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Description of two new species of Hemilophini (Coleoptera, Cerambycidae, Lamiinae)

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Keywords:

Coleoptera; *Tyrinthia*;
Cerambycidae; *Eranina*;
Lamiinae; taxonomy;
longhorned beetles; Panama;
Hemilophini; Guyane française.

Abstract. – Two new species are described : *Tyrinthia lidiae* sp. nov., from Panama; and *Eranina bezarki* sp. nov., from French Guiana. A key to species of *Tyrinthia* present in Panama, and a key to species of *Eranina* present in French Guiana are provided.

Devesa S. & Santos-Silva A., 2021. – Description of two new species of Hemilophini (Coleoptera, Cerambycidae, Lamiinae). *Faunitaxys*, 9(1): 1–6.

ZooBank: <http://zoobank.org/C813F3F7-6F7E-4FC5-B625-A1EE1B652985>

Introduction

The tribe Hemilophini Thomson, 1868 includes 132 genera (and one additional fossil genus) occurring only on the American continent (Tavakilian & Chevillotte 2020). This tribe was extensively studied by the late Ubirajara Ribeiro Martins de Souza, who described many genera and species with Maria Helena M. Galileo (currently retired). The South American genera and species were revised by them in two volumes of a series of books (Martins & Galileo 2014a, b).

Tyrinthia Bates, 1866 has 19 known species distributed from Honduras to southern Brazil. Of these, five are known in Panama. Currently, *Eranina* Monné, 2005 (replacement name for *Erana* Bates, 1866) includes 37 species distributed from Mexico to Argentina, of which only two are known in French Guiana.

Material and Methods

General observations and measurements (mm) were made in Spain, using an ocular micrometer adapted to an Olympus SZX7 0.8–5.6X stereomicroscope; photographs were taken with a Canon EOS 5D Mark III DSLR camera equipped with a Canon MP-E 65mm f/2.8 1–5X macro lens, controlled by Cognisys Stackshot; the photographs were stacked using Zerene Stacker AutoMontage software and processed with Aperture software. Photographs of *Tyrinthia picticornis* Martins & Galileo, 1991, and *Eranina meyeri* (Martins & Galileo, 1989) were taken in the MZSP (see below) with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65mm f/2.8 1–5X macro lens, controlled by Zerene Stacker AutoMontage software.

The acronyms used in the text are as follow:

– **MHNUSC**: Museo de Historia Natural de la Universidad de Santiago de Compostela, Spain

– **MZSP**: Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil

– **SDPC**: Sergio Devesa Private Collection, Pontevedra, Spain

Results

Tyrinthia lidiae sp. nov.

(Fig. 1–5)

ZooBank: <http://zoobank.org/DECD2E71-6BE2-4BCB-A3D3-726C82C31846>

Holotype, ♂: PANAMA, *Bocas del Toro*: 15 km SSW Changuinola (9°21'N/82°36'W), 300 m, 30.III-04.IV.2001, J. Prena leg. (MHNUSC, temporarily in SDPC).

Description of the holotype.

Coloration. – Integument mostly light yellowish brown; antennal tubercles and area surrounding upper eye lobes dark brown, almost black; mandibles black on apical half; scape dark brown; pedicel slightly brownish, with apex yellowish; antennomere III with narrow orangish-brown basal ring, narrow reddish-brown apical ring, and blackish on remaining surface; antennomeres IV–VIII with narrow yellowish-white basal ring, narrow light reddish-brown apical ring, and blackish on remaining surface; antennomeres IX–XI dark brown, almost black (base with almost indistinct, narrow yellowish-white ring). Coxae and trochanters partially pale yellow; tibiae dark reddish brown basally, dark brown on remaining surface; tarsi dark brown, except reddish-brown apex of tarsomere V and claws. Abdomen dark brown.

Head. – Frons finely, sparsely punctate; with moderately abundant light yellowish-brown pubescence not obscuring integument. Vertex and area behind upper eye lobes finely, sparsely punctate; posterior area of antennal tubercles and dark area surrounding upper eye lobes with dense dark brown pubescence; remaining surface of vertex and area behind upper eye lobes with abundant yellowish-brown pubescence partially obscuring integument, except glabrous narrow area along median groove; dark area with long, erect, sparse dark brown setae. Area behind lower eye lobes and genae with abundant yellowish-brown pubescence partially obscuring integument, except glabrous genal apex. Wide central area of postclypeus with pubescence as on frons, and long, erect, sparse setae of same color interspersed; sides glabrous. Labrum with long, moderately sparse, almost golden setae directed forward on posterior half (Fig. 4). Distance between upper eye lobes 0.17 times length of scape (0.25 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.50 times length of scape (0.75 times distance between outer margins of eyes). Antennae 1.54 times elytral length, reaching elytral apex at the end of antennomere IX. Scape with abundant, bristly dark brown pubescence, and long, erect, sparse setae of same color interspersed (erect setae more abundant ventrally); pedicel with sparse yellowish-brown pubescence, short, erect, sparse dark setae interspersed dorsally, and long, erect, sparse dark setae ventrally;

antennomeres III and IV with abundant, bristly dark pubescence dorsally laterally, and long, erect, abundant dark setae ventrally from near base to near apex, except narrow light basal area with sparse yellowish pubescence, and narrow apex almost glabrous (Fig. 1); remaining antennomeres with dark brown pubescence, moderately bristly dorsally, and long, erect, sparse dark setae ventrally (setae gradually shorter to XI), except narrow light basal area with sparse yellowish pubescence.

Antennal formula based on antennomere III:

– Scape = 0.73. – Pedicel = 0.10. – IV = 0.77. – V = 0.42. – VI = 0.35. – VII = 0.33. – VIII = 0.28. – IX = 0.27. – X = 0.24. – XI = 0.28.

Thorax. – Prothorax transverse; sides slightly, uniformly rounded. Pronotum coarsely, moderately sparsely punctate; with bristly yellowish-brown pubescence, nearly absent centrally, distinctly denser on longitudinal pubescent band on each side, from base to apex; with long, erect, sparse setae of same color interspersed. Sides of prothorax coarsely, moderately abundantly punctate; with yellowish-brown pubescence not obscuring integument, and long, erect, sparse setae of same color interspersed. Prosternum with yellowish-brown pubescence laterally, almost glabrous centrally. Narrowest area of prosternal process 0.08 times width of procoxal cavity. Ventral surface of meso- and metathorax with yellowish-brown pubescence, distinctly denser laterally. Mesoventral process with sides somewhat concave on anterior 2/3, triangular-shaped on posterior third (Fig. 5). Scutellum with bristly yellowish-brown pubescence not obscuring integument, and long, erect, sparse setae of same color interspersed.

Elytra. – Coarsely, abundantly punctate (punctures forming longitudinal rows); with yellowish-brown pubescence not obscuring integument, and long, erect, abundant setae of same color interspersed; apex subrounded.

Legs. – Femora with short, abundant yellowish-brown pubescence not obscuring integument, and a few long, erect setae of same color and a few dark brown interspersed. Tibiae mostly with dark yellowish-brown pubescence, and long, erect, moderately abundant yellowish-brown and dark brown setae interspersed. Inner teeth of tarsal claws slightly shorter than outer teeth.

Abdomen. – Subcylindrical, narrowed down to segment III and distinctly widened from fourth segment; with brownish pubescence not obscuring integument, and long, erect, sparse setae of same color interspersed (Fig. 2).

Dimensions (mm), holotype male.

- Total length, 6.5;
- Prothoracic length, 0.9;
- Anterior prothoracic width, 1.1;
- Posterior prothoracic width, 1.1;
- Maximum prothoracic width, 1.2;
- Humeral width, 1.5;
- Elytral length, 4.8.

Etymology. – The new species is named in honor of the wife of the first author, Lidia Soutullo.

Remarks. – *Tyrinthia lidiae* sp. nov. is similar to *T. picticornis* Martins & Galileo, 1991, but differs as follows:

In *Tyrinthia lidiae* sp. nov.:

- Antennomere IV without central yellowish-brown ring;
- Antennomere IV with long, erect and abundant dark setae ventrally, from near base to near apex;
- Ventral surface of metathorax yellowish-brown;
- Mesoventral process with sides somewhat concave on anterior 2/3, triangularly shaped on posterior third (Fig. 5);
- Abdomen cylindrical, gradually widened from fourth segment (Fig. 2).

In *T. picticornis* Martins & Galileo, 1991:

- Antennomere IV has a distinct yellowish-brown ring just after middle and the erect dark setae ventrally are distinctly shorter from middle to apex (Figs. 7–8);
- Ventral surface of metathorax dark brown;
- Mesoventral process uniformly narrowed from base to apex (Fig. 6);
- Abdomen not cylindrical and not gradually widened from toward apex (Fig. 11).

***Eranina bezarki* sp. nov.**

(Fig. 12–16)

ZooBank : <http://zoobank.org/D178B68B-6850-4D13-A000-5B5B3F62D9A0>

Holotype, ♂: FRENCH GUIANA: Kaw Mountain, pk 38, 200 msnm, 22.IX.2009, E. Navarro leg. (MHNUSC, temporary in SDPC).

Description of the holotype.

Coloration. – Head mostly light orangish brown, with wide black macula close to posterior margin of eyes; apex of mandibles black; scape dark brown on dorsal, outer side, and apex, reddish brown on remaining surface; antennomeres dark brown, with some areas slightly dark reddish brown on ventral surface of basal segments. Prothorax mostly light orangish brown, with large semicircular black macula on posterior half of pronotum. Ventral surface of mesothorax mostly dark brown, with some reddish-brown areas on mesoventrite, mesanepisternum, and mesepimeron, except yellowish-brown distal half of mesoventral process. Metanepisternum light orangish brown; metaventrite mostly dark brown with some areas slightly dark reddish brown. Elytra black on humeri and area around scutellum; remaining basal area dark brown, almost black on posterior quarter, yellowish brown on wide central area. Scutellum black. Coxae and femora orangish; tibiae orangish with

Key to species of *Tyrinthia* Bates, 1866 present in Panama

- | | | |
|-------|--|---|
| 1. | Central area of the pronotum at least partially black | 2 |
| – | Central area of the pronotum without black area | 5 |
| 2(1). | Black area of the central area of the pronotum triangularly expanded posteriorly. Panama | <i>T. photurina</i> Bates, 1885 |
| – | Black area of the central area of the pronotum, when present on posterior half, not triangularly expanded | 3 |
| 3(2). | Antennomere IV with sparse long setae ventrally, distinctly contrasting with dense setae on ventral surface of III. Costa Rica, Panama | <i>T. biformis</i> Bates, 1885 |
| – | Antennomere IV with abundant long setae ventrally on anterior half (density of the setae similar to that in antennomere III) | 4 |
| 4(3). | Frons with distinct projections in male; black central area of the pronotum not narrowed toward posterior region. Panama, Trinidad & Tobago, Brazil (Amapá, Amazonas, Pará, Rondônia, Maranhão), French Guiana | <i>T. scissifrons</i> Bates, 1866 |
| – | Frons without projections in male; black central area of the pronotum narrowed toward posterior region. Nicaragua, Costa Rica, Panama | <i>T. xanthe</i> Bates, 1881 |
| 5(1). | Antennomere IV with yellowish central ring, and long and erect setae on ventral surface only reaching about middle; mesoventral process gradually narrowed from base. Panama | <i>T. picticornis</i> Martins & Galileo, 1991 |
| – | Antennomere IV without yellowish central ring, and long and erect setae on ventral surface almost reaching apex; mesoventral process expanded posteriorly. Panama | <i>T. lidiae</i> sp. nov. |



Figures 1–5) *Tyrinthia lidiae* sp. nov., holotype, ♂.

1) Dorsal habitus; 2) Ventral habitus; 3) Lateral habitus; 4) Head, frontal view; 5) Mesoventral process.

Figures 6–11) *Tyrinthia picticornis* Martins & Galileo, 1991.

6) Mesoventral process, paratype, ♂; 7) Antennomeres III–IV, holotype, ♂; 8) Antennomeres III–IV, paratype, ♂; 9) Dorsal habitus, holotype, ♂; 10) Dorsal habitus, paratype, ♂; 11) Ventral habitus, Paratype, ♂. [Figures 7 and 9 by Eugenio H. Nearn (from Lingafelter *et al.* 2020)].

brownish apical area. Tarsomere I reddish brown with apex darkened (reddish-brown area lighter toward metatarsi); tarsomere II brownish; tarsomere III reddish brown, darker on metatarsi; tarsomeres IV–V brownish, darker on apex of V. Abdominal ventrite I dark brown; II dark brown with large orangish-brown macula on each side of posterior half (not reaching apex); III dark brown basally, apically, and laterally, orangish-brown on remaining surface; IV with dark brown macula on each side of basal 2/3, brownish at apex, orangish brown on remaining surface; V almost entirely orangish brown.

Head. – Frons finely, shallowly, moderately abundantly punctate; with abundant yellowish-white pubescence not obscuring integument; with a few long, erect brownish setae close to eyes, and a few long, erect yellowish-white setae close to genae. Vertex and light area behind eyes with abundant light yellowish-brown pubescence obscuring integument, except glabrous area along median groove; black area behind eyes with almost black pubescence obscuring integument; with long, erect, sparse brownish setae close to eyes. Area behind lower eye lobes with dark pubescence not obscuring integument on black region, with yellowish-brown pubescence not obscuring integument on light area. Genae with abundant yellowish-white pubescence not obscuring integument, except glabrous apex, and a few long, erect setae of same color interspersed. Postclypeus with pubescence as on frons, and long, erect, sparse yellowish-white setae interspersed on wide central area. Labrum with abundant, long, erect, moderately thick, almost golden setae directed forward (Fig. 15). Distance between upper eye lobes 0.25 times length of scape (0.22 times distance between outer margins of eyes); in frontal view, distance between lower eye lobes 0.64 times length of scape (0.56 times distance between outer margins of eyes). Antennae 1.65 times elytral length, reaching elytral apex at middle of antennomere IX. Scape cylindrical on basal third, moderately widened on posterior 2/3 (sub pedunculate-clavate); with yellowish-white pubescence not obscuring integument, and long, erect, sparse setae of same color interspersed (erect setae longer ventrally) (Fig. 15). Pedicel, antennomere III and basal third of IV with yellowish-white pubescence not obscuring integument; posterior 2/3 of IV and remaining antennomeres with minute yellowish-white pubescence not obscuring integument, appearing to be darker due to the integument color, especially on distal segments; pedicel and antennomeres III–VIII with long, erect dark brown setae ventrally, more abundant on III, gradually sparser toward VIII; dorsal surface of antennomeres with moderately long, erect, sparse yellowish-white setae (setae distinctly shorter than on ventral surface, gradually shorter and sparser toward apex of antennae).

Antennal formula based on antennomere III:

– Scape = 0.73. – Pedicel = 0.11. – IV = 0.51. – V = 0.32. – VI = 0.30. – VII = 0.27. – VIII = 0.26. – IX = 0.22. – X = 0.20. – XI = 0.23.

Thorax. – Prothorax transverse; sides roundly expanded centrally. Pronotum coarsely, moderately sparsely punctate; with abundant yellowish-brown pubescence on light area, partially obscuring integument, abundant, almost black pubescence on black area. Sides of prothorax coarsely, moderately abundantly punctate (punctures coarser, more distinct and abundant posteriorly); with abundant yellowish-brown pubescence not obscuring integument, gradually becoming yellowish-white toward prosternum (Fig. 12). Prosternum moderately finely, sparsely punctate on posterior 2/3, transversely sulcate, nearly smooth on anterior third; with yellowish-white pubescence not obscuring integument, distinctly sparser on center of anterior third. Narrowest area of prosternal process 0.06 times width of procoxal cavity. Ventral surface of meso- and metathorax with abundant yellowish-white pubescence not obscuring integument, sparser on center of mesoventrite, absent on center of metaventrite, yellower on some areas, and

long, erect setae of same color interspersed on metaventrite (Fig. 13). Scutellum with yellowish-brown pubescence not obscuring integument, more abundant laterally, especially on apex.

Elytra. – Coarsely, abundantly punctate except posterior quarter with punctures sparser and slightly distinct; with yellowish-brown pubescence not obscuring integument on anterior 3/4 (pubescence appearing to be darker on dark area due to the integument color), and dark pubescence not obscuring integument on posterior quarter; with long, erect brownish setae throughout, darker, more abundant on sides of posterior quarter.

Legs. – Pro- and mesofemora subfusiform; metafemora strongly narrowed ventrally on posterior third (Fig. 16); inferior margins of mesofemora somewhat crenulate; femora with abundant yellowish-white pubescence not obscuring integument dorsally and laterally, glabrous ventrally, and long, erect, moderately abundant setae of same color interspersed on sides. Tibiae with long, decumbent, moderately abundant yellowish-white setae not obscuring integument, gradually more yellowish-brown toward apex; protibiae with long, erect brownish setae dorsally (Fig. 15); meso- and metatibiae with long, erect yellowish-white setae dorsally, gradually brownish toward apex. Metatarsomere I slightly longer than II–III together.

Abdomen. – Ventrites with abundant yellowish-white pubescence not obscuring integument, and long, erect, sparse setae of same color interspersed; apical margin of ventrite V concave.

Dimensions (mm).

- Total length, 6.7;
- Prothoracic length, 1.3;
- Anterior prothoracic width, 1.4;
- Posterior prothoracic width, 1.4;
- Maximum prothoracic width, 1.5;
- Humeral width, 1.9;
- Elytral length, 4.6.

Etymology. – The new species is named in honor of our friend Larry G. Bezark (California, USA), for his friendship and constant help.

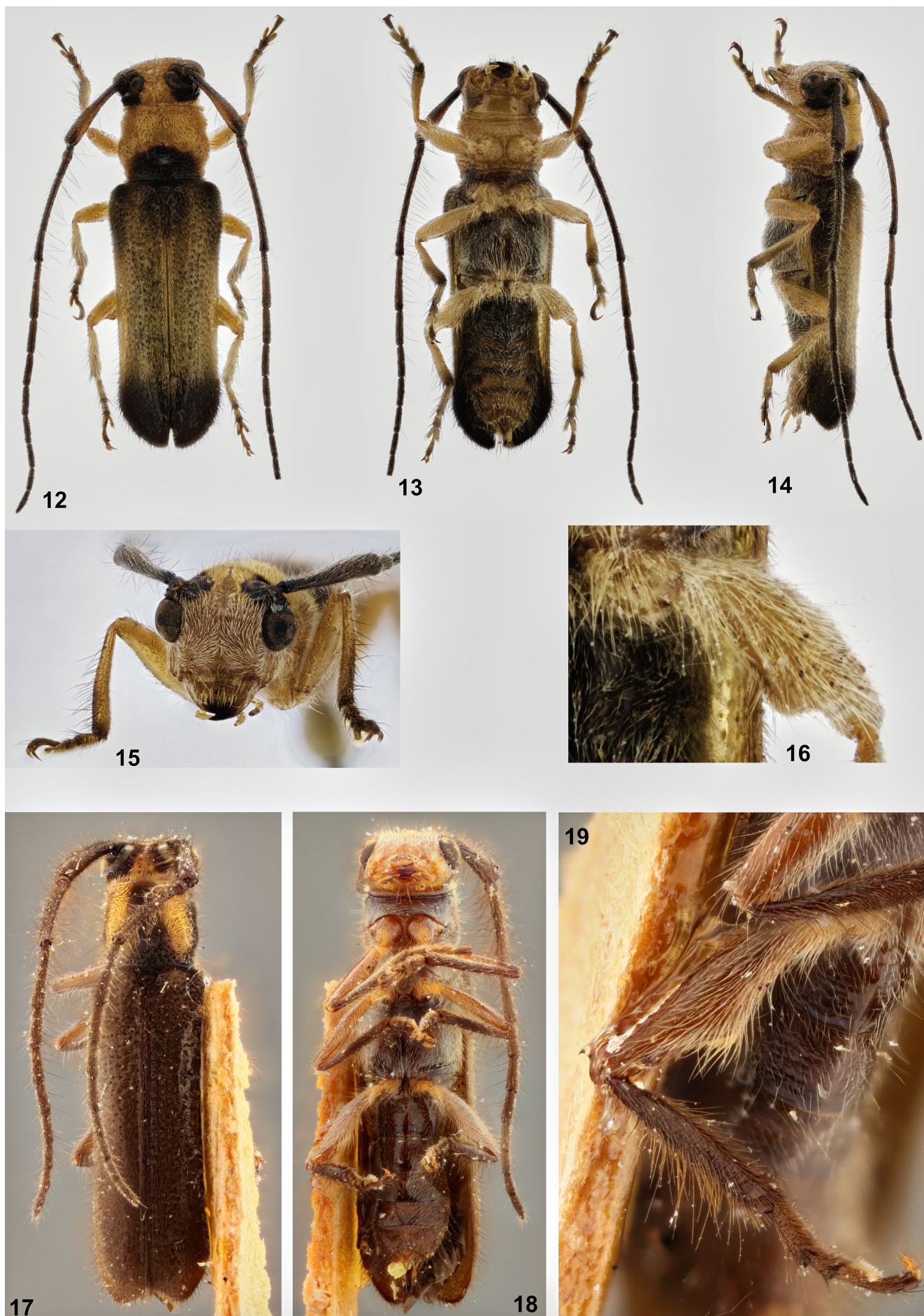
Remarks. – In *Eranina* Monné, 2005 (and nearly all Hemilophini) the metafemora in males are sublinear (Martins & Galileo 2014a). The only known exception is the male of *Eranina meyeri* (Martins & Galileo, 1989), which has the metafemora with a distinct subtriangular projection ventrally (Fig. 19).

However, the males of the following species are unknown:

- *E. argentina* (Bruch, 1911);
- *E. diana* (Martins & Galileo, 1989);
- *E. nigrita* (Galileo & Martins, 1991);
- *E. pallida* Galileo & Martins, 2013;
- *E. pallidula* (Martins & Galileo, 1989);
- *E. piterpe* Galileo & Martins, 2007;
- *E. porangaba* (Galileo & Martins, 1998);
- *E. suavissima* (Bates, 1881);
- *E. tauaira* (Martins & Galileo, 1993);
- *E. univittata* (Bates, 1881).

Key to species of *Eranina* Monné, 2005 present in French Guiana

1. Antennomeres lacking distinct light areas; elytra with wide central area yellowish brown; metafemora in male strongly and abruptly narrowed posteriorly. French Guiana ***E. bezarki* sp. nov.**
- Antennomeres with distinct light areas (some antennomeres may be entirely yellowish or whitish); elytra without wide central area yellowish brown; metafemora in male not strongly and abruptly narrowed posteriorly **2**
- 2(1). Pronotum entirely orangish. French Guiana, Brazil (Mato Grosso) ***E. cendira*** (Martins & Galileo, 1993)
- Pronotum with wide black central area. French Guiana, Brazil (Amazonas) ***E. cincticornis*** (Bates, 1866)



Figures 12–16) *Eranina bezarki* sp. nov., holotype, ♂.

12) Dorsal habitus; 13) Ventral habitus; 14) Lateral habitus; 15) Head, frontal view; 16) Metafemora.

Figures 17–19) *Eranina meyeri*, holotype, ♂.

17) Dorsal habitus; 18) Ventral habitus; 19) Metafemora.

Additionally, the sex of the holotype of *E. humeralis* (Martins & Galileo, 1989), the only known specimen, is unknown. The new species described here is the second species of the genus with the metafemora distinctly modified.

Eranina bezarki sp. nov. differs from *E. meyeri* (Figs. 17–19) as follows:

In *Eranina bezarki* sp. nov.:

- Pronotum with a semicircular black macula on the posterior half;
- Elytra bicolorous;
- Metafemora gradually widened ventrally from base to posterior third, then somewhat abruptly narrowed.

In *E. meyeri* (Martins & Galileo, 1989):

- Pronotum with a longitudinal central black macula, from base to apex;
- Elytra unicolorous;
- Ventral surface of the metafemora with a subtriangular projection.

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Résumé

Devesa S. & Santos-Silva A., 2021. – Description de deux nouveaux Hemilophini (Coleoptera, Cerambycidae, Lamiinae). *Faunitaxys*, 9(1): 1 – 6.

Deux nouvelles espèces sont décrites : *Tyrinthia lidiae* sp. nov. du Panama et *Eranina bezarki* sp. nov. de Guyane française. Une clé des *Tyrinthia* du Panama ainsi qu'une clé des *Eranina* de Guyane française sont proposées.

Mots clés. – Coleoptera, Cerambycidae, Lamiinae, longicornes, Hemilophini, *Tyrinthia*, *Eranina*, taxonomie, Panama, Guyane française.

References

- Lingafelter S.W., Monné M.A. & Nearn E.H., 2020. – Online Image Database of Cerambycoid Primary Types of the Smithsonian Institution. Available from: <http://smithsoniancerambycidae.com/> (Last accessed on 21 November 2020.)
- Martins U.R. & Galileo M.H.M., 2014a. – Subfamilia Lamiinae. Hemilophini Thomson, 1868. Parte I. In: Martins, U.R. (org.), *Cerambycidae Sul-Americanos (Coleoptera). Taxonomia*. Vol. 13. Sociedade Brasileira de Entomologia, Curitiba, 231 pp.
- Martins U.R. & Galileo M.H.M., 2014b. – Subfamilia Lamiinae. Hemilophini Thomson, 1868. Parte II. In: Martins, U.R. (org.), *Cerambycidae Sul-Americanos (Coleoptera). Taxonomia*. Vol. 14. Sociedade Brasileira de Entomologia, Curitiba, 296 pp.
- Tavakilian G.L. & Chevillotte H., 2020. – Titan: base de données internationales sur les Cerambycidae ou Longicornes. Version 3.0. Available from: <http://titan.gbif.fr/> (Last accessed 28 November 2020.)

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Derniers articles publiés

- Delaunay L. & Melle A., 2013. – Descriptions de six nouvelles *Meira* J. du Val, 1852, des Cévennes (France méridionale) et note sur *Meira sedilloti* Ch. Brisout, 1882, bona species (Coleoptera Curculionidae Entiminae). *Faunitaxys*, 1(1) : 1 – 22.
- Robiche G., 2014. – Les coléoptères Tenebrionidae du Parc National Dzanga-N’Doki, République centrafricaine (Projet Sangha). 1^{ère} partie. *Faunitaxys*, 2(1) : 1 – 16.
- Limbourg P. & Coache A., 2014. – Liste commentée et descriptions de deux nouveaux Rutelinae de la République du Bénin (Coleoptera, Scarabaeoidea, Melolonthidae). 1^{ère} partie. *Faunitaxys*, 2(2) : 1 – 20.
- Delaunay L., Coache A. & Rainon B., 2015. – Contribution à la connaissance de la biodiversité entomologique africaine. I. – *Pericordus goergeni* n. sp. du Bénin (Coleoptera, Brentidae). *Faunitaxys*, 3(1) : 1 – 4.
- Lassalle B. & Roux P., 2015. – Description de cinq nouveaux Trigonotomi (Coleoptera, Pterostichidae). *Faunitaxys*, 3(2) : 1 – 8.
- Bosuang S., Audibert C. & Porion T., 2015. – Two new *Polydictya* from Borneo (Hemiptera: Fulgoromorpha: Fulgoridae). *Faunitaxys*, 3(3) : 1 – 4.
- Lassalle B., 2015. – Deux nouveaux carabiques d’Asie (Coleoptera, Carabidae, Chlaeniini). *Faunitaxys*, 3(4) : 1 – 5.
- Lassalle B., 2015. – Premiers éléments pour un inventaire des Cicindèles de la République du Tchad (Coleoptera Cicindelidae). *Faunitaxys*, 3(5) : 1 – 12.
- Audibert C. & Porion T., 2016. – Une remarquable nouvelle espèce du Guatemala : *Enhydria monzoni* n. sp. (Hemiptera : Fulgoromorpha : Fulgoridae). *Faunitaxys*, 4(1) : 1 – 5.
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- Porion T., Audibert C. & Nagai S., 2016. – Notes sur le groupe de *Scamandra sanguiflua* (Stål, 1863) avec la description d’une nouvelle espèce de Sulawesi : *Scamandra marcellae* n. sp. (Hemiptera, Fulgoromorpha, Fulgoridae). *Faunitaxys*, 4(3) : 1 – 9.
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- Audibert C., Porion T. & Nagai S., 2016. – Une deuxième espèce pour le genre *Hariola* Stål, 1863 : *H. claryi* n. sp., de Papouasie (Hemiptera, Fulgoromorpha, Fulgoridae). *Faunitaxys*, 4(5) : 1 – 5.
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- Delahaye N. & Camiade D., 2016. – Un Acanthophorini nouveau de São Tomé (Coleoptera, Cerambycidae, Prioninae). *Faunitaxys*, 4(10) : 1 – 4.
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- Lacroix M. & Coache A., 2017. – *Paraholotrichia rainoni* n. g., n. sp., un nouveau Rhizotrogini d’Afrique (Coleoptera, Scarabaeoidea, Melolonthidae). *Faunitaxys*, 5(4) : 1 – 4.
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- Vienna P., Gomy Y. & Coache A., 2017. – Contributo alla conoscenza degli Histeridae del Senegal (Insecta: Coleoptera) (II). *Faunitaxys*, 5(6) : 1 – 25.
- Cumming R. T. & Le Tirant S., 2018. – *Phyllium shurei* n. sp., a third species of leaf insect from Java, Indonesia (Phasmida, Phylliidae). *Faunitaxys*, 6(1) : 1 – 5.
- Cumming R. T., Le Tirant S. & Teemsma S. N., 2018. – On the *Phyllium* of Peninsular Malaysia and Sumatra, Indonesia, with range expansions for currently known species, description of the previously unknown *Phyllium* (*Pu.*) *abulfatahi* Seow-Choen female, and description of the new species *Phyllium* (*Ph.*) *rubrum* n. sp. from Peninsular Malaysia (Phasmida: Phylliidae). *Faunitaxys*, 6(2) : 1 – 21.
- Audibert C. & Porion T., 2018. – Note sur quelques *Scamandra* de Sulawesi, avec la description de deux nouvelles espèces : *Scamandra multimaculata* n. sp. et *Scamandra silighinii* n. sp. (Hemiptera, Fulgoromorpha, Fulgoridae). *Faunitaxys*, 6(3) : 1 – 5.
- Lassalle B., Schnell R. & Bendanillo F., 2018. – Note sur les Trigonotomi de Cebu (Philippines) et description d’une espèce nouvelle (Coleoptera, Pterostichidae, Trigonotomi). *Faunitaxys*, 6(4) : 1 – 4.

- Robiche G. & Coache A., 2018. – Découverte du mâle de *Cossyphus (Acontodactylus) longepilis* Ardoin, 1960 (Coleoptera, Tenebrionidae, Lagriinae). *Faunitaxys*, 6(5) : 1 – 4.
- Vives E., 2018. – Descripción de una nueva especie del género *Pelossus* Thomson (Coleoptera, Cerambycidae), procedente del Norte de Filipinas. (Cerambycidos nuevos o interesantes de Filipinas. Pars XVI). *Faunitaxys*, 6(6) : 1 – 3.
- Oremans P., 2018. – Contribution à l'étude des *Euphaedra* de Guinée et description d'une nouvelle espèce (Lepidoptera, Nymphalidae). *Faunitaxys*, 6(7) : 1 – 3.
- Degallier N. & Gomy Y., 2018. – Contribution à la connaissance des escarbots afro-tropicaux du genre *Chaetabraeus* Portevin, 1929 (Insecta, Coleoptera, Histeridae, Abraeinae). *Faunitaxys*, 6(8) : 1 – 40.
- Cumming R. T., Le Tirant S. & Teemsma S. N., 2018. – Northeastern Australia record of *Nanophyllium pygmaeum* Redtenbacher, 1906, now recognized as a new species, *Nanophyllium australianum* n. sp. (Phasmida, Phylliidae). *Faunitaxys*, 6(9) : 1 – 5.
- Alonso C., 2018. – A new African *Potamogethes* Delève (Coleoptera, Elmidae). *Faunitaxys*, 6(10) : 1 – 8.
- Borowiec L. & Coache A., 2018. – *Aspidimorpha (Afroaspidimorpha) rainoni*, a new species from Benin (Coleoptera, Chrysomelidae, Cassidinae). *Faunitaxys*, 6(11) : 1 – 5.
- Opitz W., 2018. – Classification, natural history, and evolution of the Korynetinae (Coleoptera: Cleridae). Part IV. The new genus *Nolafigura* Opitz, and fourteen new species of *Korynetes* Herbst. *Faunitaxys*, 6(12) : 1 – 17.
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- Gomy Y., 2018. – Description de deux *Chaetabraeus* nouveaux du sous-genre *Mazureus* Gomy, 1991 de Côte d'Ivoire (Coleoptera, Histeridae, Abraeinae). *Faunitaxys*, 6(15) : 1 – 4.
- Degallier N. & Kovarik P. W., 2018. – Description de trois espèces nouvelles de *Chaetabraeus* (s. str.) associées aux latrines du Daman des rochers en Afrique du Sud et notes sur l'écologie des espèces du genre (Insecta, Coleoptera, Histeridae, Abraeinae). *Faunitaxys*, 6(16) : 1 – 9.
- Delaunay L., Coache A. & Rainon B., 2019. – Contribution à la connaissance de la biodiversité entomique africaine. II. – *Scopaeus lescuyeri* n. sp. de la République du Bénin (Coleoptera, Staphylinidae, Paederinae). *Faunitaxys*, 7(1) : 1 – 2.
- Gomy Y., 2019. – Description de quatre *Chaetabraeus* nouveaux du sous-genre *Mazureus* Gomy, 1991 d'Afrique tropicale et équatoriale (Coleoptera, Histeridae, Abraeinae). *Faunitaxys*, 7(2) : 1 – 8.
- Limoges R. & Le Tirant S., 2019. – Nouvelle espèce du genre *Eupholus* Boisduval, 1835 de la Papouasie occidentale, Indonésie (Coleoptera : Curculionidae : Entiminae). *Faunitaxys*, 7(3) : 1 – 5.
- Cumming R. T., Le Tirant S. & Hennemann F. H., 2019. – Review of the *Phyllium* Illiger, 1798 of Wallacea, with description of a new subspecies from Morotai Island (Phasmatodea: Phylliidae: Phylliinae). *Faunitaxys*, 7(4) : 1 – 25.
- Opitz W., 2019. – Classification, natural history, and evolution of the Korynetinae (Coleoptera: Cleridae). Part V. Taxonomy of the African genera *Avena* Opitz, *Dolichopsis* Gorham, *Notostenus* Spinola, and *Pectobullus* Opitz. *Faunitaxys*, 7(5) : 1 – 13.
- Le Tirant S. & Santos-Silva A., 2019. – New records of Neotropical Cerambycidae (Coleoptera). *Faunitaxys*, 7(6) : 1 – 8.
- Vives E., 2019. – Una nueva especie del género *Parastrangalis* Ganglbauer, de Vietnam (Coleoptera, Cerambycidae). (Cerambycidos nuevos o interesantes de Vietnam. Pars IX). *Faunitaxys*, 7(7) : 1 – 2.
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- Delahaye N., 2019. – Un nouvel Anacolini de Colombie (Coleoptera, Cerambycidae, Prioninae). *Faunitaxys*, 7(9) : 1 – 2.
- Vives E., 2019. – Descripción de un nuevo género de Oemini de Vietnam (Coleoptera, Cerambycidae). (Cerambycidos nuevos o interesantes de Vietnam. Pars X). *Faunitaxys*, 7(10) : 1 – 2.
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- Porion T. & Audibert C., 2019. – Un nouveau Fulgoridae d'Indonésie : *Scamandra pocerattui* n. sp. (Hemiptera : Fulgoromorpha). *Faunitaxys*, 7(12) : 1 – 3.
- Cumming R. T., Le Tirant S. & Hennemann F. H., 2019. – A new leaf insect from Obi Island (Wallacea, Indonesia) and description of a new subgenus within *Phyllium* Illiger, 1798 (Phasmatodea: Phylliidae: Phylliinae). *Faunitaxys*, 7(13) : 1 – 9.
- Santos-Silva A. et al., 2019. – New synonymy and new records in South American Cerambycidae (Coleoptera). *Faunitaxys*, 7(14) : 1 – 11.
- Opitz W., 2019. – Classification, natural history, and evolution of the subfamily Peloniinae Opitz (Coleoptera: Cleroidea: Cleridae). Part XV. Taxonomic revision of the new world genus *Cregya* Leconte. *Faunitaxys*, 7(15) : 1 – 126.

- Gomy Y., 2019. – Sur la présence d'un Niponiinae à Madagascar : *Lemurinius sicardi* n. gen. n. sp. (Coleoptera, Histeridae) (Cinquième contribution à la connaissance des Histeridae de Madagascar). *Faunitaxys*, 7(16) : 1 – 5.
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- Sáfián Sz., Coache A. & Rainon B., 2020. – New data on the distribution of *Iridana agneshorvathae* Collins, Larsen & Sáfián, 2008 with description of the previously unknown female (Lepidoptera, Lycaenidae, Poritiinae). *Faunitaxys*, 8(3) : 1 – 3.
- Cumming R. T. & Le Tirant S., 2020. – A new species of *Phyllium* Illiger, 1798, from the *celebicum* species group native to Laos (Phasmida: Phylliidae). *Faunitaxys*, 8(4) : 1 – 9.
- Coache A. & Rainon B., 2020. – Les Hesperidae de la forêt de Pénélan (Pénéssoulou, Bénin), avec la liste des rhopalocères rencontrés (Lepidoptera, Papilionoidea, Hesperidae). *Faunitaxys*, 8(5) : 1 – 17.
- Cumming R. T., Baker E., Le Tirant S. & Marshall J. A., 2020. – On the *Phyllium* Illiger, 1798 of Palawan (Philippines), with description of a new species (Phasmida: Phylliidae). *Faunitaxys*, 8(6) : 1 – 9.
- Fleck G., 2020. – *Onychogomphus* (*Siriusonychogomphus*) *louissiriusi*, a new species and new subgenus from Thailand (Odonata: Anisoptera: Gomphidae). *Faunitaxys*, 8(7) : 1 – 9.
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