

Pollution by anthropogenic microfibers in North-West Mediterranean Sea and efficiency of microfiber removal by a wastewater treatment plant

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Pollution by Anthropogenic Microfibers in North-West Mediterranean Sea and 1 2 Efficiency of Microfiber Removal by a Wastewater Treatment Plant 3 M.L. Pedrotti^{a*}, S. Petit^{ae*}, B. Eyheraguibel^b, M.E. Kerros^a, A. Elineau^a, J.F. Ghiglione^c, J.F. 4 5 Loret^d, A. Rostan^f, G. Gorsky^a, 6 7 ^aSorbonne Université, CNRS, Laboratoire d'Océanographie de Villefranche, LOV, F-O6230 8 Villefranche-sur-mer, France 9 ^bUniversité Clermont Auvergne, CNRS, SIGMA Clermont, Institut de Chimie (ICCF), F-63000 Clermont-Ferrand, France 10 ^cSorbonne Université, CNRS, Laboratoire d'Océanographie Microbienne, UMR 7621, 11 12 Observatoire Océanologique de Banyuls, Banyuls-sur-Mer, France 13 ^dSUEZ Groupe, CIRSEE, 78 230 Le Pecq, France 14 ^eSUEZ International, 183, avenue du 18 juin 1940, 92500 Rueil-Malmaison – France 15 ^fRégie Eau d'Azur, Rimiez, Nice, France 16 * Equally contributed 17 18 **KEY WORDS** 19 20 Microplastic, microfibers, airborne, wastewater treatment plant, urban environment, 21 Mediterranean Sea 22 **HIGHLIGHTS** 23 24 MFs were detected and analyzed in airborne and from urban areas to the open sea 25 • Contamination can be a major source of error in MFs detection and quantification • Shed of laundry fibers from WWTP are important sources of MFs in marine 26 27 environment • From 14 to 50% of marine MFs were synthetic and most of them were PES and PA 28 29 • Impacts of MFs on marine environment should include the atmospheric compartment

ABSTRACT

The widespread pollution from the release of microfibers is an emerging concern as they are a potential threat to the environment. Their identification in samples in terms of quantity and pathways remain a challenge as contamination can be a major source of error. A systematic study of synthetic microfibers (MFs) has been carried out in different environmental compartments of an urban area and in the surface waters of the northwestern Mediterranean.

The quantity, size and type of polymer of MFs were recorded in air, in waste water from a domestic washing machine, at the inlet and outlet of the Haliotis urban wastewater treatment plant (WWTP) in Nice (Provence Alpes Côte-d'Azur, France) and in a variety of coastal and offshore areas. The results showed that MFs released by clothes during washing (on average of 13×10^6 MFs per m³) are an important emitter of microplastics. Despite its high removal efficiency (87.5% to 98.5%) by Haliotis, a large number of MFs, estimated at 4.3 billion, enter the marine environment daily from the treatment plant. The attenuated total reflectance Fourier-transform infrared spectroscopy (ATR-FTIR) characterization of the raw materials showed that 14 to 50% of fibers are synthetic, mostly polyester and polyamide, the remaining 35 to 72% being natural polymers (cotton, wool) or manufactured by processing natural polymers (especially cellulose). MFs were found in all environmental compartments studied and appear to be widespread in coastal and offshore surface waters with concentrations varying from 2.6×10^3 to 3.70×10^4 m $^{-3}$. The sources of MFs in the marine environment are multiple, with laundry fibers discharges from WWTP and the atmospheric transport of urban fibers are among the main pathways.

1. Introduction

It is estimated that 8300 million metric tons (Mt) of virgin plastic have been produced since the 1950s and production is expected to increase by 4% per year (Geyer et al., 2017, Plastic Europe 2018). In 2010, poor household or municipal waste management generated 5 to 13 million tons of plastic waste in the oceans (Jambeck et al., 2015). Dumped or transported by sewage, rivers or tides, or simply by wind or urban surface runoff, about 80% of ocean plastic leakage comes from land-based sources (Andrady, 2011, Lebreton et al., 2017).

According to the latest estimates, at least 5,000 billion particles of plastic debris weighing 268,940 tons are currently floating in the oceans, with microplastics accounting for 92.4% of the total number of particles (Cozar et al., 2014; Eriksen et al., 2014). However, these estimates, based on currently used sampling methods (using a Manta or Neuston net with a mesh size of 333 mm), underestimate the true concentrations of microplastics because they do not account for the smaller particles and microfibers from synthetic textiles (Conkle et al., 2018). Recent studies suggest that smaller plastics (<0.3 mm) are 5 to 7 orders of magnitude higher than microplastic concentrations between 0.3 and 5 mm (Brandon et al., 2019).

Synthetic MFs can be considered a distinct category of microplastics because of their ubiquitous distribution, commonly found in aquatic and terrestrial environments, in human skin, as well as component of air pollution, occurring mainly as fibrous particles in dust and in the atmosphere (Browne et al., 2011; Dris et al., 2016). They are suspected to be a major contributor to plastic pollution at sea and are observed in higher concentrations than granules or fragments (Gago et al., 2018). The textile industry is of great economic importance, with 65 million tons of plastic used for the production of textile fibers in 2016 with polyester, polyamide, acrylic and polyolefin being the most common (The Fiber Year, 2017). MFs are not exclusively synthetic polymers such as nylon, polyester or polypropylene, but include also

natural (cotton, wood, yarn) and artificial fibers (rayon, viscose, cellulose acetate) (Houck, 2009). Their release into the environment, whether of synthetic or natural materials, has become an emerging pollution concern (Barrows et al., 2018, Suaria et al., 2020).

MFs entering the environment through urban runoff or discharged from textiles and transported by wastewater to coastal zones are considered as major sources of microplastics (Browne et al., 2011, Almroth et al., 2018, Galafassi et al., 2019). Wastewater treatment plants (WWTP) are effective waste management infrastructures and a strategic tool to combat marine pollution. However, according to the literature, even in high-income countries, not all MFs are retained by sewage treatment and large quantities are found both near the coasts and in the open sea (van Cauwenberghe et al., 2013, Thompson et al., 2004; Yu et al., 2018). Also, recent studies show that atmospheric deposition may also play an important role in the delivery of MFs to the marine environment (Enyoh et al., 2019, Windsor et al., 2019, Allen et al., 2019).

Like other types of microplastics, MFs are vectors for contaminants, absorbing persistent organic pollutants (POPs) such as hydrocarbons and polychlorinated biphenyl PCBs and releasing chemical additives into the environment and/or into the organisms that ingest them with multiple unknown consequences for animal and human health (Teuten et al., 2009, Hüffer and Hofmann 2016; Baini et al., 2017). In addition, the ingestion of MFs by marine life leads to physical and chemical damage such as internal abrasions or disruption of the digestive system, reduced growth and reproduction. (Derraik 2002; Moore, 2008; Mathalon and Hill, 2014; Galloway and Lewis, 2016, Torre et al., 2016). With respect to human exposure to microplastics, the main routes suggested are ingestion and inhalation (Smith et al., 2018), with ingestion being low compared to exposure via atmospheric deposition of household fibers (Catarino et al., 2018).

Due to the high risk of contamination during sampling and laboratory processing, fibers are often excluded from microplastic analyses (Foekema et al., 2013, Woodall et al., 2016; Torre et al., 2016). In addition, in the majority of studies, fibers are not sampled due to the difficulty of collecting them or their abundance is underestimated by the use of large mesh size nets (Gago et al., 2018).

To date, few studies have been conducted on the impact of microfibers in various environments (Driss et al., 2015, Gago et al., 2018, Stanton et al., 2019). Here we report on a comprehensive and systematic survey of the occurrence of synthetic MFs in an urban area and in the surface waters of the northwestern Mediterranean, one of the most polluted seas with highly variable concentrations of plastic (Kaandorp et al., 2020, Llorca, et al., 2020). We have established methods and quantified synthetic MFs in the air, production by domestic washing machines, at the inlet and outlet of the Haliotis wastewater treatment plant in Nice (Provence Alpes Côte-d'Azur, France) and concentrations in coastal and offshore areas.

This research aims to enrich existing knowledge on the sources and transport of these polymers into the marine environment. This is of paramount importance for the application of preventive and corrective solutions for European citizens, communities and industries (Carr, 2017).

2. Material and methods

2.1. Minimize microfiber contamination

To minimize the amount of contamination in our samples a quality control approach was undertaken according to the protocol previously described by Woodall and colleagues (2015) and Dris and colleagues (2016). First of all, to collect and process the samples, plastic

material has been replaced by glass or stainless-steel material except for a 50-liter Nalgene cylinder and a 500-ml Nalgene pressure bottle due to technical constraints or safety at sea. For each in situ sampling site or laboratory experiment, clean material was used, previously washed with alkaline and acidic detergents and rinsed three times before each handling with 0.2 µm (1.8 Ohm, Merck Millipore, Germany) milli-Q microfiltered water. A cotton coat was worn during sampling and further processing. During the automated sampling of waste and treated water, the samples remained in contact with the ambient air for 24 h. To account this potential source of contamination, a control consisting of 2 L of milli-Q water was maintained in the autosamplers for the same period in all experiments and then filtered through a cellulose acetate filter (Whatman®). During sample collection and laboratory analysis an ambient air blank was made by placing wet GF/F glass microfiber filter (Whatman®) in a clean 47 mm glass Petri dish kept open during the whole experiment. Finally, on the laboratory bench, blanks were also used to control all materials and fluids used for microfiber quantification. For this purpose, 300 mL of milli-Q water were passed through the equipment used and filtered through a cellulose acetate filter (Whatman®). The controls were examined using the same procedure as for the experimental samples and the count values were systematically removed from the calculated MFs results.

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2.2. Sources, Sites and sampling procedures

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2.2.1 Production of MFs by domestic washing machines

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A Miele Pro WS5425 domestic washing machine was used to quantify the release of synthetic MFs into wastewater. To be as close as possible to actual washing conditions, a commercial detergent was used. The washing program used was a "delicate synthetic" cycle

at 30 °C for 40 min with a spin speed of 600/800 rpm. The effluent was recovered at the machine outlet pipe in two 50 L Nalgene jerrycans (Dutscher, France). Seven runs of washes were carried out with different clothing compositions ranging from 100% synthetic to a mixture of natural, artificial and synthetic textiles (Table S1). For each wash, the composition and weight of the garments were recorded and 70 L of wash was collected and sieved through a stainless-steel sieve of porosity 20 µm. In order to minimize MFs contamination remaining from previous washes, five rinse and drain cycles at 600/800 rpm of 15 min each were performed without clothes at the beginning of the experiment and two cycles between washing experiments. This is based on preliminary tests which have shown that from 2 rinses onwards, the number of fibers is reduced by a factor of 10 and that this number remains constant.

These concentrated MFs issue from the washing and rinsing experiments were recovered with 75 to 575 ml of $0.2~\mu m$ milli-Q water (Merck Millipore, Germany) and stored in amber glass bottles until analysis. When the results were calculated, the remaining fiber contamination between two runs was subtracted from the next washing experiment. The diversity of natural and synthetic fibers allowed us to build a library of MFs for further analyses.

2.2.2 Air compartment

The number of MFs in aerosols was estimated by collecting atmospheric particles at two sampling sites (Fig. 1): A) at Saint Jean Cap Ferrat (locality with 1645 inhabitants), using the Atmospheric platform of the French national program MOOSE (Mediterranean Ocean Observing System for the Environment). Three experiments were conducted in June 2014 with a sampling period ranging from 6 to 11 days depending on prevailing weather

conditions. The mean aerosol volume of 393m³ with the atmospheric particles was filtered through cellulose acetate filters (Sartorius SM 11106, porosity 0.45 µm, diameter 47 mm) using a Central Partisol 2000 air sampler configured for PM10 sampling. In the site B, located on the rooftop of Haliotis treatment plant (Nice, a city of 377,328 inhabitants), two experiments were conducted in May 2016, the atmospheric particles (mean aerosol volume of 109 m³) were filtered with a cellulose acetate filter (Whatman®, porosity 0.45 µm and diameter 47 mm) using a sampling system consisting of a vacuum pump and a filter holder placed 2.5 meters high on the roof of the treatment plant. At the same time, the atmospheric deposition of MFs was measured by recovering all synthetic particles and MFs adhering to the funnel (Dris et al., 2016). For this purpose, a stainless-steel funnel (0.0735 m²) was placed on top of a 15 L stainless steel bucket. Samples were collected during 4-6 days and then filtered through cellulose acetate filters with a porosity of 0.45 µm.

2.2.3. Urban and treated wastewaters

The Nice Haliotis WWTP processes 120,000 m³ of wastewater per day and discharges its effluent into the Mediterranean Sea, 1.2 km from coast and at 100 meters depth. The treatment includes a pre-treatment unit (screening, removal of gravel and oils), primary treatment (sieving and lamellar settling), biological treatment (activated sludge), sludge treatment, the offshore pumping station and an air treatment unit.

In order to establish the sampling protocol, preliminary tests for the detection and analysis of MFs were conducted from December 2013 to March 2014 at the Haliotis WWTP. Subsequently, seasonal monitoring was carried out from March to November 2014 and six sampling campaigns of 3 days each were conducted. For each campaign, three replicate samples of 2 L of water were taken, with a sampling frequency of 1 hour, using an

autosampler that provided a 24-hour average sample. At the inlet of the treatment plant the samples were taken after the pre-treatment unit, at the outlet the treated water was sampled after the clarifier (Fig. S1). A total of 36 samples were collected and sieved through a stainless-steel sieve of porosity 20 μ m and stored at + 4 °C in amber bottles until their analysis. In order to evaluate the physical and environmental parameters that may influence the efficiency of microfiber treatment, the concentration of suspended solids (according to the European standard NF EN 872), the flow rates of the treatment plants and precipitation events were recorded.

2.2.4. In situ surface sampling in the Mediterranean Sea

A field survey was conducted from February to October 2014 in the northwestern Mediterranean Sea. A total of 15 sites were sampled and 34 water samples were taken (Fig. 1). Seven sites were seasonally monitored in spring, summer and fall with the RV *The Alchemy*, an 11m long sailing ship. First, a grid of 5 sampling points was set-up in the vicinity of the Haliotis WWTP, upstream and downstream of the outfall. Second, sampling was undertaken at Point B (permanent stations at the entrance of the Bay of Villefranche-sur-Mer) where surface and 10m depth samples were collected. Third, we sampled the offshore permanent sampling site called Dyfamed (42.6 Km offshore from Villefranche). The two permanent stations are characterized by low anthropogenic contamination (for more details see: https://www.imev-mer.fr/web/). The other 8 sites were sampled with the RV Tara a 36 m long oceanographic schooner in summer in the coastal zones of French Riviera, near Bastia and Calvi (Corsica) and in the Bay of Naples (Italy) during the Tara Mediterranean expedition (https://oceans.taraexpeditions.org/en/tara-mediterranee-expedition/). While sampling the Ligurian Sea, Tara sampled also the Pt. B and Dyfamed stations.

Surface samples were collected using a stainless-steel bucket held by a sheathed steel wire. To minimize contamination, all equipment was rinsed three times at each point with the sampled water. At each station fifty liters of surface seawater were collected. For the 10m depth seasonal samples at Pt. B, a 10 L Niskin bottle was used. On the boat, the samples were filtered through the stainless-steel sieve 20 μ m and resuspended in 200 mL of 0.2 μ m filtered seawater, stored

at + 4 °C in amber bottles until their analysis.

2.3. Detection and quantification of synthetic MFs

2.3.1 Sample preparation for analysis

Digestion protocols were either inefficient or damaged the MFs structures at the time of the study (2014), therefore we decided to use a dilution protocol. Preliminary tests were performed to evaluate the filtered aliquots used for each experiment to avoid clogging the filters and to ensure that sufficient material was present and that organic material did not affect the Fourier Transform Infrared Spectroscopy analysis (FTIR) analysis. The filtered volumes for the washing experiment were 750 ml, for wastewater (inlet) and treated water (outlet) 125 ml and 500 ml respectively, and for seawater 6.25 L of surface water. All filters showed a considerable amount of fibers. With exception of the inlet, FTIR analysis was performed at all the different steps of water treatment.

In the laboratory, samples were filtered by vacuum filtration in duplicate on a $0.2 \mu m$ Anodisc membrane (aluminum oxide, 47 mm). Control and experimental fibers were counted by the same individual under a stereo microscope (Olympus SZX10, X4.725 - X94.5) after filters being placed in a Dollfus cell (with 200 squares of 25 mm²). For the control, aerosol

and outlet samples the totality of the filters was counted. For the washing machine experiments, half of the filters (100 squares) were randomly counted (median 3906; min 2000; max 5164). For the inlet and seawater samples at least 100 squares were randomly counted in order to have a minimum of 40 fibers in each filter (median 60; min 40; max 260).

2.3.2 Visual observation, size measurements and FTIR analysis

Visual characterization of the fibers was performed using a stereomicroscope coupled with polarized light to improve the efficiency of fiber identification (Santana et al., 2016). The following criteria were used to classify synthetic fibers: the absence of cellular structures and scales on the surface, a uniform surface, the presence of equal thickness along its entire length, curved shape, coloration and solid strands (Hidalgo-Ruz et al., 2012, Wright et al., 2013, Barrows et al., 2018). The visual characterization of fibers is still an important tool given the considerable limitation of the use of FTIR spectroscopy for the analysis of textile fibers (Stanton et al., 2019). We have taken great care to avoid the overestimation of synthetic microfibers, therefore, we also analyzed images from the different washing experiments to distinguish between synthetic and non-synthetic fibers. All samples and particles that did not matched the above criteria were not considered as microplastics (about 40-60%) and were discarded from data analyzes. Consequently, in this study abundance and size measurement are then given only for the synthetic microfibers (MFs).

Size measurements were carried out in a subset of the samples. For each filter 200 images were made corresponding to the Dollfus cell squares and the synthetic fibers that followed the above criteria were selected and the length was analyzed using ImageJ v.1.5 software (Schneider et al., 2012). A total of 2673 MF was measured.

In order to validate the visual identification of MFs the chemical analysis of a subsample of particles was conducted by FTIR. Analyses were carried out using a Nicolet ™ iN10 MX infrared imaging microscope (ThermoFisher Scientific) to locate MFs on filters and obtain their IR spectrum. To screen the filters within a reasonable time and obtain a representative distribution of chemical nature of fibers, the number of MFs to be analyzed was determined using calculation methods developed by Kedzierski et al., (2019). Considering the heterogeneity in the total numbers of fibers between samples, the minimum number of fibers to be analyzed for each sample was defined to reach at least 15% of confidence interval. MFs were randomly picked on the filters and analyzed by FTIR.

The analyses were performed with a variable aperture adapted to the size of each fiber, on MFs up to a diameter of 5 µm and on surfaces up to 300 µm² (60 µm long). To improve the contact with the fiber and to optimize the acquisition of the FTIR signal, the spectra were collected in ATR (attenuated total reflectance) mode using 16 scans of 4000 cm⁻¹ at 400 cm⁻¹, with a resolution of 4 cm⁻¹. Spectra processing and fiber identification was performed using OmnicTM Specta software (ThermoFisher Scientific) to characterize the proportion of synthetic and natural MFs and to assess the predominant polymers. An automated baseline correction was performed on each spectrum and the corrected spectra were compared with 2 different polymer libraries: a commercial database (Hummel polymers library) and an open access polymer database published by Primpke et al., (2018). The hit quality index for identification of fibers was set at 70%. Indeed, the use of a single commercial polymer database (Hummel polymer and additives) can provide low matching scores or inaccurate identification. The comparison with a second library containing spectra from a wider type of component based on polymers, plant types and animal furs and considering the oxidized nature of environmental samples clearly improved the identification of the nature of the MFs (Primpke et al, 2018).

Manual assessment was performed to refine the chemical identification when the spectrum signal lacked intensity and provided a low matching score with the libraries. Characteristic absorption bands of reference synthetic polymers were sought in the sample spectra. The polymer identification was validated using a minimum of four matching absorption bands (Jung et al., 2018).

As commonly observed in other studies, the shape, length and positioning of the MFs do not always allow the acquisition of accurate and intense IR spectra (Mintening et al., 2017). The small diameters of the MFs made it impossible to target a zone of analysis restricted to the fiber, which complicated the acquisition of the spectrum. Indeed, it is difficult to go below a filter surface of 225 square microns (e.g., MFs of 15 x 15 microns) and obtain an easily identifiable spectrum for MFs smaller than 15 μ m in diameter. In addition, the fiber configuration and 3D positioning on the filter surface did not always allow analysis on the same focal plane, which limited the recorded signal and did not allow the acquisition of spectra by automatic methods. Therefore, the acquisition of spectra was performed individually, fiber by fiber, on subsamples with a total of 104 fibers out of 295 analyzed.

2.4. Statistical Analyses

Statistical analysis were performed using the statistical software R-v.2.15.0 (R Development Core Team, 2009). The Kruskal-Wallis (KW) was used to test the significance in MFs concentrations as a function of season and sampling locations. The Spearman correlation test was performed to study the correlation between the amount of MFs by season, between MFs and suspended solids, and between the amount of MFs and distance to land. One-Way ANOVA was used to verify significant differences in MFs size among samples as data were normal distributed (Shapiro-Wilk normality test; W = 0.99 to 0.98) and the Tukey

HSD test analyses were applied. The statistical significance threshold was set at a p-value of 0.05.

3. Results and discussion

3.1. Contamination control

The number of MFs observed in the laboratory blank control ranged from 2 to 14 items per filter analyzed. The average contamination observed in fluids passing through the experimental devices was 6.3 ± 4.6 MFs L⁻¹. The air checks carried out over 24 hours using automatic samplers placed at the inlet and outlet of the Haliotis plant gave an average contamination of 15.6 ± 9.4 MFs L⁻¹ (Table 1A). These high values compared to laboratory controls are probably due to the fact that the samples are exposed to the air for a longer period of time. The resulted blank data were systematic subtracted from the respective results.

Overall, blank test results indicate that contamination can be a significant source of error (14% median value). This underlines the importance of a correct evaluation of the sources of contamination in order to better take into account this category in routine microplastic analysis. In order to correctly determine the distribution and occurrence of LF, systematic quality control must be performed during sampling and processing and taken into consideration when processing the data.

3.2. Microfiber characterization

The FTIR fiber analysis of textile washes considered 100% synthetic showed that 17.4% of the microfibers were natural or derived from the transformation of natural polymers.

Although this percentage is related to the number of fibers identified as synthetic/natural in the set of fibers analyzed, we can conclude that significant contamination by non-synthetic fibers may occur during the analyses. At the end of the treatment process of the Haliotis plant due to the high removal efficiency, we were unable to identify the plastic polymers, probably due to the small number of MFs observed on the filters. A cross-check of airborne contamination suggests that values were below 10³ MFs per cubic meter and near the detection limit. Nevertheless, we kept the estimates from microscope counts, knowing that this could create a potential underestimate of the non-synthetic microfibers, which were not fully classified in the initial analysis.

In the marine samples, the overall results showed that 14-50% of the raw materials analyzed were synthetic, with the highest rate observed in surface waters at the Haliotis outfall site. The remaining 35-72% are of natural origin (cotton, wool) or made by processing natural polymers (especially cellulose) and the rest are a mixture of different materials or could not be identified (14-21%) as explained above (Table 2; Table S2). This is slightly higher than the results of textile production, where more than half of the world's production is based on plastic polymers with about 49% of the world's clothing is made of polyester (Sundt et al., 2014). The relative high presence of natural fibers is consistent with the results obtained by Suaria et al., (2020) who found high proportion of animal- and plant-based fibers throughout world's oceans with values of 6.8% for the Mediterranean Sea. Although, compared to our results this estimate is lower. Among synthetic fibers, polyamide (30%) and polyester (22%) are the most important polymers in urban wastewater and it should be noted that this material is also predominant in offshore sites with polyester reaching 62% at the Dyfamed site where the anthropogenic impact should be reduced. The first observations concerning the characterization of microplastics at the outlet of wastewater treatment plants have shown that polyester fibers are the main source of contamination and contribute to environmental pollution to a larger extent than microplastics resulting from fragmentation of larger plastics (Napper and Thompson, 2016). This is probably because of its low cost and versatile use. Indeed, polyester are the most widely used and the dominant types of polymer clothing including high-performance outdoor wear but also in-home furnishings, and as a reinforcing fiber in tires, belts, and hoses (IEEE GlobalSpec, 2019). Polyamide density ranges from 1.02 to 1.15 g cm⁻³ (Hidalgo-Ruz et al., 2012) so we can expect that only less dense PA could float explaining the lower concentration of this polymer in seawater (Reisser et al., 2013, Cozar et al., 2014). However, the higher density of Mediterranean waters (usually > 1.026 g cm⁻³) could also explain the relatively high proportion of polyamide found in its surface waters (Pedrotti et al., 2016). Polypropylene, polystyrene and polyvinyl chloride were only found at the Haliotis outfall site, while polyvinyl acetate was only found at the Point B station. Polyvinyl acetate is known as a major constituent of fishing nets and nets are widely used in the region.

In our samples, we were able to identify several types of synthetic MFs representing up to fifty percent of total fibers collected. We also showed that un important fraction of fibers is from natural origin (35-72%). The most striking results showed the presence not only of polyester and polyamide, but also of polypropylene and acrylic MFs, albeit in smaller quantity. Further studies should include size and shape for characterization of MFs in air and seawater and improve the spectral intensity and resolution of the analysis down to small size in order to understand their sources, transfers and persistence in the environment.

3.3. MFs measurement in the different compartments

The majority of the synthetic fibers collected in this study are sub-millimetric in size.

The overall length distribution is asymmetric with a fiber frequency increase with decreasing

length suggesting the presence of shorter fibers (Fig. 2). The lower size limit of MFs analyzed is 25 µm in aerosols samples and washing machine samples and maximum values 7240 µm are detected in washing machine experiments. In the marine compartment, the length of the marine MFs, irrespective of the location (coastal stations, Pt. B and offshore Dyfamed), was not significantly different (F 0.25; p > 0.8) and were pooled together for further analyses. Comparison of MFs length revealed significant differences between different environments sampled (air, washing machine, WWTP outlet and marine, ANOVA; p = 0.0001) (Table S3). The smaller size length was observed in aerosols (median 83µm) with predominant fibers lengths of 50-200 µm (75%) suggesting different sources than the other compartments (Fig. 2). A similar distribution was found for total atmospheric deposition fibers (Dris et al., 2016, 2017). This suggests that smaller microplastics may persist and be transported in the atmosphere, deposited or inhaled continuously, with potential long-term threats to ecosystems and human health (Gasperi et al 2018; Allen et al., 2019, Lui et. al., 2019). The median length of MFs collected in the washing machine and in the outlet of the WWTP are respectively 287 and 364 µm. The length of synthetic fibers released during washing experiments depends on the fiber type and fabric characteristics, in the washing experiments Napper and Thompson (2016) found longer lengths for polyester, polyester/cotton blend and acrylic 5.0 to 7.8 mm suggesting differentiated retention in WWTP. This could explain the significant difference between size of MFs transiting in the Haliotis WWTPs. MFs in the marine compartment have median length values of 589 µm and the largest range of size from 56 to 5,263 µm. The predominant fiber length is between 150 and 550 µm (48%). The upper class of MFs (>1000 µm), which is rare in aerosols, in washing machine effluents or at the outlet of sewage treatment plants, indicates a source other than wastewater treatment.

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3.4. Air compartment

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In the aerosol experiments, only fibers dominate the shape of the microplastics (Table 1B). The aerosol fibers could not be analyzed by FTIR, the results come from visual observations. Their average concentration in the area of Nice was 0.043 ± 0.026 MFs m⁻³, while the average concentration in Cap Ferrat, 25 km away, was three times higher $(0.140 \pm 0.120 \text{ MFs m}^{-3})$ although not significantly different. These urban areas are not industrialized nor densely populated and the concentration values obtained are an order of magnitude lower (0.3 and 1.5 fibers m⁻³) than those found in an outdoor area outside from Paris (Dris et al., 2017). On the other hand, the average atmospheric deposition rate of MFs in Nice (average 392 ± 141 particles m⁻²day⁻¹) is higher than that reported in a densely populated urban area of Paris (118 and $110 \pm 96 \text{ m}^{-2}\text{day}^{-1}$, Dris et al., 2015, 2016) and in Dongguan, China $(36 \pm 7 \text{ m}^{-2}\text{day}^{-1}, \text{Cai})$ et al., 2017) and a slightly lower than in the megacity of London (712 \pm 162 microplastics m⁻² day⁻¹, Wright et al., 2020). As in our survey, in all the above-mentioned sites, the fibrous form accounted for the vast majority of microplastics. Results similar to ours were found in a pristine mountain catchment (French Pyrenees) (365 m² day⁻¹, Allen et al., 2019) and in Hambourg metropolitan region (275 microplastics m² day 1; Klein and Fischer 2019) with the dominant shape of microplastics identified as fragments. Air mass trajectories and winds were suggested as a possible transport route for microplastics to these sparsely inhabited areas (Allen et al., 2019). The different methodologies used in the previous work as well as the detection of different shapes of particles at different thresholds make comparisons difficult. However, these results suggest that although cities are a source of airborne microplastics, higher population density alone does not seem to explain their abundance of and that factors other than proximity to cities, such as weather and precipitation rates and climate, also play an important role in the transport, dispersion and deposition of MFs in the wider environment (Dris et al., 2016, Allen et al., 2019).

Data presented here, while corroborating other studies, should be interpreted with caution as only a few experiments were conducted and because fiber identification was based solely on visual inspection giving place to a possible overestimation of synthetic fibers. The transport of urban atmospheric microplastics can be an important pathway for contamination of the marine environment and future research should include this compartment in monitoring activities.

3.5. Microfiber release from domestic washing machines

The MFs observed after cleaning the washing machine with two wash cycles before each run represented only 0.3% of the MFs collected from the samples. We can therefore conclude that there is no contamination from the empty machine. Our results showed that during conventional washing, from 3.2×10^6 to more than 3.9×10^7 MFs per cubic meter (average of 12.8×10^6 MFs m⁻³) could be released (Table 3). This corresponds to 3 to 99 fibers per gram of the garments released during washing (Table S1). This clearly indicate that the shedding of fibers from clothing during washing is an important emitter of microplastics. The design of garments, including the type of fabric used, and the washing treatment can significantly affect the amount of fibers released (Napper and Thompson, 2016; De Falco et al., (2019). In addition, the age of the textile can be another factor to consider (Hartline et al., 2016). Given this diversity, a daily wash was simulated with different types of garments and different load sizes to assess the number of MFs shed in a conventional wash. Our results corroborate reported levels of shedding from fabrics found by Pirc et al., (2016) and Almroth et al., (2018) and are higher than those observed by Napper and Thompson (2016) who estimate that over

7.0 x 10⁵ MFs fibers could be released from an average 6 kg wash load of acrylic fabric. As garments are mixed during most washing experiments, we cannot evaluate the relationship between the type of fabric and the number of MFs released but according to Almroth et al., (2018) no significant difference was found between nylon, acrylic, and polyester knits however these authors found that polyester fleece fabrics release a significantly greater number of fibers than other knits. This corroborates our finding of the two washing experiments using 100% polyester (Table S1); there is an order of magnitude difference between polyester fleece fabrics (R1) compared to the smarter polyester (R2) used for ski jackets and pants.

Our results allow us to estimate the expected concentration of textile microfibers resulting from the washing of clothing conveyed in sewage to and treated in the Haliotis plant in Nice. If we consider a population of 358,000 inhabitants connected to the WWTP and waste water from an average of 0.3 laundries per inhabitant per day (statistics published on www.planetoscope.com) with an average volume of 75 liters of effluent discharged by each machine, the average flow treated by the Haliotis plant per day from the washing machines is approximately $8.1 \times 10^3 \text{ m}^3$. This effluent corresponds to 6% of the average flow treated daily by the Haliotis plant. An average of 1.03×10^{11} MFs textiles enter the treatment plant daily (min 2.62×10^{10} ; max 3.19×10^{11}). With a daily flow of $120,000 \text{ m}^3$, we therefore expect an average of 8.58×10^5 MF textiles m⁻³ (min 2.17×10^5 ; max 2.66×10^6) of wastewater entering the Haliotis treatment plant daily (Table 3). MFs disseminated by domestic washing constitute an important part of microplastics in the environment; given the increase in the production and consumption of synthetic textiles, this pollution has become an urgent issue (Cesa et al., 2017).

3.6. Efficiency of the WWTP and estimated discharge of synthetic MFs in the marine environment

Large quantities of MFs were detected at the treatment plant inlet with an average of 5.31 x 10⁵ items m⁻³. These concentrations are consistent with those estimated from the domestic washing experiments (8.56 x 10⁵ MFs m⁻³) validating the consistency of the methodology used (Table 3). At the outlet of the WWTP Haliotis an average concentration of 3.6 x 10⁴ synthetic MFs per cubic meter was counted (Min 6 x 10³; Max 6.9 x 10⁴), (Table 3). The concentration of suspended solids at the inlet of the treatment plant varies from 212 to 293 μ g m⁻³ (average 257.14 \pm 24.47 μ g m⁻³) and at the outlet from 8 to 30 μ g m⁻³ (17.5 \pm 6.8 μ g m⁻³), (Table S4). MFs concentrations varies significantly with the amounts of suspended solids in the inlet of the treatment plant (r_s = -0.84, p = 0.006) while in the outlet no correlation was observed probably because of the small number of particles. This large variation highlighted that sediments and large particles in wastewater could efficiently aggregate with synthetic MFs and act as a pre-filter in the sewer system. It is assumed that, in rainy weather, this natural elimination of MFs will not occur.

Wastewater treatment plants are thus very effective in removing microplastic particles from the wastewater stream and the Nice Haliotis treatment plant has a significant removal percentage ranging from 87.5%, to 98.5% (average 91.87 \pm 5.52%) (Fig. 3). These abatements are in the same range as the WWTP found at the Detroit WWTP (95.6%) (Michielssen et al., 2016) and at the Glasgow (98.41%) (Murphy et al., 2016) using activated sludge as secondary treatment. We observed a significant seasonal effect (p = 0.003) in the concentration of MFs collected at the inlet of the Haliotis with higher concentrations in early spring (Fig. S2.A). This result is consistent with clothing customs since technical and warmer clothing is mainly made of synthetic and plastic MFs. However, no seasonal differences were

detected in the number of synthetic MFs leaving the treatment plant (Fig. S2.B), highlighting the efficiency of treatment and the sensitivity of detection.

Despite the high removal efficiency of the Haliotis plant, based on the large volume of treated wastewater and effluent discharged into surface waters (120,000 m³ per day), we have estimated that 4.3 billion fibers enter daily the marine environment from the treatment plant. This value is in line with values reported by Michielssen and his colleagues (2016) where 8.9 billion MFs seem to be discharged daily from the Detroit plant. WWTPs are also known sources of microplastics in the freshwater environment, given their incomplete removal during treatment (Estahbanati and Fahrenfeld, 2016). As machine filters and WWTPs are not specifically designed to retain all fibers, this means that the washing of synthetic garments is likely to be a significant source of fiber to the environment.

Although wastewater treatment plants are effective at removing microfiber particles from the wastewater stream, effluent discharges often contain high concentrations of MFs. These fibers are found in the environment at concentrations of up to thousands of particles per cubic meter, being available to be ingested by a broad range of species. Other problems remain unanswered, as a large proportion of these particles will settle in the sludge, they may become an environmental polluter that should be monitored.

3.7. Spatial-temporal patterns of microfiber contamination in surface waters of the Mediterranean Sea

During the two campaigns from February to October 2014, MFs were found in all samples collected with an overall mean concentration of $1.06 \times 10^4 \pm 6.77 \times 10^3$ MFs m⁻³ (median 1.07×10^4 MFs m⁻³). Maximum MFs concentration was found offshore at Dyfamed station and in the Bay of Naples (respectively 2.54 and 3.70 $\times 10^4$ MFs m⁻³) while lowest

concentration was found in Bastia station (3.31 x 10³ MFs m⁻³), (Fig. 1). Results of seasonal monitoring in front of the Haliotis WWTP (outfall, upstream and downstream the outfall, east and west of the outfall), showed comparable mean concentrations of MFs (1.32 x 10⁴ \pm 1.35 x 10³ m⁻³; Fig. 1), and no significant differences were observed between Haliotis WWTP, Point B and Dyfamed during the year (Fig. 4). In addition, we found no geographical differences in MFs abundance along the sampling sites in NW Mediterranean Sea characterized by different levels of anthropisation, nor at a depth of 10 m at point B (1.63 x 10⁴ \pm 1.0⁵ x 10⁴ MFs m⁻³). Finally, no relationship was found between MFs concentrations and distance to land (r_s = -0.0853), probably due to their rapid dilution and transport.

Our values are several orders of magnitude higher than those reported in previous studies where a wide range of concentrations ranging from 0 to more than 450 MFs m⁻³ was found (Gago et al., 2018). These studies are however difficult to compare because of the heterogeneity of approaches used. The majority of studies focus on the surface layer and most of them use 333 μ m manta nets where most fibers are not sampled and even the larger size are underestimated due to the difficulty of collecting and identify them properly (Gago et al., 2018).

Few studies have used a mesh size of 20 µm to study marine MFs. Our study, with that of Suaria et al., (2020), are the only ones available for the Mediterranean Sea. By compiling a global dataset of fibers collected in six ocean basins, Suaria et al., (2020) showed that the highest concentration of fibers is found in Mediterranean Sea with a median of 4.2 fibers L⁻¹. Although the proportion of synthetic and non-synthetic products differs between the two studies, the abundance values obtained in this study are higher than those obtained in the previous one. Microfiber pollution is a complex and worrisome threat (Carr et al., 2017) as it is present in the majority of samples collected from the sea surface, from filter-feeders' gut contents (Lusher et al., 2013, Devriese et al., 2015), from the deep sea and the sediments

(Sanchez-Vidal et al., 2018). It is likely that fibers collected in the vicinity of Haliotis are mainly from urban sources, the size categories of fibers that WWTP Haliotis send to the ocean is within 25-600 µm (73%). Smaller fibers observed in situ are likely to come from airborne fibers and their continuous atmospheric deposition may contribute to maintaining the large reservoir observed. The larger fibers (>1000 µm) found mainly in this environment most likely come from marine activities such as fishing, aquaculture and shipping. Almost the whole global fishing activities use synthetic materials as polyamide and polyolefin, where some of the most common lost or discarded objects, constituted of textile fibers, comprise ropes, lines and fishing nets (Cole et al., 2011; Sundt et al., 2014). As revealed in a study in the Great Lakes (Baldwin et al., 2016), the estimation of microfiber concentrations in aquatic environments is higher than expected from sanitation systems only. These results underscore that the release of laundry fibers, as previously assumed (Browne et al., 2011; Eriksen et al., 2013; Mason et al., 2016), is not the only route of dissemination of synthetic MF in the aquatic environment (Raju et al., 2018). A significant fraction of this leakage originates also from rivers that transport plastic waste from further inland and concentrate the litter following extreme rainfall rains and floods events (Axelsson and van Sebille, 2017). Our results showed that MFs seem to be ubiquitous and widespread found in concentrations of the same order of magnitude in urban areas as in offshore surface waters of the northwestern Mediterranean. The sources of MFs in the marine environment are multiple, with laundry fibers discharges from WWTP and atmospheric transport of urban fibers being among the important pathways.

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4. Final remarks

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We implemented a relevant methodology to study the transport dynamics of synthetic MFs from urbanized areas to the open sea. We analyzed and quantified atmospheric

contribution, so far, only a few studies have investigated atmospheric microplastics. Our study contributes to the construction of a global assessment of MPs across different environmental compartments.

Figure 5 summarizes the flow of synthetic MFs from the atmosphere to the surface of the western Mediterranean Sea. The MFs from washing machines represent an important part (average of 12.8×10^6 MFs m³) of microplastics entering the WWTP with large quantities detected in the inlet (5.31×10^5 MFs m⁻³). Although Haliotis is very effective (up to 98.5%) in removing microfibers, large concentrations (2.6×10^3 to 3.70×10^4 m⁻³) are found in nearshore and offshore marine areas.

Understanding the sources and dispersion of MFs is a major issue for policy development and prevention of this emerging pollution. Coastal urban areas have sanitation systems whose efficiency varies greatly according to the types of pollution. Hydraulic regimes (rainfall, floods) may cause massive discharges into the sea (Fisher et al., 2015). Once removed from wastewater, fibers end up as biosolids a nutrient-rich waste used as a fertilizer for soils that can persist for decades (Bayo et al., 2016; Dris et al., 2015). The application of biosolids to soils likely results in the particles ending up in rivers and lakes and being exported to the aquatic environment (Crossman et al., 2020).

Preventive solutions are needed to deal with the impact of man-made MFs on marine life and human health. In order to reduce fiber contamination in the environment, future research should be directed towards upstream techniques using strategies that can mitigate this problem. As suggested by Almroth et al (2018), making textiles smarter, reducing fiber losses in manufacturing or using more efficient filters in domestic washing machines are possible solutions. However, the ultimate solution to curb this pollution, which is entirely of human origin, can only be achieved through a collective effort to modify use of plastics.

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Tables and figure legends

- **Table 1.** Summary of the concentration of synthetic microfibers counted. A- In the laboratory control experiments and in the controls of the autosampler of the Haliotis WWTP (MFs L⁻¹).
- B- In atmospheric samples in St-Jean Cap-Ferrat and in Nice-Haliotis site (MFs m⁻³) and in the atmospheric fallout in Nice-Haliotis site (MFs m⁻² day ⁻¹). N: number of experiments.

Table 2. ATR- FTIR analysis of microfibers collected in urban and marine waters with the respective polymer composition of synthetic fibers (%).

Table 3. Synthetic microfibers concentrations (MFs m⁻³) released during a conventional machine washing, estimation of washing input in WWTP and concentrations observed at the inlet and outlet of the Haliotis WWTP.

Figure 1. Concentrations of synthetic microfibers (MFs m⁻³) in surface waters of the northwestern Mediterranean Sea in the study conducted from February to October 2014. The 15 sampling sites are highlighted with the zoomed white square in the upper right corner corresponding to the grid of 5 sampling sites in front of the Nice-Haliotis wastewater treatment plant, the coastal station off Nice, Point B and the station off Villeneuve Loubet. The points with the yellow crosses are the two inland stations at Cap Ferrat and Haliotis, where atmospheric fallout was studied. The colored circles represent the concentrations of MFs (m⁻³) in marine samples: Blue 3.3 - 9.1 10³ MFs m³; Orange 1.2 - 1.5 10⁴ MFs m⁻³ and Red 2.3 - 2.5 10⁴ MFs m⁻³ respectively.

Figure 2. The frequency distribution of synthetic microfiber lengths. A- In atmospheric fallout (n = 351). B- In washing machine effluents with 100% synthetic textiles (n = 1898). C- At the outlet of the Haliotis treatment plant (n = 14) and D- In the *in situ* samples (380). The initial bins are 25 μ m and 50 μ m, then they are grouped together by 100 μ m bins.

Figure 3. Seasonal concentrations of synthetic microfibers (MFs m⁻³) at the inlet (dark grey) and outlet (light grey) of the Haliotis WWTP. The percentage (%) corresponds to the disposal efficiency of the WWTP Haliotis.

Figure 4. Concentrations of synthetic microfibers (MFs m⁻³) recovered from surface samples from February to October 2014 in front of the Haliotis WWTP, at Point B station, and in Dyfamed station. Boxes show the 25-75 percentiles, with median value as a central line; whiskers denote upper and lower inner fences values.

Figure 5. Pathway of synthetic microfibers (MFs) from atmospheric fallout and domestic wash to the WWTP and to the Mediterranean Sea. The arrows symbolize the flow of MFs.

The figures are average densities values for each compartment. Boxes are concentrations of MFs (m⁻³) from domestic washing, wastewater treatment plant outlet and the Mediterranean Sea surface, it shows the 25-75 percentiles, with median value as a central line; whiskers indicated upper and lower inner interval values, and the outer values are indicated by crosses.

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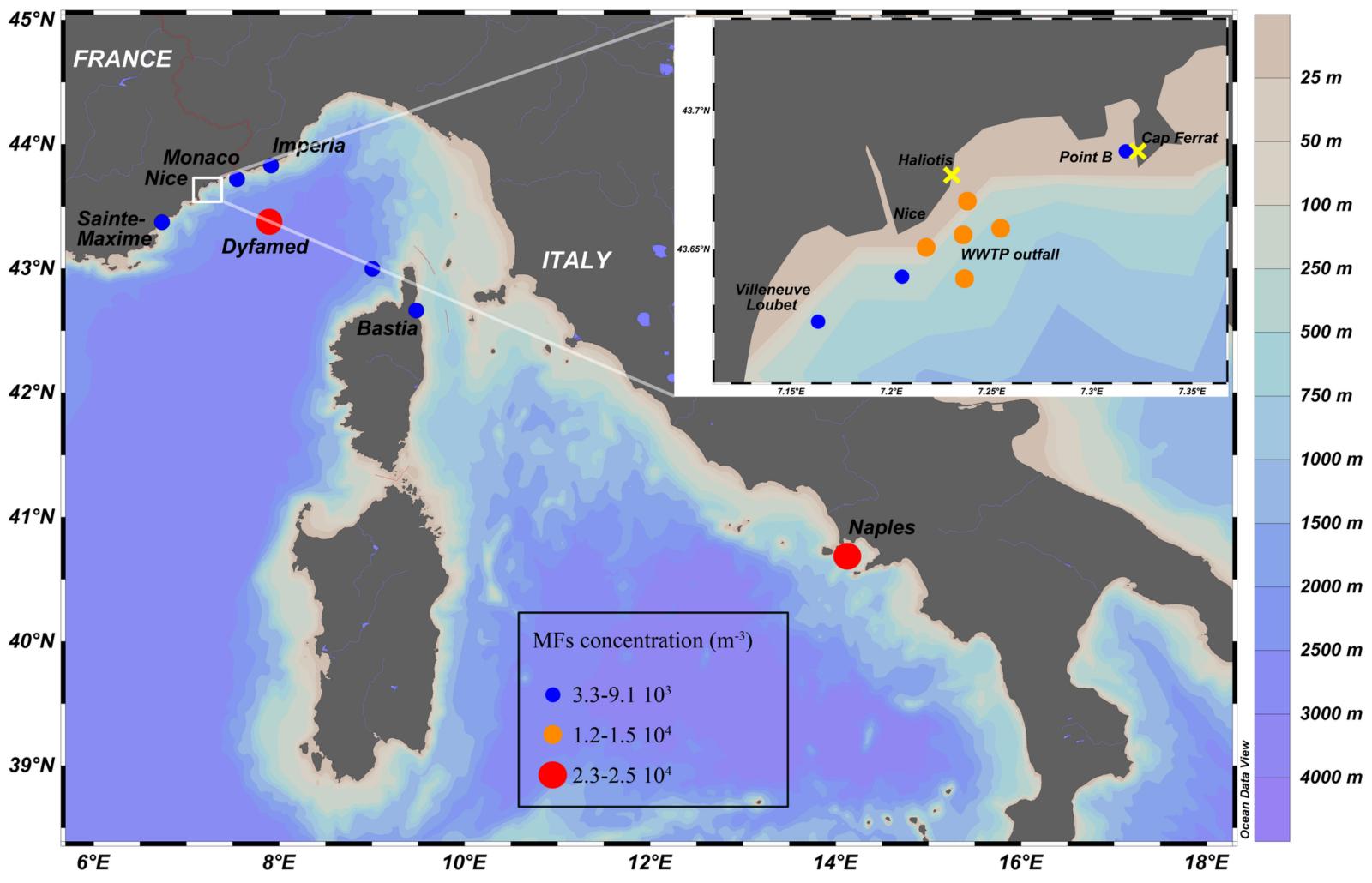
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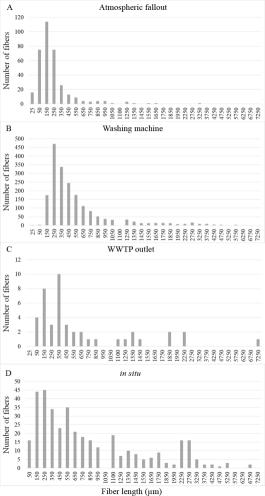
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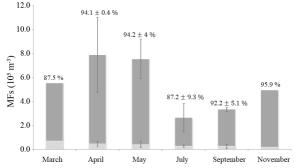
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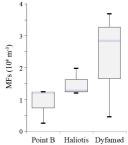
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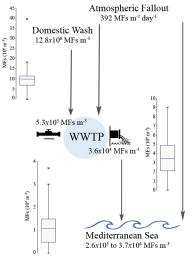


Table 1. Summary of the concentration of synthetic microfibers counted. A- In the laboratory control experiments and in the controls of the autosampler of the Haliotis WWTP (MFs L^{-1}). B- In atmospheric samples in St-Jean Cap-Ferrat and in Nice-Haliotis site (MFs m^{-3}) and in the atmospheric fallout in Nice-Haliotis site (MFs m^{-2} day $^{-1}$). N: number of experiments.

A	N	MFs (L ⁻¹)
Airborne contamination (lab control)	14	$6.3 \times 10^3 \pm 4.6$
Airborne contamination (autosampler)	4	$15.6 \times 10^3 \pm 9.4$
В	N	$MFs (m^{-3})$
Aerosol (St-Jean Cap-Ferrat)	3	0.140 ± 0.120
Aerosol (Nice-Haliotis site)	2	0.043 ± 0.026
		MFs $(m^{-2} day^{-1})$
Atmospheric fallout (Nice-Haliotis site)	2	392 ± 141

Table 2. ATR- FTIR analysis of microfibers collected in urban and marine waters with the respective polymer composition of synthetic fibers (%).

	Total Fibers (nb)	Analyz fibers (Analyzed %	Unl	known %	Natura %	l Synthetic %
Haliotis Outfall	65	27		41	15	5	35	50
Point B	23	15		65	14	1	72	14
Bastia	32	24		75	21		58	21
Dyfamed	178	38		21	18	3	47	34
	PA	PAN	PES	PP	PS	PVA	PVC	
Haliotis Outfall	30	13	22	9	17		9	
Point B	17	17	33			33		
Bastia	60		40					
Dyfamed	38		62					

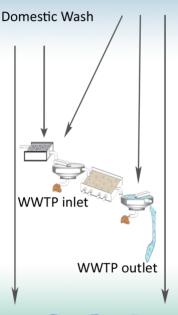
PA Polyamide; PAN Acrylic; PES Polyester; PP Polypropylene; PS Polystyrene; PVA Polyvynil acetate; VA Vinyl acetate

Table 3. Synthetic microfibers concentrations (MFs m⁻³) released during a conventional machine washing, estimation of washing input in WWTP and concentrations observed at the inlet and outlet of the Haliotis WWTP.

	Conventional washing	Estimated washing input in WWTP	Inlet Haliotis WWTP	Outlet Haliotis WWTP
Min	3.2×10^6	2.17×10^5	1.57×10^5	6×10^3
Mean	12.8×10^6	8.56×10^5	5.31×10^5	3.6×10^4
Max	3.9×10^7	2.66×10^6	1.02×10^6	6.9×10^4







Mediterranean Sea