

# Factors associated with coinfections in invasive aspergillosis: a retrospective cohort study

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## 1 Factors associated with coinfections in invasive aspergillosis: a retrospective cohort study

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- 50 **Running head:** Coinfections in invasive aspergillosis

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- 52 Preliminary results have been presented as an oral communication at ECCMID 2019.
- Keywords: Invasive Fungal Infections; Coinfections; Fungal; Leukaemia; Mortality; Risk Factors

58 Abstract

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59 **Objectives:** To describe the coinfections in invasive aspergillosis, to identify factors associated with

coinfections and to evaluate the impact of coinfection on mortality.

Patients and methods: We conducted a monocentric retrospective study of consecutive putative,

62 probable, or proven invasive aspergillosis that occurred from 1997 to 2017. All coinfections, with

an onset within 7 days before or after the first sign of aspergillosis, were identified. Factors

associated with coinfections and mortality were analysed by multivariable analysis.

Results: Among the 690 patients with IA included in the study, median age was 57 years (range: 7

days-90 years). A coinfection was diagnosed in 272/690 (39.4%, 95%CI [35.8-43.2]) patients. The

location of this coinfection was pulmonary only in 131/272 patients (48%), bloodstream only in

66/272 patients (24%) and other/multiple sites in 75/272 patients (28%). Coinfection were

bacterial (n=110/272 patients, 40%), viral (n=58/272, 21%), fungal (n=57/272, 21%), parasitic

(n=5/272, 2%) or due to multiple types of pathogens (n=42/272, 15%). Factors associated with a

coinfection in adjusted analysis were: allogeneic haematopoietic stem cell transplantation (OR=2.3

[1.2-4.4]), other haematological malignancies (OR=2.1 [1.2-3.8]), other underlying diseases

(OR=4.3 [1.4-13.6); lymphopenia (OR=1.7 [1.1-2.5]); C-reactive protein >180 mg/l (OR=1.9 [1.2-

3.0]); fever (OR=2.4 [1.5-4.1]); tracheal intubation (OR=2.6 [1.5-4.7]); isolation of  $\geq 2$  different

Aspergillus species (OR=2.7 [1.1-6.3]); and presence of non-nodular lesions on chest computed

tomography (OR=2.2 [1.3-3.7] and OR=2.2 [1.2-4.0]). Coinfections were independently associated

with a higher mortality at week 12 (adjusted HR 1.5 [1.1-1.9], p<0.01).

**Conclusions**: Coinfections were frequent in IA patients and were associated with higher mortality.

# Introduction

Invasive aspergillosis (IA) affects mostly severely immunocompromised patients. Despite improvements in diagnosis and treatment, 12-week mortality exceeds 30% [1-6]. High rates of coinfections have been described in patients with IA in a few studies. Georgiadou *et al.* showed 49% of documented pulmonary coinfections in 126 patients with haematological malignancy (HM) and IA [7]. In another monocentric study in Israel, 21% of the HM patients had a polymicrobial pneumonia diagnosed on bronchoalveolar lavage fluid (BALF) [8]. *Aspergillus* was the most common pathogen identified (87%) and in-hospital mortality was higher in the coinfected group. Recently, IA occurring after influenza virus infections was shown to have a 50% mortality rate [9, 10]. Despite their high frequency, the risk factors for coinfections in IA and the impact on mortality have not yet been well studied. Our aim was to describe the coinfections associated with IA in a large cohort of patients with putative, probable or proven aspergillosis, to identify the factors associated with coinfections and to analyse the mortality.

## Methods

We conducted a retrospective study of consecutive IA cases that occurred in the University Hospital of Strasbourg from 1997 to 2017. The study is part of a retrospective, non-interventional study on opportunistic infections in immunosuppressed and frail patients. Institutional ethical committee has approved the study. According to French regulations, the database has been declared to the *Commission Nationale de l'Informatique et des Libertés*. The study has been registered in ClinicalTrials.gov (NCT03920735). All patients alive were asked to give their consent after receiving an information letter. In agreement with ethical committee recommendations, informed consent was not requested from the relatives of patients who had died. Patients who

had been enrolled in a prospective epidemiologic, diagnostic, or therapeutic clinical trial at time of their infection had all signed the appropriate informed consent form.

Suspected cases of IA were identified from the mycology laboratory database and from the hospital diagnosis database. Selection process is detailed in supplementary material. Patients included in a previous study on prognosis factors in IA are also part of this study [11]. Patients were included if they had been diagnosed with IA meeting the updated European Organization for Research and Treatment of Cancer/Mycosis Study Group (EORTC/MSG) criteria for proven or probable IA in cancer and severely immunosuppressed patients [12] or the clinical algorithm for putative IA in critically ill patients [13]. These criteria were modified by using a cut-off for galactomannan index of ≥0.5 in serum or BAL, based on manufacturer's recommendations (Bio-Rad, Marne-La-Coquette, France). Pulmonary IA were not included if no chest computed tomography (CT) was available.

Date of first clinical, radiological or mycological sign attributed to IA was defined as Day 0 (D0). We collected all variables available on D0 or at earliest time point around D0 with the exception of mycological data that where collected over the full course of the IA to allow a definite diagnosis of this infection. Two groups were considered and compared: IA without coinfection and IA with a coinfection. A coinfection was defined by the presence of clinical and/or radiological signs of infection and the identification of a relevant pathogen in significant amount, within 7 days before or after D0 of IA [7]. Details and definitions appear in supplementary material [6, 7, 12, 14-16].

Categorical variables are described with numbers and percentages. For the study of factors associated with coinfections, and as a patient could suffer several episodes of IA, comparisons between groups were made using a mixed logistic regression model including a random patient

effect. All variables with a p-value <0.2 in univariate analysis (Table 1) were included in a multivariable model. Continuous variables were dichotomized using clinically relevant thresholds. Results are presented as odds ratios (OR) with their 95% confidence intervals (95%CI) calculated using the Wald method. Multicollinearity was assessed using the variance inflation factor (VIF) ensuring that no factor had a VIF greater than 2. The number of events per variable was greater than 10. Goodness of fit of the multivariable logistic model was assessed using the Hosmer-Lemeshow test. The rate of missing data was low (6.5% for multivariable model) and no imputation method was used. The effect of coinfection on mortality was assessed using a multivariable Cox regression model including a random patient effect. The adjustment variables included in the multivariable model were those with a p-value <0.2 in univariate analysis. Proportional hazards assumption was assessed using Schoenfeld residuals. Results are presented as hazards ratios (HR). Confidence intervals were computed using a robust variance. A p-value <0.05 was considered statistically significant. All analyses were performed using R software version 3.6.0 (2019) (R Foundation for Statistical Computing, Vienna, Austria. URL https://www.Rproject.org/).

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## Results

## 1) Global population characteristics

Among 1075 suspicions of IA, we have identified 690 IA [549/690 (80%) probable, 29/690 (4%) putative, and 112/690 (16%) proven] in 661 patients (Figure 1). Median age was 57 years (range 7 days - 90 years) and 65% (n=451/690) of patients were men (Table 1). Primary underlying diseases and other predisposing factors are listed in Table 1.

Microscopy, culture, or histopathology was positive in 404/690 (59%) patients. Galactomannan detection test was performed in serum in 678/690 (98%) episodes and was positive in 395/678 (58%). BAL was performed in 449 (69%) of the 651 lower respiratory tract IA. Galactomannan detection test was performed in BALF in 396 episodes and was positive in 266/396 (67%) of them.

Overall BALF analysis (positive culture or galactomannan test) was the only mycological documentation of IA in 121 (19%) of the 651 lower respiratory tract IA.

Site of IA was lower respiratory tract only, another single organ and disseminated in respectively 586/690 (85%), 28/690 (4%) and 76/690 (11%) cases. Among the 651 lower respiratory tract IA, chest CT findings were nodule(s) only in 138/651 (21%) episodes, nodule(s) with other lesions in 329/651(51%) and other lesions without nodule in 174/651 (27%). Ten patients with endoscopy-confirmed bronchial or tracheal lesions without lung involvement had a normal CT.

# 2) Coinfections characteristics

A coinfection was diagnosed in 272/690 (39%) episodes (Table 2). The site of coinfection was pulmonary only in 131/272 patients (48%), bloodstream infection in 66/272 patients (24%) and other or multiple sites in 75/272 patients (28%). Coinfection with bacteria only was identified in 110/272 patients (40%), viruses in 58/272 (21%), fungi in 57/272 (21%) and parasites in 5/272 (2%). In 42/272 cases (15%), ≥2 different types of copathogens were involved. The descriptions of the pathogens involved in coinfections are presented in Table 2 and supplementary Table 2. The proportion of BAL performed in lower respiratory tract IA with or without coinfection was similar (72% (n=184/257) and 67% (n=265/394) respectively; p=0.26).

We stratified our data in two periods (<2009 and ≥2009), corresponding to the implementation of multiplex respiratory virus PCR. After 2009, 158 coinfections in 347 (46%) IA patients were

documented compared to 114 in 343 (33%) IA before 2009 (p<0.01) (Supplementary Table 1). Viral coinfections were more frequently diagnosed after 2009 (n=66/347; 19%) compared to before (n=22/343; 6%) (p<0.01) whereas no differences were observed for other type of pathogens. Due to these differences, the diagnosis period (<2009 versus  $\geq$ 2009) was entered in the multivariable analysis.

# 3) Factors associated with a coinfection

Univariable analysis results appear in Table 1. In multivariable analysis, factors associated with more coinfections were alloHSCT, other HM, the group of other underlying diseases, presence of non-nodular lesions on CT, low lymphocyte count, high C-reactive protein levels, fever, tracheal intubation and isolation of  $\geq 2$  different species of *Aspergillus*. (Table 3). Age >65 years was associated with a lower rate of coinfection.

# 4) Impact of coinfections on survival

Survival in the global population was 56% (n=306/690 deaths) at week 12 and 35% (n=446/690 deaths) at 1 year (Figure 1). Twelve-week survival rate was 46% (n=146/272 deaths) in patients with a coinfection compared to 62% (n=160/418 deaths) in patients without coinfection (Figure 2). In univariable analysis, coinfection with bacteria (HR 1.4, 95% CI [1.02–1.9], fungi (HR 1.8 [1.2–2.6], parasites (HR 5.8 [2.8–12.1]) or  $\geq$ 2 types of copathogens (HR 2.9 [2.0–4.2]), but not viral coinfection (HR 1.0 [0.7–1.6]), had higher risk of death compared to the absence of coinfection. Pulmonary (HR 1.4 [1.03–1.8]), bloodstream (HR 1.6 [1.1–2.3]) and other/multiple sites (HR 2.2 [1.6–3.0]) were also associated with an increased risk of death compared to no coinfection. In multivariable analysis, coinfection was independently associated with a higher twelve-week

mortality (adjusted HR 1.5 [1.1–1.9], p<0.01). Other factors associated with mortality are in Supplementary Table 3.

# Discussion

We described a large case series of putative, probable, or proven IA over 21 years in a tertiary care centre. We documented a high rate (39%) of coinfections with a broad spectrum of copathogens. Coinfections were associated with increased mortality at week 12. We identified factors associated with coinfections.

This study describes one of the largest monocentric cohort of IA ever published [17-19]. Our analysis provides real-life data in non-selected patients with mycological documentation of IA. Underlying conditions predisposing to IA cover a large spectrum of diseases, broader than usually reported in randomised clinical trials which focus more on specific subpopulations [1, 20-22]. As expected, HM and HSCT or SOT are predominant conditions predisposing to IA. However, we also identified cases in patients with non-classical risk factors such as systemic autoimmune or inflammatory diseases, liver failure, trauma, surgery, chronic pulmonary disease, aspiration pneumonitis, near-drowning, and prematurity. More than half of these latter patients have received steroids or other immunosuppressive drugs or were neutropenic. Distribution of underlying conditions are in accordance with the multicentre surveillance study conducted between 2005 and 2007 in France with, however, a higher proportion of SOT in our cohort due to the large SOT activity in Strasbourg [23].

We identified that 39% of IA cases in this global population had concomitant coinfections. BAL was performed in 69% of the patients with a lower respiratory tract involvement. This rate is high

compared to published data and facilitated the identification of pulmonary coinfections in our cohort [24, 25]. These data confirm the critical role of BAL in the investigation of pulmonary CT abnormalities in immunocompromised patients: it improved the accurate diagnosis rate of pulmonary IA with one in five cases diagnosed by BALF analysis only and identified copathogens.

Our results on the prevalence of coinfections are in concordance with those found in specific populations. Georgiadou *et al.* showed that 62 (49%) of the 126 patients with HM diseases had documented pulmonary coinfections [7]. Bacteria and viruses were also the most frequent copathogens in their cohort. Even higher rates of coinfections (65% and 70%) have been reported in two paediatric series IA [26, 27]. Two autopsy series respectively over a 15-year period (1989-2003) and 20-year period (1989-2008) have shown 42% and 46% of bacterial and fungal coinfections [28, 29]. The impact of influenza virus infections on both the occurrence of IA and the outcome has been recently highlighted [9, 10].

In our study, we have included all coinfections irrespective of their location. Although the type of coinfections and the copathogens were therefore heterogeneous, it reflects real-life and underlines the severity of immunosuppression in patients developing IA. The proportion of viruses increased after the use of multiplex PCR in respiratory samples, notably in BALF.

Interestingly, we were able to identify factors that were associated with coinfections and that may help practitioners to identify these coinfections. Among underlying diseases, alloHSCT, other HM and other underlying diseases were associated with increased rates of coinfection compared to acute leukaemia patients. The lower risk in acute leukaemia patients may be explained by the

environmental protective methods, antimicrobial prophylaxis and early empiric antibacterial therapy in these patients.

Age ≥65 years was a protective factor, which may be linked to less aggressive chemotherapy and lower rate of alloHSCT in elderly patients. BAL was performed at the same frequency in patients younger or older than 65 years (66% and 65% respectively). Paediatric studies reported rates of coinfection up to 70% also suggesting lower age is more frequently associated with multiple infections [27].

Among the other factors, tracheal intubation may be a gateway for multiple lung infections but also a marker of disease severity. Low lymphocyte count reflects immunosuppression and might favour the coinfections. The coinfections were more symptomatic with more frequent fever and higher C-reactive protein levels. Chest CT patterns were associated with coinfections and should be used as a red flag to suspect coinfections. Indeed, presence of non-nodular lesions such as consolidations, micronodules, alveolar or interstitial infiltrates were more often associated with coinfection than presence of nodules only.

A study conducted in Israel in HM patients with 21% of polymicrobial pneumonia, identified alloHSCT and graft versus host disease as risk factors for polymicrobial infections [8]. Polymicrobial infections were also more symptomatic with more dyspnoea, haemoptysis and required more respiratory support. Another study focusing on coinfections in HM patients did not find any difference in terms of subpopulation and radiological characteristics between patients with or without coinfections [7]. In IA patients with fever, high C-reactive protein and pulmonary CT

showing other lesions than nodule(s), we suggest to actively search for coinfections to avoid delay in treatments. BAL is the cornerstone for an appropriate identification of multiple pathogens.

Coinfections were independently associated with increased mortality in our study. Hospital mortality was also higher in the polymicrobial group (49 versus 19%, p<0,001) in the study conducted in Israel [8]. In 67 patients with HM admitted to an intensive care unit in France for acute respiratory failure related to IA, concomitant bacterial infections were documented in 24 (36%) patients and were associated with higher mortality [30].

There are limitations in our study. The analysis is retrospective over 21 years, and despite our efforts to identify the IA cases, it is likely we have missed a few of them. Clinical practices in diagnosis, classification and treatment have changed over year. We added the period of diagnosis of IA (<2009 and ≥2009) in the multivariable analysis to limit this bias. The EORTC/MSG classification has been updated in 2019. We have therefore retrospectively validated the cases with current criteria, modified by the use of a galactomannan cut-off above 0.5 according to manufacturer's recommendations. While classification of putative, probable, proven IA is based on strict criteria, the discrimination between bacterial coinfection and colonization is not always easy. To reduce this risk, we only accepted results from quantitative cultures of BALF and endotracheal aspirate and applied a cut-off of 10⁴ CFU/ml and 10⁵ CFU/ml, respectively. These cut-offs provide a high specificity for community acquired, hospital acquired and ventilator associated pneumonia [14-16]. Higher mortality in patients with co-infection may be explained by multiple factors including the treatment of this coinfection. Unfortunately, we did not collect data on therapy of the coinfection to assess its appropriateness. Finally, the monocentric design of the study limits its

extrapolation as the incidence and the diagnostic procedures for IA and coinfection might change
across different sites.

In conclusion, this analysis in a large and global cohort of patients with IA confirmed the high rate
of coinfections and their poor outcome. We identified factors associated with coinfection that
should act as warning for the clinician about this potential risk, to avoid treatment delay.

## **Collaborators**:

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# **Author's contribution:**

François Danion, Céline Duval, Raoul Herbrecht: conception and design of the work, acquisition, analysis, and interpretation of data; drafting of the work; final approval; agreement to be accountable

François Séverac: analysis and interpretation of data; drafting of the work; final approval;

agreement to be accountable

Julie Denis, Valérie Letscher-Bru, Marcela Sabou: acquisition and interpretation of data; critical review of the work; final approval; agreement to be accountable

Philippe Bachellier, Ermanno Candolfi, Vincent Castelain, Raphael Clere-Jehl, Laurence Dillenseger, Eric Epailly, Justine Gantzer, Blandine Guffroy, Yves Hansmann, Jean-Etienne Herbrecht, Pierre Leyendecker, Pauline Le Van Quyen, Pierre-Olivier Ludes, Guillaume Morel, Bruno Moulin, Catherine Paillard, Benjamin Renaud-Picard, Francis Schneider, Morgane Solis, Charles Tacquard, Emilie Talagrand-Reboul, Francis Veillon, Marie-Pierre Ledoux, and Célestine Simand: acquisition

of data; critical review of the work; final approval; agreement to be accountable

# 321 **Conflict of interest:** 322 François Danion reports personal fees from Gilead outside the submitted work. 323 Justine Gantzer reports personal fees from Chugai and Ipsen outside of the submitted work. 324 Blandine Guffroy reports personal fees from Shire, outside the submitted work. 325 Yves Hansmann reports personal fees from Pfizer, MSD, and Astellas, outside the submitted work. 326 Guillaume Morel reports personal fees from Pfizer, outside the submitted work. 327 Benjamin Renaud-Picard reports personal fees from Elivie outside of the submitted work. 328 Marcela Sabou reports personal fees from Pfizer outside the submitted work. 329 Marie-Pierre Ledoux reports personal fees from Daiichi-Sankyo, Pfizer, Chugai, Gilead and Novartis outside of the submitted work. 330 331 Raoul Herbrecht reports personal fees from Amgen, Astellas, Basilea, Gilead, MSD, Novartis, and 332 Pfizer outside the submitted work and research grants from Gilead and Novartis outside the submitted work 333 334 Célestine Simand, Céline Duval, François Séverac, Philippe Bachellier, Ermanno Candolfi, Vincent 335 Castelain, Laurence Dillenseger, Raphael Clere-Jehl, Julie Denis, Eric Epailly, Jean-Etienne Herbrecht, Valérie Letscher-Bru, Pierre Leyendecker, Pauline Le Van Quyen, Pierre-Olivier Ludes, 336 337 Guillaume Morel, Bruno Moulin, Catherine Paillard, Anne-Claude Roche, Francis Schneider, 338 Morgane Solis, Emilie Talagrand-Reboul, and Francis Veillon report no conflict of interest. 339 340 **Funding:** This study was supported in part by Alsace contre le Cancer. The funder has no implication in the 341 342 design, analysis or publication of the study. 343 344

346	Figure legends
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348	Figure 1.
349	Patient flow diagram
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352	Figure 2.
353	Twelve-week probability of survival: (a) all patients; (b) according to the presence or not of a
354	coinfection; (c) according to the type of copathogen; (d) according to the site of coinfection.
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Table 1. Characteristics of patients with and without a coinfection and factors associated with the presence of a coinfection in univariate analysis

	Coinfection		p value
	No (n=418)	Yes (n=272)	
Age, years, n (%)			0.05
≤40	66 (16%)	46 (17%)	
41-64	223 (53%)	165 (61%)	
≥65	129 (31%)	61 (22%)	
Sex, male, n (%)	271 (65%)	180 (66%)	0.72
Active or history of smoking, n (%)	206 (49%)	130 (48%)	0.58
Prior respiratory disease, n (%)	125 (30%)	79 (29%)	0.81
Primary underlying disease, n (%)	,	, ,	<0.01
Acute leukaemia	135 (32%)	50 (18%)	
Allogeneic HSCT	52 (12%)	46 (17%)	
Autologous HSCT	24 (6%)	9 (3%)	
Other haematological malignancy <sup>a</sup>	87 (21%)	69 (25%)	
Immunologic or benign haematological disease b	23 (6%)	16 (6%)	
Solid organ transplantation	67 (16%)	49 (18%)	
Solid tumour	20 (5%)	16 (6%)	
Other <sup>c</sup>	10 (2%)	17 (6%)	
Diabetes mellitus, n (%)	84 (20%)	67 (25%)	0.16
T-cell immunosuppressant, n (%)	142 (34%)	96 (35%)	0.72
Systemic steroids, n (%)	207 (50%)	151 (56%)	0.72
Previous invasive aspergillosis episode, n (%)	26 (6%)	21 (8%)	0.13
	20 (0%)	108 (39%)	< <b>0.44</b>
Antifungal prophylaxis, n (%) Antibiotics, n (%)	211 (30%) 277 (66%)		0.86
		182 (67%)	
Fever ≥38.5, n (%)	305 (76%)	215 (83%)	0.02
Tracheal intubation, n (%)	38 (9%)	64 (24%)	<0.01
Neutrophils <500/μl, n (%)	225 (54%)	122 (45%)	0.02
Lymphocytes (/μl), n (%)	(===:()		0.18
<500	216 (52%)	158 (58%)	
500-4000	182 (44%)	99 (36%)	
>4000	20 (5%)	15 (6%)	
Monocytes <100/μl, n (%)	206 (49%)	123 (45%)	0.30
C-reactive protein >180 (mg/l), n (%)	77 (19%)	76 (29%)	<0.01
Fibrinogen ≥5 (g/l), n (%)	242 (59%)	133 (50)	0.03
Creatinine clearance <50 (ml/min), n (%)	79 (19%)	69 (26%)	0.05
Chest CT, n (%)			<0.01
Nodule(s) only	105 (25%)	33 (12%)	
Nodule(s) with other lesion(s) d	188 (45%)	141 (52%)	
Other lesion(s) d without nodule	92 (22%)	82 (30%)	
Extra-pulmonary invasive aspergillosis <sup>e</sup>	33 (8%)	16 (6%)	
Pleural effusion	166 (40%)	139 (51%)	<0.01
Isolation of Aspergillus species, n (%)			<0.01
None	207 (50%)	105 (39%)	
Aspergillus fumigatus complex	170 (41%)	127 (47%)	
Other <i>Aspergillus</i> sp.	28 (7%)	19 (7%)	
≥2 different <i>Aspergillus</i> sp.	13 (3%)	21 (8%)	
Galactomannan in serum	, ,	. ,	0.67
Positive	245 (59%)	150 (55%)	
Negative	166 (40%)	117 (43%)	
Not done	7 (2%)	5 (2%)	

#### Table 1 (continued)

Galactomannan in BALF			0.70
Positive	156 (37%)	110 (40%)	
Negative	81 (19%)	49 (18%)	
Not done	181 (43%)	113 (42%)	
Proven invasive aspergillosis, n (%)	69 (17%)	43 (16%)	0.81
Site of invasive aspergillosis, n (%)			0.23
Lower respiratory tract only <sup>e</sup>	360 (86%)	226 (83%)	
Other single sites	17 (4%)	11 (4%)	
Disseminated	41 (10%)	35 (13%)	
Diagnosis after the year 2009, (%)	189 (45%)	158 (58%)	<0.01

Abbreviations: BALF: bronchoalveolar lavage fluid; CT: computed tomography; HSCT:

361 haematopoietic stem cell transplantation.

<sup>a</sup> Lymphoma, chronic lymphocytic leukaemia, multiple myeloma, chronic myeloproliferative neoplasm, non-transformed myelodysplastic syndrome.

<sup>b</sup> Systemic auto-immune or inflammatory diseases (13), aplastic anaemia (6), other auto-immune or toxic cytopenia (7), inherited immune deficiency (5), HIV infection (4), other acquired immune deficiency (2), macrophage activation syndrome (2).

<sup>c</sup> Trauma or post-surgery (7), liver failure (6), chronic pulmonary disease (5), aspiration pneumonitis (3), near-drowning (2), prematurity (2), diabetes (1), steroid therapy for chronic sinusitis (1).

<sup>d</sup> Other lesions included consolidations, ground-glass opacities or micronodules.

<sup>e</sup> Ten patients had only endoscopy-confirmed bronchial or tracheal lesions with normal chest CT.

Table 2. Copathogens and sites of the coinfections in 272 episodes of invasive aspergillosis

Characteristics	Number (%)
Sites of coinfections <sup>a</sup>	272 (100%)
Pulmonary only	131 (48%)
Bloodstream only	66 (24%)
Other or multiple sites	75 (28%)
Types of coinfections <sup>a</sup>	272 (100%)
Bacterial coinfection only	110 (40%)
Viral coinfection only	58 (21%)
Fungal coinfection only	57 (21%)
Parasitic coinfection only	5 (2%)
≥2 types of coinfection	42 (15%)
Pathogens involved in coinfections <sup>a</sup>	378 (100%)
Bacteria	178 (47%)
Staphylococcus aureus	17 (4%)
Coagulase-negative staphylococci	26 (7%)
Streptococcus and Enterococcus	28 (7%)
Escherichia coli	27 (7%)
Klebsiella pneumoniae	11 (3%)
Other enterobacteria	18 (5%)
Pseudomonas aeruginosa	23 (6%)
Other non-fermenting Gram-negative bacilli	9 (2%)
Other bacteria	13 (3%)
Mycobacteria	6 (2%)
Viruses	101 (27%)
Influenza virus	24 (6%)
Other respiratory viruses	33 (9%)
Cytomegalovirus	12 (3%)
Herpes simplex virus	21 (6%)
Other	11 (3%)
Fungi	89 (23%)
Candida spp.	26 (7%)
Cryptococcus spp.	4 (1%)
Other yeasts	2 (1%)
Mucorales	25 (7%)
Fusarium spp.	5 (1%)
Scedosporium spp.	4 (1%)
Other filamentous fungi	8 (2%)
Pneumocystis jirovecii	15 (4%)
Parasites	10 (3%)
Toxoplasma gondii	8 (2%)
Plasmodium falciparum	2 (1%)

<sup>&</sup>lt;sup>a</sup> A patient could have more than one copathogen, more than one site of coinfection and more than one species of bacteria, virus or fungus.

Table 3. Multivariable analysis of factors associated with coinfection

	OR	IC95	p value
Age (years)			
≤40	ref		
41-64	0.99	0.59-1.67	0.984
≥65	0.47	0.25-0.90	0.022
Underlying disease			
Acute leukaemia	ref		
Allogeneic HSCT	2.30	1.21-4.37	0.011
Autologous HSCT	1.10	0.44-2.78	0.836
Other haematological malignancy	2.12	1.17-3.84	0.013
Immunologic or benign haematological disease	1.95	0.78-4.84	0.153
Solid organ transplantation	1.04	0.45-2.37	0.929
Solid tumour	1.03	0.42-2.54	0.949
Other	4.28	1.35-13.55	0.013
Diabetes mellitus	1.05	0.67-1.66	0.823
Systemic steroids	0.98	0.64-1.49	0.918
Antifungal prophylaxis	0.77	0.50-1.17	0.217
Fever ≥38.5	2.45	1.45-4.12	<0.001
Tracheal intubation	2.64	1.47-4.72	0.001
Neutrophils <500/μl	0.82	0.51-1.34	0.434
Lymphocytes (/μΙ)			
500-4000	ref		
<500	1.67	1.11-2.50	0.014
>4000	1.74	0.73-4.17	0.210
C-reactive protein >180 mg/l	1.90	1.21-2.97	0.005
Fibrinogen ≥5 g/l	0.77	0.52-1.13	0.182
Creatinine clearance <50 ml/min	1.16	0.72-1.87	0.548
Chest CT-scan			
Nodule(s) only	ref		
Nodule(s) with other lesion(s)	2.23	1.34-3.71	0.002
Other lesion(s) without nodule	2.22	1.25-3.96	0.007
Extra-pulmonary invasive aspergillosis	1.60	0.65-3.99	0.309
Pleural effusion	1.19	0.81-1.74	0.388
Isolation of Aspergillus species			
None	ref		
Aspergillus fumigatus complex	1.08	0.71-1.65	0.720
Other <i>Aspergillus</i> sp.	1.33	0.62-2.83	0.465
≥2 different <i>Aspergillus</i> sp.	2.67	1.12-6.35	0.026
Diagnosis after the year 2009	1.88	1.26-2.80	0.002

Abbreviations: Abbreviations: BAL: bronchoalveolar lavage; CT: computed tomography; HSCT:

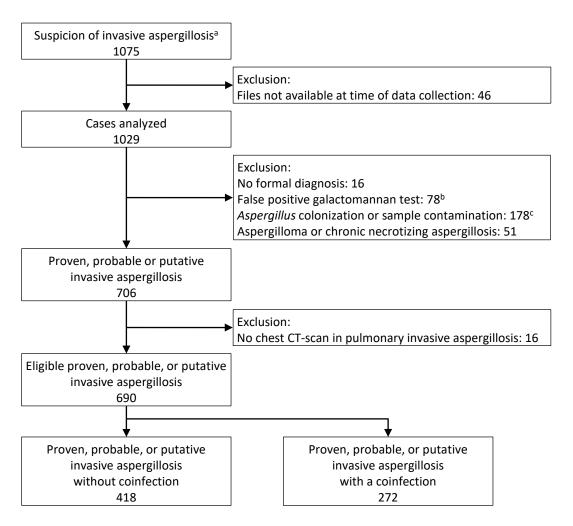
haematopoietic stem cell transplantation; ref, reference group.

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<sup>&</sup>lt;sup>a</sup> based on a positive culture, galactomannan in serum, bronchoalveolar lavage fluid or cerebrospinal fluid, or PCR in serum, bronchoalveolar lavage fluid or tissue and at least one of the following condition: proven aspergillosis, presence of an host factor as defined by the EORTC /MSG criteria or hospitalized in an intensive care unit.

<sup>&</sup>lt;sup>b</sup> False positive galactomannan in serum (n=71) or in bronchoalveolar lavage fluid (n=7).

<sup>&</sup>lt;sup>c</sup> 185 species in 178 patients.

