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# New clustering method to infer prototypes covering the 3D structures of nucleic acid fragments

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In structural biology, fragment-based 3D modeling methods make use of fragment libraries. The library associated with one fragment represents the whole set of possible 3D structures (conformations) that it can adopt (with a chosen precision). Given the computational constraints, deriving libraries of minimal cardinality appears as a strong requirement. This amounts to deriving  $\varepsilon$ -nets whose cardinalities are as close as possible to the corresponding covering numbers. A heuristic to derive such  $\varepsilon$ -nets is to cluster the observed conformations under appropriate constraints, and keep only the representatives/prototypes. In the framework of interest, the main difficulty encountered is to implement the clustering with the RMSD as dissimilarity measure. Indeed, the computation of this measure follows a superimposition, the nature of which has both biological and mathematical consequences.

We introduce such a method as a variant of the Hierarchical Agglomerative Clustering (HAC) algorithm. Compared to HAC, it makes it possible to reduce the number of prototypes, while maintaining an acceptable computation time.

# **Algorithm:**

# **Example:**

 $\mathcal{X} =$ 

Input :  $\mathcal{X} := \{x_i : 1 \leq i \leq n\}$ # set of conformers # threshold value t > 0**Output:**  $\mathcal{P} := \{p_i : 1 \leq i \leq s\}$ # set of prototypes Initialisation: # number of prototypes s := n# set of prototypes  $\mathcal{P} := \mathcal{X}$  $R := (r_{i,j}) \in \mathbb{R}^{s \times s}$ # matrix of RMSD of prototypes after one-against-one superimposition  $R^{mask} := (\mathbf{r}_{i,i}^{mask}) \in \{0,1\}^{s \times s}$  $\mathcal{E} := \{ \{x_i\}_{1 < j < s} : 1 \leqslant i \leqslant n \}$ # set of clusters fusion := True;while fusion do fusion := False; $R^{mask} := False;$ # initialized at *False* tag := False;while (not tag and not fusion) do  $i^*, j^* := \operatorname{argmin}(r_{i,j} : r_{i,j}^{mask} == False);$  $1 \leq i < j \leq s$  $mean := (p_{i^*} + p_{j^*})/2;$  $\mathcal{S}_{union} := \mathcal{E}_{i^*} \cup \mathcal{E}_{i^*};$ # superimposed on mean Compute the *center* of the smallest englobing ball associated with  $\mathcal{S}_{union}$ ;



if  $(\max_{x_i \in S_{union}} (d_{superimposed}(x_i, center)) \leq t)$  then  $\mid \mathcal{P} \setminus \{p_{i^*}, p_{j^*}\} \cup \{center\};$ 

 $\mid \begin{array}{c} \mathcal{P} \setminus \{p_{i^*}, p_{j^*}\} \cup \\ \text{Update } R; \end{array}$ 

Update  $\mathcal{E}$ ;

s := s - 1;

fusion := True;

 $\mathbf{else}$ 

$$\begin{array}{||c|} \hline r_{i_{min},j_{min}}^{mask} := True; \\ if (\forall 1 \leq i < j \leq s : r_{i,j}^{mask} == True) \text{ then} \\ | tag := True; \\ end if \\ end if \\ end while \\ end while \\ return \mathcal{P}; \end{array}$$





Two superimpositions of trinucleotides, RMSD of 1Å (top) and 3Å (bottom). 1Å corresponds to the fragment libraries giving the best docking results.

## **Conclusion and ongoing research:**

# **Analysis:**

The algorithm is based on the HAC [1], the main difference is the linkage method. We defined a linkage method which is the "smallest enclosing balls". The computation of those balls is a quadratic problem and to solve it we are using the Frank-Wolfe algorithm [3].

The algorithm was applied to trinucleotides of RNA. We compared the results obtained, for the sequence AAA, to a classic HAC with the "complete" linkage method. For the "complete" linkage we obtain 2817 clusters, and with our "smallest enclosing balls" we obtain 2421 prototypes (coming from a set of 11,813 conformations). The difference is that the "complete" linkage is not adapted to calculate  $\varepsilon$ -nets, it is merging clusters when all members are at maximum 1Å from other members, and it does not give any prototypes.

In the fragment-based method for the docking of RNA on protein [2], the docking of the conformers is proportional to the number of prototypes.

• Our method performs better than the state-of-the-art one;

• On the application of trinucleotides, the cardinality directly translate into a gain in computation time;

• We are planning to apply our method to other biological problems;

 Statistical analysis is in progress to derive generalization error bounds and excess risk bounds.

### **References:**

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3. M. Frank, and P. Wolfe. An algorithm for quadratic programming. Naval Res. Logist. Quart., 3(1):95-110, 1956.