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1 Spontaneous rotations in epithelia as an interplay between cell polarity and 2 boundaries

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20

21 ABSTRACT

22

23 Directed flows of cells *in vivo* are essential in morphogenesis. They shape living matter
24 in phenomena involving cell mechanics and regulations of the acto-myosin
25 cytoskeleton. However the onset of coherent motion is still poorly understood. Here we
26 show that coherence is associated with spontaneous alignments of cell polarity by
27 designing cellular rings of controlled dimensions. A tug-of-war between polarities
28 dictates the onset of coherence, as assessed by tracking live cellular shapes and
29 motions in various experimental conditions. In addition, we identify an internally driven
30 constraint set by cellular acto-myosin cables at boundaries as essential to ensure
31 coherence, and active force is generated as evaluated by the high RhoA activity. The
32 cables are required to trigger coherence as shown by our numerical simulations based
33 on a novel Vicsek-type model including free active boundaries. We quantitatively
34 reproduce *in silico* coherence onsets and we predict criteria leading to coherence.
35 Altogether, spontaneous coherent motion results from basic competitions between cell
36 orientations and active cables at boundaries.

37

38 INTRODUCTION

39

40 Morphogenesis is driven by the interplay between Rho GTPases and the cytoskeleton
41 ¹⁻⁴. In particular, the acto-myosin network drives morphogenesis at the cellular level by
42 elongation or neighbor exchange ^{5,6}, and the cross-talk between Rho signaling and
43 cellular mechanics has emerged as a generic property of morphogenetic systems.
44 Interestingly, these dynamics are often associated with directed motions of cells.
45 Directional flows in epithelia are reported during development in a variety of systems
46 ranging from *Drosophila* to zebrafish ⁷⁻⁹. For example, during the development of
47 spherical mammary acini or during the egg chamber elongation in *Drosophila*, cells
48 exhibit *coherent* rotation on large scales with speed up to $20\mu\text{m}\cdot\text{h}^{-1}$ ¹⁰⁻¹³. These
49 motions are proposed to be important for the robust morphogenesis of embryos.

50

1 The collective motion of cells is difficult to understand based on a simple analysis of
2 migrations at the single cell level. Individually, cells can move randomly or directionally.
3 However the epithelial layers which connect cells through adherens junctions may or
4 may not coordinate the motion of the group^{14–16}. The understanding of emergence of
5 collective effects requires theoretical approaches in tight comparison with experiments.
6 In addition, although the interplay between RhoA activity and the acto-myosin
7 cytoskeleton is documented at the single cell level^{1,17,18}, it is not yet fully characterized
8 at the multi-cellular scale. In this context of potential collective effects, the
9 understanding of coherent motion requires to study cellular assembly in simple
10 geometries *in vitro* compared quantitatively to numerical simulations.

11
12
13 To this aim, we generated rings of epithelial cells, a geometry proposed by Turing in
14 his 1952 article¹⁹. This initial condition with boundaries is distinct from former work^{20,21}
15 and led to the formation of rings of connected cells, a simple and reproducible periodic
16 configuration with no confinement. This allows the characterization of the spontaneous
17 evolution of the cellular ring. We varied the ring diameter to identify the inherent
18 coherence length of our epithelial MDCK cells. We report that below a threshold
19 perimeter, rings undergo spontaneous rotations. We next sought to extract the
20 ingredients generating this coherence. We show that an internal tug-of-war between
21 cell polarities within the ring determines the onset of coherence, as shown by the initial
22 polarity distributions and the time required to reach coherence. Tracking of cell division
23 as polarity breaker supports this framework. We also report that coherence is
24 associated with two supracellular acto-myosin cables at the inner and outer boundaries
25 of the monolayer: these continuous structures act as autonomous confinements as
26 they exhibit high RhoA activity and are shown to prevent cells from spreading out and
27 breaking coherence. Finally, we test *in silico* our central mechanism of polarity tug-of-
28 war coupled to the acto-myosin cables with a Vicsek-based model writing the
29 fundamental laws of dynamics at cellular levels for polarity, velocity and including
30 forces at boundaries. After calibration, we show that this computational approach
31 predicts the dynamics of all rings with no ad hoc adjustments. We also calculate phase
32 diagrams for coherence as a function of cellular parameters combinations. Our results
33 show that activity at boundaries – supported by RhoA activity in experiments – is
34 indispensable to ensure cellular coherence. We propose that cell polarity tug-of-war
35 together with self-generated confinement from spontaneously assembled active cables
36 set the tempo for coherent movements in epithelial monolayers.

37 38 **RESULTS**

39 40 **Obtaining coherent motion**

41
42 We first needed to identify the natural length over which cells migrate directionally –
43 namely the maximal coherence length ξ_{max} – characterizing our system. A circular
44 array of cells is expected to undergo rotation if the maximal coherence length ξ_{max}
45 matches the perimeter p . If the coherence length is smaller than the perimeter, non-
46 coherent motion is expected^{21,22}. To obtain ξ_{max} , we prepared multicellular rings with
47 free boundaries and decreasing diameters by using micro-contact printing fibronectin
48 regions and differential adhesion (see Methods) : 1000 μ m, 300 μ m and 180 μ m (see
49 Movies S1 to S3, respectively), and we measured the 12-hours velocity fields (see
50 Methods). Without confinement, cells could spread out on passivated and non-coated

1 regions, and rings had different behaviors: large rings – 1000 μm – exhibited growth of
2 cellular fingers inwards and outwards whereas smaller rings – 300 μm and 180 μm –
3 behaved like cellular assemblies during wound healing with centripetal closure (see
4 Fig.1a and Movies S1-S3). For 1000 μm and 300 μm rings, we saw local coherent flows
5 of length ξ (Fig. 1b) but no global coherence. However, at 180 μm diameter, about a
6 quarter of rings spontaneously rotated with a velocity of $\sim 20\mu\text{m}\cdot\text{h}^{-1}$, which suggested
7 that we approached the perimeter value potentially leading to coherent motion. To find
8 this perimeter, we plotted the correlation function of tangential velocity, and we
9 extracted the maximal coherence length ξ_{max} (see Fig.1c, Fig.S1a and Methods) ²³.
10 We show that ξ_{max} , obtained from the x-intersect of the tangent at origin of the
11 correlation function, is similar in every condition – $\xi_{max} = 315\mu\text{m}$ (Fig. 1c and Fig. S1a)
12 –, demonstrating that ξ_{max} is an inherent length of our cellular system.

13
14 Based on this ξ_{max} of 315 μm , we designed and focused on rings with 80 μm diameter
15 corresponding to 250 μm perimeter below ξ_{max} . As expected, we “rectified” the motion
16 of cells around the ring and we ended up with 83% of rings spontaneously rotating at
17 a velocity of 20-25 $\mu\text{m}\cdot\text{h}^{-1}$ (see Movie S4, Fig. 1d and Fig. S1b). All results are then
18 reported in Figure 1e, where $\frac{\xi_{max}}{p}$, the coherence length over the perimeter, is defined
19 as an index of global coherence: it is equal to 1 when the whole ring rotates and gets
20 close to 0 as the global coherence decreases. Interestingly, there is a bias in clockwise
21 rotation which suggests a spontaneous cellular bias as reported in previous works ^{24,25}
22 (Fig. S1c).

23
24 If coherence was obtained for this 80 μm ring diameter, the time needed to reach this
25 complete rotation (defined as $\frac{\xi}{p} = 1$, see Materials and Methods) varied from 1 hour to
26 10 hours and lasted from 1 hour to 10 hours (see Fig. S1d). This large distribution
27 impeded quantification of the transition from non-coherent to coherent motions.
28 Therefore, we needed to rescale the time needed to reach coherence starting from the
29 initial condition of continuous ring of cells. We decided to align all plots with respect to
30 the onset of coherence defined as $\frac{\xi}{p} = 1$. We could then determine the quantitative
31 behavior of coherence acquisition. This pseudo-time representation allowed to show
32 that coherence acquisition is a process lasting about 1h30 with minor variability among
33 rings (Fig. 1f). This timescale is fast considering the 12-hours duration of the
34 experiment and this can be viewed as a quick switch between the two states. At
35 coherence, a continuum of behaviours appeared among the rings populations from
36 rings with persistent rotation over hours to rings with transient coherent motion shorter
37 than one hour (Fig. 1f and Fig. S1d-e). To understand the origin of these dynamics, we
38 measured next the ring velocity and persistence of rotation, and saw their correlations
39 (Fig. S1f). This suggests that collective cell speed might be involved in maintaining
40 coherence even though inertia is negligible at this scale.

41 42 **Tug-of-war between cell polarities sets coherence**

43
44 Next we sought to identify the cellular mechanism at play for the onset of coherent
45 rotation. Cell polarity is a natural readout for its key role in setting direction of motion.
46 We thus looked at the distribution of cell polarities inside rings. For this, we used
47 labeled lamellipodia (see Fig. 2a, see Methods for the measurement of polarity). We
48 observed selected lamellipodia orientation and direction. Initial polarities showed a bi-

1 modal distribution peaked around tangential directions with two main orientations, *i.e.*
2 0° and 180° (see Fig. 2b). These two states suggest that a tug-of-war between
3 polarities could happen between cells of opposite directions within the same ring. We
4 reasoned that initial polarities could then play a role in the time needed to reach
5 coherent rotation. A large number of cells with the same polarities would minimize the
6 tug-of-war, and would decrease the time to coherence. We then followed each cell
7 polarity defined by tight-junction organisation (see Fig. 2c and Movie S5). We had
8 checked that the direction of shape anisotropy defined by cell geometry correlates with
9 the direction of the lamellipodia (Fig. S2Aa-b). Our measurements of percentage of
10 cells with aligned polarities as a function of time to coherence show a decreasing
11 relationship (Fig. 2d). This further indicates that distribution of single cell polarities
12 imposes a tug-of-war and determines the time needed to enter coherence. [We tested](#)
13 [this statement by inhibiting lamellipodia formation with the Arp2/3 inhibitor CK666 and](#)
14 [this prevented coherence \(Fig. S2B\) by suppressing the tug-of-war between polarities.](#)
15

16 The co-existence of opposite polarities is expected to generate stretching within the
17 ring. From tight junctions contours, we could also extract cell elongation (Fig. 2c and
18 Fig. S2A). Using the pseudo-time as the reference (Fig. 2e), we report three phases:
19 (i) 2.5 hours before coherence, cells elongate until (ii) their tangential stretch reach
20 constant values sustained for about 1 hour; (iii) coherence starts shortly within 1h
21 followed by a rapid drop in elongation (see Fig. S2). This is distinct from non coherent
22 rings (see Fig. S2Ac). These measurements substantiate our framework: cell
23 lamellipodia “pull” in opposite directions leading to cellular stretch and resolution of this
24 competition induces cell stretch relaxation visualized by the rapid drop in stretch.
25

26 **Cell divisions as polarity breakers impede coherence**

27
28 To further test the central role for polarity in setting coherence, we tracked cell division
29 events within rings. Right after mitosis, cells have opposite polarities and thus are
30 expected to challenge polarity alignments. These opposite polarities indeed happened
31 at cell division within rings (see Figure 2f). We quantified the effect by plotting the
32 cumulative number of cell divisions within rings. We saw that rings where division
33 regularly occurred were related to poorly coherent motion (Fig. 2g, bottom). In contrast,
34 when a burst of many cell divisions occurred within a short time period (typically 2
35 hours over a total time of experiment of 12 hours), coherence was maintained in its
36 optimal value (Fig. 2g, top). [Divisions generate cells with opposite polarities and](#)
37 [impede coherence \(see Fig. S2Ae,g,h\).](#) Division as a polarity breaker acts as a
38 coherence breaker.
39

40 41 **Acto-myosin cables as internally self-assembled boundaries imposing** 42 **tangential cell polarity**

43
44 So far, we reported that cells were polarized within the ring and we next sought to
45 further understand why tangential orientation was selected (Fig. 2a-b). If the rings were
46 confined by physical walls, this orientation would be favored²⁰. However, in our assay,
47 cells were free to move in any directions suggesting that an additional phenomenon
48 biases cell polarities. We first stained for actin and myosin since acto-myosin cables
49 were reported to self-assemble within cells at epithelial colony boundaries^{17,26,27} (Fig.
50 S3Aa). This generic feature was therefore expected to appear as well in our rings and

1 we hypothesized that these structures could contribute to orient cell polarity. We show
2 in Figure 3a the presence of these acto-myosin cables and we propose that cells
3 internally set their “walls” which in turn could potentially make them be oriented along
4 the perimeter rather than radially.

5
6 To test this idea, we evaluated the mechanical contributions of these cables and their
7 roles in cell confinement. First with live observations, we saw that the inner cable was
8 contractile and led first to the closure of rings within hours prior to extension ‘out’ of the
9 cellular disk now formed (Fig. 3b, Fig. S3Ab and Movies S3-4). This suggests that
10 indeed inner cable pulls inwards as expected for a contracting acto-myosin cable. [This](#)
11 [dynamic was distinct from larger rings of 1mm \(see Fig. 1 and Movie S1\) which](#)
12 [suggests that cable curvature plays a role in cell motion](#)²⁸. [This curvature dependence](#)
13 [was further substantiated by experiments with elliptical geometries \(see Fig. S3B\).](#)
14 Then, we reasoned that the outer ring may act the same way by pushing cells inwards.
15 To evaluate whether these structures were contractile, we used laser ablation
16 experiments²⁹. First we observed a fast and large opening of both cables within
17 seconds with a similar extent of ~ 12µm (see Figure 3c and 3d and Movie S6, see also
18 Fig. SA3b, and Methods). This demonstrates that cables are more tensile than cell-cell
19 junctions also tested in these experiments which yielded small openings (~ 1µm). In
20 addition, we measured the opening velocity after laser ablation as a read-out for
21 comparing stress between inner and outer cables assuming they bear the same
22 damping coefficients²⁹. Our results suggest that acto-myosin cables indeed apply
23 contractile constraints on the boundary cells and with similar values (see Fig. 3d).

24
25 Next, we tested the impact of this contraction at boundaries on cell polarity. We
26 followed cells transfected with Lifeact within non labeled rings. We report in Fig. 3e and
27 Movie S7 the interplay between cell polarity and the outer acto-myosin cable: cells
28 could build their own acto-myosin cables while the structure imposes lamellipodia to
29 be oriented tangentially. This supports the notion that acto-myosin cables and
30 tangential polarity are correlated.

31
32 To further test this coupling, we sought to remove locally the influence of acto-myosin
33 cables. We included within rings single cells expressing caldesmon (CaD) (Fig. 3f), a
34 calmodulin binding protein inhibiting Myosin ATPase³⁰. Indeed this triggered a local
35 decrease in myosin activity and in cable contractility. These caldesmon expressing
36 cells extended out of the rings with a radial motion (Movie S8). This further confirms
37 the key role played by the cables in confining cell polarity tangentially. This perturbation
38 also impaired rotations in the majority of cases (see Fig. 3g and Suppl. Fig. 3d). On
39 average, we found that coherence was lower on CaD mosaic rings (Fig. 3g) and this
40 was correlated to the amount of CaD levels (Fig. S3Ae). Altogether, this suggests that
41 continuity of the acto-myosin rings is important for tangential motion and coherence. In
42 conclusion, cables are essential to confine and orient cells polarities tangentially and
43 further trigger rotations.

44
45 **Force is active at boundaries and cell velocity is corelated with RhoA level.**

46
47 Observations and measurements on cell dynamics could hide signaling effects
48 associated to regulations of the acto-myosin cytoskeleton by the small GTPase
49 RhoA^{31,32}. To evaluate this potential contribution, we analysed the correlation between
50 spatio-temporal dynamics of RhoA activity and cell dynamics. [For this, we used a](#)

1 [FRET sensor which shows distributions of active RhoA in space and time](#) (Fig. 4a and
2 Ref.^{18,33}, see Methods and Fig. S4a-c). In MDCK rings, we found the largest activity at
3 inner and outer boundaries, where acto-myosin cables are assembled (Movie S9, Fig.
4 4b and Fig. 3a). This supports the contractile nature of these fibers. Accordingly, we
5 found that the closure time of rings is shorter for large FRET activities in cables (see
6 Fig. S4d-e). We can conclude that regulation by RhoA mediates active forces at
7 boundaries.

8
9 Building on this result, we sought to test whether correlation between RhoA and cell
10 velocity could be at play in the onset of coherence, knowing that cell velocity plays a
11 key role in collective cell motion. We analyzed locally and globally velocity fields and
12 their associated FRET maps (Fig. 4c and [Fig. S4h, f, g](#), see also Methods). We found
13 that high velocity regions for cells were correlated with low FRET levels (Fig. 4d,e and
14 Suppl Mat). This suggests an inverse relationship between the two quantities (see also
15 Fig S4i). Cells move faster with lower levels of FRET. This further supports the notion
16 that RhoA activity is essentially encoded in cell velocity.

17
18 We then turned to numerical simulations to check whether the onset of collective
19 motions could happen with basic interactions rules extrapolated from our experimental
20 results, i.e. active force at boundaries and cell velocity rule.

21 22 23 **Numerical simulations reproduce the ring dynamics qualitatively and** 24 **quantitatively**

25
26 [We designed numerical simulations based on an original Vicsek type model with active](#)
27 [boundaries, involving parameters accessible to experiments](#) (see Fig. 5, Fig. S5, Table
28 1 and Annex_Math). Cells are represented as particles with velocity \mathbf{v} and polarity \mathbf{p} .
29 Backed up by experimental observations (Fig. S5a-b), we assume that cell velocity
30 aligns to its polarity. Cell polarity is allowed to diffuse with a coefficient D (Fig. S5d-e)
31 and aligns with the average polarity around the ring with rate μ (Fig. S5f). Cell polarity
32 is also assumed to align with cell velocity direction with rate ν . Local springs between
33 boundary cells dynamically describe the inner and outer cables. Active and passive
34 contributions of these cables are incorporated in the polarity dynamics. Table 1
35 summarizes data taken from experiments and inserted as an input to the model. More
36 details are provided in Annex Math.

37
38 We report in Fig. 5 the comparisons in shapes and coherence values across
39 conditions. The ring phenotypes are reproduced with multicellular fingers or clusters
40 for distinct colony diameters (Fig. 5a and Movie S10-13). Similarly, the coherence
41 index is quantitatively obtained with excellent precision (see Fig. 5b) together with its
42 typical time dependence (Fig. 5c). Also, the tug-of-war between polarities correlated
43 with the time required to reach coherence for simulated rings with quantitative
44 consistency with experimental times (Fig. 5d). [We also found that closure time depends](#)
45 [on the initial cell alignment](#) (Fig. S6a).

46
47 In addition, we could probe numerically the potential contribution of active forces at
48 cables: in our model, cables have a passive component corresponding to the stretching
49 and bending moduli of the acto-myosin structure, and an active component translating
50 the contribution of cables in setting tangential polarities (as explained above and

1 shown in Figure 3). Interestingly, the active component was required for the onset of
2 coherence (Fig. 5e). We also tested the effect of having a cell with no cable within the
3 ring, thereby modelling the caldesmon experiment (Fig. 3f): the phenotype of this cell
4 escape was reproduced as well (Fig. 5f and Movie S14), substantiating the
5 confinement role of cables. [We checked that extremely stiff cable acted as walls like
6 in Jain et al. and led to coherent motion \(Fig. A10 and A11, Annex Math\).](#)

7
8 Finally, we generated phase diagrams to understand the relationship between
9 coherence and cell polarity and cables activity. We found the range of values
10 necessary to trigger coherence for the main parameters of the model (cell polarity
11 alignment, cable activity and physical properties) (Fig. 5g and 5h). We show that the
12 coherence value is essentially governed by the cell polarity alignment: a large collective
13 alignment increases coherence (Fig. 5g and [Fig. S6c-d](#)) while the bending and the
14 stretching moduli (passive terms) do not affect coherence initiation at least in the range
15 of considered parameters (Fig. 5h and [Fig. S6b](#)). However, the cables properties have
16 a strong impact on the closure time of the ring and passive and active terms have
17 antagonist effects in this regard (Fig. S5g and S5h).

18
19 Altogether our numerical experiments based on measured cellular parameters and
20 with basic interactions for single cells show that we can capture the main features for
21 the onset of coherence with no need to incorporate parameters other than polarity and
22 active cables at boundaries.

23 24 **DISCUSSION**

25
26 We show that coherence of cellular movements emerges when the system size is
27 below the coherence length inherent to MDCK cells as reported previously on epithelial
28 disks ^{21,22,34,35}. Cells align their polarities within the ring and this step sets the onset of
29 coherence. Furthermore, acto-myosin cables act as internally driven constrains
30 confining cells tangentially. RhoA activity is larger at boundaries co-localizing with
31 cables and is inversely related to the cell maximum velocity. Finally, with these
32 ingredients, experimental and numerical rings share qualitative and quantitative
33 features, supporting the relevance of our minimal framework for cell interactions. We
34 show that active forces at cables are critical to orient cell polarity and contribute to
35 coherence.

36 37 **Internally driven coherence versus externally driven coherence**

38
39 Emergence of coherent flows has been studied *in vitro* in confined situations ^{20–22}. Cells
40 were chemically or mechanically confined and cannot escape their adhesive patterns
41 ^{21,22}. This led to distinct phenomena – although with similar velocities of $20 \mu\text{m}\cdot\text{h}^{-1}$ -
42 where the coherence is not driven only by the cellular characteristics but also by the
43 boundaries of confinement as suggested by previous theoretical works ³⁶. [In Ref. 20,](#)
44 [this led to rotations even on rings with a diameter of 1mm, far above the natural](#)
45 [coherence length of the epithelial cell line. We reproduced this result numerically \(see](#)
46 [Sec. 3.4 Annex Math\) which further shows that the external walls may impose the](#)
47 [rotation for large scales. Intuitively this can be understood by the fact that cell motion](#)
48 [within a cohesive ring monolayer can only be tangential with solid boundary.](#) In our
49 case, we let cells spontaneously self-organize without confinement and we decreased
50 the pattern size to initiate the onset of coherence. This new configuration makes this

1 rotation spontaneous in essence and not guided by 3D walls which force the motion
2 along the ring. Remarkably, the symmetry of rotation clockwise-counterclockwise is
3 broken on average, in contrast to former studies in confinement²⁰ but in agreement
4 with the bias reported in other studies^{24,25,37} with potential roles for left-right asymmetry
5 during development. We make the hypothesis that chirality at the single cell level –
6 generated by molecular structures with broken symmetry²⁵ – is conserved through
7 scales and is translated in the emerging collective motion³⁸. Altogether, our
8 spontaneous broken symmetry reveals inherent cellular properties which convey
9 spatio-temporal order selected by cells.

10 11 **Coherence length and spatial scales**

12
13 It is interesting to analyze the potential origin of this natural coherence length. This
14 length is distinct from former works reporting spontaneous flows of active nematics:
15 rotation was not associated with opposite senses of rotation at boundaries, and the
16 coherence length is higher compared to these 2D confluent monolayers. Here, cell
17 motions are not impeded by cellular jamming seen in fully confluent monolayers³⁹.
18 Therefore, we propose that the coherence length found in our study is not expected to
19 be the same as that defining the crossover from rotation to active turbulence seen in
20 fully two dimensional confluent layers of cells.

21
22 In addition, the passage from single cell to tens of cells forming a new entity necessarily
23 go through adhesion mediated by cell-cell junctions and its interplay with cytoskeleton.
24 We indeed saw that rings with MDCK cells overexpressing cadherins presented a
25 larger amount of rotations compared to WT MDCK cells. This suggests that larger
26 adherence between cells either increases the coherence length or/and initiation is
27 facilitated due to higher cell polarization rates μ and ν . Therefore, we can conjecture
28 that the spontaneous length could be organ dependent and contribute to the key
29 morphogenesis events.

30 31 **RhoA activity and its inverse relation with velocity**

32
33 We report live activity of RhoA integrated over an entire multicellular system. With
34 coarse graining, we considered MDCK rings as a continuum material where RhoA has
35 its own dynamics and extracted FRET levels through time and space. Our results point
36 to a simplification in their connection : RhoA and velocity are inversely related (along
37 other work⁴⁰) and this allows to consider only velocities for the model. This
38 simplification would need to be further tested in other situations but it opens an
39 interesting framework with a simple relation between RhoA activity and cell velocity. In
40 future studies, RhoA and other Rho GTPases could be modulated through space and
41 time using photoactivable tools to further understand its implication in collective
42 coherent motion^{41,42}. This will allow to control cell polarity and cell velocity and in turn
43 characterize the coupling between Rho signaling and cell mechanics in specific
44 manner with identification of new mechanosensory proteins.

45 46 **Implication of coherent motions *in vivo***

47
48 Our velocities found during coherent motion were close to the ones found during
49 *Drosophila* egg chamber development^{10,11}. In this context, follicle cells collectively
50 rotate along the chamber and the presence of lamellipodia is reported¹⁰. This could

1 potentially mean that the phenomena described in this work on 2D rings, driven by cell
2 “cryptic” lamellipodia, act also during these *in vivo* processes. Also, coherent rotations
3 have been reported in various morphogenetic events *in vivo* in mammary acini, in egg
4 chamber and in testis rotation in *Drosophila*^{10–13,43}. We propose that these phenomena
5 could be generic and experienced by any growing tissues as suggested also by a
6 recent theoretical study⁴⁴. If right, spontaneous rotation could occur when coherence
7 lengths of each epithelial layer would be similar to dimensions of tissues. It would be
8 interesting to compare coherence lengths across model systems to evaluate whether
9 this generic property of living matter is physiological and is important for optimal
10 development.

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21 **REFERENCES**

- 22 1. Munjal, A., Philippe, J., Munro, E. & Lecuit, T. A self-organized biomechanical
23 network drives shape changes during tissue morphogenesis. (2015)
24 doi:10.1038/nature14603.
- 25 2. Nishikawa, M., Naganathan, S. R., Jülicher, F. & Grill, S. W. Controlling
26 contractile instabilities in the actomyosin cortex. *Elife* 1–21 (2017)
27 doi:10.7554/eLife.19595.
- 28 3. Sit, S. T. & Manser, E. Rho GTPases and their role in organizing the actin
29 cytoskeleton. *J. Cell Sci.* **124**, 679–683 (2011).
- 30 4. Ridley, A. J. & Hall, A. The small GTP-binding protein rho regulates the
31 assembly of focal adhesions and actin stress fibers in response to growth
32 factors. *Cell* **70**, 389–399 (1992).
- 33 5. Comelles, J. *et al.* Elongations of epithelial colony in vitro : symmetry breaking
34 through collective effects. *bioRxiv* 1–15 (2019).
- 35 6. Yam, P. T. *et al.* Actin-myosin network reorganization breaks symmetry at the
36 cell rear to spontaneously initiate polarized cell motility. *J. Cell Biol.* **178**, 1207–
37 1221 (2007).
- 38 7. Arboleda-Estudillo, Y. *et al.* Movement Directionality in Collective Migration of
39 Germ Layer Progenitors. *Curr. Biol.* **20**, 161–169 (2010).
- 40 8. Jain, A. *et al.* Regionalized tissue fluidization is required for epithelial gap
41 closure during insect gastrulation. *Nat. Commun.* **11**, (2020).
- 42 9. Münster, S. *et al.* Attachment of the blastoderm to the vitelline envelope affects
43 gastrulation of insects. *Nature* **568**, 395–399 (2019).
- 44 10. Cetera, M. *et al.* Epithelial rotation promotes the global alignment of contractile
45 actin bundles during *Drosophila* egg chamber elongation. *Nat. Commun.* **5**,
- 46
- 47
- 48
- 49
- 50

- 1 (2014).
- 2 11. Haigo, S. L. & Bilder, D. Global tissue revolutions in a morphogenetic
3 movement controlling elongation. *Science (80-.)*. **331**, 1071–1074 (2011).
- 4 12. Wang, H., Lacoche, S., Huang, L., Xue, B. & Muthuswamy, S. K. Rotational
5 motion during three-dimensional morphogenesis of mammary epithelial acini
6 relates to laminin matrix assembly. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 163–
7 168 (2013).
- 8 13. Tanner, K., Mori, H., Mroue, R., Bruni-Cardoso, A. & Bissell, M. J. Coherent
9 angular motion in the establishment of multicellular architecture of glandular
10 tissues. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 1973–1978 (2012).
- 11 14. Selmeçzi, D., Mosler, S., Hagedorn, P. H., Larsen, N. B. & Flyvbjerg, H. Cell
12 Motility as Persistent Random Motion : Theories from Experiments. *Biophys. J.*
13 **89**, 912–931 (2005).
- 14 15. Vedula, S. R. K. *et al.* Emerging modes of collective cell migration induced by
15 geometrical constraints. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 12974–12979
16 (2012).
- 17 16. Ladoux, B. & Mège, R. M. Mechanobiology of collective cell behaviours. *Nat.*
18 *Rev. Mol. Cell Biol.* **18**, 743–757 (2017).
- 19 17. Reffay, M. *et al.* Interplay of RhoA and mechanical forces in collective cell
20 migration driven by leader cells. *Nat. Cell Biol.* **16**, 217–223 (2014).
- 21 18. Pertz, O., Hodgson, L., Klemke, R. L. & Hahn, K. M. Spatiotemporal dynamics
22 of RhoA activity in migrating cells. *Nature* **440**, 1069–1072 (2006).
- 23 19. Turing, A. M. The chemical basis of morphogenesis. *Philos. Trans. R. Soc.*
24 *London* **237**, 37–72 (1952).
- 25 20. Jain, S. *et al.* The role of single-cell mechanical behaviour and polarity in
26 driving collective cell migration. *Nat. Phys.* **16**, 802–809 (2020).
- 27 21. Deforet, M., Hakim, V., Yevick, H. G., Duclos, G. & Silberzan, P. Emergence of
28 collective modes and tri-dimensional structures from epithelial confinement.
29 *Nat. Commun.* 1–9 (2014) doi:10.1038/ncomms4747.
- 30 22. Doxzen, K. *et al.* Guidance of collective cell migration by substrate geometry.
31 *Integr. Biol. (United Kingdom)* **5**, 1026–1035 (2013).
- 32 23. Petitjean, L. *et al.* Velocity fields in a collectively migrating epithelium. *Biophys.*
33 *J.* **98**, 1790–1800 (2010).
- 34 24. Chin, A. S. *et al.* Epithelial cell chirality revealed by three-dimensional
35 spontaneous rotation. *Proc. Natl. Acad. Sci. U. S. A.* **115**, 12188–12193 (2018).
- 36 25. Tee, Y. H. *et al.* Cellular chirality arising from the self-organization of the actin
37 cytoskeleton. *Nat. Cell Biol.* **17**, 445–457 (2015).
- 38 26. Nier, V. *et al.* Tissue fusion over nonadhering surfaces. *Proc. Natl. Acad. Sci.*
39 *U. S. A.* **112**, 9546–9551 (2015).
- 40 27. Ravasio, A. *et al.* Regulation of epithelial cell organization by tuning cell-
41 substrate adhesion. *Integr. Biol. (Camb)*. **7**, 1228–1241 (2015).
- 42 28. Ravasio, A. *et al.* Gap geometry dictates epithelial closure efficiency. *Nat.*
43 *Commun.* **6**, (2015).
- 44 29. Mayer, M., Depken, M., Bois, J. S., Jülicher, F. & Grill, S. W. Anisotropies in
45 cortical tension reveal the physical basis of polarizing cortical flows. *Nature*
46 **467**, 617–621 (2010).
- 47 30. Helfman, D. M. *et al.* Caldesmon inhibits nonmuscle cell contractility and
48 interferes with the formation of focal adhesions. *Mol. Biol. Cell* **10**, 3097–3112
49 (1999).
- 50 31. Ridley, A. J., Paterson, H. F., Johnston, C. L., Diekmann, D. & Hall, A. The

- 1 small GTP-binding protein rac regulates growth factor-induced membrane
2 ruffling. *Cell* **70**, 401–410 (1992).
- 3 32. Hodgson, L., Shen, F. & Hahn, K. Biosensors for characterizing the dynamics
4 of Rho family GTPases in living cells. *Curr. Protoc. Cell Biol.* 14111–141126
5 (2010) doi:10.1002/0471143030.cb1411s46.
- 6 33. MacHacek, M. *et al.* Coordination of Rho GTPase activities during cell
7 protrusion. *Nature* **461**, 99–103 (2009).
- 8 34. Segerer, F. J., Thüroff, F., Piera Alberola, A., Frey, E. & Rädler, J. O.
9 Emergence and persistence of collective cell migration on small circular
10 micropatterns. *Phys. Rev. Lett.* **114**, 1–5 (2015).
- 11 35. Zorn, M. L., Marel, A. K., Segerer, F. J. & Rädler, J. O. Phenomenological
12 approaches to collective behavior in epithelial cell migration. *Biochim. Biophys.*
13 *Acta - Mol. Cell Res.* **1853**, 3143–3152 (2015).
- 14 36. Soumya, S. S. *et al.* Coherent Motion of Monolayer Sheets under Confinement
15 and Its Pathological Implications. *PLoS Comput. Biol.* **11**, 1–30 (2015).
- 16 37. Naganathan, S. R. a., Fürthauer, S., Nishikawa, M., Jülicher, F. & Grill, S. W.
17 Active torque generation by the actomyosin cell cortex drives left-right
18 symmetry breaking. *Elife* **3**, e04165 (2014).
- 19 38. Tee, Y. H. *et al.* Actin polymerisation and crosslinking drive left-right asymmetry
20 in single cell and cell collectives. *Nat. Commun.* **14**, 776 (2023).
- 21 39. Garcia, S. *et al.* Physics of active jamming during collective cellular motion in a
22 monolayer. *Proc. Natl. Acad. Sci. U. S. A.* **112**, 15314–15319 (2015).
- 23 40. Gupta S, Duszyc K, Verma S, Budnar S, Liang X, Gomez GA, Marcq P,
24 Noordstra I, Y. A. Enhanced RhoA signalling stabilizes E-cadherin in migrating
25 epithelial monolayers. *J. Cell Sci.* **17**, (2021).
- 26 41. Juan, G. R. R., Oakes, P. W. & Gardel, M. L. Contact guidance requires spatial
27 control of leading-edge protrusion. *Mol. Biol. Cell* **28**, 1043–1053 (2017).
- 28 42. Wu, Y. I. *et al.* A genetically encoded photoactivatable Rac controls the motility
29 of living cells. *Nature* **461**, 104–108 (2009).
- 30 43. Chougule, A. *et al.* The Drosophila actin nucleator DAAM is essential for left-
31 right asymmetry. *PLoS Genet.* **16**, 1–23 (2020).
- 32 44. Maitra, A. & Lenz, M. Spontaneous rotation can stabilise ordered chiral active
33 fluids. *Nat. Commun.* **10**, 1–6 (2019).
- 34 45. Beutel, O., Maraspini, R., Pombo-García, K., Martin-Lemaitre, C. &
35 Honigmann, A. Phase Separation of Zonula Occludens Proteins Drives
36 Formation of Tight Junctions. *Cell* **179**, 923-936.e11 (2019).
- 37 46. Etournay, R. *et al.* TissueMiner: A multiscale analysis toolkit to quantify how
38 cellular processes create tissue dynamics. *Elife* **5**, 1–28 (2016).
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1 MATERIAL AND METHODS

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3 Micro-contact printing

4

5 Ring motifs are patterned on glass coverslips using micro-contact printing.
6 Polydimethylsiloxane (PDMS; 1:10 w/w cross-linker:pre-polymer) (Sylgard 184 kit,
7 Dow Corning, cat. DC184-1.1). Stamps are produced with UV-photolithography.
8 Stamps are rinsed with 70% ethanol and then rendered hydrophilic by exposure to
9 oxygen plasma (30s) (Diener Electronic, cat. ZeptoB). They are next incubated with a
10 10 µg/mL rhodamine-labelled fibronectin solution (Cytoskeleton, cat. FNR01-A) for 1h
11 and then dried at room temperature for about 5min. In the meantime, glass coverslips
12 are functionalized by vapour phase for 1h with 3-(mercapto)propyltrimethoxysilane
13 (FluroChem). After incubation of both the coverslip and PDMS, the stamp is deposited
14 onto the glass slide and kept in contact during 30min. To ensure a proper transfer
15 between the two surfaces, a 50g weight is placed on the top of the stamp. Finally, the
16 stamp is removed and the printed coverslip is washed with PBS 1X and stored in Milli-
17 Q water. Non-printed areas are passivated with 0.1 mg/mL PLL-g-PEG (in 1mM
18 HEPES pH 7.4, SuSoS AG, cat. SZ33-15) for 20min at room temperature. [This](#)
19 [approach consists in setting differential coating on the surface allowing to plate cells](#)
20 [and select cells adhering to the fibronectin printed ring.](#)

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22 Cell Culture

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24 MDCK cells stably transfected with E-cadherin-DsRED are cultured in low glucose
25 Dulbecco's Modified Eagle's (DMEM) with 10% Foetal Bovine Serum (FBS) and 1%
26 Penicillin-Streptomycin. Cells are kept subconfluent (at ~ 70%). Prior experiments,
27 cells are detached with Trypsin-0.25% EDTA (Fisher Scientific, cat. 11570626) and
28 centrifuged at 500rpm during 3min. The pellet is then re-suspended in DMEM 1% FBS.
29 This low serum concentration will further impede attachment on the passivated surface
30 during the first hours following seeding (differential adhesion). After 1h incubation, the
31 printed surface is carefully washed out to remove cells from the non-printed area. After
32 washout, DMEM is replaced by L-15 Leibovitz Medium (Fisher Scientific, cat.
33 11540556) supplemented with 1% FBS and 1% Penicillin-Streptomycin and the
34 sample is incubated 1h under the microscope at 37°C before acquisition. This last
35 incubation allows cells to spread on patterns and ensures that rings are closed at the
36 onset of the experiment.

37

38 For laser ablation experiments, MDCK stably transfected with GFP-Myosin Regulatory
39 Light Chain (MRLC) were used. Tight junctions were imaged with a MDCK cell line
40 genetically engineered with CRISPR-Cas9 to express ZO1-GFP⁴⁵. We generated a
41 MDCK cell line stably expressing the RhoA FRET biosensor using Piggy-Bac
42 engineering. Briefly, cells were transfected with the biosensor plasmid together with a
43 transposase. After integration into the genome, fluorescent cells were sorted with flow
44 cytometry and kept under antibiotics pressure for 3 weeks.

45

46 Transfection

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48 Plasmids for mosaic experiments such as Actin-GFP or Caldesmon-GFP were
49 incubated with cells using Lipofectamin (Invitrogen, cat. 11668030). One day prior
50 experiment, cells at 50% confluency in a 6-well plate were incubated for 4-6h with 1µg

1 of DNA and 10 μ L of lipofectamin mixed in Opti-MEM medium. After incubation, wells
2 are washed out and the medium is replaced by fresh DMEM 10% FBS.

3

4 **Drug experiments**

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6 Cells were incubated with CK666 50 μ M (Sigma-Aldrich, cat. SML0006) during the
7 whole course of the experiment (12 hours).

8 For RhoA inhibition, cells were incubated 2 hours with C3-transferase (Cytoskeleton
9 Inc., cat. CT03-A) at a concentration of 1 μ g/mL. Culture medium was washed out
10 before the start of the experiment.

11

12 **Standard time-lapse microscopy**

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14 Phase contrast images were acquired through a 10x objective (NA = 0.25) with a frame
15 rate of 1 image every 10min. Patterns were systematically checked prior the time-lapse
16 start with a standard epifluorescence lamp (FluoArc Hg Lamp) coupled with a
17 rhodamine fluorescence filter. The very same set-up was used to acquire cells during
18 mosaic experiments (actin-GFP and caldesmon-GFP). Tight junctions (ZO1-GFP)
19 were imaged with a confocal microscope (Leica SP8 inverted) and an autofocus
20 system. All z-planes were taken and finally projected onto one single plane.

21

22 **Biosensor FRET activity**

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24 Images were acquired with a confocal microscope (Leica SP8 inverted) with the
25 pinhole opened to the maximum in order to increase slice thickness and the signal to
26 noise ratio for the analysis. We used two Photo Multiplier Tubes (PMTs) as detectors.
27 Bandwidths were chosen as follow: 472nm-512nm for mTFP1 (CFP analog, donor)
28 and 523nm-544nm for Venus (YFP analog, acceptor). mTFP1 was excited by an argon
29 laser at 468nm with 15% of power and emission for mTFP1 (donor) and Venus (FRET)
30 were simultaneously acquired by the two PMTs. Images were then corrected and
31 processed with Biosensor Package from Danuser Lab (available on the following link:
32 <https://github.com/DanuserLab/Biosensor>). Because it is a unimolecular biosensor, no
33 bleedthrough correction was done. Prior analysis, histograms were manually resliced
34 in order to remove high and close-to-zero boundary artefacts.

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36 To check whether the FRET sensor was reliable, we performed additional tests:
37 incubation with 1.0 μ g/ml C3-transferase (RhoA inhibitor, Cytoskeleton, cat. CT04) and
38 with 1.0 μ g/ml of RhoA activator (Cytoskeleton, cat. CN03). Drugs led to decrease and
39 increase of activities respectively (Fig. S4a-b). We also used a mutant sensor – F39A,
40 unable to interact with the RBD domain¹⁸ – with the same acquisition parameters
41 detailed above. This was associated with a significant decrease in the FRET signal
42 (Fig. S4c).

43

44 **Laser ablation**

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46 Laser ablations on acto-myosin cables were done using the FRAP module from a Leica
47 SP-5 inverted microscope. The sample was imaged and ablated through a 40x oil
48 objective (NA = 1.25) and an 800nm infrared laser (80 MHz, pulsed).

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Velocity fields and coherence length

Velocity fields were generated using PIVlab 1.4 and performing cross-correlations. Windows of about a cell size, adjusted to the magnification (32x32 or 64x64 in PIVlab) were taken. No smoothing was performed on the velocity fields.

Coherence lengths ξ were extracted from velocity fields with a custom-made Matlab code by computing the spatial correlation function of the tangential velocity v_θ :

$$C_{v_\theta}(\delta x, t) = \frac{\langle \vec{v}_\theta(\vec{x} + \delta \vec{x}, t) \cdot \vec{v}_\theta(\vec{x}, t) \rangle}{\langle v_\theta(\vec{x}, t)^2 \rangle}$$

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Exponential decays are generated from the computations of C_{v_θ} and the intersections between the tangents at origin and x-axis give ξ . Because C_{v_θ} was not always converging to 0, we made linear fits on the 3 first points of C_{v_θ} in order to extract the tangent at origin.

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We selected ξ_{max} to capture the largest length accessible to the cells. To extract this maximal coherence length ξ_{max} presented in Figure 1c and Fig. S1a, velocity fields were averaged over intervals of 10 frames (100min) and the time interval leading to the largest correlation function was considered. Stated differently, we isolated the time window during which the coherent motion was optimal and extracted the associated length over experimental conditions.

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Image processing

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Nuclei in RhoA ratiometric maps were removed using Ilastik and phase contrast images were processed with Fiji.

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Cell polarity was defined by its lamellipodium: the axis nucleus-lamellipodium sets direction and orientation for cell polarity. Since many factors could contribute to define physical polarity and because lamellipodia are not always observable, we decided to track ZO1 marked cells – not in their anisotropy of signal – but for the well-defined shapes. In majority of cases, cells could be associated to isosceles triangles. Therefore, cell polarity was given by the long axis of the triangle while the sense was defined by the triangle base. We confirmed that this polarity was consistent with the polarity defined by lamellipodia in the large majority of cases (Fig. S2a-b). Segmentation and extraction of single cell parameters in ZO1 experiments were done using Tissue Analyzer⁴⁶ in Fiji.

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Model

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We designed a mechanistic-based minimal “Vicsek—type model” in which cells are treated as particles with polarity \mathbf{p} and velocity \mathbf{v} , initially placed on a ring with two layers of cells matching the cell density and number tested in experiments. Their interaction rules are determined by alignment of polarities and by polarity relaxation to the direction of motion and by random fluctuations of polarities (see Annex Math for details). Velocities are also projected on the directions of polarities to the limit of volume exclusion. The inner and outer supra-cellular acto-myosin cables are modeled as

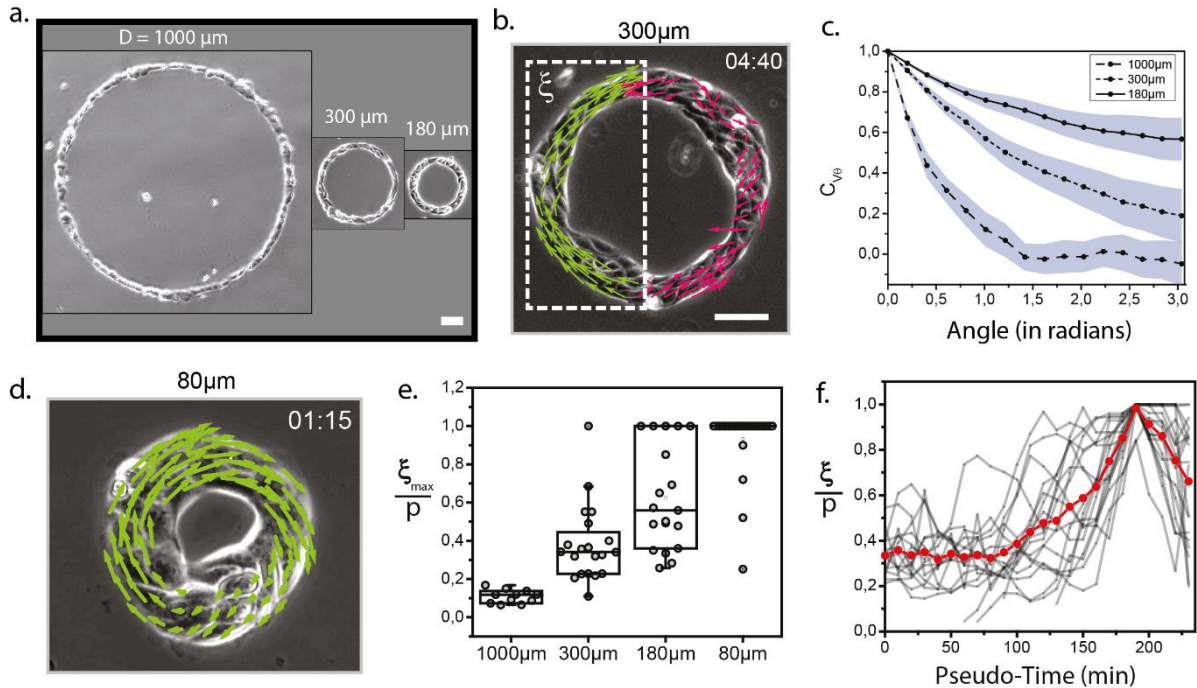
1 segments connecting boundary cells with two physical elastic properties, stretching
2 and bending energies. In addition to the force exerted by the cables on the cells and
3 its velocities, cables also act on cell polarity. The values of parameters are extracted
4 from measurements and from estimates based on references (see Annex Math and
5 Table 1).

6
7 We show (see Fig. 5e) that with this passive treatment of cables and actual
8 experimental values, we do not recapitulate coherent motion. Based on the Rho activity
9 at both cables supporting an active stress generation, we add in the model an active
10 cable force with estimates of the associated experimental values. Simulations based
11 on the theoretical model reveal that cells undergo coherent motion for small rings.
12 Moreover, when we increase the ring diameter keeping the same values for
13 parameters, we reproduce the experimental dynamics with no further adjustments.
14 These results support the relevance of our model.

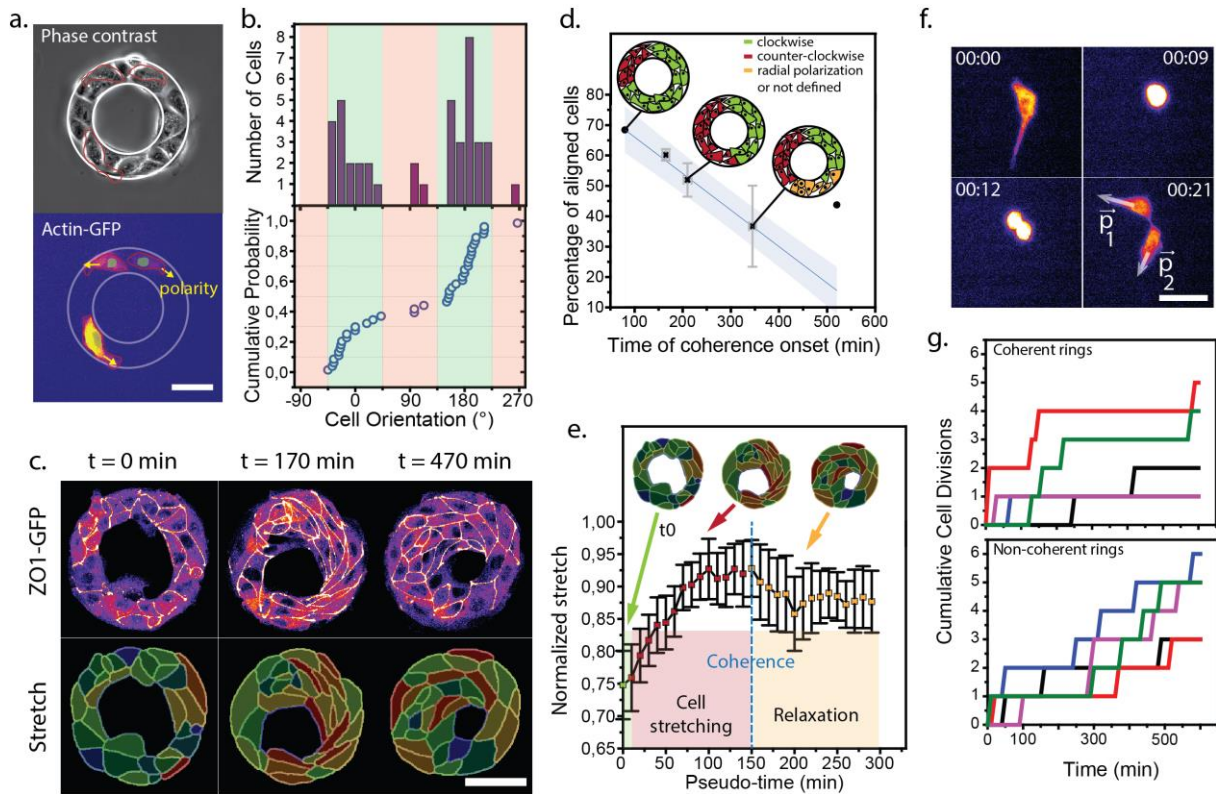
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1 **FIGURES AND MOVIE CAPTIONS**

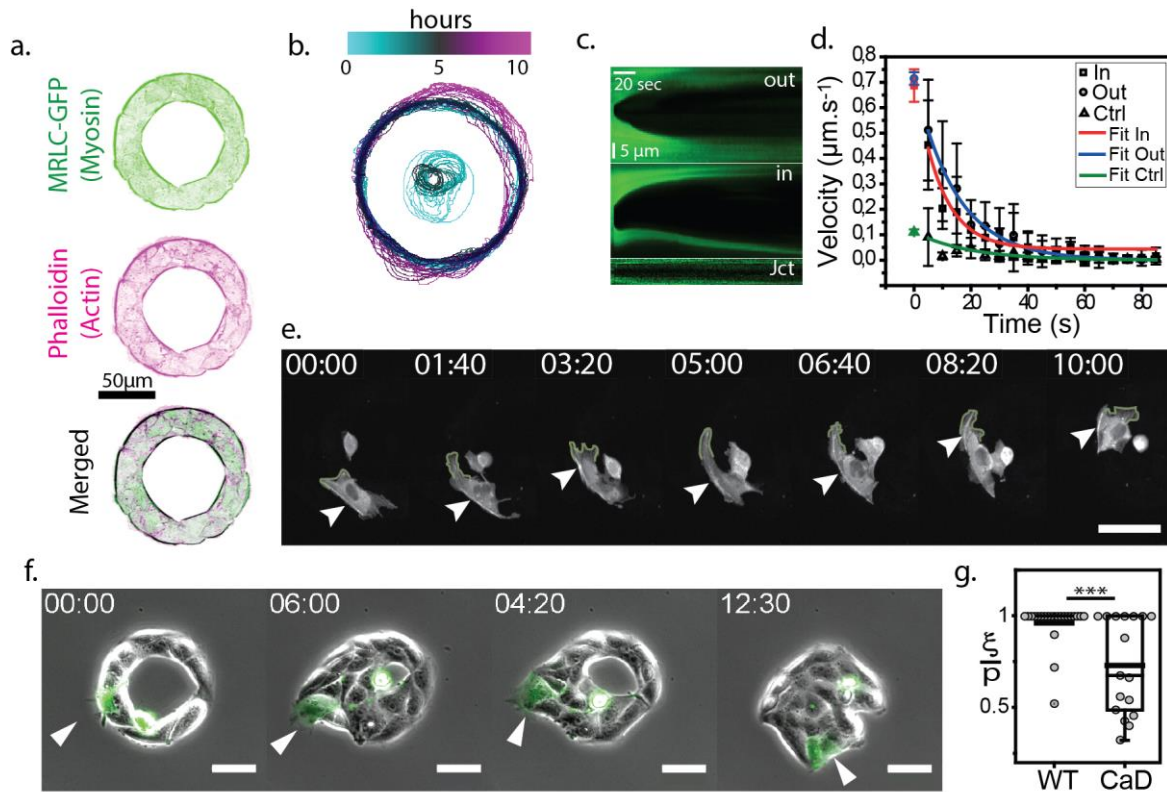
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4 **Figure 1. Acquisition of coherent motion.** (a) MDCK rings of various diameters at
5 initial time t_0 . (b) Coherent flows can be extracted from the velocity fields. A coherent
6 flow of length ξ is highlighted in green. Time in hh:mm; Scale bar = $50 \mu\text{m}$. (c) Average
7 correlation functions of tangential velocity v_θ for each ring diameter. The angle θ
8 corresponds to the polar coordinate on the ring. (d) $80 \mu\text{m}$ ring with a perimeter below
9 the coherence length undergoes spontaneous rotation. (e) Index of global coherence
10 $\frac{\xi_{max}}{p}$ for each ring diameter. $80 \mu\text{m}$ rings are “rectified” and the majority of rings are
11 coherent ($\frac{\xi_{max}}{p} = 1$). $n_{1000} = 11$; $n_{300} = 20$; $n_{180} = 20$; $n_{80} = 24$. Medians are shown as
12 lines in the boxplot. (f) Temporal sequences are aligned on the coherence acquisition
13 ($\frac{\xi}{p} = 1$) along a new pseudo-time. The red line corresponds to the mean of all
14 experiments. Coherence is a non-linear process taking place within $\sim 1\text{h}30$.
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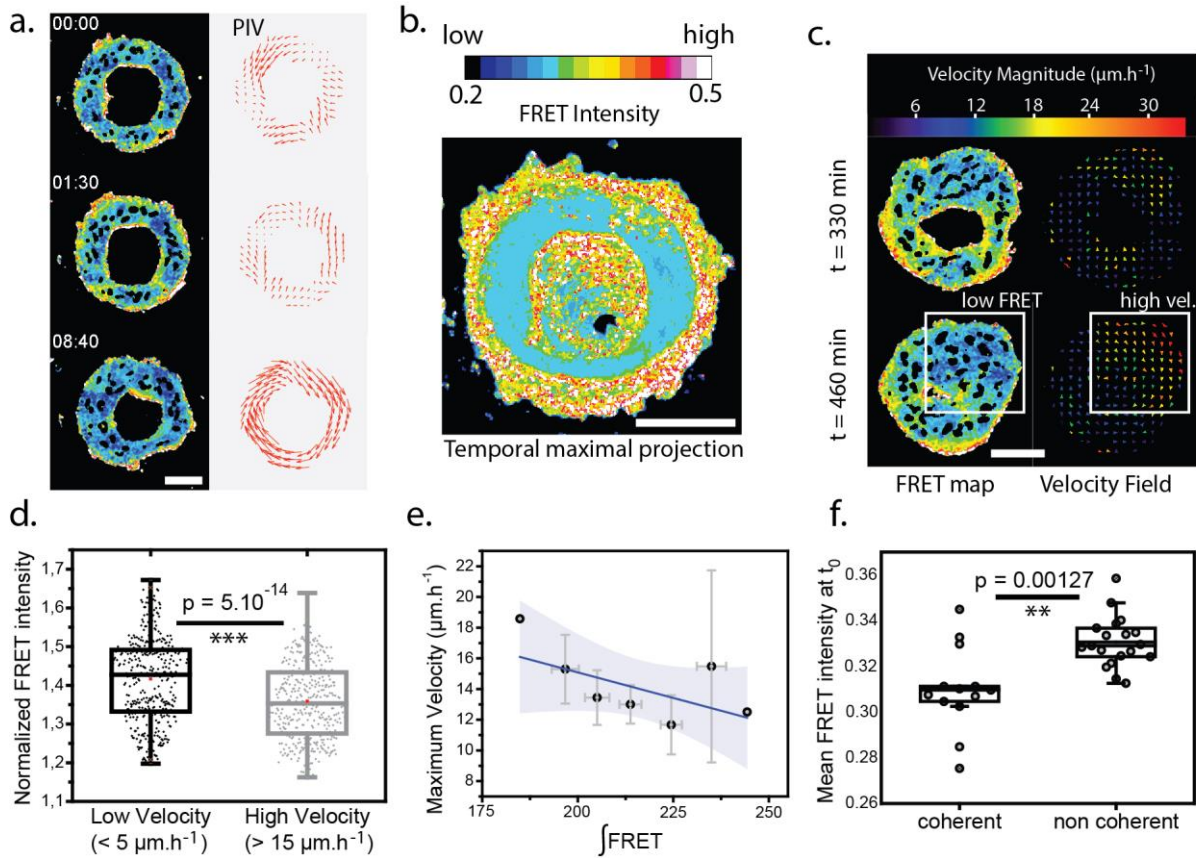


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2 **Figure 2. Cell polarity and density determine dynamics of coherent motion.** (a)
3 80µm “mosaic” ring. Arrows indicate cell polarity based on the direction/orientation of
4 cell lamellipodium. Scale bar = 50µm. (b) Distribution of cell initial orientations within
5 the rings. 0° (counter clockwise) and 180° (clockwise) correspond to tangential
6 orientations. $n_{\text{rings}} = 12$, $n_{\text{cells}} = 42$. (c) Segmentation of multicellular rings based on ZO1
7 distributions. Single cell parameters such as cell elongation (stretch) can be extracted,
8 red and blue largest and lowest stretch respectively. Scale bar = 50µm. (d) Percentage
9 of aligned cells based on the polarity given by ZO1 distributions. Data are binned with
10 respect to the time of coherence onset. Data are represented as Mean \pm SD. N = 10.
11 (e) Average normalized stretch (initial/max) aligned along a new pseudo-time. The
12 dashed blue line corresponds to coherence acquisition. Data are represented as Mean
13 \pm SD. N = 10. (f) Division of an actin-GFP labeled cell within a 80µm ring. Cell division
14 generates daughter cells with opposite polarities p_1 and p_2 . Scale bar = 20µm. (g)
15 Cumulative cell divisions as a function of time. Coherent rings exhibit bursts of divisions
16 with large periods without divisions whereas non-coherent rings undergo regular
17 divisions throughout the course of the experiment. For clarity, only 5 representative
18 rings are shown for each case. Full population is available in Fig. S2A.
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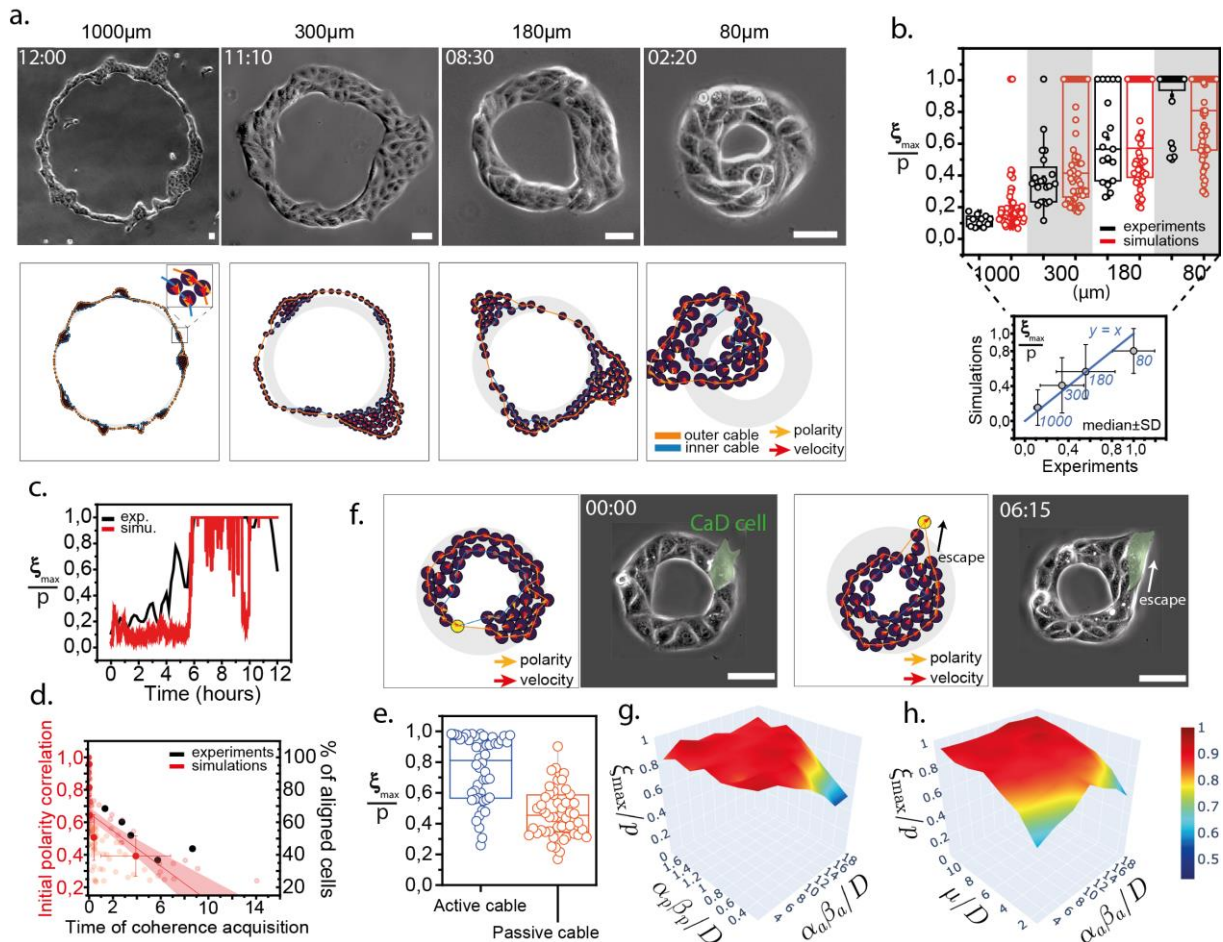
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Figure 3. Acto-myosin cables as internally-driven constraints. (a) Immunostaining of a 80µm ring. Myosin and actin are visualized. (b) Tracking of the ring boundaries. The color code indicates time. The ring first closes inwards before extending outwards. (c) Kymographs showing acto-myosin ring retraction after laser ablation and cell-cell junction opening (as a control) after ablation. (d) Retraction velocity as a function of time after laser ablation. Exponential fits give estimates of v_0 defining the relative tension stored in the ring. (e) Actin visualization (lifeAct) of a transfected cell within a 80µm ring. The acto-myosin cable is self-generated by the cell during motion and constrains the polarity in a tangential orientation. White arrow indicates the cable, green contour underlines the lamellipodium boundary. Scale bar = 20µm. (f) Time-lapse showing a CaD transfected cell (white arrow) escaping the multicellular ring and challenging coherence. Scale bar = 50µm. (g) Coherence acquisition for non-transfected ring and rings with CaD transfected cells. There is a significant decrease in coherence ($p = 7.10^{-4}$) in the CaD rings.



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Figure 4. RhoA activity correlates with different ring behaviors. (a) RhoA activity on a $80\mu\text{m}$ ring as described by the FRET ratiometric map (left) associated to a velocity field (right). Scale bar = $50\mu\text{m}$. (b) Temporal maximal projection showing highest FRET signals at ring boundaries. Scale bar = $50\mu\text{m}$. (c) FRET map for each frame is represented with its associated velocity field. Scale bar = $50\mu\text{m}$. Comparison is shown in (d) where high FRET regions correlate with low velocity. (e) Maximum velocity of the ring (with both radial and tangential components) is represented as a function of the averaged FRET level integrated over the entire experiment. (f) FRET intensity at t_0 averaged over the cellular “bulk” excluding cables. Initial levels of RhoA activity are plotted for different coherent behaviors. High activity is correlated with low coherence. Data in (e) are binned; full distributions are shown in Fig. S4h. Same color codes for (a), (b) and (c) for FRET signals.



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Figure 5. Theoretical modelling of multicellular rings. Experiments are compared with simulations. (a) Rings of different diameters are represented (top) when typical motifs appear, fingering for large diameters, clusters for intermediate rings, closure for smaller rings. Morphology is reproduced by numerical simulations (bottom). Scale bar = 50 μm. Time in hh:mm; simulations times (in hh:mm, from left to right): 05:24; 17:56; 04:00; 04:57. (b) Coherence index as a function of ring diameters for simulations and experiments, with an inset comparing median values. (c) Dynamics of coherence acquisition *in silico* compared with a typical MDCK ring. (d) Initial polarity correlation as a function of the time of coherence acquisition. The decay observed in experiments is similar to simulated rings. (e) Coherence index for two different populations of MDCK rings: *activity* within cable facilitates acquisition of coherence. (f) Caldesmon experiment and cellular ‘escape’ are reproduced with simulations. The yellow particle represents the CaD cell and its cable parameters are set to zero. Scale bar = 50 μm. Time in hh:mm. Simulation times (in hh:mm, from top to bottom): 05:20; 13:03. (g-h) *Phase diagrams for coherence with respect to the dimensionless parameters associated with the model (active cable force - active alignment, active cable force - passive cable force). Index p refers to the passive component of the cable while a corresponds to the active one. On each plot, each point of the surface corresponds to the average value of 50 simulations.*

