

Mucormycosis treatment: Recommendations, latest advances, and perspectives

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1 Mucormycosis treatment: recommendations, latest advances, and perspectives. 2 3 ^{1,2,3}Kévin Brunet and ^{1,2,4}Blandine Rammaert 4 5 ¹ INSERM U1070, Poitiers, France 6 ² Université de Poitiers, Faculté de médecine et pharmacie, Poitiers, France 7 ³ CHU Poitiers, service de Mycologie-Parasitologie, Département des agents infectieux, Poitiers, 8 France 9 ⁴ CHU Poitiers, service de maladies infectieuses et tropicales, Poitiers, France 10 11 Corresponding author: 12 Dr Kévin Brunet 13 INSERM U 1070 14 UFR de Médecine et Pharmacie Université de Poitiers 15 16 Pôle Biologie Santé 17 1 rue Georges Bonnet 18 86022 Poitiers cedex 19 Tel: (33) 5 49 36 64 37 20 21 Conflict of interest 22 BR received travel grants for conferences from Pfizer and MSD, speaker's fees from MSD, Basilea, 23 Gilead, Astellas, Igone. 24 KB received travel grants for conferences from Pfizer, MSD and Gilead. 25

Abstract

Mucormycosis are life-threatening fungal infections especially affecting immunocompromised or diabetic patients. Despite treatment, mortality remains high (from 32 to 70% according to organ involvement). This review provides an update on mucormycosis management. The latest recommendations strongly recommend as first-line therapy the use of liposomal amphotericin B (≥ 5mg/kg) combined with surgery whenever possible. Isavuconazole and intravenous or delayed-release tablet forms of posaconazole have remained second-line. Many molecules are currently in development to fight against invasive fungal diseases but few have demonstrated efficacy against *Mucorales*. Despite *in vitro* efficacy, combinations of treatment have failed to demonstrate superiority versus monotherapy. Adjuvant therapies are particularly complex to evaluate without prospective randomized controlled studies which are complex to perform due to low incidence rate and high mortality of mucormycosis. Perspectives are nonetheless encouraging. New approaches assessing relationships between host, fungi, and antifungal drugs, and new routes of administration such as aerosols could improve mucormycosis treatment.

Keywords

Mucorales, antifungal drugs, nebulization, polyenes, azoles, prophylaxis

Introduction

Mucormycosis are life-threatening invasive fungal diseases (IFD) due to fungi belonging to Mucorales order [1]. Mucormycosis lead to many clinical manifestations, ranging from localized to disseminated infection. Pulmonary or disseminated diseases are commonly found in immunosuppressed patients (hematological malignancy, hematopoietic stem cell transplantation), rhino-orbito-cerebral form in diabetic patients, and cutaneous forms in patients having trauma [2]. Other localizations (gastrointestinal, endocarditis, osteoarticular or isolated cerebral infections) are less frequent. Treatment is based on surgery when possible, correction of underlying factors, and aggressive antifungal drug therapy [3]. In contrast to other fungi, few molecules are active. Amphotericin B (AmB), posaconazole (PSZ), and isavuconazole (ISZ) have shown in vitro efficacy while voriconazole (VCZ) and echinocandins are inefficient [4,5]. The reference method for antifungal susceptibility testing (AST) is broth microdilution method with the methodology of the European Committee for Antimicrobial Susceptibility Testing (EUCAST) [6] or the Clinical and Laboratory Standards Institute (CLSI) [7]. However, a major concern with AST is the lack of clinical breakpoints. Prognosis remains poor, mortality ranging from 32 to 70%, and is linked to underlying diseases and clinical forms [2]. Therapeutic improvement is therefore mandatory [8]. The aim of this review is to focus on latest recommendations, advances and perspectives on mucormycosis treatment.

Recent and new antifungal drugs

Current recommendations

The European Conference on Infections in Leukemia (ECIL) published mucormycosis treatment guidelines in 2017 [9] and the European Confederation of Medical Mycology (ECMM) provided an update in 2019 [10]. Both societies strongly recommend liposomal Amphotericin B (L-AmB) for first-line treatment in adults (A II) (Table 1). Another lipid formulation, Amphotericin B lipid complex (ABLC) could be used in mucormycosis but without central nervous system (CNS) involvement according to the ECIL (B II) [9]. For neonates and pediatric population, L-AmB and ABLC were strongly recommended as first-line treatment (A II) [10].

One issue to be addressed is the dose regimen of L-AmB. The ECMM recommends 5-10 mg/kg and 10 mg/kg in the event of CNS involvement [10]. In a prospective pilot study (Ambizygo), high doses of L-AmB were tested as first-line treatment of mucormycosis [11]. Response rate was 43% (12/28) in patients who received ≥ 7.5 mg/kg/day during the first week compared with 0% (0/5) in patients who did not. A high dose of L-AmB (10mg/kg), combined with surgery in 71% of cases, led to an overall response rate of 36% at week 4 and 45% at week 12. Compared to another study using L-AmB ≥ 5 mg/kg in mucormycosis treatment, response was similar at week 4 (36% vs 40%) but was better at week 12 (45% vs 35%) [12]. Mortality rates were equivalent at week 12 (38% vs 42%). A major side effect with L-AmB high dose was creatinine level doubling in 40% of patients.

The ECMM recommended dose should not be slowly increased over several days but a full dose must be given from the first day of daily treatment [10].

ISZ per os (PO) or intravenous (IV), PSZ delayed-release (DR) tablets or IV forms have been recommended with moderate strength (B II) and PSZ oral suspension have been marginally

recommended (C II) [10]. Moreover, the ECMM strongly discouraged the use of AmB deoxycholate (AmBd) (D II).

Treatment should be started as soon as possible, as delayed AmB therapy is linked to increased mortality [13]. Moreover, it must be continued until complete response on imaging and permanent reversal of immunosuppression [10]. Treatment duration necessary to treat mucormycosis is unknown and further studies are needed to better determine it. To facilitate treatment in stable disease, ISZ PO or PSZ DR tablets are strongly recommended. Time between induction phase with AmB and introduction of azoles depends on clinical and imaging responses. Some authors recommend at least 3 weeks of induction with parenteral AmB [14].

The ECMM has addressed recommendations concerning prophylaxis (Table 2). In neutropenic or GvHD, PSZ DR tablets or IV form are moderately supported (B II, B III) and PSZ oral suspension marginally recommended (C II) while ISZ is marginally supported in neutropenic patients (C II). Finally, in solid organ transplant (SOT) recipients, PSZ and ISZ are marginally recommended in prophylaxis (C III, C II).

Recent drugs

ISZ, a new azole, was approved in the United States and in Europe in 2015 for the treatment of mucormycosis [3,15]. ISZ is available in oral and IV formulations, and presents some advantages: linear pharmacokinetics, few interactions with cytochrome P450 isoenzymes leading to few drugdrug interactions, QT decrease, no nephrotoxic cyclodextrin in the IV formulation (different from posaconazole IV form), no need for dose adjustment in kidney or liver failure and in obesity, and excellent oral bioavailability with no food requirements [3]. Although ISZ has shown higher minimal inhibitory concentrations (MIC) than posaconazole [16], it is demonstrably as effective as AmB to decrease fungal burden and to improve survival in a neutropenic mouse model of mucormycosis [17]. ISZ was tested in VITAL study, a phase 3, single-arm, open-label, non-comparative study. This study assessed safety and efficacy of ISZ in the treatment of mucormycosis [18]. Case-control analysis with historic controls treated with AmB included in the Fungiscope registry showed similar survival benefit. However, some biases are noticeable. A total of 21 patients treated with ISZ were compared to 33 matched controls who had received AmB. AmB was administered through AmBd formulation in 7 controls. This formulation is appreciably less efficient than the liposomal one. Other drawbacks could be emphasized in the use of ISZ. Breakthrough Mucorales infections in patients receiving ISZ have been reported [19]. Moreover, some authors have shown in Drosophila model of mucormycosis that preexposure to ISZ enhances the virulence of Mucorales [20,21]. In a large study including 147 patients, ISZ prophylaxis was less effective than VCZ or PSZ against IFD. Two patients who received ISZ as prophylaxis presented mucormycosis [22]. Although ISZ seems to be less hepatotoxic than other mould-active azoles and present a better tolerance profile than L-AmB [23], the ECMM recommends only moderately ISZ as first-line treatment [10].

Regarding central nervous system (CNS) infections, treatment is based on L-AmB due to clinical experience and *in vitro* data [24]. It has been demonstrated that ISZ penetrates the blood-brain barrier in animal models [25], while AmB displays limited penetration. Concentration of ISZ in the necrotic center of brain abscess has been shown low, but concentration in inflammatory brain tissue surrounding the abscess was adequate, equivalent to predicted plasma concentration [26]. A

recent retrospective study has shown that ISZ is effective in *Mucorales* CNS infections [27]. This result has to be confirmed with larger studies.

IV and DR tablets of PSZ were recently developed and lead to better bioavailability and drug exposure than previous oral solution [28–30]. This increased drug exposure has been related to increased PSZ efficiency [31]. Moreover, DR tablets lead to less variability in absorption and compared to oral suspension are not affected by food [32]. Due to higher serum level, suspension DR tablets and IV forms are moderately recommended while oral suspension is only marginally recommended by the ECMM as first-line treatment [10]. There is no safety concern compared to oral suspension for the two new forms, since there is no correlation between serum level and safety [29–32]. However, IV form is solubilized in cyclodextrin and may lead to renal issues [31]. In a matched-paired analysis of patients treated for invasive mucormycosis, new formulations of PSZ were evaluated [33]. The authors showed that PSZ new formulations are as effective as AmB as first-line treatment and as oral suspension in salvage therapy. However, these results should be interpreted with caution. Numerous biases can be noted such as small sample size, retrospective design, heterogeneity of infectious sites, lack of drug monitoring, and pre-exposure to other antifungal drugs.

Routine therapeutic drug monitoring (TDM) is strongly recommended for patients treated by PSZ [10]. Serum trough PSZ concentrations of 1 mg/L or higher are recommended. However, there is currently no conclusive evidence for routine TDM with ISZ. It could be useful in case of suspected toxicity, treatment failure, drug interactions, obesity, or after a switch from IV to PO therapy [10].

152 <u>New drugs</u>

Some new antifungal drugs are under clinical evaluation include Rezafungin, SCY-078, orolofim, and encochleated amphotericin B [34]. Rezafungin, a new echinocandin has not been tested against *Mucorales*. SCY-078, member of a new glucan synthase inhibitor subclass is poorly or not active against *Mucorales* [35]. Olorofim is a member of the orotomides, a new antifungal class inhibiting dihydroorotate dehydrogenase (DHODH), a key enzyme in pyrimidine biosynthesis. It is also poorly active against *Mucorales* [36]. Encochleated amphotericin B is a new oral formulation of amphotericin B [34]. It has been shown to be well-tolerated, and is currently tested for cryptococcosis treatment in developing countries (clinical trial NCT04031833). No studies on *Mucorales* efficacy are available.

Other antifungal drugs with activity against *Mucorales* are being developed. VT-1161 is a novel inhibitor of the fungal CYP 51 with *in vitro* activity against *Mucorales*. VT-1161 used as curative or prophylactic treatment has prolonged survival of neutropenic mice in *R. arrhizus* models [37,38]. SCH 42427 a broad-spectrum triazole was found to be effective in a murine model [39]. APX001A (Fosmanogepix) (formerly E1210) is an antifungal agent targeting protein Gwt1. Gwt1 is a surface protein of the glycosylphosphotidyl inositol post-translational modification pathway. Although MICs against *Mucorales* are high [40,41], several authors have shown that APX001A is as effective as AmB to protect mice in a *R. delamar* model [3,42]. Finally, PC1244 a new long-acting fungicidal azole, has shown antifungal activity against *Mucorales* with MICs from 0.25 to 2 mg/L [43] but has not been tested *in vivo*. Among antibiotics, colistin has presented modest *in vitro* and *in vivo* activity against *Mucorales* [44].

New therapeutic approaches

New approaches have recently emerged regarding relationship between fungus, antifungal agent, and host [45]. For example, some authors have emphasized the capacity of PSZ to accumulate within leukocyte membrane due to its lipophilic properties. Cells from HL-60 leukemia cell line differentiated to neutrophil-like phenotype have been loaded with PSZ and used in an aspergillosis mouse model to deliver PSZ directly to the infectious site [46]. However, this new approach has not been tested in a *Mucorales* model. Bioengineering has made great improvement, especially in genetically modified cytotoxic T-cells. These modified cells can specifically target beta-glucan of fungus cell wall [47]. However, this approach has only been tested in an aspergillosis model.

Recently, a *Mucorales* peptide named CotH3 was found to be linked to mucormycosis endothelial invasion by binding the endothelial cell receptor GRP78. Authors generated antibodies against CotH3 to prevent endothelial invasion. Anti-CotH3 antibodies protected neutropenic and diabetic mice from mucormycosis and acted synergistically with antifungal drugs [48]. Moreover, other authors have shown that blocking GRP78 cell receptor by GRP78-specific immune serum may protects diabetic mice from mucormycosis [49]. This peptide-receptor interaction may be a new therapeutic way of research.

New concepts may guide antifungal prophylaxis.

Several authors have hypothesized that *Mucorales*, such as *Histoplasma sp* or *Cryptococcus sp*, can remain latent in immunocompetent patients and lead to active disease when a patient becomes immunosuppressed [50,51]. Authors have shown that *Mucorales* spores might remain dormant in cutaneous granulomatous lesions in an immunocompetent rabbit model [52] and inside innate granuloma in a Zebrafish model [51]. In the event of immunodepression, spores were reactivated. In a murine model of latent mucormycosis, L-AmB was effective to prevent reactivation in *Lichtheimia corymbifera* colonized mice [53]. This concept must be verified in human to evaluate if decolonization of patients before immunosuppression could reduce the risk of reactivation.

Combinations

Combination are not currently recommended for first-line therapy due to lack of evidence of their efficacy (C II, C III) (Table 1) [54]. They could nonetheless represent a major way of increasing antifungal treatment efficacy [55].

Combinations of antifungal agents have been largely tested *in vitro*. Most combinations were indifferent, except for AmB + caspofungin (CAS), PSZ + CAS and ISZ + CAS which were synergistic [55–57]. For azoles + echinocandins, few *in vivo* studies have been performed, showing lack of synergy [58,59]. Among AmB + echinocandins, more data are available *in vivo*. *In vitro* data have been confirmed in a ketoacidotic mouse model where L-AmB and echinocandins (micafungin and anidulafungin) appeared synergistic [60]. Combination of L-AmB and echinocandins prolonged survival and decreased fungal burden of mice in an IV model of *Rhizopus arrhizus* infection. These data were confirmed with ABLC + caspofungin in a ketoacidotic mouse model with improvement of

survival but without fungal clearance in organs [61]. Case reports of combinations have been published and five retrospective clinical studies have been performed on antifungal combinations. AmB + CAS and/or PSZ combinations have been evaluated. Four of the five studies showed indifference [62–65] and one showed synergy [66]. However, the latter included only rhino-orbitocerebral forms and a small number of patients.

Some non-antifungal agents combined with antifungals drugs have shown interesting synergy. Iron chelators have shown high synergy with antifungal drugs *in vitro* and *in vivo* [59,67,68], but this has not been confirmed in patients [12]. Calcineurin inhibitors (cyclosporine A and tacrolimus), which have immunosuppressive effects, have shown synergy with AmB, PSZ or ISZ *in vitro* and *in vivo* [69–71]. Other agents such as MGCD290 (a Hos2 Histone Deacetylase Inhibitor), not yet FDA-approved, have shown synergy *in vitro* with PSZ [72]. Although lovastatin has shown synergy *in vitro* and *in vivo* with voriconazole [73], the latter is known to enhance *Mucorales* virulence [21]. Ciprofloxacin and fluconazole have shown synergy in mouse models [74], while miltefosine and azoles, and rifampicine and AmB have presented synergistic effects *in vitro* on \leq 50% [75] and 56 to 83 % [76,77] of tested strains respectively.

Randomized, placebo-controlled clinical trials are needed to determine combination therapy efficacy [78]. Added toxicity, drug interactions, and cost-benefit balance of combinations remain unclear [10].

Adjunctive treatment

Current recommendations

The ECMM and the ECIL-6 strongly recommend surgery and control of underlying disease including management of ketoacidosis and hyperglycemia in diabetic patients, modulation of corticosteroids and immunosuppressive drugs, and reduction of neutropenia duration using hematopoietic growth factor if possible (A II, AII) (Table 3) [9,10]. Granulocyte colony-stimulating factor (G CSF) and hyperbaric oxygen are moderately recommended in case of neutropenia and in diabetic patients respectively (B II), while iron chelators are strongly discouraged.

<u>Surgery</u>

Surgery remains easier to perform in rhino-orbital or cutaneous localizations than in cerebral, pulmonary or disseminated disease. Surgery is precluded in critical ill patients [79]. In patients with unifocal pulmonary mucormycosis, lobectomy or pneumonectomy have provided benefit [80]. However, in case of multifocal or close to great vessels lesions, benefit is less established and surgery is most complicated to perform. In rhino-orbito-cerebral forms, surgery is strongly linked to treatment outcome [81,82]. In a clinico-epidemiological review over 10 years, surgery was performed in 65.2 % of 184 patients but only in 21.4 % of hematological patients [83]. Surgical debridement in combination with medical therapy was associated with a better outcome than medical therapy only. Attention must be paid to the fact that only retrospective studies and epidemiological data are available. However, the benefits of surgery are presumed and is highly recommended whenever possible (A II) [10].

Adjunctive therapies

Adjunctive therapy is used to reverse immunosuppression. Granulocyte (macrophage) colony-stimulating factor (G(M)-CSF) or interferon- γ increases the activity of granulocytes against *Mucorales* such as hyphae damage [84,85]. However, several authors have shown that G-CSF or GM-CSF did not improve antifungal activity of PSZ or L-AmB in a neutropenic murine model [86,87]. Clinical data on this topic are very poor and few cases are published [88–91]. Clear benefit has yet to be established.

Iron chelators have been tested in adjunctive therapy as means of reducing iron availability and thereby inhibiting fungal growth. Deferoxamine, an iron chelator, has been associated with increased mucormycosis incidence. Deferoxamine acts as a xenosiderophore, whereas the two other iron chelators, deferiprone and deferasirox, do not [92]. Ibrahim *et al.* have shown that deferiprone protected diabetic mice from mucormycosis [93]. Deferasirox shared the same effect in diabetic and neutropenic mice and acted synergistically with AmB [67]. Triple therapy using L-AmB, micafungin, and deferasirox was also found to be effective [94]. Moreover, deferasirox increased PSZ activity in a neutropenic mouse model [59]. These promising results led to a clinical study assessing deferasirox + LAmB efficacy [12]. However, patients with mucormycosis treated with deferasirox + L-AmB had a higher mortality rate at 90 days than patients treated with L-AmB alone. However, patients treated with deferasirox had more active malignancy, neutropenia and corticosteroid therapy compared to the placebo group. Population imbalance between deferasirox and placebo groups did not allow for clear conclusions. Unlike iron chelators, zinc chelators have not shown synergy with AmB and have shown poor synergy with PSZ [95].

Hyperbaric oxygen treatment has been shown to deploy direct antimicrobial activity. It exerts a synergistic effect with antimicrobial agents, and enhances cellular immune system and tissue repair in some infectious diseases [96]. In a mucormycosis mouse model, addition of hyperbaric oxygen to AmB did not improve survival [97]. However, in this model, mice were neither immunosuppressed nor diabetic and infection was performed by IV route. In a review of 28 published cases, authors showed that while hyperbaric oxygen improved survival in diabetic patients, it was ineffective in neutropenic patients [98]. However, hyperbaric oxygen treatment failure may be underestimated due to publication bias. Up until now, there has been no randomized study with control group to evaluate efficacy of hyperbaric oxygen so far.

New routes of administration

Nebulized antifungal agents may be a new way of research to improve mucormycosis therapy. The pulmonary aerosolization of antifungal agents can theoretically increase their concentration at the infectious site, which could improve efficacy while limiting their systemic exposure and toxicity. [99]. Administration of L-AmB aerosol was evaluated in a neutropenic mouse model of *R. arrhizus* pulmonary infection [100]. The authors showed that aerosolized L-AmB is effective as a means of decreasing fungal burden and improving survival when administered from day 1 to 5 after infection compared to placebo. However, aerosolized L-AmB was not compared to systemic L-AmB. More animal studies are needed to assess aerosols efficacy alone and in

combination with systemic treatment. Few human cases of mucormycosis treated with AmB aerosols are reported in the literature [101–105]. AmBd has been the most used formulation. Dosage of nebulized AmBd ranges from 6 mg three times a day to 30 mg twice a week, in combination with AmB systemic treatment and surgical treatment. ABLC has also been used in *Rhizomucor sp* infection treatment (50 mg twice daily) [105].

Topical AmB has been used in a few clinical cases [106-108], particularly in burned patients [109]. It has been employed in different forms: washes [110], 5% sulfamylon–amphotericin B (2 µg/ml) dressings [111], daily topical infusions through dressings (50 mg L-AmB diluted in 1L of sterile water) [112], soaks [113] or gauze soaked in 0.2 % AmB solution [114]. Nystatin cream was used in one case report [115]. AmB nanoemulsion was developed to use AmB as topical route of administration, but it has not been tested on *Mucorales* [116,117]. Other authors have developed nanoemulsions containing surfactant to mechanically disrupt microbial membranes. For example, nanoemulsion NB-201, containing refined soybean oil, water, glycerol, EDTA, Tween 20, and the surfactant benzalkonium chloride has shown *in vitro* activity against *Mucorales* [118].

Other routes of administration for AmB are anecdotal: eye drop in *Mucorales* keratitis [119], oral administration in gastrointestinal mucormycosis [120], intradiaphyseal incorporation cement beads in osteomyelitis [121], intrathecal administration in cerebral abscess [122], percutaneous injection in cutaneous lesions [123].

Conclusion

Mucormycosis treatment recommendations were recently updated by the ECMM. L-AmB remains the first-line drug in mucormycosis therapy. ISZ and new PSZ formulations have been added to the guidelines but remain in second-line treatment following L-AmB due to some remaining issues. Few anti-*Mucorales* drugs are currently under development. Moreover, evidence for adjunctive therapies is scarce and doubts on their effectiveness persist due to a lack of randomized prospective controlled studies. They are particularly complex to implement in a context of low incidence disease. Few advances have been made on mucormycosis treatment. However, empowering new concepts and new routes of administration to fight this devastating disease appear promising and are to be encouraged.

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References

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- Farmakiotis D, Kontoyiannis DP. Mucormycoses. Infect Dis Clin North Am 2016;30:143–63. https://doi.org/10.1016/j.idc.2015.10.011.
- Serris A, Danion F, Lanternier F. Disease Entities in Mucormycosis. J Fungi (Basel) 2019;5. https://doi.org/10.3390/jof5010023.
- Sipsas NV, Gamaletsou MN, Anastasopoulou A, Kontoyiannis DP. Therapy of Mucormycosis. J Fungi (Basel) 2018;4. https://doi.org/10.3390/jof4030090.
- Alastruey-Izquierdo A, Castelli MV, Cuesta I, Zaragoza O, Monzón A, Mellado E, et al. In vitro activity of antifungals against Zygomycetes. Clin Microbiol Infect 2009;15 Suppl 5:71–6. https://doi.org/10.1111/j.1469-0691.2009.02984.x.
- Guinea J, Peláez T, Recio S, Torres-Narbona M, Bouza E. In vitro antifungal activities of
 isavuconazole (BAL4815), voriconazole, and fluconazole against 1,007 isolates of zygomycete,
 Candida, Aspergillus, Fusarium, and Scedosporium species. Antimicrob Agents Chemother
 2008;52:1396–400. https://doi.org/10.1128/AAC.01512-07.
- Subcommittee on Antifungal Susceptibility Testing of the ESCMID European Committee for Antimicrobial Susceptibility Testing. EUCAST Technical Note on the method for the determination of broth dilution minimum inhibitory concentrations of antifungal agents for conidia-forming moulds. Clin Microbiol Infect 2008;14:982–4. https://doi.org/10.1111/j.1469-0691.2008.02086.x.
- CLSI. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. M38-A2. Clinical and Laboratory Standards Institute, Wayne, Pa 2008.
- Prakash H, Chakrabarti A. Global Epidemiology of Mucormycosis. J Fungi (Basel) 2019;5. https://doi.org/10.3390/jof5010026.
- Tissot F, Agrawal S, Pagano L, Petrikkos G, Groll AH, Skiada A, et al. ECIL-6 guidelines for the treatment of invasive candidiasis, aspergillosis and mucormycosis in leukemia and hematopoietic stem cell transplant patients. Haematologica 2017;102:433–44. https://doi.org/10.3324/haematol.2016.152900.
- [10] Cornely OA, Alastruey-Izquierdo A, Arenz D, Chen SCA, Dannaoui E, Hochhegger B, et al.
 Global guideline for the diagnosis and management of mucormycosis: an initiative of the
 European Confederation of Medical Mycology in cooperation with the Mycoses Study Group
 Education and Research Consortium. Lancet Infect Dis 2019;19:e405–21.
 https://doi.org/10.1016/S1473-3099(19)30312-3.
- Lanternier F, Poiree S, Elie C, Garcia-Hermoso D, Bakouboula P, Sitbon K, et al. Prospective pilot study of high-dose (10 mg/kg/day) liposomal amphotericin B (L-AMB) for the initial treatment of mucormycosis. J Antimicrob Chemother 2015;70:3116–23. https://doi.org/10.1093/jac/dkv236.
- 365 [12] Spellberg B, Ibrahim AS, Chin-Hong PV, Kontoyiannis DP, Morris MI, Perfect JR, et al. The
 366 Deferasirox-AmBisome Therapy for Mucormycosis (DEFEAT Mucor) study: a randomized,
 367 double-blinded, placebo-controlled trial. J Antimicrob Chemother 2012;67:715–22.
 368 https://doi.org/10.1093/jac/dkr375.
- Chamilos G, Lewis RE, Kontoyiannis DP. Delaying amphotericin B-based frontline therapy significantly increases mortality among patients with hematologic malignancy who have zygomycosis. Clin Infect Dis 2008;47:503–9. https://doi.org/10.1086/590004.
- 372 [14] Kontoyiannis DP, Lewis RE. How I treat mucormycosis. Blood 2011;118:1216–24. 373 https://doi.org/10.1182/blood-2011-03-316430.
- Jonnelley MA, Zhu ES, Thompson GR. Isavuconazole in the treatment of invasive aspergillosis and mucormycosis infections. Infect Drug Resist 2016;9:79–86.
 https://doi.org/10.2147/IDR.S81416.

- 377 [16] Arendrup MC, Jensen RH, Meletiadis J. In Vitro Activity of Isavuconazole and Comparators 378 against Clinical Isolates of the Mucorales Order. Antimicrob Agents Chemother 2015;59:7735– 379 42. https://doi.org/10.1128/AAC.01919-15.
- Luo G, Gebremariam T, Lee H, Edwards JE, Kovanda L, Ibrahim AS. Isavuconazole Therapy
 Protects Immunosuppressed Mice from Mucormycosis. Antimicrob Agents Chemother
 2014;58:2450–3. https://doi.org/10.1128/AAC.02301-13.
- 383 [18] Marty FM, Ostrosky-Zeichner L, Cornely OA, Mullane KM, Perfect JR, Thompson GR, et al.
 384 Isavuconazole treatment for mucormycosis: a single-arm open-label trial and case-control
 385 analysis. Lancet Infect Dis 2016;16:828–37. https://doi.org/10.1016/S1473-3099(16)00071-2.
- Rausch CR, DiPippo AJ, Bose P, Kontoyiannis DP. Breakthrough Fungal Infections in Patients
 With Leukemia Receiving Isavuconazole. Clin Infect Dis 2018;67:1610–3.
 https://doi.org/10.1093/cid/ciy406.
- Wurster S, Lewis RE, Albert ND, Kontoyiannis DP. Preexposure to Isavuconazole Increases the Virulence of Mucorales but Not Aspergillus fumigatus in a Drosophila melanogaster Infection Model. Antimicrob Agents Chemother 2019;63. https://doi.org/10.1128/AAC.01896-18.
- Lamaris GA, Ben-Ami R, Lewis RE, Chamilos G, Samonis G, Kontoyiannis DP. Increased
 virulence of Zygomycetes organisms following exposure to voriconazole: a study involving fly
 and murine models of zygomycosis. J Infect Dis 2009;199:1399–406.
 https://doi.org/10.1086/597615.
- Fontana L, Perlin DS, Zhao Y, Noble BN, Lewis JS, Strasfeld L, et al. Isavuconazole prophylaxis in patients with hematologic malignancies and hematopoietic-cell transplant recipients. Clin Infect Dis 2019. https://doi.org/10.1093/cid/ciz282.
- Jenks JD, Mehta SR, Hoenigl M. Broad spectrum triazoles for invasive mould infections in adults: Which drug and when? Med Mycol 2019;57:S168–78.

 https://doi.org/10.1093/mmy/myy052.
- 402 [24] Chikley A, Ben-Ami R, Kontoyiannis DP. Mucormycosis of the Central Nervous System. J Fungi (Basel) 2019;5. https://doi.org/10.3390/jof5030059.
- Schmitt-Hoffmann A-H, Kato K, Townsend R, Potchoiba MJ, Hope WW, Andes D, et al. Tissue
 Distribution and Elimination of Isavuconazole following Single and Repeat Oral-Dose
 Administration of Isavuconazonium Sulfate to Rats. Antimicrob Agents Chemother 2017;61.
 https://doi.org/10.1128/AAC.01292-17.
- 408 [26] Lamoth F, Mercier T, André P, Pagani JL, Pantet O, Maduri R, et al. Isavuconazole brain 409 penetration in cerebral aspergillosis. J Antimicrob Chemother 2019;74:1751–3. 410 https://doi.org/10.1093/jac/dkz050.
- 411 [27] Schwartz S, Cornely OA, Hamed K, Marty FM, Maertens J, Rahav G, et al. Isavuconazole for the 412 treatment of patients with invasive fungal diseases involving the central nervous system. Med 413 Mycol 2019. https://doi.org/10.1093/mmy/myz103.
- Skiada A, Lass-Floerl C, Klimko N, Ibrahim A, Roilides E, Petrikkos G. Challenges in the diagnosis
 and treatment of mucormycosis. Med Mycol 2018;56:S93–101.
 https://doi.org/10.1093/mmy/myx101.
- 417 [29] Cornely OA, Robertson MN, Haider S, Grigg A, Geddes M, Aoun M, et al. Pharmacokinetics and safety results from the Phase 3 randomized, open-label, study of intravenous posaconazole in patients at risk of invasive fungal disease. J Antimicrob Chemother 2017;72:3406–13. https://doi.org/10.1093/jac/dkx263.
- 421 [30] Cornely OA, Duarte RF, Haider S, Chandrasekar P, Helfgott D, Jiménez JL, et al. Phase 3
 422 pharmacokinetics and safety study of a posaconazole tablet formulation in patients at risk for
 423 invasive fungal disease. J Antimicrob Chemother 2016;71:718–26.
 424 https://doi.org/10.1093/jac/dkv380.
- 425 [31] Maertens J, Cornely OA, Ullmann AJ, Heinz WJ, Krishna G, Patino H, et al. Phase 1B study of 426 the pharmacokinetics and safety of posaconazole intravenous solution in patients at risk for 427 invasive fungal disease. Antimicrob Agents Chemother 2014;58:3610–7. 428 https://doi.org/10.1128/AAC.02686-13.

- 429 [32] Duarte RF, López-Jiménez J, Cornely OA, Laverdiere M, Helfgott D, Haider S, et al. Phase 1b 430 study of new posaconazole tablet for prevention of invasive fungal infections in high-risk 431 patients with neutropenia. Antimicrob Agents Chemother 2014;58:5758–65. 432 https://doi.org/10.1128/AAC.03050-14.
- 433 [33] Salmanton-García J, Seidel D, Koehler P, Mellinghoff SC, Herbrecht R, Klimko N, et al.
 434 Matched-paired analysis of patients treated for invasive mucormycosis: standard treatment
 435 versus posaconazole new formulations (MoveOn). J Antimicrob Chemother 2019;74:3315–27.
 436 https://doi.org/10.1093/jac/dkz344.
- Van Daele R, Spriet I, Wauters J, Maertens J, Mercier T, Van Hecke S, et al. Antifungal drugs:
 What brings the future? Med Mycol 2019;57:S328–43. https://doi.org/10.1093/mmy/myz012.
- 439 [35] Lamoth F, Alexander BD. Antifungal activities of SCY-078 (MK-3118) and standard antifungal 440 agents against clinical non-Aspergillus mold isolates. Antimicrob Agents Chemother 441 2015;59:4308–11. https://doi.org/10.1128/AAC.00234-15.
- Jørgensen KM, Astvad KMT, Hare RK, Arendrup MC. EUCAST Determination of Olorofim
 (F901318) Susceptibility of Mold Species, Method Validation, and MICs. Antimicrob Agents
 Chemother 2018;62. https://doi.org/10.1128/AAC.00487-18.
- Gebremariam T, Wiederhold NP, Fothergill AW, Garvey EP, Hoekstra WJ, Schotzinger RJ, et al.
 VT-1161 Protects Immunosuppressed Mice from Rhizopus arrhizus var. arrhizus Infection.
 Antimicrob Agents Chemother 2015;59:7815–7. https://doi.org/10.1128/AAC.01437-15.
- 448 [38] Gebremariam T, Alkhazraji S, Lin L, Wiederhold NP, Garvey EP, Hoekstra WJ, et al. Prophylactic
 449 Treatment with VT-1161 Protects Immunosuppressed Mice from Rhizopus arrhizus var.
 450 arrhizus Infection. Antimicrob Agents Chemother 2017;61.
 451 https://doi.org/10.1128/AAC.00390-17.
- 452 [39] Goldani LZ, Sugar AM. Treatment of murine pulmonary mucormycosis with SCH 42427, a 453 broad-spectrum triazole antifungal drug. J Antimicrob Chemother 1994;33:369–72. 454 https://doi.org/10.1093/jac/33.2.369.
- 455 [40] Miyazaki M, Horii T, Hata K, Watanabe N-A, Nakamoto K, Tanaka K, et al. In vitro activity of E1210, a novel antifungal, against clinically important yeasts and molds. Antimicrob Agents Chemother 2011;55:4652–8. https://doi.org/10.1128/AAC.00291-11.
- 458 [41] Rivero-Menendez O, Cuenca-Estrella M, Alastruey-Izquierdo A. In vitro activity of APX001A 459 against rare moulds using EUCAST and CLSI methodologies. J Antimicrob Chemother 460 2019;74:1295–9. https://doi.org/10.1093/jac/dkz022.
- 461 [42] Gebremariam T, Alkhazraji S, Alqarihi A, Wiederhold NP, Shaw KJ, Patterson TF, et al.
 462 Fosmanogepix (APX001) Is Effective in the Treatment of Pulmonary Murine Mucormycosis
 463 Due to Rhizopus arrhizus. Antimicrob Agents Chemother 2020;64.
 464 https://doi.org/10.1128/AAC.00178-20.
- 465 [43] Colley T, Sehra G, Chowdhary A, Alanio A, Kelly SL, Kizawa Y, et al. In Vitro and In Vivo Efficacy
 466 of a Novel and Long-Acting Fungicidal Azole, PC1244, on Aspergillus fumigatus Infection.
 467 Antimicrob Agents Chemother 2018;62. https://doi.org/10.1128/AAC.01941-17.
- 468 [44] Ben-Ami R, Lewis RE, Tarrand J, Leventakos K, Kontoyiannis DP. Antifungal activity of colistin 469 against mucorales species in vitro and in a murine model of Rhizopus oryzae pulmonary 470 infection. Antimicrob Agents Chemother 2010;54:484–90. 471 https://doi.org/10.1128/AAC.00956-09.
- 472 [45] Lamoth F, Kontoyiannis DP. Therapeutic Challenges of Non-Aspergillus Invasive Mold
 473 Infections in Immunosuppressed Patients. Antimicrob Agents Chemother 2019;63.
 474 https://doi.org/10.1128/AAC.01244-19.
- 475 [46] Baistrocchi SR, Lee MJ, Lehoux M, Ralph B, Snarr BD, Robitaille R, et al. Posaconazole-Loaded 476 Leukocytes as a Novel Treatment Strategy Targeting Invasive Pulmonary Aspergillosis. J Infect 477 Dis 2017;215:1734–41. https://doi.org/10.1093/infdis/jiw513.
- 478 [47] Kumaresan PR, Manuri PR, Albert ND, Maiti S, Singh H, Mi T, et al. Bioengineering T cells to 479 target carbohydrate to treat opportunistic fungal infection. Proc Natl Acad Sci USA 480 2014;111:10660–5. https://doi.org/10.1073/pnas.1312789111.

- 481 [48] Gebremariam T, Alkhazraji S, Soliman SSM, Gu Y, Jeon HH, Zhang L, et al. Anti-CotH3
 482 antibodies protect mice from mucormycosis by prevention of invasion and augmenting
 483 opsonophagocytosis. Sci Adv 2019;5:eaaw1327. https://doi.org/10.1126/sciadv.aaw1327.
- Liu M, Spellberg B, Phan QT, Fu Y, Lee AS, et al. The endothelial cell receptor GRP78 is required for mucormycosis pathogenesis in diabetic mice. J Clin Invest 2010;120:1914–24. https://doi.org/10.1172/JCI42164.
- 487 [50] Brunet K, Alanio A, Lortholary O, Rammaert B. Reactivation of dormant/latent fungal infection. J Infect 2018;77:463–8. https://doi.org/10.1016/j.jinf.2018.06.016.
- 489 [51] Inglesfield S, Jasiulewicz A, Hopwood M, Tyrrell J, Youlden G, Mazon-Moya M, et al. Robust 490 Phagocyte Recruitment Controls the Opportunistic Fungal Pathogen Mucor circinelloides in 491 Innate Granulomas In Vivo. MBio 2018;9. https://doi.org/10.1128/mBio.02010-17.
- 492 [52] Sheldon WH, Bauer H. Activation of quiescent mucormycotic granulomata in rabbits by 493 induction of acute alloxan diabetes. J Exp Med 1958;108:171–8. 494 https://doi.org/10.1084/jem.108.1.171.
- 495 [53] Brunet T, Brunet K, Jouvion G, Cateau E, Marchand S, Rammaert B. Lichtheimia corymbifera
 496 Colonization Leading to Pulmonary Infection Can Be Prevented with Liposomal Amphotericin B
 497 in a New Murine Model. Antimicrob Agents Chemother 2019;63.
 498 https://doi.org/10.1128/AAC.02544-18.
- Jeong W, Keighley C, Wolfe R, Lee WL, Slavin MA, Chen SC-A, et al. Contemporary
 management and clinical outcomes of mucormycosis: A systematic review and meta-analysis
 of case reports. Int J Antimicrob Agents 2019;53:589–97.
 https://doi.org/10.1016/j.ijantimicag.2019.01.002.
- 503 [55] Schwarz P, Cornely OA, Dannaoui E. Antifungal combinations in Mucorales: A microbiological perspective. Mycoses 2019;62:746–60. https://doi.org/10.1111/myc.12909.
- 505 [56] Zhang S, Li R, Yu J. Drug combinations against Mucor irregularis in vitro. Antimicrob Agents Chemother 2013;57:3395–7. https://doi.org/10.1128/AAC.02612-12.
- 507 [57] Guembe M, Guinea J, Peláez T, Torres-Narbona M, Bouza E. Synergistic effect of posaconazole 508 and caspofungin against clinical zygomycetes. Antimicrob Agents Chemother 2007;51:3457–8. 509 https://doi.org/10.1128/AAC.00595-07.
- [58] Gebremariam T, Wiederhold NP, Alqarihi A, Uppuluri P, Azie N, Edwards JE, et al.
 Monotherapy or combination therapy of isavuconazole and micafungin for treating murine
 mucormycosis. J Antimicrob Chemother 2017;72:462–6. https://doi.org/10.1093/jac/dkw433.
- Lewis RE, Albert ND, Kontoyiannis DP. Comparative pharmacodynamics of posaconazole in
 neutropenic murine models of invasive pulmonary aspergillosis and mucormycosis.
 Antimicrob Agents Chemother 2014;58:6767–72. https://doi.org/10.1128/AAC.03569-14.
- Ibrahim AS, Gebremariam T, Fu Y, Edwards JE, Spellberg B. Combination echinocandin-polyene
 treatment of murine mucormycosis. Antimicrob Agents Chemother 2008;52:1556–8.
 https://doi.org/10.1128/AAC.01458-07.
- [61] Spellberg B, Fu Y, Edwards JE, Ibrahim AS. Combination therapy with amphotericin B lipid
 complex and caspofungin acetate of disseminated zygomycosis in diabetic ketoacidotic mice.
 Antimicrob Agents Chemother 2005;49:830–2. https://doi.org/10.1128/AAC.49.2.830 832.2005.
- 523 [62] Abidi MZ, Sohail MR, Cummins N, Wilhelm M, Wengenack N, Brumble L, et al. Stability in the 524 cumulative incidence, severity and mortality of 101 cases of invasive mucormycosis in high-525 risk patients from 1995 to 2011: a comparison of eras immediately before and after the 526 availability of voriconazole and echinocandin-amphotericin combination therapies. Mycoses 527 2014;57:687–98. https://doi.org/10.1111/myc.12222.
- Kyvernitakis A, Torres HA, Jiang Y, Chamilos G, Lewis RE, Kontoyiannis DP. Initial use of combination treatment does not impact survival of 106 patients with haematologic
 malignancies and mucormycosis: a propensity score analysis. Clin Microbiol Infect
 2016;22:811.e1-811.e8. https://doi.org/10.1016/j.cmi.2016.03.029.

- van Burik J-AH, Hare RS, Solomon HF, Corrado ML, Kontoyiannis DP. Posaconazole is effective as salvage therapy in zygomycosis: a retrospective summary of 91 cases. Clin Infect Dis 2006;42:e61-65. https://doi.org/10.1086/500212.
- Lanternier F, Dannaoui E, Morizot G, Elie C, Garcia-Hermoso D, Huerre M, et al. A global analysis of mucormycosis in France: the RetroZygo Study (2005-2007). Clin Infect Dis 2012;54 Suppl 1:S35-43. https://doi.org/10.1093/cid/cir880.
- Fig. 18 Reed C, Bryant R, Ibrahim AS, Edwards J, Filler SG, Goldberg R, et al. Combination polyenecaspofungin treatment of rhino-orbital-cerebral mucormycosis. Clin Infect Dis 2008;47:364– 71. https://doi.org/10.1086/589857.
- [67] Ibrahim AS, Gebermariam T, Fu Y, Lin L, Husseiny MI, French SW, et al. The iron chelator
 deferasirox protects mice from mucormycosis through iron starvation. J Clin Invest
 2007;117:2649–57. https://doi.org/10.1172/JCI32338.
- Ibrahim AS, Gebremariam T, Luo G, Fu Y, French SW, Edwards JE, et al. Combination therapy
 of murine mucormycosis or aspergillosis with iron chelation, polyenes, and echinocandins.
 Antimicrob Agents Chemother 2011;55:1768–70. https://doi.org/10.1128/AAC.01577-10.
- 547 [69] Dannaoui E, Schwarz P, Lortholary O. In vitro interactions between antifungals and 548 immunosuppressive drugs against zygomycetes. Antimicrob Agents Chemother 549 2009;53:3549–51. https://doi.org/10.1128/AAC.00184-09.
- Lewis RE, Ben-Ami R, Best L, Albert N, Walsh TJ, Kontoyiannis DP. Tacrolimus enhances the potency of posaconazole against Rhizopus oryzae in vitro and in an experimental model of mucormycosis. J Infect Dis 2013;207:834–41. https://doi.org/10.1093/infdis/jis767.
- 553 [71] Schwarz P, Schwarz PV, Felske-Zech H, Dannaoui E. In vitro interactions between
 554 isavuconazole and tacrolimus, cyclosporin A or sirolimus against Mucorales. J Antimicrob
 555 Chemother 2019;74:1921–7. https://doi.org/10.1093/jac/dkz102.
- Pfaller MA, Messer SA, Georgopapadakou N, Martell LA, Besterman JM, Diekema DJ. Activity
 of MGCD290, a Hos2 Histone Deacetylase Inhibitor, in Combination with Azole Antifungals
 against Opportunistic Fungal Pathogens. J Clin Microbiol 2009;47:3797–804.
 https://doi.org/10.1128/JCM.00618-09.
- 560 [73] Chamilos G, Lewis RE, Kontoyiannis DP. Lovastatin has significant activity against zygomycetes 561 and interacts synergistically with voriconazole. Antimicrob Agents Chemother 2006;50:96– 562 103. https://doi.org/10.1128/AAC.50.1.96-103.2006.
- 563 [74] Sugar AM, Liu XP. Combination antifungal therapy in treatment of murine pulmonary 564 mucormycosis: roles of quinolones and azoles. Antimicrob Agents Chemother 2000;44:2004– 565 6. https://doi.org/10.1128/aac.44.7.2004-2006.2000.
- 566 [75] Biswas C, Sorrell TC, Djordjevic JT, Zuo X, Jolliffe KA, Chen SC-A. In vitro activity of miltefosine
 567 as a single agent and in combination with voriconazole or posaconazole against uncommon
 568 filamentous fungal pathogens. J Antimicrob Chemother 2013;68:2842–6.
 569 https://doi.org/10.1093/jac/dkt282.
- 570 [76] Dannaoui E, Afeltra J, Meis JFGM, Verweij PE, Eurofung Network. In vitro susceptibilities of zygomycetes to combinations of antimicrobial agents. Antimicrob Agents Chemother 2002;46:2708–11. https://doi.org/10.1128/aac.46.8.2708-2711.2002.
- 573 [77] Christenson JC, Shalit I, Welch DF, Guruswamy A, Marks MI. Synergistic action of amphotericin 574 B and rifampin against Rhizopus species. Antimicrob Agents Chemother 1987;31:1775–8. 575 https://doi.org/10.1128/aac.31.11.1775.
- 576 [78] Spellberg B, Ibrahim A, Roilides E, Lewis RE, Lortholary O, Petrikkos G, et al. Combination 577 Therapy for Mucormycosis: Why, What, and How? Clin Infect Dis 2012;54:S73–8. 578 https://doi.org/10.1093/cid/cir885.
- 579 [79] Lin E, Moua T, Limper AH. Pulmonary mucormycosis: clinical features and outcomes. Infection 2017;45:443–8. https://doi.org/10.1007/s15010-017-0991-6.
- Tedder M, Spratt JA, Anstadt MP, Hegde SS, Tedder SD, Lowe JE. Pulmonary mucormycosis: results of medical and surgical therapy. Ann Thorac Surg 1994;57:1044–50.

- 583 [81] Gamaletsou MN, Sipsas NV, Roilides E, Walsh TJ. Rhino-orbital-cerebral mucormycosis. Curr 584 Infect Dis Rep 2012;14:423–34. https://doi.org/10.1007/s11908-012-0272-6.
- Vironneau P, Kania R, Morizot G, Elie C, Garcia-Hermoso D, Herman P, et al. Local control of
 rhino-orbito-cerebral mucormycosis dramatically impacts survival. Clin Microbiol Infect
 2014;20:O336-339. https://doi.org/10.1111/1469-0691.12408.
- 588 [83] Manesh A, Rupali P, Sullivan MO, Mohanraj P, Rupa V, George B, et al. Mucormycosis-A 589 clinicoepidemiological review of cases over 10 years. Mycoses 2019;62:391–8. 590 https://doi.org/10.1111/myc.12897.
- 591 [84] Gil-Lamaignere C, Simitsopoulou M, Roilides E, Maloukou A, Winn RM, Walsh TJ. Interferon-592 gamma and granulocyte-macrophage colony-stimulating factor augment the activity of 593 polymorphonuclear leukocytes against medically important zygomycetes. J Infect Dis 594 2005;191:1180–7. https://doi.org/10.1086/428503.
- Liles WC, Huang JE, van Burik JA, Bowden RA, Dale DC. Granulocyte colony-stimulating factor
 administered in vivo augments neutrophil-mediated activity against opportunistic fungal
 pathogens. J Infect Dis 1997;175:1012–5. https://doi.org/10.1086/513961.
- 598 [86] Saoulidis S, Simitsopoulou M, Dalakiouridou M, Walsh TJ, Wheat LJ, Papaioannidou P, et al.
 599 Antifungal activity of posaconazole and granulocyte colony-stimulating factor in the treatment
 600 of disseminated zygomycosis (mucormycosis) in a neutropaenic murine model. Mycoses
 601 2011;54:e486-492. https://doi.org/10.1111/j.1439-0507.2010.01958.x.
- 602 [87] Rodríguez MM, Calvo E, Mariné M, Pastor FJ, Fernandez-Ballart J, Guarro J. Efficacy of
 603 liposomal amphotericin B combined with gamma interferon or granulocyte-macrophage
 604 colony-stimulating factor for treatment of systemic zygomycosis in mice. Antimicrob Agents
 605 Chemother 2009;53:3569–71. https://doi.org/10.1128/AAC.00456-09.
- 606 [88] Ma B, Seymour JF, Januszewicz H, Slavin MA. Cure of pulmonary Rhizomucor pusillus infection 607 in a patient with hairy-cell leukemia: role of liposomal amphotericin B and GM-CSF. Leuk 608 Lymphoma 2001;42:1393–9. https://doi.org/10.3109/10428190109097768.
- Sahin B, Paydaş S, Coşar E, Biçakçi K, Hazar B. Role of granulocyte colony-stimulating factor in
 the treatment of mucormycosis. Eur J Clin Microbiol Infect Dis 1996;15:866–9.
 https://doi.org/10.1007/bf01691218.
- 612 [90] Garcia-Diaz JB, Palau L, Pankey GA. Resolution of rhinocerebral zygomycosis associated with 613 adjuvant administration of granulocyte-macrophage colony-stimulating factor. Clin Infect Dis 614 2001;32:e145-150. https://doi.org/10.1086/320767.
- 615 [91] Abzug MJ, Walsh TJ. Interferon-gamma and colony-stimulating factors as adjuvant therapy for 616 refractory fungal infections in children. Pediatr Infect Dis J 2004;23:769–73. 617 https://doi.org/10.1097/01.inf.0000134314.65398.bf.
- 518 [92] Symeonidis AS. The role of iron and iron chelators in zygomycosis. Clin Microbiol Infect 2009;15 Suppl 5:26–32. https://doi.org/10.1111/j.1469-0691.2009.02976.x.
- [93] Ibrahim AS, Edwards JE, Fu Y, Spellberg B. Deferiprone iron chelation as a novel therapy for
 experimental mucormycosis. J Antimicrob Chemother 2006;58:1070–3.
 https://doi.org/10.1093/jac/dkl350.
- [94] Ibrahim AS, Gebremariam T, Luo G, Fu Y, French SW, Edwards JE, et al. Combination therapy
 of murine mucormycosis or aspergillosis with iron chelation, polyenes, and echinocandins.
 Antimicrob Agents Chemother 2011;55:1768–70. https://doi.org/10.1128/AAC.01577-10.
- [95] Leonardelli F, Macedo D, Dudiuk C, Theill L, Cabeza MS, Gamarra S, et al. In Vitro Activity of
 Combinations of Zinc Chelators with Amphotericin B and Posaconazole against Six Mucorales
 Species. Antimicrob Agents Chemother 2019;63. https://doi.org/10.1128/AAC.00266-19.
- 629 [96] Tragiannidis A, Groll AH. Hyperbaric oxygen therapy and other adjunctive treatments for 630 zygomycosis. Clin Microbiol Infect 2009;15 Suppl 5:82–6. https://doi.org/10.1111/j.1469-631 0691.2009.02986.x.
- 632 [97] Barratt DM, Van Meter K, Asmar P, Nolan T, Trahan C, Garcia-Covarrubias L, et al. Hyperbaric oxygen as an adjunct in zygomycosis: randomized controlled trial in a murine model.

- Antimicrob Agents Chemother 2001;45:3601–2. https://doi.org/10.1128/AAC.45.12.3601-3602.2001.
- [98] John BV, Chamilos G, Kontoyiannis DP. Hyperbaric oxygen as an adjunctive treatment for
 zygomycosis. Clin Microbiol Infect 2005;11:515–7. https://doi.org/10.1111/j.1469 0691.2005.01170.x.
- [99] Takazono T, Izumikawa K, Mihara T, Kosai K, Saijo T, Imamura Y, et al. Efficacy of combination
 antifungal therapy with intraperitoneally administered micafungin and aerosolized liposomal
 amphotericin B against murine invasive pulmonary aspergillosis. Antimicrob Agents
 Chemother 2009;53:3508–10. https://doi.org/10.1128/AAC.00285-09.
- 643 [100] Mihara T, Kakeya H, Izumikawa K, Obata Y, Nishino T, Takazono T, et al. Efficacy of aerosolized 644 liposomal amphotericin B against murine invasive pulmonary mucormycosis. J Infect 645 Chemother 2014;20:104–8. https://doi.org/10.1016/j.jiac.2013.09.002.
- [101] Alfageme I, Reina A, Gallego J, Reyes N, Torres A. Endobronchial instillations of amphotericin
 B: complementary treatment for pulmonary mucormycosis. J Bronchology Interv Pulmonol
 2009;16:214–5. https://doi.org/10.1097/LBR.0b013e3181aa2583.
- Furco A, Mouchet B, Carbonnelle M, Vallerand H. [Pulmonary mucormycosis: benefit of aerosol amphotericin B?]. Rev Mal Respir 2001;18:309–13.
- 651 [103] McGuire FR, Grinnan DC, Robbins M. Mucormycosis of the bronchial anastomosis: a case of 652 successful medical treatment and historic review. J Heart Lung Transplant 2007;26:857–61. 653 https://doi.org/10.1016/j.healun.2007.05.010.
- [104] Safdar A, O'Brien S, Kouri IF. Efficacy and feasibility of aerosolized amphotericin B lipid
 complex therapy in caspofungin breakthrough pulmonary zygomycosis. Bone Marrow
 Transplantation 2004;34:467–8. https://doi.org/10.1038/sj.bmt.1704552.
- [105] Safdar A, Rodriguez GH. Aerosolized amphotericin B lipid complex as adjunctive treatment for fungal lung infection in patients with cancer-related immunosuppression and recipients of hematopoietic stem cell transplantation. Pharmacotherapy 2013;33:1035–43.
 https://doi.org/10.1002/phar.1309.
- [106] Di Pentima MC, Chan S, Powell J, Napoli JA, Walter AW, Walsh TJ. Topical amphotericin B in combination with standard therapy for severe necrotizing skin and soft-tissue mucormycosis in an infant with bilineal leukemia: case report and review. J Pediatr Hematol Oncol 2014;36:e468-470. https://doi.org/10.1097/MPH.000000000000166.
- [107] Cohen-Ludmann C, Kerob D, Feuilhade M, Chaine B, Guermazi A, Janier M, et al. Zygomycosis
 of the penis due to Rhizopus oryzae successfully treated with surgical debridement and a
 combination of high-dose liposomal and topical amphotericin B. Arch Dermatol
 2006;142:1657–8. https://doi.org/10.1001/archderm.142.12.1657.
- [108] Steve AK, Hurdle VA, Brown JY. Orbitomaxillofacial Mucormycosis Requiring Complex
 Multifactorial Management. Plast Reconstr Surg Glob Open 2018;6:e1927.
 https://doi.org/10.1097/GOX.000000000001927.
- [109] Devauchelle P, Jeanne M, Fréalle E. Mucormycosis in Burn Patients. J Fungi (Basel) 2019;5.
 https://doi.org/10.3390/jof5010025.
- Thielen BK, Barnes AMT, Sabin AP, Huebner B, Nelson S, Wesenberg E, et al. Widespread
 Lichtheimia Infection in a Patient with Extensive Burns: Opportunities for Novel Antifungal
 Agents. Mycopathologia 2019;184:121–8. https://doi.org/10.1007/s11046-018-0281-6.
- [111] Farmer AR, Murray CK, Driscoll IR, Wickes BL, Wiederhold N, Sutton DA, et al. Combat-Related
 Pythium aphanidermatum Invasive Wound Infection: Case Report and Discussion of Utility of
 Molecular Diagnostics. J Clin Microbiol 2015;53:1968–75. https://doi.org/10.1128/JCM.00410 15.
- 681 [112] Piazza RC, Thomas WL, Stawski WS, Ford RD. Mucormycosis of the face. J Burn Care Res 2009;30:520–3. https://doi.org/10.1097/BCR.0b013e3181a28d2f.
- 683 [113] Atty C, Alagiozian-Angelova VM, Kowal-Vern A. Black plaques and white nodules in a burn 684 patient. Fusarium and Mucormycosis. JAMA Dermatol 2014;150:1355–6. 685 https://doi.org/10.1001/jamadermatol.2014.2463.

- Tang D, Wang W. Successful cure of an extensive burn injury complicated with mucor wound sepsis. Burns 1998;24:72–3. https://doi.org/10.1016/s0305-4179(97)00099-5.
- [115] Constantinides J, Misra A, Nassab R, Wilson Y. Absidia corymbifera fungal infection in burns: a
 case report and review of the literature. J Burn Care Res 2008;29:416–9.
 https://doi.org/10.1097/BCR.0b013e318166da78.
- [116] Hussain A, Singh VK, Singh OP, Shafaat K, Kumar S, Ahmad FJ. Formulation and optimization of
 692 nanoemulsion using antifungal lipid and surfactant for accentuated topical delivery of
 693 Amphotericin B. Drug Deliv 2016;23:3101–10.
 694 https://doi.org/10.3109/10717544.2016.1153747.
- [117] Sosa L, Clares B, Alvarado HL, Bozal N, Domenech O, Calpena AC. Amphotericin B releasing
 topical nanoemulsion for the treatment of candidiasis and aspergillosis. Nanomedicine
 2017;13:2303–12. https://doi.org/10.1016/j.nano.2017.06.021.
- [118] Garcia A, Fan YY, Vellanki S, Huh EY, Vanegas D, Wang SH, et al. Nanoemulsion as an Effective
 Treatment against Human-Pathogenic Fungi. MSphere 2019;4.
 https://doi.org/10.1128/mSphere.00729-19.
- 701 [119] Mesa Varona D, Celis Sánchez J, Alfaya Muñoz L, Avendaño Cantos EM, Romero Moraleda L.
 702 Keratitis caused by Absidia corymbifera in an immunocompetent male with no corneal
 703 injuries. Arch Soc Esp Oftalmol 2015;90:139–41. https://doi.org/10.1016/j.oftal.2014.02.020.
 - [120] Anderson A, McManus D, Perreault S, Lo Y-C, Seropian S, Topal JE. Combination liposomal amphotericin B, posaconazole and oral amphotericin B for treatment of gastrointestinal Mucorales in an immunocompromised patient. Med Mycol Case Rep 2017;17:11–3. https://doi.org/10.1016/j.mmcr.2017.05.004.

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- 708 [121] Parize P, Mamez A-C, Garcia-Hermoso D, Dumaine V, Poirée S, Kauffmann-Lacroix C, et al.
 709 Successful Treatment of Saksenaea sp. Osteomyelitis by Conservative Surgery and
 710 Intradiaphyseal Incorporation of Amphotericin B Cement Beads. Antimicrob Agents
 711 Chemother 2019;63. https://doi.org/10.1128/AAC.01006-18.
 - [122] Grannan BL, Yanamadala V, Venteicher AS, Walcott BP, Barr JC. Use of external ventriculostomy and intrathecal anti-fungal treatment in cerebral mucormycotic abscess. J Clin Neurosci 2014;21:1819–21. https://doi.org/10.1016/j.jocn.2014.01.008.
- 715 [123] Fu M-H, Liu J, Liang G-Z, Li C-R, Zhu X-M, Wang L, et al. Successful Treatment of Eczema-Like 716 Mucormycosis in a Child by Combination of Intravenous Drip and Percutaneous Injection 717 Amphotericin B. Mycopathologia 2019;184:309–13. https://doi.org/10.1007/s11046-018-718 0273-6.

	ECIL-6 2017	Grade	ECMM 2019	Grade
First-line antifunga	ıl therapy			
Liposomal amphotericin B	5mg/kg	BII	5-10 mg/kg	ΑII
			For CNS involvement: 10 mg/kg	A III
Amphotericin B	without CNS involvement	BII	Any without CNS involvement	BII
lipid complex			SOT: 10 mg/kg	A III
Amphotericin B deoxycholate		CII		DII
Posaconazole		C III	DR tablet or IV: 300 mg b.i.d, day 1; 300 mg/d from day 2	BII
			Oral suspension (4 x 200 mg or 2 x 400 mg)	CII
Isavuconazole			200 mg t.i.d, day 1 -2 ; 200 mg/d from day 3	BII
Combination therapy		C III	Liposomal amphotericin B + caspofungin and/or posaconazole	C II - C III
Control of underlyi	ng conditions			
Diabetes	Control of diabetes	AII	Control of hyperglycaemia and Ketoacidosis	A III
Immuno- suppression	Discontinuation/tapering of steroids, reduction of immunosuppressive therapy	AII	Rapidly taper glucocorticosteroid dose to discontinue, if feasible, or reduce dose to minimum required	A II
Surgery				
	Rhino-orbito-cerebral infection	A II	Repeated surgery in addition to antifungal treatment	A II
	Soft tissue infection	ΑII		
	Localized pulmonary lesion	B III		
	Disseminated infection	C III		

SOT: solid organ transplantation. CNS: central nervous system. DR: delayed-release. IV: intravenous. b.i.d: twice a day. t.i.d: three times a day.

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Table 2: Prophylaxis recommendation from European Confederation of Medical Mycology (ECMM) (2019), adapted from [10].

ECMM 2019				
Primary	Neutropenic,	Posaconazole DR tablet (300 mg b.i.d day1, 300 mg/d from day2)	BII	
	GvHD	Posaconazole IV (300 mg b.i.d day1, 300 mg/d from day2)	B III	
		Posaconazole oral suspension (200 mg t.i.d)	CII	
	Neutropenic	Isavuconazole PO or IV (200 mg t.i.d day1-2, 200 mg/d from day3; or 200 mg/d from day1)	CII	
	SOT	Isavuconazole PO/IV (200 mg t.i.d day1-2, 200 mg/d from day3; or 200 mg/d from day1)	C II	
		Posaconazole IV (300 mg b.i.d day1, 300 mg/d from day2)	C III	
		Posaconazole oral suspension (200 mg t.i.d)	C III	
	All induction chemotherapy	Liposomal amphotericin B	DI	
	Neutropenic or GvHD	Fluconazole, itraconazole, voriconazole	DII	
Secondary		Last effective drug in the same patient	A III	

SOT: solid organ transplantation. DR: delayed-release. IV: intravenous. PO: per os. GvHD: graft versus host disease. d: day. b.i.d: twice a day. t.i.d: three times a day.

Table 3: Adjunctive therapy recommendations for treatment of invasive mucormycosis from European Conference on Infections in Leukemia 6 (ECIL-6) (2017) and European Confederation of Medical Mycology (ECMM) (2019), adapted from [9,10].

ECIL-6 2017		ECMM 2019		
Adjunctive therapy				
Against use of deferasirox	All	Deferasirox (other than haematology)	CII	
		Deferasirox (haematology)	DII	
		Deferoxamine	DII	
Hyperbaric oxygen	C III	Exposure to 100% hyperbaric oxygen (haematology)	C II	
		Exposure to 100% hyperbaric oxygen (diabetes)	BII	
Hematopoietic growth factor if neutropenia	A II	G-CSF (haematology, ongoing neutropenia)	BII	
		Granulocyte transfusion (haematology, ongoing neutropenia)	C II	
		Granulocyte transfusion + IFNy1b (haematology, ongoing neutropenia)	C III	
		GM-CSF (diabetes)	C III	
		Adoptive immunotherapy, T cells generated in response to <i>R. arrhizus</i> antigens	C III	
		Nivolumab + interferon-γ	C III	

G-CSF: granulocyte colony stimulating factor. GM-CSF: granulocyte macrophage colony stimulating factor.