

Metal pollutants have additive negative effects on honey bee cognition

Coline Monchanin, Erwann Drujont, Jean-Marc Devaud, Mathieu Lihoreau,

Andrew B. Barron

► To cite this version:

Coline Monchanin, Erwann Drujont, Jean-Marc Devaud, Mathieu Lihoreau, Andrew B. Barron. Metal pollutants have additive negative effects on honey bee cognition. Journal of Experimental Biology, 2021, 224 (12), 10.1242/jeb.241869. hal-03438756

HAL Id: hal-03438756 https://hal.science/hal-03438756v1

Submitted on 22 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Metal pollutants have additive negative effects on honey bee cognition
2	
3	Running title: Metal pollutant cocktails impair bee cognition
4	Author list
5	Coline Monchanin ^{1,2} , Erwann Drujont ¹ , Jean-Marc Devaud ¹ , Mathieu Lihoreau ¹ , Andrew B.
6	Barron ²
7	¹ Research Center on Animal Cognition (CRCA), Center for Integrative Biology (CBI); CNRS,
8	University Paul Sabatier – Toulouse III, France
9	² Department of Biological Sciences, Macquarie University, NSW, Australia
10	
11	Corresponding author: coline.monchanin@univ-tlse3.fr
12	
13	Keywords: Apis mellifera, PER conditioning, pollutant interaction, arsenic, lead, copper
14	
15	Summary statement:
16	Honey bees displayed reduced learning and memory performance following acute exposure to

arsenic, copper or lead. Exposure to combinations of these metals had additive effects.

- 18 Abstract
- 19

20 Environmental pollutants can exert sublethal deleterious effects on animals. These include 21 disruption of cognitive functions underlying crucial behaviours. While agrochemicals have 22 been identified as a major threat to pollinators, metal pollutants, which are often found in 23 complex mixtures, have so far been overlooked. Here we assessed the impact of acute exposure 24 to field-realistic concentrations of three common metal pollutants, lead, copper, arsenic, and 25 their combinations, on honey bee appetitive learning and memory. All treatments involving 26 single metals slowed down learning and disrupted memory retrieval at 24 h. Combinations of 27 these metals induced additive negative effects on both processes, suggesting common pathways 28 of toxicity. Our results highlight the need to further assess the risks of metal pollution on 29 invertebrates.

30

31 Introduction

32

33 Metal pollution is of increasing concern for both ecosystem and public health (Nriagu and Pacyna, 1988). Over the last century, the widespread use of metals in domestic, industrial and 34 agricultural applications (Bradl, 2005) has considerably elevated their concentrations in water 35 36 (Mance, 1987) and terrestrial habitats (Krämer, 2010; Su et al., 2014) to potentially toxic levels. 37 Pollinators, such as honey bees, are directly exposed to metal pollutants when foraging on contaminated nectar and pollen (Perugini et al., 2011; Xun et al., 2018), and while flying 38 39 through air containing suspended particles (Thimmegowda et al., 2020). Metals accumulate in the bodies of adults (Giglio et al., 2017) and larvae (Balestra et al., 1992), as well as in the hive 40 41 products (Satta et al., 2012). For instance, concomitant bioaccumulation of arsenic (As), copper 42 (Cu) and lead (Pb), resulting from metal production industries (Kabir et al., 2012) and mining 43 (Khaska et al., 2018; Lee et al., 2005), is common in honey bees (Badiou-Bénéteau et al., 2013; 44 Giglio et al., 2017; Goretti et al., 2020) and their honey (Pisani et al., 2008; Terrab et al., 2005). 45 The deleterious effects of metals on humans (Tchounwou et al., 2012) and some model animals (mice: Cobbina et al., 2015; flies: Do anlar et al., 2014) are well-known. As, Cu, Pb 46 and other metals have neurotoxic effects that induce neural and neuromuscular alterations, 47 sensory impairments and many related forms of behavioural dysfunctions (Chen et al., 2016). 48 49 Deficits in cognition and memory have been reported for As (e.g. humans: Tolins et al., 2014;

mice: Tyler et al., 2018; Wu et al., 2006), Pb (e.g. mice: Anderson et al., 2016; humans: Mason
et al., 2014) and Cu (e.g. mice: Lamtai et al., 2020; Pal et al., 2013; flies: Zamberlan, 2020).

2

52 Recent studies showed that low doses of Pb (Monchanin et al., 2021a) and selenium (Se)
53 (Burden et al., 2016) also impair behaviour and cognition in honey bees, suggesting a
54 widespread impact on pollinators. So far, however, very little attention has been given to the
55 potential combined effects of co-exposure to different metals (Monchanin et al., 2021b).

56 Interactions among stressors are commonly classified as antagonistic (when the effect 57 of one stressor reduces the effect of the other one), additive (when stressors have simple 58 cumulative effects) or synergistic (when stressors together have a greater effect than the sum of 59 their individual effects) (Folt et al., 1999). Additive effects of As, Cu and Pb have been 60 described for humans (Lin et al., 2016), rats (Aktar et al., 2017; Mahaffey et al., 1981; Schmolke et al., 1992) and fishes (Verriopoulos and Dimas, 1988). In rats, for instance, co-exposure to 61 Pb and As disrupted brain biogenic amine levels (Agrawal et al., 2015). In humans, it was 62 hypothesized that combined exposure to Pb and As, or other metal pollutants, have additive or 63 synergistic toxic responses leading to cognitive dysfunction (Karri et al., 2016). To our 64 65 knowledge, two studies have addressed the impact of metallic cocktails on bee physiology. Honey bees simultaneously exposed to Pb, cadmium (Cd) and Cu accumulated significant 66 levels of these metals in their body and had lower brain concentrations of dopamine compared 67 to unexposed honey bees (Nisbet et al., 2018). Cd and Cu exerted a weak synergistic effect on 68 69 honey bee survival (Di et al., 2020). However, none of these studies investigated potential 70 effects of combined exposure on cognition.

71 Here we compared the effects of an exposure to single metals or ecologically relevant 72 combinations of these metals on honey bee learning and memory. We hypothesised that 73 combinations of metals may have synergistic negative effects, as it has been found with 74 pesticides (Yao et al., 2018; Zhu et al., 2017). We tested individual honey bees in a standard 75 protocol of proboscis extension reflex (PER) conditioning following acute exposure to As, Pb 76 and Cu or a combination of them. We tested three concentrations of As, considered the most 77 toxic substance (ATSDR, 2019), and added one concentration of Cu or Pb (binary mixtures), 78 or both (tertiary mixture), to reach the molarity of the As solutions, allowing us to better assess 79 any combined effects.

80

81 Materials and methods

82

83 *Metal solutions*

Arsenic (NaAsO₂), lead (PbCl₂) and copper (CuCl₂2H₂O) were purchased from Sigma-Aldrich
Ltd (Lyon, France) and diluted in 50% (w/v) sucrose solution. Control honey bees were fed

86	50% sucrose solution. Three concentrations of As were used (Table 1): a low concentration
87	(0.13 $\mu M)$ corresponding to the maximal permissible value in drinking water (0.01 mg.L^-1)
88	(Codex Alimentarius, 2015), a high concentration (0.67 μ M) corresponding to half the maximal
89	permissible value in irrigation water (0.1 mg.L ⁻¹) (Ayers and Westcot, 1994), and an
90	intermediate concentration (0.40 μ M). This range of concentrations was reported in water
91	sampled from polluted areas (e.g. mining sites) and in honey (Table S1). For Pb and Cu, we
92	chose 0.27 μM (0.055 mg.L $^{\text{-1}}\text{of}$ Pb and 0.017 mg.L $^{\text{-1}}\text{of}$ Cu) so that the binary combinations (As
93	$0.13~\mu M$ + Cu $0.27~\mu M$ or As $0.13~\mu M$ + Pb $0.27~\mu M)$ could be compared to the As intermediate
94	concentration (0.40 μM), and the tertiary combination (As 0.13 μM + Pb 0.27 μM + Cu 0.27
95	$\mu M)$ to the As high concentration (0.67 $\mu M)$ (Table 1). These concentrations of Pb and Cu have
96	also been reported in honey samples (Table S1). The mass consumed for As and the
97	concentrations for Cu and Pb fell within sublethal ranges for the honey bee: the LD50 of
98	elemental As for NaAsO ₂ ranged from 0.330 to 0.540 μ g/bee (Fujii, 1980), the LC50 of Cu is
99	72 mg.L ⁻¹ (Di et al., 2016) and of Pb is 345 mg.L ⁻¹ (Di et al., 2016).

Treatment	Molarity (µM)	Concentration (mg.L ⁻¹)		Ingestion of 5µL (ng/bee)			
		As	Cu	Pb	As	Cu	Pb
Control	0	0	0	0	0	0	0
Low [As]	0.13	0.01	0	0	0.05	0	0
[Cu]	0.27	0	0.02	0	0	0.09	0
[Pb]	0.27	0	0	0.06	0	0	0.28
Med [As]	0.40	0.03	0	0	0.15	0	0
[As+Cu]	0.40	0.01	0.02	0	0.05	0.09	0
[As+Pb]	0.40	0.01	0	0.06	0.05	0	0.28
High [As]	0.67	0.05	0	0	0.25	0	0
[As+Cu+Pb]	0.67	0.01	0.02	0.06	0.05	0.09	0.28

Table 1: Concentrations used. Combined treatments are shown in grey.

Bee exposure to metals

We collected honey bees (*Apis mellifera*) returning from foraging trips at the entrance of five
different hives in mornings during August 2020. We anesthetised the bees on ice and harnessed
them in plastic tubes, secured with tape and a droplet of wax at the back of the head (Matsumoto

107 et al., 2012). We tested all the bees for intact proboscis extension (PER) by stimulating their

antennae with 50% sucrose. We then fed the responding honey bees 5 μ L of 50% sucrose solution (see Table 1), making sure they consumed the whole droplet, and left them to rest for 3 h in the incubator (temperature: 25±2°C, humidity: 60%). Honey bees that did not respond to

- 111 the sucrose solution were discarded.
- 112

113 Absolute learning

Prior to conditioning, we tested all honey bees for PER by stimulating their antennae with 50% 114 115 sucrose solution, and kept only those that displayed the reflex. We then performed olfactory absolute conditioning according to a standard protocol using an automatic stimulus delivery 116 system (Aguiar et al., 2018). Honey bees had to learn to respond to an olfactory conditioned 117 stimulus (CS, 1-nonanol, Sigma-Aldrich Ltd, Lyon, France) reinforced with unconditioned 118 119 stimulus (US, 50% sucrose solution), over five conditioning trials with a ten-minute inter-trial interval. Each trial (37 s in total) began when a bee was placed in front of the stimulus delivery 120 121 system, which released a continuous flow of clean air (3,300 mL.min⁻¹) to the antennae. After 122 15 s, the odour was introduced into the airflow for 4 s, the last second of which overlapped with 123 sucrose presentation to the antennae using a toothpick. This was immediately followed by 124 feeding for 4 s by presenting the toothpick to the proboscis. The bee remained for another 15 s 125 under the clean airflow. We recorded the presence or absence (1/0) of a conditioned PER in 126 response to the odorant presentation during each conditioning trial. Honey bees spontaneously 127 responding in the first conditioning trial were discarded from the analysis. The sum of 128 conditioned responses over all trials provided an individual acquisition score (between 0 and 129 4), and honey bees responding at the last trial were categorized as learners.

130

131 Long-term memory

Only honey bees that had learnt the task were kept for the analysis of memory performance. 132 After conditioning, these honey bees were fed 15 µL of 50% sucrose solution, left overnight in 133 the incubator, and fed another 5 µL of sucrose solution the following morning. Three hours 134 135 after (24 h post-conditioning), we performed the retention test, consisting of three trials similar 136 to conditioning except that no sucrose reward was presented. In addition to the odour used during the conditioning (CS), we presented two novel odours, in randomized order, to assess 137 138 the specificity of the memory: nonanal was expected to be perceived by honey bees similarly 139 to 1-nonanol, while 1-hexanol was expected to be perceived differently (Guerrieri et al., 2005). We recorded the presence or absence (1/0) of a conditioned PER to each odorant at each 140 memory retention trial. We classified honey bees according to their response patterns: response 141

to the CS only, response to the CS and the similar odour (low generalization level), response toall odours (high generalization level), no or inconsistent response.

144

145 *Statistics*

146 We analysed the data using R Studio v.1.2.5033 (RStudio Team, 2015). Raw data are available in Dataset S1. We performed binomial generalised linear mixed-effects models (GLMM) 147 148 (package lme4; Bates et al., 2015), with hive and conditioning date as random factors and treatment as fixed effect. Using the GLMMs, we evaluated whether molarity or treatment 149 150 impacted the initial response to antennal stimulation, the spontaneous response in the first 151 conditioning trial, the response in the last trial, the response to each odorant during the memory 152 test, the proportion of honey bees per response pattern in the retention test, and the survival at 153 24 h. Acquisition scores were standardised and compared with GLMMs using Template Model 154 Builder (Brooks et al., 2017). For all response variables, we compared (1) the treated groups to 155 the control, (2) groups exposed to concentrations of the same molarity (e.g. Med [As], [As+Cu] and [As+Pb]), (3) the separate and joint effects of the treatments (e.g. Low [As], [Cu] and 156 157 [As+Cu]) in order to identify interactive effects (antagonistic, additive, synergistic).

158

159 Results and discussion

160

161 *Exposure to metals did not impact appetitive motivation*

The proportion of honey bees that responded to the initial antennal stimulation with sucrose 162 was similar among treatments (GLMM: p>0.05). All honey bees responded in the other groups. 163 Therefore, treatment did not affect appetitive motivation or sucrose perception. Consistently 164 165 with our observations, the ingestion of similar concentrations of Pb and Cu had no effect on the 166 responsiveness to increasing concentrations of sucrose (Burden et al., 2019). By contrast, Di et 167 al. (2020) found that honey bees exposed to increasing concentrations of a mixture of Cu and 168 Cd exhibited a decreased ability to distinguish sucrose concentrations, but this may be 169 explained by the much higher (at least 600 times) concentrations used in that study. Thus, in 170 our conditions any impact on appetitive learning is unlikely due to a decreased motivation for 171 sucrose.

172

173 Individual and joint exposures to metals reduced learning performance

Two out of the 381 honey bees submitted to the absolute learning task spontaneously responded

to the first odour presentation and were therefore discarded. In all groups, the number of honey

176 bees showing the conditioned response increased over trials, thus showing learning (Fig. 1A). However, fewer honey bees exposed to metals learned the task when compared to controls 177 178 (GLMM: p<0.05, except for Low [As], p=0.082). Accordingly, the acquisition scores of honey 179 bees from all treatments were lower than controls (Fig. 1B). Honey bees exposed to Med [As] 180 (GLMM: -0.610±0.246, p=0.013), High [As] (GLMM: -0.639±0.241, p=0.008) and [As+Cu+Pb] (GLMM: -0.592±0.244, p=0.015) had acquisition scores significantly lower than 181 182 controls. Honey bees exposed to solutions of [As+Pb] had similar acquisition scores to Med [As] (GLMM: 0.299±0.234, p=0.201), but honey bees exposed to [As+Cu] performed better 183 184 (GLMM: 0.596±0.241, p=0.013). Honey bees exposed to High [As] and [As+Cu+Pb] exhibited 185 similar acquisition scores (GLMM: p=0.810). We found no difference in the acquisition scores 186 and the proportions of learners between honey bees treated with a single metal and mixed 187 treatments (GLMM: p>0.05), that would have indicated non-additive effects (i.e. antagonistic 188 or synergistic). Thus, exposure to metals significantly reduced learning performance, and 189 combined exposure appeared to exert simple additive deleterious effects.

190

Figure 1: Learning. A) Learning curves show changes in the percentages of honey bees 191 192 displaying the conditioned proboscis extension response (PER) over five training trials. Asterisks indicate significant differences in responses at the last trial compared to control honey 193 194 bees. B) Violin plots of acquisition score values (sum of conditioned responses for each honey 195 bee). Symbols (*circle*: single exposure; *triangle*: binary mixture; *diamond*: tertiary mixture) 196 indicate the mean score for each treatment. Significant differences between groups exposed to 197 the same molarity solutions (#) or with respect to control honey bees (*) are indicated 198 (#/*p<0.05, **p<0.01; GLMM).

199

200 Individual and joint exposures to metals reduced long-term memory specificity

201 To examine possible effects of metal exposure on memory retention, we tested the capacity for 202 long-term memory retention. Only honey bees that had learned the CS-US association at the 203 end of conditioning were tested, to evaluate retention levels independently of possible biases 204 due to slight variations in learning performance among treatments. 167 out of the 379 honey 205 bees submitted to the absolute learning task did not learn and were therefore discarded. We 206 found no effect of treatment on survival at 24 h (GLMM: p>0.05). However, long-term memory 207 was significantly affected (Fig. 2). Overall, treated honey bees responded less to the learned 208 odorant (CS) than controls, as indicated by a significant effect of exposure to metals on retention 209 levels (GLMM: p<0.05) (Fig. 2A). Yet, this decrease was not significant for honey bees exposed to Med [As] (GLMM: -0.260±0.628, p=0.679) and High [As] (GLMM: -1.023±0.570,
p=0.073). Finally, there was no clear dose effect on responses to the CS among treated groups
(GLMM: -0.576±0.579, p=0.320).

213 Yet, individual response patterns (Fig. 2B) revealed a loss of memory specificity. While 214 honey bees from all treatments responded equivalently to the similar odour (GLMM: p>0.05), 215 those exposed to higher doses responded more frequently to all odorants, indicating a higher 216 degree of response generalization (GLMM: 1.954±0.775, p=0.012). This was accompanied by a significantly lower proportion of specific (CS-only) responses for honey bees exposed to [Pb] 217 (GLMM: -1.795±0.690, p=0.009), low [As] (GLMM: -1.313±0.589, p=0.026) and 218 [As+Cu+Pb] (GLMM: -1.200±0.588, p=0.041). Exposure also significantly increased the 219 220 frequency of inconsistent responses as compared to controls (GLMM: p<0.05). This was the 221 case for each individual treatment except for Med [As], p=0.293). Thus, exposure to metals had 222 a negative impact on memory performance at 24h. The analysis of individual response patterns 223 also revealed additive effects as they neither differ among groups exposed to solutions with the 224 same molarity, nor between single and mixed metal treatments (GLMM: p>0.05). Thus, most 225 treatments reduced memory performance at 24h.

226

Figure 2: Long-term memory. A) Percentages of responses to the CS odour in the 24 hmemory retention test (mean \pm s.e.m). B) Distribution of honey bees according to their individual response pattern during the long-term memory test: response to CS only; response to CS and similar; response to all odours; no or inconsistent response. Significant differences with controls are indicated (*p<0.05, **p<0.01; GLMM).

232

233 The additive effects of metal mixtures may be explained by common pathways of toxicity

234 Although many mechanisms of metal toxicity have not yet been elucidated, some points of 235 consensus are emerging from the literature. Firstly, interactions between metals can occur 236 outside the organism (including interactions with the environment (Grobelak and Kowalska, 237 2020; Noyes and Lema, 2015)), and during uptake into the organism, leading to potentially 238 toxic processes of speciation, absorption, binding, transport and distribution (Wu et al., 2016). 239 Once metals enter an organism, they can induce, interact or inhibit a range of biological 240 responses and metabolic pathways. By mimicking other essential metals (Bridges and Zalups, 241 2005) or damaging the permeability of biological membranes (Rothshein, 1959), metals allow 242 the uptake or loss of other compounds in/out intracellular compartments (Viarengo, 1994). Metals are known to disrupt signalling and calcium homeostasis (particularly important in 243

244 neurons) by interfering with the calcium channels (Bridges and Zalups, 2005; Chavez-Crooker 245 et al., 2001; Tamano and Takeda, 2011). This might lead to dysfunction and cytotoxicity due 246 to the disruption of cell signalling and calcium homeostasis. Genotoxicity (Do anlar et al., 247 2014) may be achieved through covalent binding to DNA (Brocato and Costa, 2013; Senut et al., 2014). Eventually, oxidative stress and lipid peroxidation of the cell membrane may lead to 248 neuronal death. Additionally, metals in mixture could interact at target sites, but the potential 249 250 for modulation of that toxic impact is largely unknown (Svendsen et al., 2011). Metal mixtures 251 could change the bioavailibity, toxicokinetic and toxicodynamic of each metal, that could 252 directly impact the toxicity towards the organism (Løkke et al., 2013). Based on these shared 253 mechanisms of toxicity that include oxidative stress (Nikoli et al., 2016; Zaman et al., 1995), 254 apoptosis (Raes et al., 2000) and interference with neurotransmitters (Nisbet et al., 2018), the 255 toxic effects of metal pollutants in mixtures may be expected to be additive (von Stackelberg 256 et al., 2013).

257 These effect may affect many aspects of neural activity and brain function in honey bees, as in other species (Karri et al., 2016). Here, we focused on learning and memory of 258 259 olfactory cues because they play crucial roles in the behavioural ecology of honey bees and 260 other pollinators, for the identification of food resources. Our results in controlled laboratory 261 conditions suggest that exposure to sublethal combinations of toxic elements in the field might 262 alter individual foraging efficiency, and in turn jeopardize survival of pollinator populations. 263 Our findings call for further evaluation of the joint actions of metals (Meyer et al., 2015) to better assess the risk they pose (Nys et al., 2018; Otitoloju, 2003) and better inform regulatory 264 265 framework (European Commission, 2012). Current risk assessment guidance mainly assesses the effect of individual exposure, that fails to capture potential interactive effects. Hence, the 266 267 evaluation of metal mixture impacts and their modes of action needs to be developed (Sasso et 268 al., 2010). More generally, the study of the interactions between toxic metals and environmental 269 factors (Nagash et al., 2020) and their impacts on the toxicokinetic of other chemicals (EFSA Scientific Committee et al., 2019) (e.g. pesticides (Sgolastra et al., 2018; Singh et al., 2017), 270 271 volatile organic compounds (Sasso et al., 2010) etc.) should be implemented in this integrated 272 research framework.

273

274 Conclusion

In summary, we demonstrated that arsenic, lead, copper or combinations of these metals, at
levels found in the environment, slow down appetitive learning and reduce long-term memory
specificity in honey bees. These metals show simple additive effects as we found no differences

278	between different solutions of the same molarity suggesting possible non-linear effects					
279	(synergism or antagonism). Thus, regarding effects on learning and memory, concentration					
280	seem to be more important than identity of any specific metal. Since learning and memory of					
281	olfactory cues play crucial roles in the behavioural ecology of honey bees, acute exposure to					
282	metal pollutants mixtures could impair fundamental hive function and population growth.					
283						
284	Acknowledgments					
285	We thank Olivier Fernandez for assistance with beekeeping.					
286						
287	Competing interests					
288	The authors declare no competing or financial interests.					
289						
290	Funding					
291	This work was supported by the CNRS. CM was funded by a PhD fellowship from French					
292	Ministry of Higher Education, Research and Innovation. ABB was funded by a Future					
293	Fellowship from the Australian Research Council (FT140100452) and the Eldon and Anna					

.1 1

- Foote Trust. ML was funded by grants of the Agence Nationale de la Recherche (ANR-16-294 295 CE02-0002-01, ANR-19-CE37-0024, ANR-20-ERC8-0004-01) and the European Regional
- 296 Development Fund (project ECONECT).

- 297
- **Data availability** 298
- Raw data will be available on Dryad repository upon publication. 299
- 300
- 301 References
- Agrawal, S., Bhatnagar, P. and Flora, S. J. S. (2015). Changes in tissue oxidative stress, 302 brain biogenic amines and acetylcholinesterase following co-exposure to lead, arsenic and 303 mercury in rats. Food Chem. Toxicol. 86, 208-216. 304
- Aguiar, J. M. R. B. V., Roselino, A. C., Sazima, M. and Giurfa, M. (2018). Can honey bees 305 discriminate between floral-fragrance isomers? J. Exp. Biol. 221, jeb180844. 306
- Aktar, S., Jahan, M., Alam, S., Mohanto, N. C., Arefin, A., Rahman, A., Haque, A., 307
- 308 Himeno, S., Hossain, K. and Saud, Z. A. (2017). Individual and combined effects of arsenic
- and lead on behavioral and biochemical changes in mice. Biol. Trace Elem. Res. 177, 288-296. 309 Anderson, D. W., Mettil, W. and Schneider, J. S. (2016). Effects of low level lead exposure

310 on associative learning and memory in the rat: Influences of sex and developmental timing of 311 exposure. Toxicol. Lett. 246, 57-64. 312

ATSDR (2019). The ATSDR 2019 Substance Priority List. 313

- Ayers, R. S. and Westcot, D. W. (1994). *Water quality for agriculture*. Food and Agriculture
 Organization of the United Nations, Rome.
- 316 Badiou-Bénéteau, A., Benneveau, A., Géret, F., Delatte, H., Becker, N., Brunet, J. L.,
- **Reynaud, B. and Belzunces, L. P.** (2013). Honeybee biomarkers as promising tools to monitor environmental quality. *Environ. Int.* **60**, 31–41.
- Balestra, V., Celli, G. and Porrini, C. (1992). Bees, honey, larvae and pollen in biomonitoring
 of atmospheric pollution. *Aerobiologia* 8, 122–126.
- Bates, D., Mächler, M., Bolker, B. and Walker, S. (2015). Fitting linear mixed-effects models
 using lme4. J. Stat. Softw. 67, 1–48.
- 323 Bopp, S. K., Barouki, R., Brack, W., Dalla Costa, S., Dorne, J.-L. C. M., Drakvik, P. E.,
- **Faust, M., Karjalainen, T. K., Kephalopoulos, S., van Klaveren, J., et al.** (2018). Current EU research activities on combined exposure to multiple chemicals. *Environ. Int.* **120**, 544–
- 326 562.
- Bradl, H. B. (2005). Sources and origins of heavy metals. In *Interface Science and Technology*(ed. Bradl, H. B.), pp. 1–27. Elsevier.
- 329 Bridges, C. C. and Zalups, R. K. (2005). Molecular and ionic mimicry and the transport of 330 toxic metals. *Toxicol. Appl. Pharmacol.* **204**, 274–308.
- 331 Brocato, J. and Costa, M. (2013). Basic mechanics of DNA methylation and the unique
- landscape of the DNA methylome in metal-induced carcinogenesis. Crit. Rev. Toxicol. 43, 493–
- **333** 514.
- 334 Brooks, M., E., Kristensen, K., van Benthem, K. J., Magnusson, A., Berg, C., W., Nielsen,
- A., Skaug, H., J., Maechler, M. and Bolker, B., M. (2017). glmmTMB balances speed and
 flexibility among packages for zero-inflated generalized linear mixed modeling. *R J.* 9, 378–
 400.
- Burden, C. M., Elmore, C., Hladun, K. R., Trumble, J. T. and Smith, B. H. (2016). Acute
 exposure to selenium disrupts associative conditioning and long-term memory recall in honey
 bees (*Apis mellifera*). *Ecotoxicol. Environ. Saf.* 127, 71–79.
- 341 Burden, C. M., Morgan, M. O., Hladun, K. R., Amdam, G. V., Trumble, J. J. and Smith,
- B. H. (2019). Acute sublethal exposure to toxic heavy metals alters honey bee (*Apis mellifera*)
 feeding behavior. *Sci. Rep.* 9, 4253.
- Chavez-Crooker, P., Garrido, N. and Ahearn, G. A. (2001). Copper transport by lobster
 hepatopancreatic epithelial cells separated by centrifugal elutriation: measurements with the
 fluorescent dye Phen Green. J. Exp. Biol. 204, 1433.
- 347 Chen, P., Miah, M. R. and Aschner, M. (2016). Metals and neurodegeneration.
 348 *F1000Research* 5, 366.
- 349 Codex Alimentarius (2015). Codex general standard for contaminants and toxins in food and
 350 *feed CODEX STAN 193-1995.* Joint FAO/WHO.
- **Di, N., Hladun, K. R., Zhang, K., Liu, T.-X. and Trumble, J. T.** (2016). Laboratory bioassays on the impact of cadmium, copper and lead on the development and survival of honeybee (*Apis mellifera* L.) larvae and foragers. *Chemosphere* **152**, 530–538.
- Di, N., Zhang, K., Hladun, K. R., Rust, M., Chen, Y.-F., Zhu, Z.-Y., Liu, T.-X. and
 Trumble, J. T. (2020). Joint effects of cadmium and copper on *Apis mellifera* foragers and
- arvae. Comp. Biochem. Physiol. Part C Toxicol. Pharmacol. 237, 108839.
- **Do anlar, Z. B., Do anlar, O. and Tabakçıo lu, K.** (2014). Genotoxic effects of heavy metal mixture in *Drosophila melanogaster*: Expressions of heat shock proteins, RAPD profiles and
- 359 mitochondrial DNA sequence. *Water. Air. Soil Pollut.* 225, 2104.
- 360 EFSA Scientific Committee, More, S. J., Bampidis, V., Benford, D., Bennekou, S. H.,
- 361 Bragard, C., Halldorsson, T. I., Hernández-Jerez, A. F., Koutsoumanis, K., Naegeli, H., et
- **al.** (2019). Guidance on harmonised methodologies for human health, animal health and
- ecological risk assessment of combined exposure to multiple chemicals. *EFSA J.* 17, e05634.

- 364 European Commission (2012). Scientific Committee on Health and Environmental Risks.
 365 Toxicity and assessment of chemical mixtures.
- Folt, C. L., Chen, C. Y., Moore, M. V. and Burnaford, J. (1999). Synergism and antagonism
 among multiple stressors. *Limnol. Oceanogr.* 44, 864–877.
- Fujii, L. K. (1980). Oral dose toxicity vs. tissue residue levels of arsenic in the honey bee (*Apis mellifera* L.).
- 370 Giglio, A., Ammendola, A., Battistella, S., Naccarato, A., Pallavicini, A., Simeon, E.,
- 371 Tagarelli, A. and Giulianini, P. G. (2017). Apis mellifera ligustica, Spinola 1806 as
- 372 bioindicator for detecting environmental contamination: a preliminary study of heavy metal
- 373 pollution in Trieste, Italy. *Environ. Sci. Pollut. Res.* 24, 659–665.
- 374 Goretti, E., Pallottini, M., Rossi, R., La Porta, G., Gardi, T., Cenci Goga, B. T., Elia, A.
- 375 C., Galletti, M., Moroni, B., Petroselli, C., et al. (2020). Heavy metal bioaccumulation in
 376 honey bee matrix, an indicator to assess the contamination level in terrestrial environments.
- 377 Environ. Pollut. 256, 113388.
- 378 Grobelak, A. and Kowalska, A. (2020). Heavy metal mobility in soil under futuristic climatic
- 379 conditions. In *Climate Change and Soil Interactions* (ed. Pietrzykowski, M.) and Prasad, M. N.
- 380 V.), pp. 437–451. Elsevier.
- 381 Guerrieri, F., Schubert, M., Sandoz, J.-C. and Giurfa, M. (2005). Perceptual and neural 382 olfactory similarity in honeybees. *PLoS Biol.* **3**, e60.
- 383 Kabir, E., Ray, S., Kim, K.-H., Yoon, H.-O., Jeon, E.-C., Kim, Y. S., Cho, Y.-S., Yun, S.-
- **T. and Brown, R. J. C.** (2012). Current status of trace metal pollution in soils affected by industrial activities. *Sci. World J.* **2012**, 1–18.
- 386 Karri, V., Schuhmacher, M. and Kumar, V. (2016). Heavy metals (Pb, Cd, As and MeHg)
- as risk factors for cognitive dysfunction: a general review of metal mixture mechanism in brain. *Environ. Toxicol. Pharmacol.* 48, 203–213.
- 389 Khaska, M., Le Gal La Salle, C., Sassine, L., Cary, L., Bruguier, O. and Verdoux, P.
- 390 (2018). Arsenic and metallic trace elements cycling in the surface water-groundwater-soil
 391 continuum down-gradient from a reclaimed mine area: isotopic imprints. *J. Hydrol.* 558, 341–
 392 355.
- 393 Krämer, U. (2010). Metal hyperaccumulation in plants. *Annu. Rev. Plant Biol.* 61, 517–534.
- Lamtai, M., Zghari, O., Ouakki, S., Marmouzi, I., Mesfioui, A., El Hessni, A. and
 Ouichou, A. (2020). Chronic copper exposure leads to hippocampus oxidative stress and
 impaired learning and memory in male and female rats. *Toxicol. Res.* 36, 359–366.
- Lee, J.-S., Chon, H.-T. and Kim, K.-W. (2005). Human risk assessment of As, Cd, Cu and Zn in the abandoned metal mine site. *Environ. Geochem. Health* **27**, 185–191.
- Lin, X., Gu, Y., Zhou, Q., Mao, G., Zou, B. and Zhao, J. (2016). Combined toxicity of heavy
 metal mixtures in liver cells. *J. Appl. Toxicol.* 36, 1163–1172.
- 401 Løkke, H., Ragas, A. M. J. and Holmstrup, M. (2013). Tools and perspectives for assessing
 402 chemical mixtures and multiple stressors. *Toxicology* 313, 73–82.
- 403 Mahaffey, K. R., Capar, S. G., Gladen, B. C. and Fowler, B. A. (1981). Concurrent exposure
- to lead, cadmium, and arsenic: effects on toxicity and tissue metal concentrations in the rat. J. *Lab. Clin. Med.* 98, 463–481.
- 406 Mance, G. (1987). *Pollution threat of heavy metals in aquatic environments*. (ed. Mellanby,
 407 K.) The Netherlands: Springer Science & Business Media.
- 408 Mason, L. H., Harp, J. P. and Han, D. Y. (2014). Pb neurotoxicity: neuropsychological 409 effects of lead toxicity. *BioMed Res. Int.* 2014, 1–8.
- 410 Matsumoto, Y., Menzel, R., Sandoz, J.-C. and Giurfa, M. (2012). Revisiting olfactory
- 411 classical conditioning of the proboscis extension response in honey bees: A step toward
- 412 standardized procedures. J. Neurosci. Methods 211, 159–167.
- 413 Meyer, J. S., Farley, K. J. and Garman, E. R. (2015). Metal mixtures modeling evaluation

- 414 project: background. *Environ. Toxicol. Chem.* **34**, 726–740.
- 415 Monchanin, C., Blanc-Brude, A., Drujont, E., Negahi, M. M., Pasquaretta, C., Silvestre,
- 416 J., Baqué, D., Elger, A., Barron, A. B., Devaud, J.-M., et al. (2021a). Chronic exposure to 417 trace lead impairs honey bee learning. *Ecotoxicol. Environ. Saf.* 212, 112008.
- 418 Monchanin, C., Devaud, J.-M., Barron, A. and Lihoreau, M. (2021b). Current permissible
- 419 levels of heavy metal pollutants harm terrestrial invertebrates. *Sci. Total Environ.* **779**, 146398.
- 420 Naqash, N., Prakash, S., Kapoor, D. and Singh, R. (2020). Interaction of freshwater
- 421 microplastics with biota and heavy metals: a review. *Environ. Chem. Lett.* **18**, 1813–1824.
- 422 Nikoli , T. V., Koji , D., Or i , S., Batini , D., Vuka inovi , E., Blagojevi , D. P. and
- 423 Pura , J. (2016). The impact of sublethal concentrations of Cu, Pb and Cd on honey bee redox
- status, superoxide dismutase and catalase in laboratory conditions. *Chemosphere* **164**, 98-105.
- Nisbet, C., Guler, A., Ormanci, N. and Cenesiz, S. (2018). Preventive action of zinc against
 heavy metals toxicity in honeybee. *Afr. J. Biochem. Res.* 12, 1–6.
- 427 Noyes, P. D. and Lema, S. C. (2015). Forecasting the impacts of chemical pollution and
 428 climate change interactions on the health of wildlife. *Curr. Zool.* 61, 669–689.
- 429 Nriagu, J. O. and Pacyna, J. M. (1988). Quantitative assessment of worldwide contamination
 430 of air, water and soils by trace metals. *Nature* 333, 134–139.
- 431 Nys, C., Van Regenmortel, T., Janssen, C. R., Oorts, K., Smolders, E. and De
- 432 Schamphelaere, K. A. C. (2018). A framework for ecological risk assessment of metal
- 433 mixtures in aquatic systems: Ecological risk assessment framework for metal mixtures.
- 434 *Environ. Toxicol. Chem.* **37**, 623–642.
- 435 Otitoloju, A. A. (2003). Relevance of joint action toxicity evaluations in setting realistic
 436 environmental safe limits of heavy metals. *J. Environ. Manage.* 67, 121–128.
- 437 Pal, A., Badyal, R. K., Vasishta, R. K., Attri, S. V., Thapa, B. R. and Prasad, R. (2013).
- 438 Biochemical, histological, and memory impairment effects of chronic copper toxicity: a model
- 439 for non-wilsonian brain copper toxicosis in wistar rat. *Biol. Trace Elem. Res.* **153**, 257–268.
- Perugini, M., Manera, M., Grotta, L., Abete, M. C., Tarasco, R. and Amorena, M. (2011).
 Heavy metal (Hg, Cr, Cd, and Pb) contamination in urban areas and wildlife reserves:
- 442 honeybees as bioindicators. *Biol. Trace Elem. Res.* **140**, 170–176.
- 443 Pisani, A., Protano, G. and Riccobono, F. (2008). Minor and trace elements in different honey
 444 types produced in Siena County (Italy). *Food Chem.* 107, 1553–1560.
- Raes, H., Braeckman, B. P., Criel, G. R. J., Rzeznik, U. and Vanfleteren, J. R. (2000).
 Copper induces apoptosis in Aedes C6/36 cells. J. Exp. Zool. 286, 1–12.
- **Rothshein, A.** (1959). Cell membrane as site of action of heavy metals. *Fed. Proc.* 18, 1026–
 1038.
- 449 RStudio Team (2015). RStudio: integrated development for R. RStudio, Inc., Boston, MA
 450 URL http://www.rstudio.com/.
- 451 Sasso, A. F., Isukapalli, S. S. and Georgopoulos, P. G. (2010). A generalized physiologically-
- 452 based toxicokinetic modeling system for chemical mixtures containing metals. *Theor. Biol.*453 *Med. Model.* 7, 17.
- Satta, A., Verdinelli, M., Ruiu, L., Buffa, F., Salis, S., Sassu, A. and Floris, I. (2012).
 Combination of beehive matrices analysis and ant biodiversity to study heavy metal pollution
- 456 impact in a post-mining area (Sardinia, Italy). *Environ. Sci. Pollut. Res.* 19, 3977–3988.
- 457 Schmolke, G., Elsenhans, B., Ehtechami, C. and Forth, W. (1992). Arsenic-copper
 458 interaction in the kidney of the rat. *Hum. Exp. Toxicol.* 11, 315–321.
- 459 Senut, M.-C., Sen, A., Cingolani, P., Shaik, A., Land, S. J. and Ruden, D. M. (2014). Lead
- 460 exposure disrupts global DNA methylation in human embryonic stem cells and alters their
- 461 neuronal differentiation. *Toxicol. Sci.* **139**, 142–161.
- 462 Sgolastra, F., Blasioli, S., Renzi, T., Tosi, S., Medrzycki, P., Molowny-Horas, R., Porrini,
- 463 C. and Braschi, I. (2018). Lethal effects of Cr(III) alone and in combination with

- 464 propiconazole and clothianidin in honey bees. *Chemosphere* 191, 365–372.
- 465 Singh, N., Gupta, V. K., Kumar, A. and Sharma, B. (2017). Synergistic effects of heavy metals and pesticides in living systems. Front. Chem. 5, 70. 466
- 467 Su, C., Jiang, L. and Zhang, W. (2014). A review on heavy metal contamination in the soil worldwide: situation, impact and remediation techniques. Environ. Skept. Crit. 3, 24-38. 468
- Svendsen, C., Jager, T., Haddad, S., Yang, R., Dorne, J., Broerse, M. and Kramarz, P. 469
- (2011). Toxicokinetics and toxycodynamics. In Mixture toxicity: linking approaches from 470
- ecological and human toxicology (ed. Van Gestel, C. A.), Jonker, M.), Kammenga, J. E.), 471
- Laskowski, R.), and Svendsen, C.), pp. 47–93. New York, NY, USA: CRC press. 472
- Tamano, H. and Takeda, A. (2011). Dynamic action of neurometals at the synapse. 473 474 Metallomics 3, 656.
- Tchounwou, P. B., Yedjou, C. G., Patlolla, A. K. and Sutton, D. J. (2012). Heavy metal 475 toxicity and the environment. In Molecular, Clinical and Environmental Toxicology (ed. Luch, 476 477 A.), pp. 133–164. Basel: Springer Basel.
- Terrab, A., Recamales, A., Gonzalezmiret, M. and Heredia, F. (2005). Contribution to the 478 study of avocado honeys by their mineral contents using inductively coupled plasma optical 479 emission spectrometry. Food Chem. 92, 305-309. 480
- Thimmegowda, G. G., Mullen, S., Sottilare, K., Sharma, A., Mohanta, S. S., Brockmann, 481
- 482 A., Dhandapany, P. S. and Olsson, S. B. (2020). A field-based quantitative analysis of
- sublethal effects of air pollution on pollinators. Proc. Natl. Acad. Sci. USA 117, 20653-20661. 483
- 484 Tolins, M., Ruchirawat, M. and Landrigan, P. (2014). The developmental neurotoxicity of arsenic: cognitive and behavioral consequences of early life exposure. Ann. Glob. Health 80, 485
- 486 303-314.
- Tyler, C. R. S., Smoake, J. J. W., Solomon, E. R., Villicana, E., Caldwell, K. K. and Allan, 487
- 488 A. M. (2018). Sex-dependent effects of the histone deacetylase inhibitor, sodium valproate, on reversal learning after developmental arsenic exposure. Front. Genet. 9, 1–14. 489
- 490 Verriopoulos, G. and Dimas, S. (1988). Combined toxicity of copper, cadmium, zinc, lead, nickel, and chrome to the copepod Tisbe holothuriae'. Bull. Environ. Contam. Toxicol. 41, 378-491
- 492 384.
- Viarengo, A. (1994). Heavy metal cytotoxicity in marine organisms: effects on Ca2+ 493 homeostasis and possible alteration of signal transduction pathways. In Advances in 494 495 Comparative and Environmental Physiology: Volume 20 (ed. Arpigny, J. L.), Coyette, J.), Davail, S.), Feller, G.), Fonzé, E.), Foulkes, E. C.), Frère, J.-M.), Fujii, R.), Génicot, S.), 496
- 497 Gerday, Ch.), et al.), pp. 85–110. Berlin, Heidelberg: Springer Berlin Heidelberg.
- 498 von Stackelberg, K., Guzy, E., Chu, T. and Henn, B. C. (2013). Mixtures, metals, genes and pathways: a systematic review.pp. 1–67. Harvard Center for Risk Analysis. 499
- 500 Wu, C., Gu, X., Ge, Y., Zhang, J. and Wang, J. (2006). Effects of high fluoride and arsenic 501 on brain biochemical indexes and learning-memory in rats. Fluoride 39, 274-279.
- Wu, X., Cobbina, S. J., Mao, G., Xu, H., Zhang, Z. and Yang, L. (2016). A review of toxicity 502 and mechanisms of individual and mixtures of heavy metals in the environment. Environ. Sci. 503
- 504 Pollut. Res. 23, 8244-8259.
- Xun, E., Zhang, Y., Zhao, J. and Guo, J. (2018). Heavy metals in nectar modify behaviors of 505 pollinators and nectar robbers: consequences for plant fitness. Environ. Pollut. 242, 1166-1175. 506
- 507 Yao, J., Zhu, Y. C., Adamczyk, J. and Luttrell, R. (2018). Influences of acephate and
- 508 mixtures with other commonly used pesticides on honey bee (Apis mellifera) survival and
- 509 detoxification enzyme activities. Comp. Biochem. Physiol. Part C Toxicol. Pharmacol. 209, 9-510 17.
- Zaman, K., MacGill, R. S., Johnson, J. E., Ahmad, S. and Pardini, R. S. (1995). An insect 511
- 512 model for assessing oxidative stress related to arsenic toxicity: arsenic toxicity and oxidative stress. Arch. Insect Biochem. Physiol. 29, 199-209. 513

- **Zamberlan, D. C.** (2020). Copper decreases associative learning and memory in *Drosophila melanogaster. Sci. Total Environ.* 9, 1–14.
- 516 Zhu, Y. C., Yao, J., Adamczyk, J. and Luttrell, R. (2017). Synergistic toxicity and
- 517 physiological impact of imidacloprid alone and binary mixtures with seven representative
- 518 pesticides on honey bee (*Apis mellifera*). *PLOS ONE* **12**, e0176837.

519