



HAL
open science

Physiological functions of malate shuttles in plants and algae

Ousmane Dao, Franziska Kuhnert, Andreas P.M. Weber, G. Peltier, Yonghua Li-Beisson

► **To cite this version:**

Ousmane Dao, Franziska Kuhnert, Andreas P.M. Weber, G. Peltier, Yonghua Li-Beisson. Physiological functions of malate shuttles in plants and algae. Trends in Plant Science, 2021, 10.1016/j.tplants.2021.11.007 . hal-03453418

HAL Id: hal-03453418

<https://hal.science/hal-03453418>

Submitted on 28 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution - NonCommercial - NoDerivatives 4.0 International License

1 **Physiological functions of malate shuttles in plants and algae**

2

3 Ousmane Dao¹, Franziska Kuhnert², Andreas P. M. Weber², Gilles Peltier¹, Yonghua Li-
4 Beisson^{1*}

5

6 ¹Aix Marseille Univ, CEA, CNRS, BIAM, Institut de Biosciences et Biotechnologies Aix-
7 Marseille, CEA Cadarache, Saint Paul-Lez-Durance 13108, France

8 ²Institute of Plant Biochemistry, Cluster of Excellence on Plant Science (CEPLAS), Heinrich
9 Heine University, 40225 Düsseldorf, Germany

10

11

12 *: Author for correspondence: Yonghua Li-Beisson (email: yonghua.li@cea.fr; Tel:
13 +33(0)442254483)

14

15 **ORCID IDs:** 0000-0002-7040-5770 (O.D.); 0000-0002-0374-6671 (F.K.); 0000-0001-5545-
16 7583 (A.P.M.W.); 0000-0002-2226-3931 (G.P.); 0000-0003-1064-1816 (Y.L.-B.)

17

18

19

20

21

22 **Key words:**

23 Dicarboxylate translocator; Redox Trafficking; Malate Dehydrogenase; CO₂ concentration
24 mechanism; Photorespiration; Reactive Oxygen Species

25

26

1 **Glossary**

2 **CO₂ Concentrating Mechanism (CCM):** CCM, as the name implies, is a process of
3 concentrating CO₂ to the active site of rubisco, the major protein of the carbon photo-
4 reduction cycle (i.e. often called Calvin-Benson-Bassham cycle). It is therefore considered a
5 mechanism to reduce the rate of photorespiration. Depending on species, CCM could refer to
6 the dicarboxylic acid cycle in C₄ plants, CAM-based CCM in CAM plants, carboxysome
7 based-CCM in cyanobacteria and the biophysical CCM occurring in algae (the conversion of
8 CO₂ to bicarbonate and its transport to the pyrenoid).

9

10 **Cyclic Electron Flow (CEF):** CEF refers to the recycling of electron from the acceptor side
11 of photosystem I through the cytb₆/f complex, increasing the proton motive force (*pmf*). The
12 *pmf* is then used to drive ATP synthesis. The CEF promote ATP synthesis without additional
13 NADPH production and is therefore considered a way to increase the ATP/NADPH ratio in
14 response to metabolic or environmental adaptations. Several pathways of CEF have been
15 characterized: (1) the plant type I NADPH dehydrogenase complex (NDH); (2) the type II
16 NDH (NDA2) unique to algae; and (3) the proton gradient regulator 5 (PGR5)/PGR like 1
17 (PGRL1)-mediated CEF pathway, which is common between plants, algae and cyanobacteria.

18

19 **Fatty Acid β -Oxidation:** One of the major mechanisms of fatty acid degradation. Oxidation
20 of fatty acids can occur either at the α -, β - or ω - carbon position of a given fatty acid, among
21 which β -oxidation is found as a major pathway in plants and algae, therefore termed fatty acid
22 β -oxidation. This pathway ultimately breaks down fatty acids to acetyl-CoAs, which are
23 further metabolized either for energy production through the mitochondrial electron transport
24 chain, or for synthesis of sugars when coupled to glyoxylate and gluconeogenesis pathways.

25

26 **Glyoxylate Cycle:** An anabolic pathway occurring widely in plants, bacteria, fungi, and
27 algae. It allows the conversion of acetyl-CoA to succinate for the synthesis of carbohydrates,
28 therefore allowing microorganisms to utilize two carbon compounds as carbon source. For
29 example, it plays an essential role in the heterotrophic growth of green algae on acetate. It
30 shares several steps of tricarboxylic acid (TCA) cycle but bypasses the steps that release CO₂.

31

32 **Malate Shuttle:** Malate shuttle refers to the means of redox trafficking from one subcellular
33 compartment to another through malate transport. It is made of two major protein components
34 i.e. malate dehydrogenases and di- or tri-carboxylate translocators.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15

Photorespiration: a metabolic pathway occurring in organisms performing oxygenic photosynthesis where the major enzyme rubisco can fix either CO₂ or O₂ depending on their stoichiometric ratio. The oxygenation reaction produces 2-phosphoglycolate, a toxic metabolite, which is detoxified by conversion to the CBC intermediate 3-phosphoglycerate. This conversion involves a set of reactions occurring in four subcellular compartments i.e. chloroplast, cytosol, peroxisome, and mitochondria, and it entails the loss of CO₂, NH₄⁺ and energy in the process.

Pseudo-Cyclic Electron Flow (CEF): Pseudo-CEF is a pathway where electrons generated from water splitting at PSII are used to reduce O₂ to H₂O, thereby driving photosynthetic electron flow and generating a proton gradient, without NADPH production. In green algae, O₂-photoreduction is mostly mediated by the flavodiiron (FLV) proteins.

1 **Abstract:**

2

3 Subcellular compartmentalization confers evolutionary advantage to eukaryotic cells but entails
4 the need for efficient inter-organelle communication. Malate functions as redox carrier and
5 metabolic intermediate. It can be shuttled across membranes through translocators. The
6 interconversion of malate and oxaloacetate mediated by malate dehydrogenases requires
7 oxidation/reduction of NAD(P)H/NAD(P)⁺, therefore malate trafficking serves to transport
8 reducing equivalents, termed ‘malate shuttle’. Although the term “malate shuttle” has been
9 coined more than 50 years ago, novel functions are still emerging. This review highlights recent
10 findings on the functions of malate shuttles in photorespiration, fatty acid β-oxidation, inter-
11 organelle signaling and its putative role in CO₂-concentrating mechanisms. We compare and
12 contrast knowledge in plants and algae, thereby providing an evolutionary perspective on redox
13 trafficking in photosynthetic eukaryotes.

14

15 **Malate as a major cellular redox carrier**

16 Eukaryotic cells are compartmentalized, and distinct subcellular organelles house specific
17 subsets of metabolic reactions that are physically separated from each other by biological
18 membranes. Exchange of information and energy between compartments mostly mediated by
19 metabolites are indispensable to achieve whole-cell homeostasis and ensure optimal growth
20 [1,2]. While some of these exchanges occur through passive diffusion, most of them are
21 mediated by transporters [3]. Understanding the complex interplay of energy and information
22 between subcellular compartments is key towards the domestication of photosynthetic
23 organisms for tailor-made production of valuable compounds.

24 Malate is a C4 dicarboxylic acid and is involved in a number of metabolic pathways,
25 including the tricarboxylic acid (TCA), C4-dicarboxylic acid, and **glyoxylate cycles** in plants
26 and algae [4–6]. Malate can be produced by malate synthase (MAS) and metabolized by two
27 types of enzymatic reactions: (i) malic enzyme (ME), or (ii) malate dehydrogenase (MDH)
28 (**Box 1**). Amongst these, only MDH catalyzes a reversible redox reaction that is coupled to the
29 reduction/oxidation of NAD(P)⁺/NAD(P)H, while inter-converting malate and oxaloacetate
30 (OAA). In contrast to the main cellular redox-couple, i.e. the pyridine nucleotides NAD(P)(H),
31 malate can be efficiently transported across membranes through di- or tri-carboxylate
32 translocators. Although specific transporters of NAD⁺ are present in the membrane of
33 chloroplast, mitochondria, or peroxisome [7,8], their activity is restricted in importing NAD⁺
34 into organelles and probably important in de novo loading of organelles with NAD⁺ pool. But

1 these transporters have shown a very low affinity for NAD(P)H [9], therefore probably not
2 sufficient for redox exchange between compartments. Malate trafficking thus serves as an
3 indirect but efficient way of transporting reducing equivalents.

4 Trafficking of malate between different subcellular compartments has been extensively
5 studied in plants (**Box 2**) [10]. The inner membrane of chloroplast and mitochondria of plant
6 and algal cells are equipped with several types of di- and tri-carboxylate translocators catalyzing
7 malate transport in counter-exchange with, e.g., OAA, glutamate, or aspartate. It can function
8 in either direction depending on the concentration gradient resulting from a metabolic demand
9 [11,12]. Collectively, the trafficking of malate through specialized translocators from one
10 compartment to another combined with the malate processing enzymes (MDHs) is termed
11 “**malate shuttle**” [10].

12 Although the concept of “malate shuttle” has been coined over 50 years ago,
13 components of this pathway remain hypothetical in microalgae, whereas considerable progress
14 has been made in land plants [10]. Depending on the type of carboxylates being transported,
15 several variations of “malate shuttle” occur, i.e. malate/OAA shuttle, malate/aspartate shuttle
16 (**Box 3**), or malate/2-oxoglutarate shuttle. These various types of malate shuttles, their
17 subcellular locations and involvement in different metabolic pathways are illustrated in **Figure**
18 **1**.

19 Here, we review some of the most recent discoveries on the roles of malate shuttles in
20 connecting carbon, nitrogen, and energy metabolism in plants and algae. While isoforms of
21 MDHs and di-/tri-carboxylate translocators have been well characterized in land plants,
22 evidence is just emerging regarding their functions in algae. We provide a comparative analysis
23 of the corresponding proteins between the model plant arabidopsis (*Arabidopsis thaliana*) and
24 the model green alga chlamydomonas (*Chlamydomonas reinhardtii*) (**Table 1**). We discuss the
25 malate shuttle’s physiological importance in photosynthesis, **photorespiration**, and **fatty acid**
26 **β-oxidation** and we point to its possible implication in algal **CO₂ concentrating mechanisms**
27 (CCM). Finally, we highlight the role of malate shuttles in inter-organelle communication
28 through interaction with reactive oxygen species (ROS)-mediated signal transduction.

30 **Malate shuttle as a valve for photosynthetic electron dissipation**

31 During the linear electron flow (LEF) of photosynthesis, light energy is converted by two
32 photosystems (PSII and PSI) into chemical energy in the form of NADPH and ATP. ATP and
33 NADPH are mostly used to drive metabolic reactions, particularly CO₂ fixation by Rubisco and
34 its conversion into triose phosphates through the Calvin-Benson-Bassham (CBB) cycle [13].

1 However, the LEF is known to produce an insufficient amount of ATP (as compared to
2 NADPH) than required for optimal CO₂ photo-reduction, leading to an imbalance [14]. The
3 imbalance in ATP and NADPH could be further accentuated by changes in environmental
4 conditions. For example, under CO₂ limiting conditions, photorespiration in plants or a CCM
5 in algae requires additional ATP [14,15]. Excess reducing equivalents could potentially lead to
6 ROS over-production [16]. Coordination between production and usage of ATP and NAD(P)H
7 is therefore essential to cell fitness and survival in a changing environment.

8 ATP and NADPH balance can be achieved by increasing ATP production in the
9 chloroplast or through import, or by lowering NADPH level through enhancing sink strength
10 or export to other compartments (**Figure 2**). Several mechanisms are involved in the supply of
11 extra-ATP, which include **cyclic electron flow** (CEF) around PSI, or the O₂-photoreduction by
12 flavodiiron proteins (FLV) also known as **pseudo-CEF**, or ATP import from mitochondria
13 [17]. On the other hand, NADPH dissipation can occur either through enhanced metabolic
14 demand (e.g. boosting de novo fatty acid synthesis which has high need for reducing
15 equivalents), or through its export to other subcellular compartments by the “malate shuttle”
16 [18].

17 Despite having been hypothesized for several decades, the role of malate shuttle in
18 photosynthetic electron exportation (as a “malate valve”) has only recently been demonstrated.
19 In arabidopsis, the knock-out mutants in plastidial NADP-MDH (*nadp-mdh*) possessed a more
20 reduced plastoquinone pool but with only slight impairment in growth under high light (HL)
21 [19,20]. Strong growth defects, however, were observed under fluctuating light in a mutant in
22 which one of its regulatory redox switches at its C-terminus was deleted by genome editing
23 [21]. The carboxylate translocator involved in the ‘malate valve’ (AtOMT1) has also been
24 characterized in arabidopsis. AtOMT1 is a high-affinity OAA translocator, whose absence led
25 to the hyper-accumulation of reducing power in the stroma, photoinhibition, and impaired
26 growth under HL [22]. Taken together, above studies demonstrate the essential role of the
27 malate valve in chloroplast energy homeostasis, which is particularly important to balance the
28 chloroplast redox status during fluctuating conditions.

29 In algae, none of the components of the chloroplast ‘malate valve’ have been
30 characterized at a physiological level. The chlamydomonas CrMDH5, the homolog of the
31 arabidopsis NADP-MDH, has been studied in vitro [23]. The thioredoxin (TRX)-dependent
32 redox regulation of the CrMDH5 activity is much faster (responding in an all-or-nothing way)
33 than the plant homolog but the latter is fine-tuned due to the presence of an additional N-
34 terminus disulfide regulatory domain. Even though its physiological function has not been

1 shown directly, it was proposed that the “malate valve” may operate in the chlamydomonas
2 mutant lacking the proton gradient like 1 (PGRL1)-mediated CEF in conditions of strong
3 ATP/NADPH imbalance [24]. It was hypothesized that the malate valve helps to evacuate
4 excess reducing equivalents towards cytoplasm and mitochondria. Nevertheless, direct
5 evidence on the operation of the malate valve in algae is yet to come.

6 It is worth bearing in mind that the “malate valve” in the chloroplast could also function
7 in importing reducing power into the chloroplast. Responses of photosynthesis and chloroplast
8 metabolism to changes in redox state in other subcellular compartments (mitochondria or
9 peroxisome) seem to support such a possibility [25,26]. Our understanding of redox
10 homeostasis in the chloroplast is further complicated by the presence of both NAD⁺- and
11 NADP⁺-dependent MDHs and the co-existence of two pyridine nucleotide pools (NADH versus
12 NADPH) (discussed in **Outstanding Questions**).

13

14 **The role of malate shuttle in plant photorespiration**

15 Photorespiration, initiated by the oxygenation reaction of Rubisco, is inevitable in an oxygen
16 containing atmosphere. The rate of photorespiration is reduced in land plants performing C4
17 photosynthesis and in algae having a CCM [27]. Photorespiration consists of multiple metabolic
18 reactions distributed over four subcellular compartments: chloroplast, cytosol, peroxisome and
19 mitochondria, and requires therefore intimate inter-organelle communication. Photorespiratory
20 reactions have a strong impact on the cellular energetics, producing NADH in the mitochondria
21 where two glycine molecules are converted to serine, but consuming NADH in the peroxisome
22 where hydroxypyruvate is converted to glycerate (**Figure 1**). Therefore, NADH homeostasis in
23 mitochondria and peroxisome is critical for optimal functioning of photorespiration. Further,
24 changes in leaf internal CO₂ concentration, as a consequence of altered stomatal aperture, can
25 impact redox status in the respective organelles through photorespiration. Indeed, in land plants,
26 malate shuttle components (i.e. pNADP-MDH, pNAD-MDH, DiT1 and DiT2.1) are
27 upregulated upon acclimation to low CO₂, and downregulated under elevated CO₂ (where
28 photorespiration is suppressed) suggesting their possible implication in the adaptation of
29 cellular metabolism in response to fluctuating CO₂ availability [28,29].

30 The malate shuttle contributes to photorespiration at multiple levels (**Figure 1**). It is
31 involved in the transamination of glyoxylate into glycine coupled with the conversion of
32 glutamate to 2-oxoglutarate in the peroxisome catalyzed by the glutamate: glyoxylate
33 aminotransferase (GGT). Plastidial malate/2-oxoglutarate shuttle provides glutamate from
34 chloroplast that serves as a donor of NH₃ for the transamination of glyoxylate [30]. Arabidopsis

1 DiT2.1, but not DiT2.2, is a glutamate/malate translocator involved in photorespiration [31].
2 And moreover, DiT2.1 and DiT1 (AtpOMT1) work in concert for the optimal functioning of
3 photorespiration, and the lack of either DiT2.1 or DiT1 led to growth impairment under
4 photorespiratory conditions [22,31,32].

5 The malate shuttle is further involved in the export of NADH produced during glycine
6 decarboxylation in mitochondria, thus recycling NAD⁺ and ensuring proper functioning of
7 photorespiration. The decrease of mNAD-MDH activity in the *mmdh1* knockout mutant had a
8 strong effect on photorespiratory flux, leading to a reduced growth under photorespiration-
9 promoting conditions (i.e. very low-CO₂) [33]. In fact, the *mmdh1* mutant displayed a high
10 glycine/serine ratio indicative of impaired glycine decarboxylation due to the lack of the
11 electron acceptor NAD⁺ and an over-reduction in the mitochondria. The homeostasis of
12 NAD⁺/NADH can be achieved either by increasing the consumption of NADH by
13 mitochondrial reactions or by exporting the excess NADH to other compartments. A likely
14 route for exporting NADH from glycine decarboxylation is through a malate/aspartate shuttle
15 toward peroxisomes where NADH could be consumed by the hydroxypyruvate reductase
16 (HPR), another key reaction of photorespiration (**Figure 1**). The aspartate/glutamate
17 translocators involved in this malate/aspartate shuttle have been recently characterized in
18 arabidopsis as belonging to the uncoupling protein (UCP) family [11]. Both AtUCP1 and
19 AtUCP2 are able to efficiently counter-exchange aspartate and glutamate [11]. Similar to
20 *mmdh1*, mutants of AtUCP1 are impaired in photorespiration with a reduced rate of glycine
21 oxidation in the mitochondria [11]. Therefore, AtUCP1&2 and mNAD-MDH1&2 work
22 together in photorespiration by forming the malate/aspartate shuttle that connects glycine
23 decarboxylation in the mitochondria with hydroxypyruvate reduction in the peroxisome. We
24 note that redox equivalents derived from photorespiratory glycine decarboxylation in
25 mitochondria may not be necessarily shuttled to the peroxisome but could potentially fuel
26 nitrate and/or hydroxypyruvate reduction in the cytoplasm [34,35]. Antisense repression of
27 plastidial OMT1/DiT1 in tobacco however indicates a prominent role of the plastidial malate
28 shuttle in supplying nitrate reduction with NADH [36]. Therefore, the contribution of
29 mitochondria derived NADH in nitrate reduction in the cytosol could only be minor.

30 Finally, at the step of hydroxypyruvate reduction into glycerate by the peroxisomal
31 HPR, pNAD-MDH can oxidize malate imported from the cytosol to provide additional reducing
32 equivalents needed for HPR. Double mutants of pNAD-MDH1 & 2 were however viable under
33 photorespiratory conditions (i.e. ambient CO₂), but, the stoichiometry of photorespiratory CO₂
34 generated per Rubisco oxygenation was increased by 50% as compared to wild type [37]. This

1 was explained by an alternative pathway which was thought to be a non-enzymatic conversion
2 of hydroxypyruvate to glycolate thereby releasing extra CO₂. Timm et al [35] showed that
3 *HPR2* encodes for a cytosolic hydroxypyruvate reductase activity (i.e., not peroxisomal) and
4 that reduction of hydroxypyruvate can flexibly shift between peroxisomes and cytoplasm.

6 **A putative role of malate shuttle in algal CCM**

7 To cope with the low CO₂ to O₂ ratio, unlike plants where photorespiration plays a significant
8 role, microalgae frequently employ a CCM. The algal biophysical CCM is an energetic
9 mechanism that pumps and sequesters atmospheric CO₂ into the pyrenoid close to the active
10 site of Rubisco, key to algae's proliferation in their natural habitat where CO₂ level could be
11 extremely low [27]. In the past 10 years, enzymes (carbonic anhydrases), transcription factors
12 (CCM1) and also several inorganic carbon (Ci) transporters involved in CCM have been
13 identified and characterized in chlamydomonas [38,39]. Known transporters include the HL
14 activated 3 (HLA3) protein that is localized at the plasma membrane, a low-carbon inducible
15 protein (LCIA) in the chloroplast envelope, and recently the thylakoid localized bestrophin-like
16 (BST) transporters [40–43]. However, not until recently has the energization mechanism of the
17 CCM begun to be unveiled.

18 By studying double mutants of chlamydomonas lacking both PGRL1 and FLV-B
19 mediating the CEF and pseudo-CEF respectively, Burlacot et al [15] showed that double
20 mutants are impaired in growth under air (i.e. low-CO₂ condition) as was the reduced affinity
21 of photosynthesis of the double mutants for Ci. The authors concluded that the proton motive
22 force produced through the combined action of CEF and pseudo-CEF mechanisms favors the
23 conversion of bicarbonate anions transported by the thylakoid localized BSTs into CO₂ at the
24 active site of the carboxylating enzyme Rubisco. By showing that inhibitors of mitochondrial
25 respiration also decrease the affinity of photosynthesis for Ci, Burlacot et al [15] further
26 proposed that mitochondrial respiration serves as an alternative source of energy powering
27 distant transporters involved in CCM (located either at the plasma membrane or at the
28 chloroplast envelope). Considering that CCM is a light driven process, it is thus highly plausible
29 that the malate shuttle contributes to CCM through mediating redox exchange between
30 chloroplast and mitochondria (**Figure 2**).

31 Furthermore, implication of the malate shuttle in CCM is supported by RNAseq-based
32 transcriptome analyses, revealing the upregulation of putative chloroplastic 2-OG/malate
33 translocators and MDHs in chlamydomonas and also in four phytoplankton species when
34 transferred from high to low CO₂ condition [44–46]. Nevertheless, molecular actors and

1 experimental evidence for such a function is yet to come, e.i., by investigating the Ci affinity in
2 mutants impaired in malate shuttle mediating the redox exchange between chloroplast and
3 mitochondria.

5 **Malate shuttle mediates inter-organelle signaling through ROS**

6 Inter-compartmental exchange of signals is a key for cellular homeostasis and the acclimation
7 of photosynthetic organisms in fluctuating environments [47]. ROS signaling has been
8 considered a powerful system for regulation of gene expression, leading to a cascade of
9 physiological adjustments, such as induction of programmed cell death (PCD) [48,49]. ROS
10 can be generated in four major subcellular compartments, i.e., chloroplast, peroxisome,
11 mitochondria and cytosol. ROS are produced when O₂ serves as alternative electron acceptor
12 during photosynthesis or mitochondrial respiration electron transfer and intimately linked to
13 peroxisome-based fatty acid β-oxidation or photorespiration. ROS signaling has therefore been
14 found to connect organelle redox state between chloroplast and mitochondria, and between
15 chloroplast and peroxisome [19,49,50]. Below we discuss such examples.

16 ROS generation in the mitochondria was shown to trigger PCD in the arabidopsis mutant
17 *mod1* (for *mosaic death 1*) affected in the chloroplast enoyl-ACP reductase (ENR) catalyzing
18 the NADH-dependent enoyl-ACP reduction in the fatty acid synthase (FAS) complex [51]. A
19 screen for *mod1* suppressors identified various components of the malate shuttle involved in
20 chloroplast to mitochondrion communication, including pINAD-MDH, DiT1 and mNAD-
21 MDH1 [50]. Fatty acid biosynthesis is a major sink for reductant in the chloroplast [52].
22 Therefore, impairment in ENR leads to over-accumulation of NADH in the chloroplast. The
23 pINAD-MDH in conjunction with DiT1 allow the export of reducing power in the form of
24 malate. The translocator responsible of the malate import in mitochondria is still unknown.
25 Nevertheless, once inside mitochondrion, malate oxidation by the mNAD-MDH1 generates
26 NADH to fuel mitochondrial complex 1, ultimately resulting in the production of ROS [50].
27 This mitochondria-generated ROS has been considered as the signaling molecule in triggering
28 PCD in the *mod1* mutant cells [50].

29 Inter-organelle communication from chloroplast to peroxisome has also been proposed
30 to play a role in retrograde signaling in arabidopsis [19]. Under conditions where the
31 photosynthetic chain becomes over-reduced due to the limitations of the electron acceptor
32 NADP⁺, molecular oxygen (O₂) can be used to accept electron coming from PSI [53].
33 Photoreduction of O₂ can occur via two routes, i.e. the non-enzymatic reaction (also known as
34 Mehler reaction) [54,55] and the enzymatic reduction of O₂ [56,57]. In the Mehler reaction, the

1 transfer of electrons from PSI acceptor side to O₂ is associated with the generation of superoxide
2 O₂⁻ that could be converted to H₂O₂ by superoxide dismutase (SOD) and then detoxified by the
3 peroxidase located in the chloroplast such as ascorbate peroxidase (APX). H₂O₂ can also diffuse
4 out into other compartments where it can be detoxified, for instance, in the peroxisome by
5 catalase (CAT) [58]. H₂O₂ is a cellular messenger and has been shown to play an important role
6 as signaling agent [59]. However, to take up such a role, the intracellular concentration of H₂O₂
7 must be sufficient to induce the expression of H₂O₂-responsive nuclear genes. The reversible
8 inactivation of CAT in the peroxisome, allowing a burst in H₂O₂, requires the import of
9 reducing equivalents originating from the chloroplast through malate shuttle to peroxisome
10 [19,60]. This reversible inactivation of CAT was abolished in the arabidopsis *nadp-mdh*
11 mutants [19], indicating the important role of the light malate valve (i.e. plNADP-MDH) in the
12 regulation of CAT activity by transmitting the redox state of the chloroplast to the peroxisome.

13 A H₂O₂-based signaling pathway from peroxisome to chloroplast has also been
14 proposed recently in chlamydomonas. The chlamydomonas mutants deficient in the
15 peroxisomal MDH2 exhibited a higher rate of photosynthesis, starch, and *de novo* fatty acid
16 biosynthesis in the chloroplast during photoautotrophic nitrogen deprivation [25].
17 Communication between the two organelles is proposed to be mediated by a H₂O₂/malate-
18 dependent signaling pathway as supported by an elevated amount of H₂O₂ and malate in the
19 *mdh2* knockout mutants. Exogenously supplied H₂O₂ has a strong positive effect on the
20 expression of starch- and lipid- related genes in wild-type cells of chlamydomonas [61],
21 corroborating that the increase in starch and lipid in the *mdh2* mutants resulted from the increase
22 in H₂O₂ intracellular concentration [25]. Collectively, the malate shuttle represents a redox-
23 poise system communicating the redox-state between compartments involving H₂O₂-mediated
24 signaling pathway.

25

26 **Malate shuttle connects fatty acid catabolism to chloroplast metabolism**

27 Fatty acid β-oxidation, photorespiration, and glyoxylate cycle occur either totally or partially
28 in the peroxisome, making it a third subcellular compartment involved in energy metabolism
29 after chloroplast and mitochondrion [62–64]. In addition, peroxisomes house reactions that
30 produce H₂O₂. Fatty acid degradation and glyoxylate cycle generate reducing equivalents as
31 NADH inside the peroxisome, whereas photorespiration consume it. Therefore, CO₂
32 availability, by modulating the activity of photorespiration, can potentially impact these
33 metabolic pathways. Plants and algae have therefore evolved several mechanisms that regulate
34 NADH homeostasis in the peroxisome. This could be achieved either indirectly through the use

1 of metabolite translocators involving the malate shuttle or directly through the import of NAD⁺
2 via the NAD⁺ carrier or through NAD⁺ regeneration by local antioxidant system [7,65,66]. In
3 germinating castor bean, the transfer of NADH generated by fatty acid β -oxidation from
4 peroxisome to the mitochondria was postulated to occur through a malate/aspartate shuttle [67].
5 In arabidopsis, both direct or indirect routes occur, which include a peroxisomal NAD⁺ carrier
6 (PXN) belonging to the mitochondria carrier family (MCF) [7], the two peroxisomal MDHs
7 (pMDH1, pMDH2) [68], the photorespiratory HPR [69], or the components of the ascorbate
8 peroxidase/monodehydroascorbate reductase (APX/MDAR) electron transfer system [70]. The
9 occurrence of these multiple mechanisms suggests the importance of the redox balance control
10 in peroxisomes. Indeed, mutants deficient in either of these pathways have difficulty in seed
11 germination, seedling rigor or optimal growth.

12 The occurrence of plant-like peroxisomes has initially been debated in chlamydomonas,
13 and this was mostly built on two observations i) the lack of a crystalloid core under electron
14 microscope, typically observed in plant- or animal-type of peroxisomes; and ii) the likely
15 location of catalase in mitochondria based on organelle purification studies [71]. But recent
16 data have firmly established that chlamydomonas does have classical peroxisomes, and
17 evidence are i) peroxisomes have been observed in chlamydomonas both under confocal as well
18 as by electron microscopy [72,73]; ii) the typical peroxisomal targeting signal (PTS1/2) is
19 functional in Chlamydomonas [73]; iii) the subcellular localization studies for enzymes of
20 glyoxylate cycle [4]; iv) the identification of a H₂O₂-producing activity i.e. acyl-CoA oxidase
21 in peroxisome and its role in fatty acid β -oxidation [63]; v) catalase 1 is found to be peroxisomal
22 in chlamydomonas [74]; and finally genes encoding homologs of the arabidopsis proteins
23 involved in redox homeostasis (MDH, PXN, HPR, and MDAR) or in fatty acid β -oxidation are
24 present [75].

25 That said, except for MDH2, experimental evidence for the function of the other proteins
26 is still lacking. The chlamydomonas *mdh2* mutants, similar to its arabidopsis counterpart (the
27 *pmdh1pmdh2* mutants), were compromised in oil remobilization during nitrogen recovery
28 following a period of N deprivation [25], firmly establishing its contribution to fatty acid β -
29 oxidation in green algae. Interestingly, in addition to peroxisome metabolism, metabolic
30 responses have also been observed in the chloroplast, i.e., enhanced photosynthetic activity,
31 and enhanced starch and *de novo* fatty acid biosynthesis under high CO₂ condition (2% of CO₂
32 supplementation to the air). It is worth mentioning here that the phenotypes of the
33 chlamydomonas *mdh2* mutants were suppressed under atmospheric CO₂ level (photorespiratory
34 condition), pointing to the complex contribution of diverse pathways to redox management and

1 homeostasis. Nevertheless, the exact route or the molecular identity of the peroxisomal
2 translocators involved remain to be identified (see **Outstanding Questions**).

3

4 **Concluding remarks and future perspectives**

5 Because variations in environmental parameters may differentially affect the different cellular
6 functions involved in bioenergetics of photosynthetic cells, which are located in different
7 subcellular compartments, plants and algae have evolved efficient trafficking of reducing
8 equivalents between subcellular compartments to maintain redox homeostasis. Among these
9 mechanisms, the “malate shuttle” enables efficient transport of reducing equivalents between
10 chloroplasts, cytosol, mitochondria and peroxisomes. Research in the past decades has shed
11 light on the role of malate shuttle in key subcellular pathways including photosynthesis,
12 photorespiration, fatty acid β -oxidation, nitrogen assimilation, and inter-organelle signaling,
13 but has also raised new questions (detailed in **Outstanding Questions**). Beyond physiology
14 and energy homeostasis, emerging literature suggests a critical role for the malate shuttle in the
15 CO₂ concentration mechanism [15] and carbon storage [25].

16 Future investigations towards detailed understanding of malate shuttle mechanism in
17 plants and algae should involve the combination of cutting-edge molecular genetics tools for
18 creating combinatory mutations in different genes, overexpressing multiple isoforms combined
19 with the use of ¹³C labeling techniques to monitor changes in metabolic fluxes. We envision
20 that emerging genetically encoded biosensors to monitor real-time changes in subcellular redox
21 and ATP status will be valuable to monitor in real time redox status in subcellular compartments
22 [76,77]. Taken together, by connecting energy homeostasis to carbon and nitrogen metabolism,
23 the malate shuttle represents an exciting target for engineering the cellular energetic status for
24 better climate resilience and for improved production of plant and algal biomass for food, fuel
25 and biomaterials, key to a sustainable and greener future.

26

27 **Acknowledgements:**

28 O. Dao thanks CEA for a PhD scholarship. G. Peltier and Y. Li-Beisson thanks the continuous
29 financial support of CEA (LD-power, CO2Storage) for supporting research in her laboratory.
30 F. Kuhnert and A.P.M. Weber acknowledge funding by the Deutsche Forschungsgemeinschaft
31 (DFG, German Research Foundation) – Project-ID 267205415 – CRC 1208 and under
32 Germany’s Excellence Strategy EXC-2048/1, Project ID 390686111.

33

1 **Table 1. Malate shuttle main components in arabidopsis and chlamydomonas^a**

Protein family	Chlamydomonas	Phytozome ID	Arabidopsis	TAIR ID	Localization	Refs.
Malate dehydrogenases	CrMDH1	Cre03.g194850	cpNAD-MDH	AT3G47520	Chloroplast (arabidopsis) Peroxisome* (chlamydomonas)	[4,50,78–80]
	CrMDH2	Cre10.g423250	pMDH1	AT2G22780	Peroxisome	[25,37,68]
			pMDH2	AT5G09660		
	CrMDH3	Cre02.g145800	cyNAD-MDH1	AT1G04410	Cytosol	[81,82]
			cyNAD-MDH2	AT5G43330		
			cyNAD-MDH3	AT5G56720		
	CrMDH4	Cre12.g483950	mMDH1	AT1G53240	Mitochondria	[33,83,84]
mMDH2			AT3G15020			
CrMDH5	Cre09.g410700	cpNADP-MDH	AT5G58330	Chloroplast	[19,20,23]	
Malate translocators	CrOMT1	Cre17.g713350	DiT1 (OMT1)	AT5G12860	Chloroplast	[22,32,50]
	CrOMT2	Cre17.g713200				
	CrLCI20	Cre06.g260450	DiT2.1 (DCT1)	AT5G64290		
			DiT2.2 (DCT2)	AT5G64280		
	CrMiTC14	Cre16.g672650	DIC1	AT2G22500	Mitochondria	[11,85,86]
			DIC2	AT4G24570		
			DIC3	AT5G09470		
DTC			AT5G19760			

2 Abbreviations: Cr, *Chlamydomonas reinhardtii*; ‘pl’, plastidial; ‘p’, peroxisome; ‘cy’, cytosol
3 and ‘m’, mitochondria. * Refers to a putative localization of CrMDH1 in the peroxisome
4 based on fusion of N-terminus transit peptide with fluorescent tag.

5 ^aHomology between chlamydomonas and arabidopsis proteins were identified through
6 reversible BlastP search of TAIR (for arabidopsis) and Phytozome v13 (for chlamydomonas)
7 proteins. Arabidopsis protein sequences were used first as baits. Note that the function
8 assigned to mitochondrial malate translocators in arabidopsis is only based on in vitro
9 experiments except for DIC2 which has recently been characterized in vivo [87]. Other
10 players in the malate shuttle can be found in the **Box 2 and 3**.

11
12
13
14
15
16
17

1

2 **Box 1. Malate metabolism in photosynthetic cells**

3 Malate is a versatile compound and their cellular metabolism involve malate synthase (MAS),
4 malic enzyme (ME), or malate dehydrogenase (MDH) (**Figure I**). MAS catalyzes the formation
5 of malate from acetyl-CoA and glyoxylate, which is a key step in the glyoxylate cycle [4,88].
6 The glyoxylate cycle allows cells to utilize two-carbon compounds (C2) such as acetate, and it
7 is essential for the heterotrophic growth of some algae such as chlamydomonas [89]. The
8 concerted action of MAS and isocitrate lyse (ICL) in the glyoxylate cycle forms succinate that
9 enters the mitochondrial TCA cycle, bypassing the steps that result in carbon loss [4,64].
10 ME catalyzes the decarboxylation of malate to pyruvate, resulting in a production of NAD(P)H
11 and CO₂. This reaction has been shown as an important source of reducing equivalents for de
12 novo fatty acid synthesis as well as desaturation in fungi [90], plants [91] and more lately in
13 algae [92]. It is also a crucial reaction in C4 and CAM (Crassulacean acid metabolism)
14 photosynthetic metabolism [93].

15 MDHs are the only enzymes catalyzing a reversible reaction, important for its function
16 in modulating redox status. In arabidopsis, nine genes encoding MDHs are present in the
17 genome and activities of MDH could be detected in most subcellular compartments including
18 mitochondrion (mNAD-MDH1, 2), peroxisome (pNAD-MDH1, 2), cytosol (cyNAD-MDH1,
19 2, 3) and plastid (pNAD-MDH and pNADP-MDH) [10]. In contrast to land plants, five
20 isoforms of MDH (MDH1-5) are encoded in the genome of Chamydomonas [94]. CrMDH5
21 (the only NADP⁺ requiring MDH) is predicted to be chloroplastic, CrMDH2 peroxisomal
22 [4,63], CrMDH3 is considered cytosolic and CrMDH4 mitochondrial [95]. The situation is
23 more complex with CrMDH1 which is predicted as chloroplast-based on PredAlgo [95], but
24 found peroxisomal when its 25 aa of its N-terminus is fused to the yellow fluorescent protein
25 (YFP) [4]. Therefore, subcellular localization(s) of CrMDH1 using full-length protein fusion
26 or immunogold labeling would be needed to clarify its location.

27

28 **Box 2, Di- and tri-carboxylate translocators**

29 In addition to MDHs, the malate shuttle requires translocators of malate/or other di- or
30 tricarboxylates from one compartment to another. Different translocators have been predicted
31 and experimentally verified in arabidopsis chloroplasts and mitochondria [12,96]. The transfer
32 of malate through the chloroplast envelope membrane can be mediated by at least three different
33 translocators. The 2-oxoglutarate (OG)/malate-translocator 1 (OMT1 or DiT1) catalyzes the
34 export of malate in exchange for 2-OG [31]. AtOMT1 was also shown to have high affinity for

1 OAA in reconstituted liposomes [22]. The dicarboxylate translocators 1 and 2 (DCT1 & 2 also
2 named sometimes as DiT2.1 & 2.2) are glutamate/malate-translocators importing malate in
3 counter-exchange with glutamate [31]. Very likely also malate uniporters exist in the
4 chloroplast envelope membrane. However, the corresponding genes have not yet been
5 identified.

6 The transfer of malate across the mitochondrial inner membrane is mediated by at least
7 three dicarboxylate carriers (DIC1, DIC2, DIC3) and one dicarboxylate-tricarboxylate carrier
8 (DTC) belonging to the mitochondrial carrier family (MCF) [85,86]. The recombinant DICs,
9 when introduced into liposomes, catalyze the transport of a wide range of dicarboxylates such
10 as malate, OAA, succinate but with low affinity for 2-OG. These DICs were proposed to be
11 involved in mitochondrial malate-OAA shuttle [86]. Very recently, the physiological function
12 of DIC2 has been demonstrated in arabidopsis as being a high affinity malate–citrate antiporter
13 in the mitochondria, through evidence from the reverse genetic approach in combination with
14 comprehensive *in vitro*, *in organello*, and *in vivo* analyses [87]. On the other hand, in addition
15 to dicarboxylates (malate, OAA, 2-OG and succinate), DTC was shown to transport also
16 tricarboxylates (such as citrate or isocitrate) [85]. Therefore, DTC can serve as a malate/2-OG
17 translocator. A malate/aspartate shuttle has been proposed by [67] connecting mitochondria to
18 peroxisome, which requires coordinated functions of two types of translocators i.e. malate/2-
19 OG carrier (likely DTC) and a glutamate/aspartate carrier.

20 Genes encoding putative proteins homologous to the plant di- or tri-carboxylate
21 translocators are present in the chlamydomonas genome (**Table 1**). Three putative plastidial 2-
22 OG/malate translocators (annotated as OMT1, OMT2 and low carbon inducible20 - LCI20),
23 and one mitochondrial translocator known as MiTC14 and belonging to the MCF family, have
24 been identified [94]. The protein sequence of LCI20 shares 55% and 60% of similarity with
25 DiT2.1 and DiT2.2 respectively whereas AtDiT1 shares 63% of similarity with both CrOMT1
26 and CrOMT2 suggesting that they are evolutionary related dicarboxylate translocators.

27

28 **Box 3, Other players in the malate shuttle**

29 Apart from MDH and the malate translocators listed in Table 1, other enzymes could also be
30 involved in functioning of the variants of malate shuttle. One notable example is the malate-
31 aspartate shuttle, which require aspartate aminotransferases (AspAT), sometimes named as
32 glutamate-oxaloacetate transaminases (GOT). AspATs are ubiquitous enzymes encoded by a
33 multigenic family catalyzing the reversible interconversion between aspartate and 2-
34 oxoglutarate with glutamate and OAA. In arabidopsis, five isoforms of eukaryotic type AspAT

1 are present i.e. AspAT1 (mitochondrial), AspAT2 and AspAT4 (cytosolic), AspAT3
2 (peroxisomal), AspAT5 (plastidial) and one prokaryotic type AspAT (PTA) localized in the
3 chloroplast [97,98]. Activity of AspAT was also detected in chlamydomonas [99] and five
4 genes *AST1-AST5* (phytozome v13) encode for AspAT proteins [94] respectively. AST1 and
5 AST3 (mitochondrial), AST2 (chloroplastic), AST4 (mitochondrial by homology) and AST5
6 (cytosolic). The plant peroxisomal and mitochondrial isoforms of AspAT have been proposed,
7 in conjunction with MDHs, to play a role in shuttling reducing equivalents between both
8 organelles through malate-aspartate shuttle during photorespiration [100], but none of the algal
9 proteins has been studied.

10

11 **Figure Legends:**

12

13 **Figure 1. Known and putative steps of malate trafficking in a subcellular context.**

14 The known di-/tri-carboxylate translocators based on either in vivo or in vitro characterization
15 are reported in **table 1**. Transporter name in white refers to arabidopsis proteins and black refers
16 to the chlamydomonas putative transporters (*) based on homology without any functional data.
17 “?” represent the putative translocators involved in the malate shuttle. Dashed lines indicate
18 reactions with several transformation steps. Created with BioRender.com.

19 **Abbreviations:** ADP, adenosine diphosphate; ATP, adenosine triphosphate; Asp, aspartate;
20 AspAT, aspartate aminotransferase (**Box 3**); β -Ox., β -Oxidation; CBBC, Calvin-Benson-
21 Bassham cycle; DIC/DTC, mitochondrial dicarboxylate/ dicarboxylate-tricarboxylate carrier;
22 DiT1/2, dicarboxylate translocator 1/2; FA, fatty acid; FdGOGAT, ferredoxin-dependent
23 glutamine 2-oxoglutarate aminotransferase; G3P, glyceraldehyde-3-phosphate; GDC, glycine
24 decarboxylase; GGAT, glutamate-glyoxylate aminotransferase; Gln, glutamine; Glu,
25 glutamate; GS, glutamine synthase; HPR, hydroxypyruvate reductase; Mal, malate; MiTC14,
26 mitochondrial substrate carrier protein 14; MDH, malate dehydrogenase; mETC, mitochondrial
27 electron transport chain; NAD(P)H, nicotinamide adenine dinucleotide (phosphate); OAA,
28 oxaloacetate; 2-OG, 2-oxoglutarate; OH-Pyr, hydroxypyruvate; OMT1/2, 2-
29 oxoglutarate/malate translocator; OPPP, oxidative pentose phosphate pathway; 2-PG, 2-
30 phosphoglycolate; 3-PGA, 3-phosphoglycerate; PS, photosystems; RuBisCO, ribulose-1,5-
31 biphosphate carboxylase/oxygenase; TCA, tricarboxylic acid; UCP, uncoupling protein.

32

33 **Figure 2. Photosynthetic redox production, management and carbon metabolism in algae.**

1 The photosynthetic linear electron flow (LEF) occurring in the thylakoid membrane produces
2 ATP and NADPH for the fixation of CO₂ in the Calvin Benson Bassham (CBB) cycle to yield
3 carbohydrates. The G3P, ATP and NAD(P)H are then used for starch and de novo fatty acid
4 biosynthesis (blue arrows). Alternative electron pathways are represented by cyclic electron
5 flow (CEF; orange arrows) and pseudo-CEF (PCEF; orange arrows). Chloroplast to
6 mitochondrial communication (CTM) is represented in dark green arrows. Excess NAD(P)H is
7 used by plNAD(P)-MDHs to reduce OAA to malate that is shuttled through chloroplastic
8 malate translocators to fuel the mitochondrial electron transport chain (mETC) for the synthesis
9 of ATP. The CCM is represented in the chloroplast where atmospheric CO₂ is pumped then
10 sequestered into the pyrenoid. Part of energy needed for CCM comes from the proton motive
11 force generated by photochemical reactions and another part is likely to be derived from mETC
12 (dashed black arrow). Note that most of the reactions shown in this figure are also present in
13 higher plants except the algal CCM. Created with BioRender.com.

14 **Abbreviations:** ACCase, acetyl-CoA carboxylase; ADP, adenosine diphosphate; AGPase,
15 ADP-glucose pyrophosphorylase; ATP, adenosine triphosphate; CBB, Calvin Benson
16 Bassham; CCM, CO₂-concentrating mechanism; CEF, cyclic electron flow; Cyt *b₆f*,
17 cytochrome *b₆f* complex; ER, enoyl-acyl carrier protein (ACP) reductase; FA, fatty acid; FAS,
18 fatty acid synthase; Fd, ferredoxin; FLV, flavodiiron protein; FNR, ferredoxin-NADP⁺
19 reductase; G1P, glucose-1-phosphate; G3P, glucose-3-phosphate; G1P, glucose-1-phosphate;
20 KAR, ketoacyl-ACP Reductase; Mal, malate; MDH, malate dehydrogenase; mETC,
21 mitochondrial electron transport chain; NAD(P)H, nicotinamide adenine dinucleotide
22 (phosphate); NDA2, NAD(P)H dehydrogenase 2; OAA, oxaloacetate; PC, plastocyanin; PCEF,
23 pseudo-CEF; PETC, photosynthetic electron transport chain; PGR5, proton gradient regulation
24 5; PGRL1, proton gradient like 1; PQ/PQH₂, plastoquinone/plastoquinol; PTOX, plastoquinone
25 terminal oxidase; PS, photosystem; Q_A, quinone A; ROS, reactive oxygen species; TAG,
26 triacylglycerol; TCA, tricarboxylic acid.

27

28 **Figure I, in Box1: Metabolic reactions that involve malate.**

29 MDH, ME and MAS enzymes are found in higher plants and algae. MAS catalyzes the
30 biosynthesis of malate by condensing glyoxylate and acetyl-CoA. This reaction is part of the
31 glyoxylate cycle. ME uses either NADP⁺ or NAD⁺ to decarboxylate malate into pyruvate
32 releasing CO₂ and present in several subcellular compartments. MDH catalyzes the reversible
33 interconversion between malate and oxaloacetate using either NAD(P)⁺ or NAD(P)H as
34 cofactor. This reaction occurs in the plastid, cytosol, peroxisome, and mitochondria.

1 **Abbreviations:** CoA, Co-enzyme A; MDH, malate dehydrogenase; ME, malic enzyme;
2 NAD(P)⁺, nicotinamide adenine dinucleotide (phosphate).

3 * Denote that MDH reaction is reversible.

4 **References:**

- 5 1 Raghavendra, A.S. and Padmasree, K. (2003) Beneficial interactions of mitochondrial metabolism
6 with photosynthetic carbon assimilation. *Trends Plant Sci.* 8, 546–553
- 7 2 Weber, A.P.M. and Linka, N. (2011) Connecting the Plastid: Transporters of the Plastid Envelope
8 and Their Role in Linking Plastidial with Cytosolic Metabolism. *Annu. Rev. Plant Biol.* 62, 53–77
- 9 3 Neuhaus, H.E. and Wagner, R. (2000) Solute pores, ion channels, and metabolite transporters in
10 the outer and inner envelope membranes of higher plant plastids. *Biochimica et Biophysica Acta*
11 (*BBA*) - *Biomembranes* 1465, 307–323
- 12 4 Lauersen, K.J. *et al.* (2016) Peroxisomal microbodies are at the crossroads of acetate assimilation
13 in the green microalga *Chlamydomonas reinhardtii*. *Algal Research* 16, 266–274
- 14 5 Ludwig, M. (2016) The Roles of Organic Acids in C₄ Photosynthesis. *Front. Plant Sci.* 7,
- 15 6 Eisenhut, M. *et al.* (2019) Mechanistic understanding of photorespiration paves the way to a new
16 green revolution. *New Phytologist* 223, 1762–1769
- 17 7 van Roermund, C.W.T. *et al.* (2016) The Peroxisomal NAD Carrier from Arabidopsis Imports NAD
18 in Exchange with AMP1[OPEN]. *Plant Physiol* 171, 2127–2139
- 19 8 Hashida, S. and Kawai-Yamada, M. (2019) Inter-Organelle NAD Metabolism Underpinning Light
20 Responsive NADP Dynamics in Plants. *Front. Plant Sci.* 10,
- 21 9 Palmieri, F. *et al.* (2009) Molecular Identification and Functional Characterization of Arabidopsis
22 thaliana Mitochondrial and Chloroplastic NAD⁺ Carrier Proteins *. *Journal of Biological Chemistry*
23 284, 31249–31259
- 24 10 Selinski, J. and Scheibe, R. (2019) Malate valves: old shuttles with new perspectives. *Plant Biol*
25 (*Stuttg*) 21, 21–30
- 26 11 Monné, M. *et al.* (2018) Uncoupling proteins 1 and 2 (UCP1 and UCP2) from Arabidopsis thaliana
27 are mitochondrial transporters of aspartate, glutamate, and dicarboxylates. *Journal of Biological*
28 *Chemistry* 293, 4213–4227
- 29 12 Lee, C.P. and Millar, A.H. (2016) The Plant Mitochondrial Transportome: Balancing Metabolic
30 Demands with Energetic Constraints. *Trends in Plant Science* 21, 662–676
- 31 13 Allen, J.F. (2002) Photosynthesis of ATP—Electrons, Proton Pumps, Rotors, and Poise. *Cell* 110,
32 273–276
- 33 14 Kramer, D.M. and Evans, J.R. (2011) The Importance of Energy Balance in Improving
34 Photosynthetic Productivity¹[W]. *Plant Physiol* 155, 70–78
- 35 15 Burlacot, A. *et al.* (2021) Alternative electron pathways of photosynthesis drive the algal CO₂
36 concentrating mechanism. *bioRxiv* DOI: 10.1101/2021.02.25.432959
- 37 16 Erickson, E. *et al.* (2015) Light stress and photoprotection in *Chlamydomonas reinhardtii*. *The Plant*
38 *Journal* 82, 449–465
- 39 17 Alric, J. and Johnson, X. (2017) Alternative electron transport pathways in photosynthesis: a
40 confluence of regulation. *Current Opinion in Plant Biology* 37, 78–86
- 41 18 Burlacot, A. *et al.* (2019) Subcellular Energetics and Carbon Storage in *Chlamydomonas*. *Cells* 8,
42 1154
- 43 19 Heyno, E. *et al.* (2014) Putative role of the malate valve enzyme NADP–malate dehydrogenase in
44 H₂O₂ signalling in Arabidopsis. *Philosophical Transactions of the Royal Society B: Biological*
45 *Sciences* 369, 20130228
- 46 20 Hebbelmann, I. *et al.* (2012) Multiple strategies to prevent oxidative stress in Arabidopsis plants
47 lacking the malate valve enzyme NADP-malate dehydrogenase. *J Exp Bot* 63, 1445–1459
- 48 21 Yokochi, Y. *et al.* (2021) Redox regulation of NADP-malate dehydrogenase is vital for land plants
49 under fluctuating light environment. *PNAS* 118,

- 1 22 Kinoshita, H. *et al.* (2011) The chloroplastic 2-oxoglutarate/malate transporter has dual function
2 as the malate valve and in carbon/nitrogen metabolism. *The Plant Journal* 65, 15–26
- 3 23 Lemaire, S.D. *et al.* (2005) NADP-Malate Dehydrogenase from Unicellular Green Alga
4 *Chlamydomonas reinhardtii*. A First Step toward Redox Regulation? *Plant Physiology* 137, 514–521
- 5 24 Dang, K.-V. *et al.* (2014) Combined Increases in Mitochondrial Cooperation and Oxygen
6 Photoreduction Compensate for Deficiency in Cyclic Electron Flow in *Chlamydomonas reinhardtii*.
7 *The Plant Cell* 26, 3036–3050
- 8 25 Kong, F. *et al.* (2018) Interorganelle Communication: Peroxisomal MALATE DEHYDROGENASE2
9 Connects Lipid Catabolism to Photosynthesis through Redox Coupling in *Chlamydomonas*. *The*
10 *Plant Cell* 30, 1824–1847
- 11 26 Massoz, S. *et al.* (2015) Isolation of *Chlamydomonas reinhardtii* mutants with altered
12 mitochondrial respiration by chlorophyll fluorescence measurement. *J Biotechnol* 215, 27–34
- 13 27 Wang, Y. *et al.* (2015) The CO₂ concentrating mechanism and photosynthetic carbon assimilation
14 in limiting CO₂: how *Chlamydomonas* works against the gradient. *The Plant Journal* 82, 429–448
- 15 28 Backhausen, J.E. and Scheibe, R. (1999) Adaptation of tobacco plants to elevated CO₂: influence
16 of leaf age on changes in physiology, redox states and NADP-malate dehydrogenase activity.
17 *Journal of Experimental Botany* 50, 665–675
- 18 29 Li, Y. *et al.* (2014) Was low CO₂ a driving force of C₄ evolution: Arabidopsis responses to long-term
19 low CO₂ stress. *Journal of Experimental Botany* 65, 3657–3667
- 20 30 Takahashi, S. *et al.* (2007) Impairment of the Photorespiratory Pathway Accelerates
21 Photoinhibition of Photosystem II by Suppression of Repair But Not Acceleration of Damage
22 Processes in Arabidopsis. *Plant Physiology* 144, 487–494
- 23 31 Renné, P. *et al.* (2003) The Arabidopsis mutant *dct* is deficient in the plastidic glutamate/malate
24 translocator DiT2. *The Plant Journal* 35, 316–331
- 25 32 Taniguchi, M. *et al.* (2002) Identifying and Characterizing Plastidic 2-Oxoglutarate/Malate and
26 Dicarboxylate Transporters in Arabidopsis thaliana. *Plant Cell Physiol* 43, 706–717
- 27 33 Lindén, P. *et al.* (2016) Reduced mitochondrial malate dehydrogenase activity has a strong effect
28 on photorespiratory metabolism as revealed by ¹³C labelling. *Journal of Experimental Botany* 67,
29 3123–3135
- 30 34 Bloom, A.J. *et al.* (2010) Carbon Dioxide Enrichment Inhibits Nitrate Assimilation in Wheat and
31 Arabidopsis. *Science* 328, 899–903
- 32 35 Timm, S. *et al.* (2008) A Cytosolic Pathway for the Conversion of Hydroxypyruvate to Glycerate
33 during Photorespiration in Arabidopsis. *The Plant Cell* 20, 2848–2859
- 34 36 Schneidereit, J. *et al.* (2006) Antisense repression reveals a crucial role of the plastidic 2-
35 oxoglutarate/malate translocator DiT1 at the interface between carbon and nitrogen metabolism.
36 *The Plant Journal* 45, 206–224
- 37 37 Cousins, A.B. *et al.* (2008) Peroxisomal Malate Dehydrogenase Is Not Essential for Photorespiration
38 in Arabidopsis But Its Absence Causes an Increase in the Stoichiometry of Photorespiratory CO₂
39 Release. *Plant Physiol* 148, 786–795
- 40 38 Moroney, J.V. *et al.* (2011) The carbonic anhydrase isoforms of *Chlamydomonas reinhardtii*:
41 intracellular location, expression, and physiological roles. *Photosynth Res* 109, 133–149
- 42 39 Mackinder, L.C.M. *et al.* (2017) A Spatial Interactome Reveals the Protein Organization of the Algal
43 CO₂-Concentrating Mechanism. *Cell* 171, 133-147.e14
- 44 40 Duanmu, D. *et al.* (2009) Knockdown of limiting-CO₂-induced gene HLA3 decreases HCO₃-
45 transport and photosynthetic C_i affinity in *Chlamydomonas reinhardtii*. *PNAS* 106, 5990–5995
- 46 41 Wang, Y. and Spalding, M.H. (2014) Acclimation to Very Low CO₂: Contribution of Limiting CO₂
47 Inducible Proteins, LCIB and LCIA, to Inorganic Carbon Uptake in *Chlamydomonas reinhardtii*. *Plant*
48 *Physiology* 166, 2040–2050
- 49 42 Mukherjee, A. *et al.* (2019) Thylakoid localized bestrophin-like proteins are essential for the CO₂
50 concentrating mechanism of *Chlamydomonas reinhardtii*. *Proc Natl Acad Sci USA* 116, 16915–
51 16920

- 1 43 Kono, A. and Spalding, M.H. (2020) LCI1, a Chlamydomonas reinhardtii plasma membrane protein,
2 functions in active CO₂ uptake under low CO₂. *The Plant Journal* 102, 1127–1141
- 3 44 Hennon, G.M.M. *et al.* (2017) Diverse CO₂-Induced Responses in Physiology and Gene Expression
4 among Eukaryotic Phytoplankton. *Front. Microbiol.* 8,
- 5 45 Yamano, T. *et al.* (2008) Expression Analysis of Genes Associated with the Induction of the Carbon-
6 Concentrating Mechanism in Chlamydomonas reinhardtii. *Plant Physiol* 147, 340–354
- 7 46 Miura, K. *et al.* (2004) Expression Profiling-Based Identification of CO₂-Responsive Genes
8 Regulated by CCM1 Controlling a Carbon-Concentrating Mechanism in Chlamydomonas
9 reinhardtii. *Plant Physiol* 135, 1595–1607
- 10 47 Czarnocka, W. and Karpiński, S. (2018) Friend or foe? Reactive oxygen species production,
11 scavenging and signaling in plant response to environmental stresses. *Free Radical Biology and*
12 *Medicine* 122, 4–20
- 13 48 Dietz, K.-J. *et al.* (2016) Redox- and Reactive Oxygen Species-Dependent Signaling into and out of
14 the Photosynthesizing Chloroplast. *Plant Physiology* 171, 1541–1550
- 15 49 Zhao, Y. *et al.* (2020) Malate Circulation: Linking Chloroplast Metabolism to Mitochondrial ROS.
16 *Trends in Plant Science* 25, 446–454
- 17 50 Zhao, Y. *et al.* (2018) Malate transported from chloroplast to mitochondrion triggers production
18 of ROS and PCD in Arabidopsis thaliana. *Cell Research* 28, 448–461
- 19 51 Mou, Z. *et al.* (2000) Deficiency in Fatty Acid Synthase Leads to Premature Cell Death and Dramatic
20 Alterations in Plant Morphology. *The Plant Cell* 12, 405–417
- 21 52 Li-Beisson, Y. *et al.* (2019) The lipid biochemistry of eukaryotic algae. *Progress in Lipid Research* 74,
22 31–68
- 23 53 Allen, J.F. (1975) Oxygen reduction and optimum production of ATP in photosynthesis. *Nature* 256,
24 599–600
- 25 54 Mehler, A.H. (1951) Studies on reactions of illuminated chloroplasts: I. Mechanism of the
26 reduction of oxygen and other hill reagents. *Archives of Biochemistry and Biophysics* 33, 65–77
- 27 55 Curien, G. *et al.* (2016) The Water to Water Cycles in Microalgae. *Plant and Cell Physiology* 57,
28 1354–1363
- 29 56 Chaux, F. *et al.* (2017) Flavodiiron Proteins Promote Fast and Transient O₂ Photoreduction in
30 Chlamydomonas. *Plant Physiology* 174, 1825–1836
- 31 57 Gerotto, C. *et al.* (2016) Flavodiiron proteins act as safety valve for electrons in Physcomitrella
32 patens. *PNAS* 113, 12322–12327
- 33 58 Mubarakshina, M.M. *et al.* (2010) Production and diffusion of chloroplastic H₂O₂ and its
34 implication to signalling. *Journal of Experimental Botany* 61, 3577–3587
- 35 59 Gadjev, I. *et al.* (2006) Transcriptomic Footprints Disclose Specificity of Reactive Oxygen Species
36 Signaling in Arabidopsis. *Plant Physiology* 141, 436–445
- 37 60 Michelet, L. *et al.* (2013) Down-regulation of catalase activity allows transient accumulation of a
38 hydrogen peroxide signal in Chlamydomonas reinhardtii. *Plant, Cell & Environment* 36, 1204–1213
- 39 61 Blaby, I.K. *et al.* (2015) Genome-wide analysis on Chlamydomonas reinhardtii reveals the impact
40 of hydrogen peroxide on protein stress responses and overlap with other stress transcriptomes.
41 *The Plant Journal* 84, 974–988
- 42 62 Lazarow, P.B. (2016) Peroxisomes. In *Encyclopedia of Cell Biology* (Bradshaw, R. A. and Stahl, P.
43 D., eds), pp. 248–272, Academic Press
- 44 63 Kong, F. *et al.* (2017) Chlamydomonas carries out fatty acid β -oxidation in ancestral peroxisomes
45 using a bona fide acyl-CoA oxidase. *The Plant Journal* 90, 358–371
- 46 64 Plancke, C. *et al.* (2014) Lack of isocitrate lyase in Chlamydomonas leads to changes in carbon
47 metabolism and in the response to oxidative stress under mixotrophic growth. *The Plant Journal*
48 77, 404–417
- 49 65 Gakière, B. *et al.* (2018) NAD⁺ Biosynthesis and Signaling in Plants. *Critical Reviews in Plant*
50 *Sciences* 37, 259–307
- 51 66 Charton, L. *et al.* (2019) Plant peroxisomal solute transporter proteins. *Journal of Integrative Plant*
52 *Biology* 61, 817–835

- 1 67 Mettler, I.J. and Beevers, H. (1980) Oxidation of NADH in Glyoxysomes by a Malate-Aspartate
2 Shuttle. *Plant Physiology* 66, 555–560
- 3 68 Pracharoenwattana, I. *et al.* (2007) Arabidopsis peroxisomal malate dehydrogenase functions in
4 β -oxidation but not in the glyoxylate cycle. *The Plant Journal* 50, 381–390
- 5 69 Pracharoenwattana, I. *et al.* (2009) Fatty acid beta-oxidation in germinating Arabidopsis seeds is
6 supported by peroxisomal hydroxypyruvate reductase when malate dehydrogenase is absent.
7 *Plant Mol Biol* 72, 101
- 8 70 Eastmond, P.J. (2007) MONODEHYROASCORBATE REDUCTASE4 Is Required for Seed Storage Oil
9 Hydrolysis and Postgerminative Growth in Arabidopsis. *The Plant Cell* 19, 1376–1387
- 10 71 Kato, J. *et al.* (1997) Characterization of catalase from green algae *Chlamydomonas reinhardtii*.
11 *Journal of Plant Physiology* 151, 262–268
- 12 72 Hayashi, Y. *et al.* (2015) Increase in peroxisome number and the gene expression of putative
13 glyoxysomal enzymes in *Chlamydomonas* cells supplemented with acetate. *J Plant Res* 128, 177–
14 185
- 15 73 Hayashi, Y. and Shinozaki, A. (2012) Visualization of microbodies in *Chlamydomonas reinhardtii*. *J.*
16 *Plant Res.* 125, 579–586
- 17 74 Kato, N. *et al.* (2021) Subcellular Localizations of Catalase and Exogenously Added Fatty Acid in
18 *Chlamydomonas reinhardtii*. *Cells* 10, 1940
- 19 75 Kong, F. *et al.* (2018) Lipid catabolism in microalgae. *New Phytologist* 218, 1340–1348
- 20 76 Voon, C.P. *et al.* (2018) ATP compartmentation in plastids and cytosol of Arabidopsis thaliana
21 revealed by fluorescent protein sensing. *PNAS* 115, E10778–E10787
- 22 77 Elsässer, M. *et al.* (2020) Photosynthetic activity triggers pH and NAD redox signatures across
23 different plant cell compartments. *bioRxiv* DOI: 10.1101/2020.10.31.363051
- 24 78 Beeler, S. *et al.* (2014) Plastidial NAD-Dependent Malate Dehydrogenase Is Critical for Embryo
25 Development and Heterotrophic Metabolism in Arabidopsis. *Plant Physiology* 164, 1175–1190
- 26 79 An, Y. *et al.* (2016) Purification and characterization of the plastid-localized NAD-dependent
27 malate dehydrogenase from Arabidopsis thaliana. *Biotechnology and Applied Biochemistry* 63,
28 490–496
- 29 80 Schreier, T.B. *et al.* (2018) Plastidial NAD-Dependent Malate Dehydrogenase: A Moonlighting
30 Protein Involved in Early Chloroplast Development through Its Interaction with an FtsH12-FtsHi
31 Protease Complex. *The Plant Cell* 30, 1745–1769
- 32 81 Huang, J. *et al.* (2018) Self-protection of cytosolic malate dehydrogenase against oxidative stress
33 in Arabidopsis. *Journal of Experimental Botany* 69, 3491–3505
- 34 82 Liszka, A. *et al.* (2020) Three cytosolic NAD-malate dehydrogenase isoforms of Arabidopsis
35 thaliana: on the crossroad between energy fluxes and redox signaling. *Biochemical Journal* 477,
36 3673–3693
- 37 83 Tomaz, T. *et al.* (2010) Mitochondrial Malate Dehydrogenase Lowers Leaf Respiration and Alters
38 Photorespiration and Plant Growth in Arabidopsis[W][OA]. *Plant Physiol* 154, 1143–1157
- 39 84 Sew, Y.S. *et al.* (2016) Loss of Mitochondrial Malate Dehydrogenase Activity Alters Seed
40 Metabolism Impairing Seed Maturation and Post-Germination Growth in Arabidopsis1[OPEN].
41 *Plant Physiol* 171, 849–863
- 42 85 Picault, N. *et al.* (2002) Identification of a Novel Transporter for Dicarboxylates and Tricarboxylates
43 in Plant Mitochondria: BACTERIAL EXPRESSION, RECONSTITUTION, FUNCTIONAL
44 CHARACTERIZATION, AND TISSUE DISTRIBUTION *. *Journal of Biological Chemistry* 277, 24204–
45 24211
- 46 86 Palmieri, L. *et al.* (2008) Molecular identification of three Arabidopsis thaliana mitochondrial
47 dicarboxylate carrier isoforms: organ distribution, bacterial expression, reconstitution into
48 liposomes and functional characterization. *Biochemical Journal* 410, 621–629
- 49 87 Lee, C.P. *et al.* (2021) The versatility of plant organic acid metabolism in leaves is underpinned by
50 mitochondrial malate–citrate exchange. *The Plant Cell* DOI: 10.1093/plcell/koab223
- 51 88 Nogales, J. *et al.* (2004) Functional analysis and regulation of the malate synthase from
52 *Chlamydomonas reinhardtii*. *Planta* 219, 325–331

- 1 89 Bogaert, K.A. *et al.* (2019) Metabolic, Physiological, and Transcriptomics Analysis of Batch Cultures
 2 of the Green Microalga *Chlamydomonas* Grown on Different Acetate Concentrations. *Cells* 8, 1367
 3 90 Wynn, J.P. and Ratledge, C. (1997) Malic Enzyme is a Major Source of NADPH for Lipid
 4 Accumulation by *Aspergillus Nidulans*. *Microbiology* 143, 253–257
 5 91 Zell, M.B. *et al.* (2010) Analysis of Arabidopsis with Highly Reduced Levels of Malate and Fumarate
 6 Sheds Light on the Role of These Organic Acids as Storage Carbon Molecules. *Plant Physiology* 152,
 7 1251–1262
 8 92 Zhu, B.-H. *et al.* (2018) The Role of Malic Enzyme on Promoting Total Lipid and Fatty Acid
 9 Production in *Phaeodactylum tricornutum*. *Front. Plant Sci.* 9,
 10 93 Schlüter, U. and Weber, A.P.M. (2020) Regulation and Evolution of C4 Photosynthesis. *Annual*
 11 *Review of Plant Biology* 71, 183–215
 12 94 Merchant, S.S. *et al.* (2007) The *Chlamydomonas* genome reveals the evolution of key animal and
 13 plant functions. *Science* 318, 245–250
 14 95 Tardif, M. *et al.* (2012) PredAlgo: A New Subcellular Localization Prediction Tool Dedicated to
 15 Green Algae. *Mol Biol Evol* 29, 3625–3639
 16 96 Weber, A.P.M. and Linka, N. (2011) Connecting the Plastid: Transporters of the Plastid Envelope
 17 and Their Role in Linking Plastidial with Cytosolic Metabolism. *Annu. Rev. Plant Biol.* 62, 53–77
 18 97 Torre, F. de la *et al.* (2006) Identification and functional analysis of a prokaryotic-type aspartate
 19 aminotransferase: implications for plant amino acid metabolism. *The Plant Journal* 46, 414–425
 20 98 Schultz, C.J. and Coruzzi, G.M. (1995) The aspartate aminotransferase gene family of Arabidopsis
 21 encodes isoenzymes localized to three distinct subcellular compartments. *The Plant Journal* 7, 61–
 22 75
 23 99 Muñoz-Blanco, J. *et al.* (1988) Characterization of an L-aspartate aminotransferase activity in
 24 *Chlamydomonas reinhardtii*. *Physiologia Plantarum* 74, 433–439
 25 100 Journet, E.-P. *et al.* (1981) Role of Glutamate-oxaloacetate Transaminase and Malate
 26 Dehydrogenase in the Regeneration of NAD⁺ for Glycine Oxidation by Spinach leaf Mitochondria
 27 1. *Plant Physiology* 67, 467–469
 28
 29
 30

31 Outstanding Questions

- 32 • *Does redox exchange occur between NADH and NADPH pools in the chloroplast?* Both
 33 plant and algal chloroplasts contain two pools of pyridine nucleotides (NADH versus
 34 NADPH). Possible interaction between the two redox pools (NADH versus NADPH)
 35 in the chloroplast is not known but likely mediated by the NAD(P)⁺ transhydrogenase.
 36 • *Do *Chlamydomonas* chloroplasts contain a NAD-MDH?* CrMDH1 is found
 37 peroxisomal based on fusion of the first 25 amino acids at the N-terminus to the yellow
 38 fluorescent protein, but it is predicted to be chloroplastial by PredAlgo. A key question
 39 is the subcellular location(s) of CrMDH1. The location of this enzyme could have
 40 implications for our understanding of metabolism and redox homeostasis not only for
 41 the chloroplast but also for the peroxisome.
 42 • *Is there any connection between the enzymatic and non-enzymatic function of the*
 43 *plastidial NAD-dependent MDH?* It was recently shown that the NAD-dependent MDH

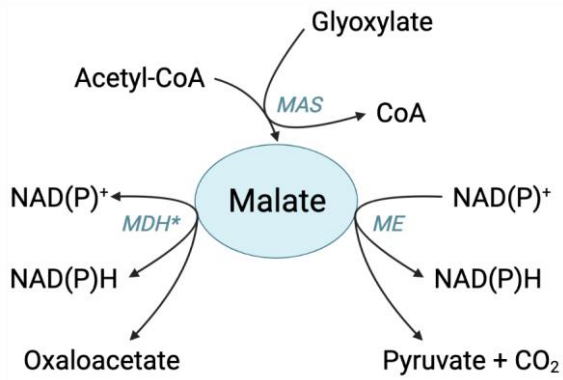
1 plays a moonlighting function in chloroplast metabolism and biogenesis. The
2 moonlighting function is an interesting feature of some proteins to have more than one
3 function in the cell. In addition to its enzymatic function as malate dehydrogenase, the
4 plastidial NAD-dependent MDH plays a central role in chloroplast biogenesis through
5 its physical interaction with the FtsH12-FtsHi protease complex. However, it is still not
6 clear whether these two functions are connected or what is the biological significance
7 of grouping these two functions (enzymatic and structural) in one widespread protein
8 such as MDH?

- 9 • *What is the identity of peroxisomal malate translocator?* This is probably one of the
10 most elusive components of peroxisome redox metabolism. Putative candidates are
11 emerging with modern proteomics analysis of isolated peroxisomes.
- 12 • *What is the interaction between redox trafficking through malate shuttle and carbon
13 storage in microalgae?* Starch and fatty acid biosynthesis are major sink of cellular
14 carbon and energy. Their synthesis requires a carbon source, ATP and NAD(P)H but at
15 different ratio. Alterations of redox trafficking and cell energetic metabolism could have
16 direct consequences for biomass productivity and composition.
- 17 • *What is the nature of the malate uniport mechanism in C4 plants?* Using isolated
18 chloroplasts from a range of C4 plants two types of malate uniporters have been found,
19 one coupled to sodium symport (in most C4 species tested), the other coupled to proton
20 symport (in maize, sorghum, and sugarcane). In contrast to the metabolite translocators,
21 this type of malate transporter would allow for net malate transport for, e.g., the malic
22 enzyme reaction.

23
24
25
26
27
28
29
30
31
32

1 Figure in Box 1:

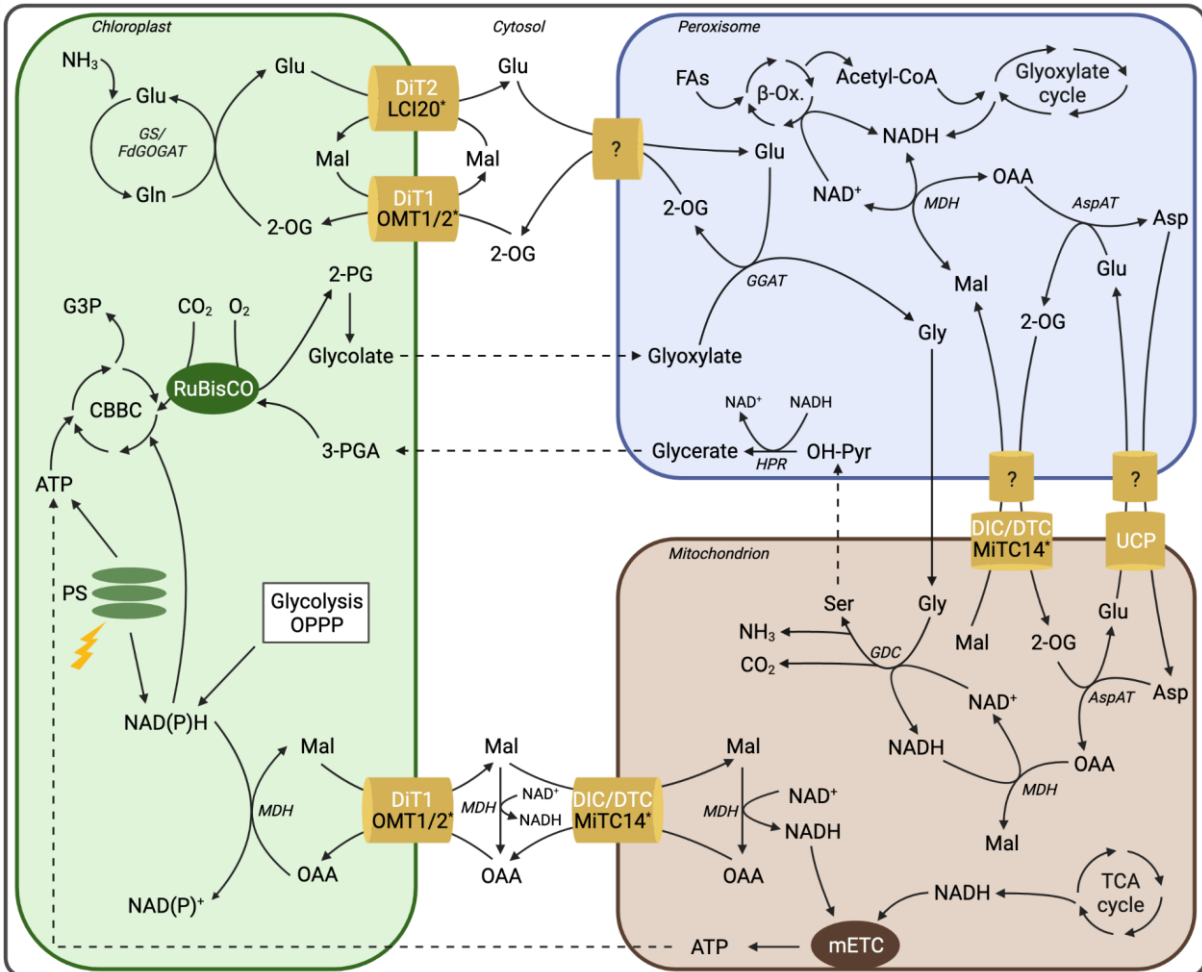
2



3

4

5 **Figure 1:**



6

7

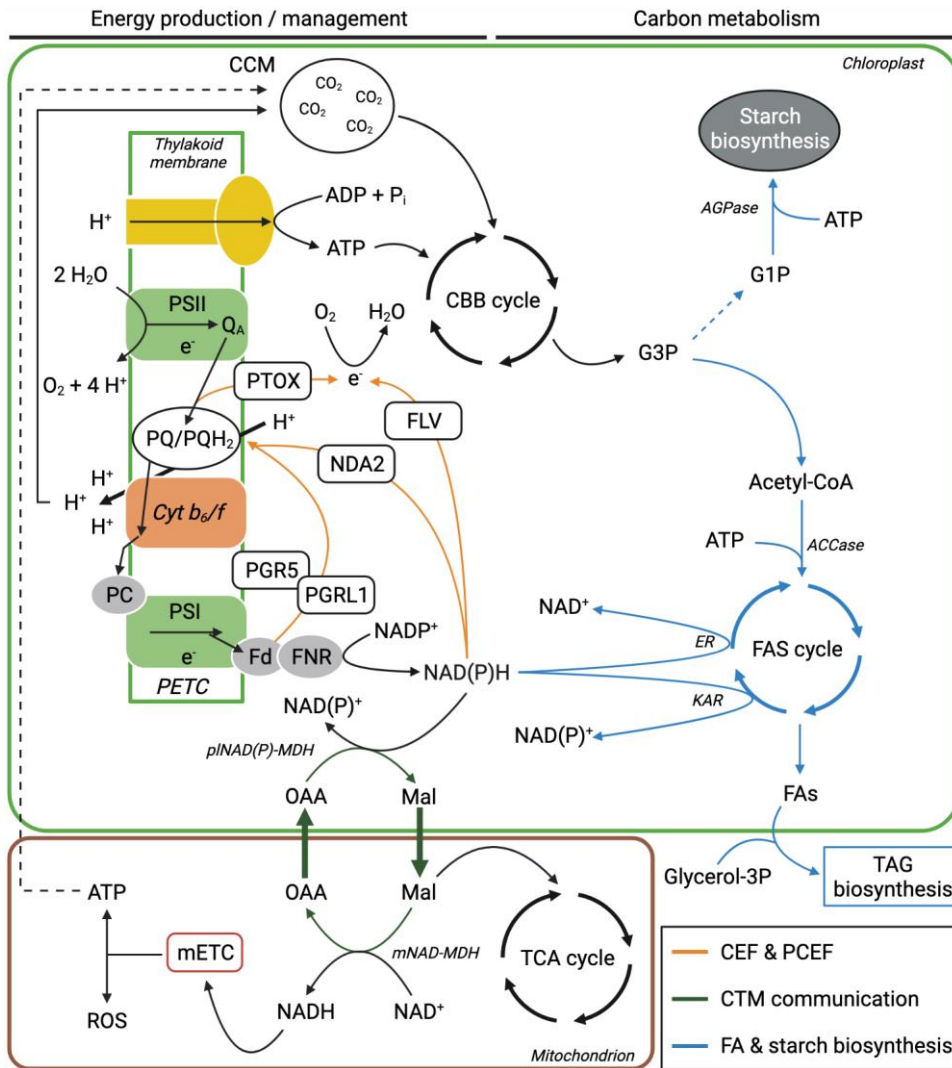
8

9

1

2 **Figure 2:**

3



4