounier, J.R. Fillard, E. Barbiche, and C. Hauswirth:
259 Time-course of changes in inflammatory response after whole-body cryotherapy
260 multi exposures following severe exercise. *PLoS One* 6 (2011) e22748.

261 [9] J. Rehman, P. Mills, S. Carter, J. Chou, J. Thomas, and A. Maisel: Dynamic exercise leads to
262 an increase in circulating ICAM-1: further evidence for adrenergic modulation of cell
263 adhesion. *Brain Behav Immun* 11 (1997) 343-351.

264 [10] P.M. Ridker, C.H. Hennekens, B. Roitman-Johnson, M.J. Stampfer, and J. Allen: Plasma
265 concentration of soluble intercellular adhesion molecule-1 and risks of future
266 myocardial infarction in apparently healthy men. *Lancet* 351 (1998) 88-92.

267

268 Figure 1.

269

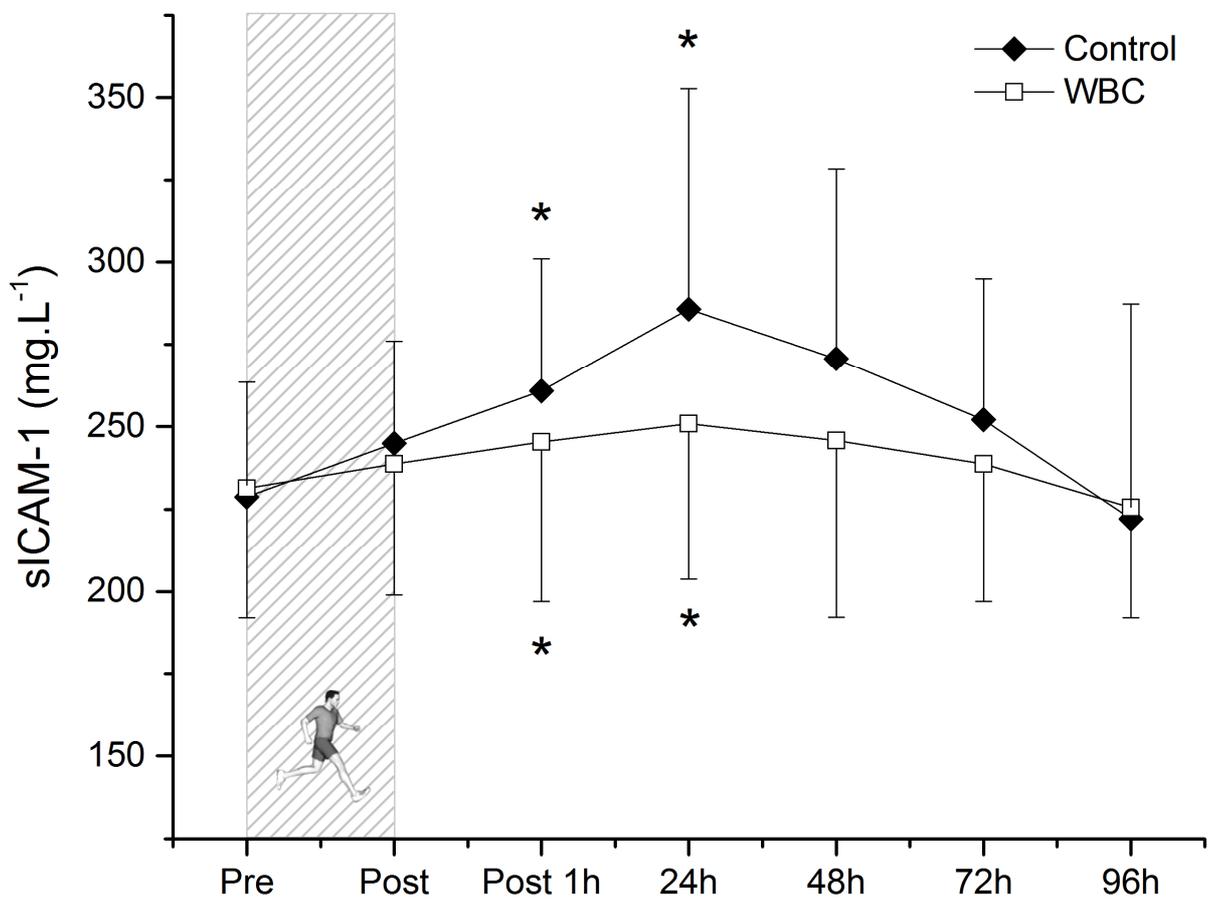
270 Changes in circulating sICAM-1 ($\text{ng}\cdot\text{ml}^{-1}$) from basal concentration to recovery after
271 exhausting physical exercise without (diamond) or with (square) using a 3min
272 cryostimulation exposure in trained athletes. Asterisk indicates a significant increase
273 ($P<0.01$) in comparison to basal circulating concentrations.

274

275

276

277



278

279

280

281

282