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# The stimulation by ethylene of the UDP glucose-flavonoid 3-O-

## glucosyltransferase (UFGT) in grape tissues is independent from the

### **MybA transcription factors**

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Introduction: Grapes are regarded as non-climacteric fruit (CHERVIN et al., 2004 and references herein), in which ethylene evolution is very low and for which the ripening process seems to occur independently of ethylene production. However a recent study has shown that grape berry tissues have a fully functional pathway for ethylene synthesis, that this pathway is activated just before veraison when red berries start to accumulate anthocyanins, and that ethylene perception is critical for some berry changes associated with ripening, including anthocyanin accumulation (CHERVIN et al., 2004). This last conclusion was supported by the fact that anthocyanin accumulation in the berry skins was inhibited when a specific inhibitor of ethylene receptors, 1-methylcyclopropene was applied to berries just before veraison, at the same time as ethylene production was activated in the berry.

The control of anthocyanin accumulation during the ripening phase in red grape berries is thought to be greatly dependent on UFGT activity (KOBAYASHI *et al.*, 2001; YAKUSHIJI *et al.*, 2006). This enzyme plays a key role in stabilising the aglycone moiety of the anthocyanins (PIFFAUT *et al.*, 1994) and may be essential for their transport to the vacuole. We have shown previously that exogenous ethylene could induce grape *ufgt* 

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expression (EL-KEREAMY *et al.*, 2003), confirming observations about the commercial use of ethylene precursor to boost grape skin colour.

The anthocyanin biosynthesis in model plants for the anthocyanin pathway such as petunia and maize is controlled by Myc and Myb transcription factors, and indeed, one transcription factor of this family, MybA1, was shown to be critical for the regulation of *ufgt* expression (YAKUSHIJI *et al.*, 2006 and refs therein) and anthocyanin accumulation in grape tissues. Walker et al. (2007) showed that two isoforms of this transcription factor, MybA1 and MybA2 are involved in anthocyanin accumulation in grapevine.

The question remains whether the ethylene signal is acting on grape ufgt expression through the activation of mybA expression or via ethylene responsive transcription factors.

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Materials and methods: Cabernet Sauvignon grapevines used for the ethylene experiments are grafted on 110 Richter rootstocks and grown in Toulouse, South-West of France, in a non-irrigated vineyard. Full bloom occurred around mid-June. The ethylene was applied for 24 h at different times after full bloom, at an initial concentration of 40 µl.l<sup>-1</sup>, three biological replicates were then collected and stored at -80°C. All other sampling processes and the RNA extractions were performed as described in EL-KEREAMY et al. (2003). Suspensions of purple grape cells, cv. Gamay were grown as described previously (TRIANTAPHYLIDES et al., 1993). Cells were vacuum-filtered onto Whatman n°1 filter paper and set upon 5 ml fresh MS-based cell culture medium in small Petri dishes. Cells were then sprayed with one ml of a sterile solution of 7 mM 2-chloroethylphosphonic acid (2-CEPA), a precursor of ethylene, the controls were sprayed with sterile water. Incubation time was 6 hours before freezing samples for RNA extraction, performed as described above. The qRT-PCR analyses were performed according to EL-SHARKAWY et al. (2005), with the following modifications. Oligonucletide sequences of mybA primers were VvmybA1c(F), 5'-GCAAGCCTCAGGACAGAA -3' VvmybA1c(R), 5'and

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- 1 AAGCCCACATCAAATGGAAAA-3'. They would amplify both *VvmybA1* (AB097923)
- and VvmybA2 (AB097924) isoforms. Oligonucletide sequences for the ufgt primers were
- 3 Vvufgt(F) 5'-GGCTTTTGTCACACATTGCG-3' and Vvufgt(R) 5'-
- 4 AAAAAGGCCTGCAAATCAA-3'. The myb et ufgt mRNA levels were expressed as
- 5 transcript accumulation indexes relative to a control gene  $efl \alpha$ , as previously shown
- 6 (TERRIER et al., 2005). The LSD value between treatments was calculated at the 5% level
- 7 using a one way ANOVA (SigmaStat, Systat Software Inc., San Jose, CA).

8 The ethylene cis-elements were estimated by homology search using PLACE

database, http://www.dna.affrc.go.jp/PLACE/signalscan.html or PlantCARE database,

http://bioinformatics.psb.ugent.be/webtools/plantcare/html/.

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**Results and Discussion:** The application of gaseous ethylene significantly stimulated the accumulation of *ufgt* mRNA in berries when applied 8 and 9 weeks after full bloom (Fig. 1a). These observations made in Cabernet Sauvignon berry tissues were confirmed in another cultivar, under different experimental conditions, the Gamay cell suspension sprayed with the 2-CEPA, an ethylene precursor (Fig. 1c). These results confirmed previous observations, either the enhancement of anthocyanin accumulation after spraying 2-CEPA onto the grape clusters (EL-KEREAMY *et al.*, 2003), or the inhibition of anthocyanin accumulation following a treatment of the clusters with the ethylene inhibitor, 1-methylcyclopropene (CHERVIN *et al.*, 2004).

However no significant difference was seen for the accumulation *mybA* mRNAs in the same experimental conditions (Fig. 1b and 1c). This absence of responsiveness to an ethylene signal was further confirmed by the analysis of the *VvmybA1* promoter, GenBank AB242302 (the sequence of the *VvmybA2* promoter was not available when we wrote this article), which showed three ethylene cis-elements (ethylene response elements, ERE) in the

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-2 kb upstream region (Fig. 2a). The closest GCCGCC box, one of the most potent EREs 1 2 (FUJIMOTO et al., 2000), locates around –2100 bp. The closest ATTTnAAA box, the other 3 common ERE, locates at -850 bp from mybA1 start codon (Fig 2). In comparison there are 4 seven EREs in the *ufgt* promoter, GenBank AY955269, (Fig. 2). The GCCGCC sequence 5 was found at -160 bp from the *ufgt* start codon, a good place to be active (Dr Ohme-Takagi, 6 pers. comm. and analyses of ethylene responsive promoters, data not shown). Two other 7 ERE sequences, ATTTnAAA are reasonably close to the gene start codon, located between -8 450 and -350 bp. In another ethylene responsive promoter, they were shown to be active 9 when placed at -500 bp (Itzhaki et al., 1994). Here we used the sequence of the Shiraz ufgt 10 promoter, which has 97 to 99 % homology to the grapevine ufgt promoter sequences isolated 11 from the Kyoho, Italia, Ruby Oku, Muscat of Alexandria and Flame Muscat cultivars 12 (KOBAYASHI et al., 2001), and 95 % to Cabernet Sauvignon (GenBank AY919624). The 13 alignment of various ufgt promoter sequences has already been showed (KOBAYASHI et al., 14 2001).

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So we conclude that the stimulation of *ufgt* expression by ethylene is independent from *mybA* expression. But it does not preclude the importance of MybA in controlling the *ufgt* expression. Indeed, Dr Goto-Yamamoto found that the MybA protein binds to a region of the *ufgt* promoter around -428 to -303 bp from the start codon (pers. comm.) and recent works confirm the role of MybA in stimulating anthocyanin accumulation (Yakushiji et al., 2006; Walker et al., 2007).

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To our knowledge this is the first report addressing the responsiveness of *myb* expression to an ethylene signal.

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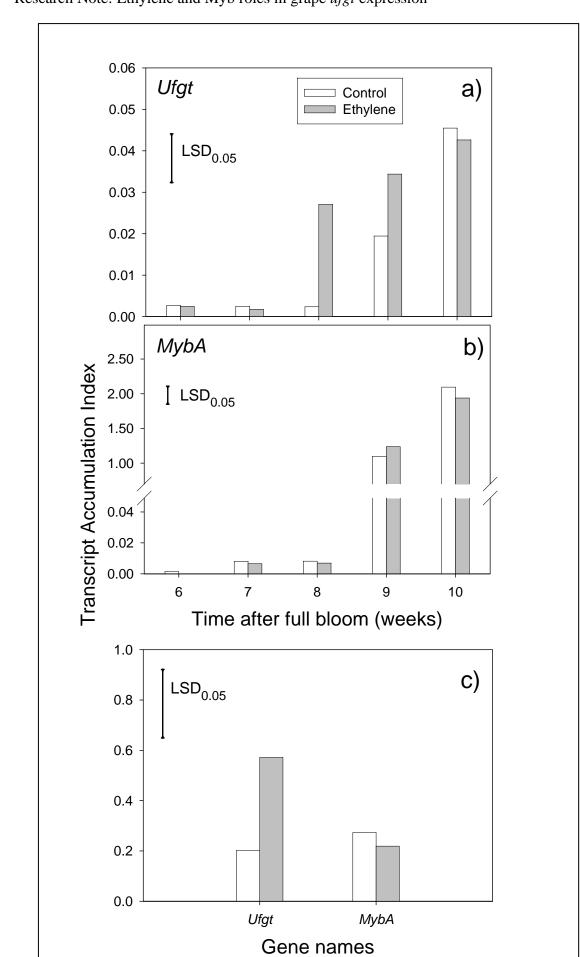
- 1 (Régie Agricole de la Ville de Toulouse) for the grapes and the team of CRGS platform of
- 2 Genopole Toulouse where the qRT-PCR analyses were performed.

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1 Hamdi, S.; 2005: Isogene specific oligo arrays reveal multifaceted changes in gene 2 expression during grape berry (Vitis vinifera, L.) development. Planta 222, 832–847. 3 Triantaphylidès, C.; Nespoulous, L.; Chervin, C.; 1993: Ammonium requirement for radiation-4 induced accumulation of polyamines in suspension cultured grape cells. Physiol. Plant. 5 **87**, 389-395. 6 Walker A.R.; Lee, E.; Bogs, J.; McDavid, D. A. J.; Thomas, M. R.; Robinson, S. P.; 2007. 7 White grapes arose through the mutation of two similar and adjacent regulatory genes. 8 Plant J. **49**, 772–785. 9 Yakushiji, H.; Kobayashi, S.; Goto-Yamamoto, N.; Tae Jeong, S.; Sueta, T.; Mitani, N.; Azuma, A.; 2006: A skin color mutation of grapevine, from black-skinned Pinot Noir 10 11 to white-skinned Pinot Blanc, is caused by deletion of the functional VvmybA1 allele. 12 Biosci. Biotech. Biochem. 70, 1506-1508. 13 14 **Figure captions** 15 Figure 1: Effects of exogenous ethylene to a) the ufgt mRNA levels, to b) the mybA1 mRNA 16 levels in grape berries, cv. Cabernet Sauvignon according to time after full bloom, and to c) 17 mRNAs of both genes in grape cell suspensions, cv. Gamay. The mRNA levels were 18 determined by qRT- PCR analyses and are expressed as the transcript accumulation index 19 relative to  $efl-\alpha$ . Each bar represents the mean of three biological replicates, and LSD bars 20 were calculated at the 0.05 level for differences between the treatments (control or ethylene). 21 22 Figure 2: Locations of ethylene responsive elements (cis-acting) on the promoter of mybA1 23 (AB242302) and on the promoter of ufgt (AY955269) of Vitis vinifera. The nucleotide 24 numbering is relative to the start codon (0). Cis-elements were estimated by homology 25 search using PLACE and PlantCARE databases.

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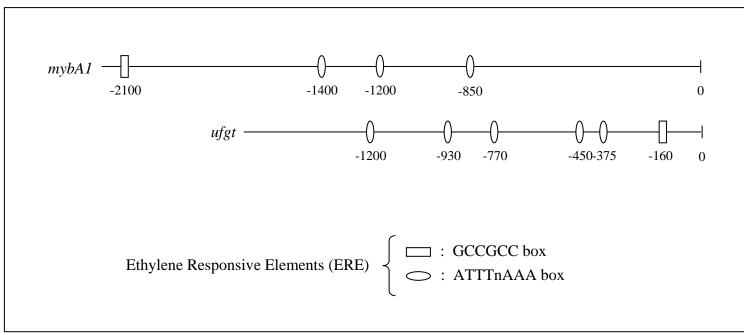


Figure 2