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1 First description of bacteremia caused by Oscillibacter valericigenes in a patient

- 2 hospitalized for leg amputation.
- 3 Lauranne Broutin^{1,*}, Luc Deroche^{1,*}, Anthony Michaud¹, Gwenaël Le Moal², Christophe
- 4 Burucoa¹, Louis-Etienne Gayet², Chloé Plouzeau^{1,+}, Maxime Pichon^{1,+,∞}
- ⁵ ¹ University Hospital of Poitiers, Bacteriology and Infection Control Laboratory, Infectious
- 6 Agents Department, Poitiers, France
- ⁷² University Hospital of Poitiers, Department of Infectious Diseases, CHU, Poitiers, France.
- 8 ³ University Hospital of Poitiers, Department of Orthopedic Surgery and Traumatology, CHU,
- 9 Poitiers, France.
- 10 * These authors contributed equally to the present work
- 11 + These authors supervised equally the present work
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13 ∞ Corresponding author

- 14 Maxime PICHON, Bacteriology and Infection Control Laboratory, Infectious Agents
- 15 Department, University Hospital of Poitiers, 2 rue de la Milétrie, 86021 Poitiers Cedex,
- 16 France ; Tel : +33 (0)5.49.44.41.43 ; Fax : +33 (0)5.49.44.38.88 ; e-mail :
- 17 maxime.pichon@chu-poitiers.fr

18 Highlights :

- Culture of Oscillibacter valericigenes, a Gram-negative rod, remains very difficult
- This Bacterium was, for the first time, isolated during bacteremia
- 16S rDNA sequencing is not of interest to predict disease severity

22 Abstract

Initially isolated from the alimentary canal of a Japanese corbicula clam, *Oscillibacter valericigenes* is a Gram-negative rod, of which culture remains very difficult. Herein we
present the first case of bacteremia due to *Oscillibacter valericigenes*, in humans.

A 55-year-old man was hospitalized for clinical management of multiple neglected leg
wounds (colonized with maggots) that had occurred during a motorcycle accident. Following
radiological confirmation of the bone infection, a transfemoral amputation was performed to
limit the risk of extended infection.

30 During hospitalization, before the amputation, the patient experienced fever, biological inflammation justifying the sampling of multiple blood cultures. Anaerobic blood culture was 31 positive after 34 hours, without identification by routine procedure (MALDI-TOF), justifying 32 identification by 16S DNA sequencing. In the absence of possible subculture, antibiotic 33 sensitivity testing could not be performed. A pre-emptive treatment by piperacillin-34 35 tazobactam was introduced for 14 days. The evolution was good, except for a local disunion. Complete phylogenic analysis of the clinical strain showed that it significantly differed from 36 37 the reference strain, which is distantly related to the Clostridia cluster IV. Due to the culture 38 conditions and specialized identification method by sequencing, prevalence of O. 39 valericigenes may be underestimated. Optimization of blood culture procedures and utilization of 16S rRNA gene sequencing are tools needed for identification of rare pathogens 40 that could help to optimize clinical management of infected patients. 41

42 Introduction

The endospore-forming anaerobic Gram-positive clostridial bacteria are extremely
heterogeneous phylogenetically. They form nineteen clusters based on their 16S rDNA gene
sequence [1]. Within these clusters, the fourth (Clostridial cluster IV) is also phenotypically
heterogeneous. The bacteria belonging to this cluster are present mostly in the alimentary
canal and feces of numerous hosts as wood-feeding termites, anaerobic sewage sludge and
anaerobic digesters.

Oscillibacter valericigenes is a Gram-negative rod, motile due to a peritrichous flagella that
optimally grows at 30°C (range 15-35°C) [2]. This bacterium remains very difficult to culture.
This could explain the underrepresentation of its clinical impact in humans.

52 Herein we present the first case of bacteremia due to Oscillibacter valericigenes in humans.

53 Case Study

A 55-year-old man was hospitalized in the orthopedic department (University Hospital of Poitiers, Poitiers, France) for clinical management of multiple chronic wounds on the left leg that had occurred one year before during a motorcycle accident. Mostly on account of his depression and alcoholism, these wounds were neglected, leading to multiple necrosis and fibroses. Given the overall aspect of the leg and radiological confirmation of the bone infection, it was decided to perform a transfemoral amputation to limit the risk of bacteremia and extension of the infection.

During the hospitalization for pre-resection check-up, it was locally observed that the chronic
wounds were colonized with maggots. Periosteal tibial and fibular affections (evocative of
bone infection) were observed and vascular echography was abnormal, with decreased arterial
flow. Before the amputation, the patient experienced fever (39°C) and biological

inflammation (increase in neutrophil counts ie. 15220 neutrophils per mm³ for 17700 65 leukocytes per mm³, elevation of C-Reactive protein *ie* 302 mg/l, associated with 66 inflammatory anemia *ie* 5g/L, necessitating multiple blood transfusions). Given this septical 67 presentation, multiple blood cultures were sampled and anaerobic blood vials were positive 68 after 34 hours of culture. Bacteria could not be identified by routine procedure using MALDI-69 70 TOF (bioMérieux, Marcy-l'étoile, France), justifying the need for 16S DNA sequencing. 71 Given subculture failure, Antimicrobial Susceptibility Testing could not be performed. A 72 probabilistic treatment by Piperacillin-Tazobactam (4 grams three times a day) was introduced for a 14-day post-operative period. Evolution was good, except for local disunion 73 74 of the scar due to the patient's agitation. 75 Sanger sequencing of the 16S rDNA V1-V3 regions was performed on 3500 Dx Genetic 76 Analyzer (ThermoFisher Waltham, MA, USA). Obtained sequences were then compared to other available ones, using the LeBibi website to identify Oscillibacter valericigenes [3]. 77

A complete phylogenic analysis of the strain based on 16S analysis showed that this clinical 78 strain differs from the reference strain (Sjm18-20^T) with a pairwise alignment score of 79 7862963, with dissimilarities on 9 nucleotides (2.34%) out of 384 nucleotide coordinates (24 80 nucleotides for Oscillispira guillermondi on the same coordinates). Phylogenic analysis is 81 82 summarized on Figure 1. Unlike some other pathogens, no differentiation could be observed between pathogenic or symbiotic strains based on exclusively phylogenic analyses of the 16S 83 rDNA gene [4]. Sequences were archives on Sequence Read archives under the accession 84 number n° SRR9717275. 85

86 Discussion

We described herein the first case of severe human infection caused by a very rare bacterium,*Oscillibacter valericigenes*.

Described for the first time in 2007, in Japan (Shimane Prefecture), the Clostridial bacterium 89 90 is a Gram-negative, non-sporulating strictly anaerobic rod that belongs to the Firmicutes phylum and the Oscillospiraceae family [2,5]. A member of the Clostridial cluster IV, one of 91 the clusters based on 16SrRNA gene sequences, this bacterium is distantly related to other 92 members (similarity values less than 91%, on the whole-sequenced genome of more than 4.41 93 Megabases), and most similar to those of uncultured bacteria in animal digestive tracts and 94 95 feces [1]. It is very interesting to note that phylogenetic analysis using protein-coding genes (such as *ileS*, *valS*, *gyrB* and *rplKLM*) allowed classification of *O*. *valericigenes* reference 96 strain Sjm18-20^T within cluster IV. As the most closely neighboring anaerobic species 97 98 described up until now (in our phylogenic analysis of the clinical strain), Oscillispira guillermondii differ from Oscillibacter sp. strain due to several properties, particularly the 99 capacity to generate multicellular structure and endospore, which is impossible for 100 101 Oscillibacter valericigenes [6].

This bacterium was initially isolated from the alimentary canal of a Japanese corbicula clam
(*Corbicula japonica*) sampled on a seacoast. Similarly, the habitat of anaerobic bacteria
belonging to Clostridial cluster IV consists in the alimentary canal and feces of numerous
organisms including wood-feeding termites, anaerobic sewage sludge and anaerobic digesters. *O. valericigens* has also been detected physiologically in human intestinal microbiota. Its
association with Crohn's disease suggests its use in clinical diagnosis and/or therapies [7,8].

In the present case, the wounds of the patient were colonized with maggots, which may havebeen the inoculation source of the infection. Bacteraemia involving rare bacteria (like

110 Ignatzschineria indica or Wohlfahrtiimonas chitiniclastica) have previously been described in

111 patients with chronic wounds colonized by maggots [9,10]. Poor personal hygiene,

112 alcoholism, vascular disease, chronic wound or maggot colonisation have been identified as

113 risk factors for infections due to these bacteria. As described in our case study, identification

and culture of *O. valericigenes* is difficult, as 16S rDNA was mandatory to identify the
bacteria, and its high susceptibility to oxygen led to non-recovery of a live bacterial strain.
Databases of the MALDI-TOF instruments commonly used for rapid diagnosis should be
improved in order to allow laboratories not using costly methods (16s rRNA gene
sequencing...) to make the diagnosis.

In conclusion, due to the difficult culture conditions and specialized identification method by 119 sequencing, the prevalence of O. valericigenes bacteremia may be underestimated. Medical 120 biologists and clinicians should pay more attention to rare bacteria such as these in order to 121 clearly determine their clinical significance. Finally, optimization of blood culture procedures 122 (allowing study in an anaerobic atmosphere using an automatic device) and utilization of 16S 123 124 rRNA gene sequencing are powerful tools for rare pathogen identification from blood and 125 other sterile body fluids; as such, they should be widely utilized in view of optimizing clinical management of infected patients. 126

127 Conflict of interest: none

128 Authors' contributions :

129 L.B.: analyzed and wrote the manuscript; L.D.: analyzed and wrote the manuscript; A.M.:

130 ensured clinical management of the patient ; G.L.M.: ensured clinical management of the

- 131 patient ; C.B.: ensured clinical management of the patient ; L-E.G.: ensured clinical
- 132 management of the patient ; C.P.: collected the data, wrote the manuscript ; M.P.: collected
- 133 the data, analyses, wrote the manuscript
- 134 All the authors approved the final version of the article.

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139 **References**

- 140 [1] M.D. Collins, P.A. Lawson, A. Willems, J.J. Cordoba, J. Fernandez-Garayzabal, P.
- 141 Garcia, J. Cai, H. Hippe, J.A. Farrow, The phylogeny of the genus Clostridium: proposal
- 142 of five new genera and eleven new species combinations, Int. J. Syst. Bacteriol. 44
- 143 (1994) 812–826. https://doi.org/10.1099/00207713-44-4-812.
- 144 [2] T. Lino, K. Mori, K. Tanaka, K. -i. Suzuki, S. Harayama, Oscillibacter valericigenes
- gen. nov., sp. nov., a valerate-producing anaerobic bacterium isolated from the
- alimentary canal of a Japanese corbicula clam, Int. J. Syst. Evol. Microbiol. 57 (2007)

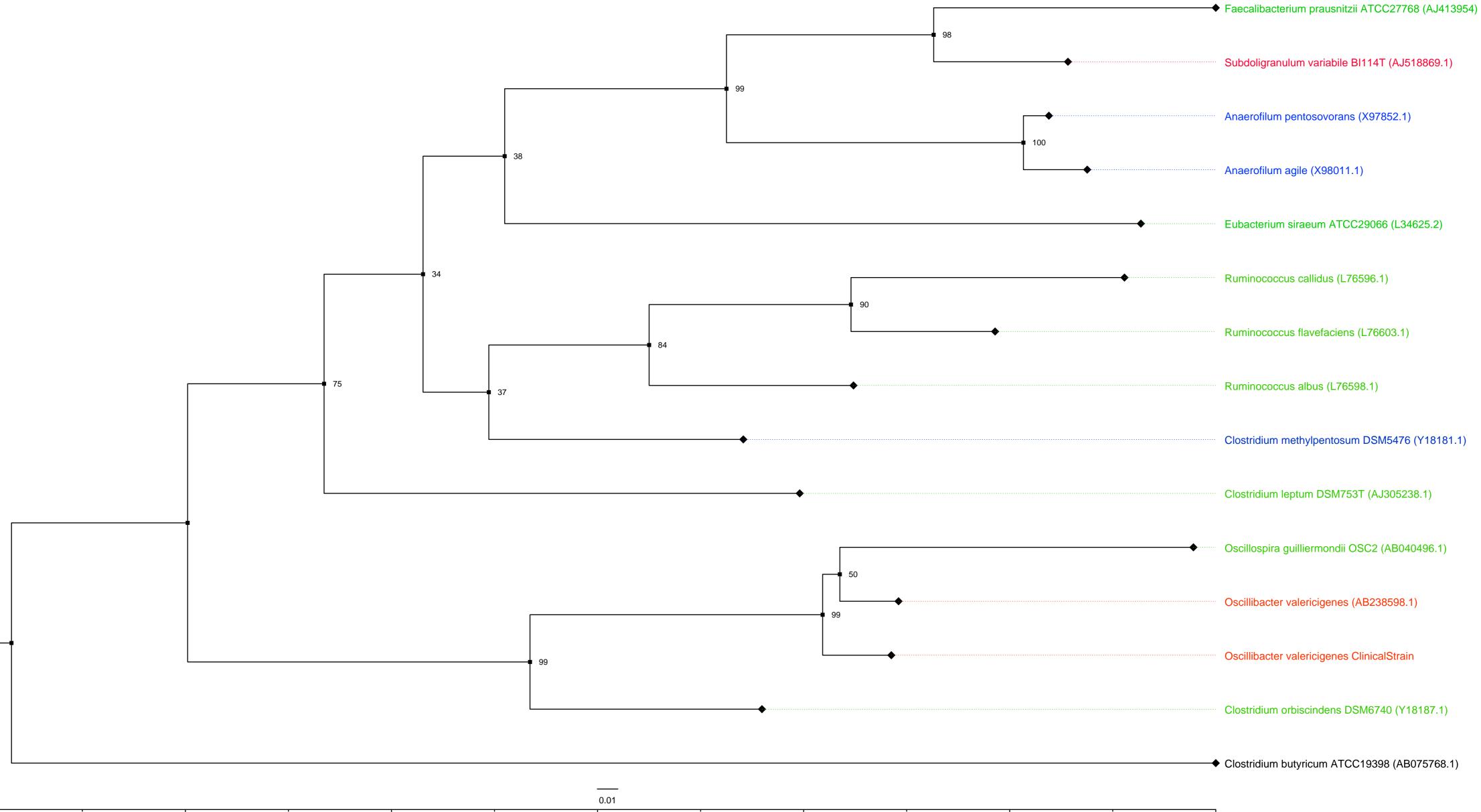
147 1840–1845. https://doi.org/10.1099/ijs.0.64717-0.

- 148 [3] J.-P. Flandrois, G. Perrière, M. Gouy, leBIBIQBPP: a set of databases and a webtool for
- automatic phylogenetic analysis of prokaryotic sequences, BMC Bioinformatics. 16
- 150 (2015) 251. https://doi.org/10.1186/s12859-015-0692-z.
- 151 [4] B. Simon, M. Pichon, M. Valette, G. Burfin, M. Richard, B. Lina, L. Josset, Whole
- 152 Genome Sequencing of A(H3N2) Influenza Viruses Reveals Variants Associated with
- 153 Severity during the 2016⁻2017 Season, Viruses. 11 (2019).
- 154 https://doi.org/10.3390/v11020108.
- 155 [5] Y. Katano, S. Fujinami, A. Kawakoshi, H. Nakazawa, S. Oji, T. Iino, A. Oguchi, A.
- 156 Ankai, S. Fukui, Y. Terui, S. Kamata, T. Harada, S. Tanikawa, K.-I. Suzuki, N. Fujita,
- 157 Complete genome sequence of Oscillibacter valericigenes Sjm18-20(T) (=NBRC
- 158 101213(T)), Stand. Genomic Sci. 6 (2012) 406–414.
- 159 https://doi.org/10.4056/sigs.2826118.

- 160 [6] K. Yanagita, A. Manome, X.-Y. Meng, S. Hanada, T. Kanagawa, T. Tsuchida, R.I.
- 161 Mackie, Y. Kamagata, Flow cytometric sorting, phylogenetic analysis and in situ
- detection of Oscillospira guillermondii, a large, morphologically conspicuous but
- uncultured ruminal bacterium, Int. J. Syst. Evol. Microbiol. 53 (2003) 1609–1614.
- 164 https://doi.org/10.1099/ijs.0.02541-0.
- 165 [7] S. Mondot, S. Kang, J.P. Furet, D. Aguirre de Carcer, C. McSweeney, M. Morrison, P.
- 166 Marteau, J. Doré, M. Leclerc, Highlighting new phylogenetic specificities of Crohn's
- disease microbiota, Inflamm. Bowel Dis. 17 (2011) 185–192.
- 168 https://doi.org/10.1002/ibd.21436.
- 169 [8] S.M. Man, N.O. Kaakoush, H.M. Mitchell, The role of bacteria and pattern-recognition
- receptors in Crohn's disease, Nat. Rev. Gastroenterol. Hepatol. 8 (2011) 152–168.
- 171 https://doi.org/10.1038/nrgastro.2011.3.
- 172 [9] H. Muse, R.L. Jenkins, M.B. Oliver, S. Kim, R.L. Grantier, B.K. Malhotra, J.J. Parham,
- 173 K.R. Stover, A Case of Ignatzschineria indica Bacteremia following Maggot
- 174 Colonization, Case Rep. Infect. Dis. 2017 (2017) 3698124.
- 175 https://doi.org/10.1155/2017/3698124.
- 176 [10] Y. Katanami, S. Kutsuna, M. Nagashima, S. Takaya, K. Yamamoto, N. Takeshita, K.
- 177 Hayakawa, Y. Kato, S. Kanagawa, N. Ohmagari, Wohlfahrtiimonas chitiniclastica
- 178 Bacteremia Hospitalized Homeless Man with Squamous Cell Carcinoma, Emerg. Infect.
- 179 Dis. 24 (2018) 1746–1748. https://doi.org/10.3201/eid2409.170080.

180 Table and Figure

Figure 1. Phylogenic analysis highlighting the position of the clinical strain relative to 181 182 other representative strains within the Clostridial cluster IV. The phylogenic tree was constructed by the ML-Method based on the alignment of 1,614 informative sites of 16S 183 rDNA gene sequences. Corresponding NCBI accession numbers are shown in parentheses. 184 Numbers at nodes indicate support values obtained from 1,000 bootstrap replications. 185 186 Clostridium butyricum does not belong to Clostridia cluster IV and was used as an outgroup. The analysis was performed using the V1-V3 consensus sequence obtained by sequencing the 187 188 cultured strain of the bacteria. In blue, the bacterial strains never associated with a human clinical outcome or pathological status are represented. The bacterial strains associated with a 189 beneficial or detrimental outcome are represented in green and red respectively. 190



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