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1	The stimulation by ethylene of the UDP glucose-flavonoid 3-O-
2	glucosyltransferase (UFGT) in grape tissues is independent from the
3	MybA transcription factors
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11	Introduction: Grapes are regarded as non-climacteric fruit (CHERVIN et al., 2004 and
12	references herein), in which ethylene evolution is very low and for which the ripening
13	process seems to occur independently of ethylene production. However a recent study has
14	shown that grape berry tissues have a fully functional pathway for ethylene synthesis, that
15	this pathway is activated just before veraison when red berries start to accumulate
16	anthocyanins, and that ethylene perception is critical for some berry changes associated with
17	ripening, including anthocyanin accumulation (CHERVIN et al., 2004). This last conclusion
18	was supported by the fact that anthocyanin accumulation in the berry skins was inhibited
19	when a specific inhibitor of ethylene receptors, 1-methylcyclopropene was applied to berries
20	just before veraison, at the same time as ethylene production was activated in the berry.
21	The control of anthocyanin accumulation during the ripening phase in red grape
22	berries is thought to be greatly dependent on UFGT activity (KOBAYASHI et al., 2001;

24 the anthocyanins (PIFFAUT *et al.*, 1994) and may be essential for their transport to the 25 vacuole. We have shown previously that exogenous ethylene could induce grape ufgt

YAKUSHIJI et al., 2006). This enzyme plays a key role in stabilising the aglycone moiety of

expression (EL-KEREAMY *et al.*, 2003), confirming observations about the commercial use
 of ethylene precursor to boost grape skin colour.

The anthocyanin biosynthesis in model plants for the anthocyanin pathway such as petunia and maize is controlled by Myc and Myb transcription factors, and indeed, one transcription factor of this family, MybA1, was shown to be critical for the regulation of *ufgt* expression (YAKUSHIJI *et al.*, 2006 and refs therein) and anthocyanin accumulation in grape tissues. Walker et al. (2007) showed that two isoforms of this transcription factor, MybA1 and MybA2 are involved in anthocyanin accumulation in grapevine.

9 The question remains whether the ethylene signal is acting on grape *ufgt* expression 10 through the activation of *mybA* expression or via ethylene responsive transcription factors.

11

12 Materials and methods: Cabernet Sauvignon grapevines used for the ethylene 13 experiments are grafted on 110 Richter rootstocks and grown in Toulouse, South-West of 14 France, in a non-irrigated vineyard. Full bloom occurred around mid-June. The ethylene was 15 applied for 24 h at different times after full bloom, at an initial concentration of 40 μ l.l⁻¹, 16 three biological replicates were then collected and stored at -80°C. All other sampling 17 processes and the RNA extractions were performed as described in EL-KEREAMY et al. 18 (2003). Suspensions of purple grape cells, cv. Gamay were grown as described previously 19 (TRIANTAPHYLIDES et al., 1993). Cells were vacuum-filtered onto Whatman n°1 filter paper 20 and set upon 5 ml fresh MS-based cell culture medium in small Petri dishes. Cells were then 21 sprayed with one ml of a sterile solution of 7 mM 2-chloroethylphosphonic acid (2-CEPA), 22 a precursor of ethylene, the controls were sprayed with sterile water. Incubation time was 6 23 hours before freezing samples for RNA extraction, performed as described above.

The qRT-PCR analyses were performed according to EL-SHARKAWY *et al.* (2005), with the following modifications. Oligonucletide sequences of *mybA* primers were *VvmybA1c*(F), 5'-GCAAGCCTCAGGACAGAA -3' and *VvmybA1c*(R), 5'-

AAGCCCACATCAAATGGAAAA-3'. They would amplify both VvmybA1 (AB097923) 1 2 and VvmybA2 (AB097924) isoforms. Oligonucletide sequences for the ufgt primers were 3 5'-GGCTTTTGTCACACATTGCG-3' *Vvufgt*(F) and *Vvufgt*(**R**) 5'-4 AAAAAGGGCCTGCAAATCAA-3'. The myb et ufgt mRNA levels were expressed as 5 transcript accumulation indexes relative to a control gene $efl\alpha$, as previously shown 6 (TERRIER et al., 2005). The LSD value between treatments was calculated at the 5% level 7 using a one way ANOVA (SigmaStat, Systat Software Inc., San Jose, CA).

8 The ethylene cis-elements were estimated by homology search using PLACE 9 database, http://www.dna.affrc.go.jp/PLACE/signalscan.html or PlantCARE database, 10 http://bioinformatics.psb.ugent.be/webtools/plantcare/html/.

11

12 **Results and Discussion:** The application of gaseous ethylene significantly 13 stimulated the accumulation of *ufgt* mRNA in berries when applied 8 and 9 weeks after full 14 bloom (Fig. 1a). These observations made in Cabernet Sauvignon berry tissues were 15 confirmed in another cultivar, under different experimental conditions, the Gamay cell 16 suspension sprayed with the 2-CEPA, an ethylene precursor (Fig. 1c). These results 17 confirmed previous observations, either the enhancement of anthocyanin accumulation after 18 spraying 2-CEPA onto the grape clusters (EL-KEREAMY et al., 2003), or the inhibition of 19 anthocyanin accumulation following a treatment of the clusters with the ethylene inhibitor, 20 1-methylcyclopropene (CHERVIN et al., 2004).

However no significant difference was seen for the accumulation *mybA* mRNAs in the same experimental conditions (Fig. 1b and 1c). This absence of responsiveness to an ethylene signal was further confirmed by the analysis of the *VvmybA1* promoter, GenBank AB242302 (the sequence of the *VvmybA2* promoter was not available when we wrote this article), which showed three ethylene cis-elements (ethylene response elements, ERE) in the

-2 kb upstream region (Fig. 2a). The closest GCCGCC box, one of the most potent EREs 1 2 (FUJIMOTO et al., 2000), locates around -2100 bp. The closest ATTTnAAA box, the other 3 common ERE, locates at -850 bp from mybA1 start codon (Fig 2). In comparison there are 4 seven EREs in the ufgt promoter, GenBank AY955269, (Fig. 2). The GCCGCC sequence 5 was found at -160 bp from the *ufgt* start codon, a good place to be active (Dr Ohme-Takagi, 6 pers. comm. and analyses of ethylene responsive promoters, data not shown). Two other 7 ERE sequences, ATTTnAAA are reasonably close to the gene start codon, located between -8 450 and -350 bp. In another ethylene responsive promoter, they were shown to be active 9 when placed at -500 bp (Itzhaki et al., 1994). Here we used the sequence of the Shiraz ufgt 10 promoter, which has 97 to 99 % homology to the grapevine *ufgt* promoter sequences isolated 11 from the Kyoho, Italia, Ruby Oku, Muscat of Alexandria and Flame Muscat cultivars 12 (KOBAYASHI et al., 2001), and 95 % to Cabernet Sauvignon (GenBank AY919624). The 13 alignment of various ufgt promoter sequences has already been showed (KOBAYASHI et al., 14 2001).

15

16 So we conclude that the stimulation of *ufgt* expression by ethylene is independent 17 from mybA expression. But it does not preclude the importance of MybA in controlling the 18 *ufgt* expression. Indeed, Dr Goto-Yamamoto found that the MybA protein binds to a region 19 of the *ufgt* promoter around -428 to -303 bp from the start codon (pers. comm.) and recent 20 works confirm the role of MybA in stimulating anthocyanin accumulation (Yakushiji et al., 21 2006; Walker et al., 2007).

22

To our knowledge this is the first report addressing the responsiveness of myb expression to an ethylene signal.

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2	Genopole Toulouse where the qRT-PCR analyses were performed.
3	
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13	
14	Figure captions
15	Figure 1: Effects of exogenous ethylene to a) the ufgt mRNA levels, to b) the mybA1 mRNA
16	levels in grape berries, cv. Cabernet Sauvignon according to time after full bloom, and to c)
17	mRNAs of both genes in grape cell suspensions, cv. Gamay. The mRNA levels were
18	determined by qRT- PCR analyses and are expressed as the transcript accumulation index
19	relative to $ef1-\alpha$. Each bar represents the mean of three biological replicates, and LSD bars
20	were calculated at the 0.05 level for differences between the treatments (control or ethylene).
21	

<u>Figure 2:</u> Locations of ethylene responsive elements (cis-acting) on the promoter of *mybA1*(AB242302) and on the promoter of *ufgt* (AY955269) of *Vitis vinifera*. The nucleotide
numbering is relative to the start codon (0). Cis-elements were estimated by homology
search using PLACE and PlantCARE databases.





Figure 2