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Title: Soft, skin-interfaced sweat stickers for cystic fibrosis diagnosis and management

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One Sentence Summary: A soft, wearable microfluidic platform collects and quantitatively analyzes sweat biomarkers to improve the diagnosis of cystic fibrosis.

Abstract: The concentration of chloride in sweat remains the most robust biomarker for confirmatory diagnosis of cystic fibrosis (CF), a common life-shortening genetic disorder. Early diagnosis via quantitative assessment of sweat chloride allows prompt initiation of care and is critically important to extend life expectancy and improve quality of life. The collection and analysis of sweat using conventional wrist-strapped devices and iontophoresis can be cumbersome, particularly for infants with fragile skin, who often have insufficient sweat production. Here, we introduce a soft, epidermal microfluidic device (“sweat sticker”) designed for the simple and rapid collection and analysis of sweat. Intimate, conformal coupling with the skin supports nearly perfect efficiency in sweat collection without leakage. Real-time image analysis of chloride reagents allows for quantitative assessment of chloride concentrations using a smartphone camera, without requiring extraction of sweat or external analysis. Clinical validation studies involving patients with CF and healthy subjects, across a spectrum of age groups, support clinical equivalence compared to existing device platforms in terms of accuracy and demonstrate meaningful reductions in rates of leakage. The wearable microfluidic technologies and smartphone-based analytics reported here establish the foundation for diagnosis of CF outside of clinical settings.

Introduction

Cystic fibrosis (CF) is among the most common life-shortening genetic disorders, affecting 1 out of every 3,300 births in the United States and 70,000 people worldwide(1, 2). CF arises from mutations to the cystic fibrosis transmembrane conductance regulator (*CFTR*), an essential regulatory epithelial chloride channel in the sweat glands, lungs, intestines, pancreas, and kidneys(3). With over 2000 *CFTR* mutations identified to date(2), individuals with CF exhibit wide variability in the presentation and severity of clinical symptoms, such as chronic pulmonary infection, progressive loss of lung function, and pancreatic insufficiency(2, 4, 5). Clinical treatment involves complex, symptomatic therapy plans based in centers accredited by the Cystic Fibrosis Foundation (CFF) (6). Treatment efficacy depends primarily upon the age and mode of diagnosis as well as the overall quality of care(1, 4, 5). Although recently developed molecular therapeutics that target the basic *CFTR* defect have substantial potential(7), current pharmacological agents can only alter rather than reverse disease progression(8–12). As such, early diagnosis and management of CF progression continues to be critically important for patients with CF (5, 12).

Initial diagnosis in the United States and other developed countries is based on newborn screening using dried blood spots. This typically involves measurement of immunoreactive trypsinogen (IRT), a pancreatic protein that is elevated in newborns with CF, followed by *CFTR* mutation testing in infants with elevated IRT(2, 5, 6). This test captures almost all infants with CF, but it suffers from several limitations including a high false positive rate(2, 13); an inability to ascertain rare *CFTR* mutations, deletions, and duplications(6); and, in the case of DNA-based mutation testing, poor sensitivity in detecting mutations in infants from ethnic and racial minority groups(14). These limitations necessitate the use of quantitative sweat testing to confirm a CF diagnosis even in cases where only 1 *CFTR* mutation is found (often indicating a heterozygote

carrier); in some regions, a very high IRT concentration is considered a positive screening test. Elevated sweat chloride is a robust indicator of the loss of *CFTR* function that is independent of the type of mutation (15) and remains the gold standard for diagnosis of CF. As a result, quantitative evaluation of chloride in sweat represents the only method for testing *CFTR* function that is widely available in accredited care centers, as approved by the US Food and Drug Administration (FDA) and other regulatory agencies(16) and confirmed by recent CFF guidelines(17).

Developed almost 60 years ago(5), testing involves stimulation of sweat glands by the cholinergic agonist pilocarpine, passed transdermally by iontophoresis. There are two accepted approaches for sweat collection: (i) the traditional Gibson-Cooke (GC) method, introduced in 1959, that uses gauze pads (18) and (ii) a more modern technique, introduced in 1986, that relies on coiled tubes in plastic housings (Macroduct; MSCS, ELITech Group) (19). Both schemes use pilocarpine iontophoresis to stimulate sweat production followed by a 30-minute collection period. A coulometric chloridometer enables quantitative analysis of the collected samples. In practice, the collection of sufficient volumes of sweat for analysis can be challenging—especially in infants(20–25). Clinical and Laboratory Standards Institute (CLSI) guidelines specify the collection of at least 15 μL of sweat within a 30-minute time period for accurate analysis using the MSCS(6). An insufficient sweat volume is reported as “quantity not sufficient” (QNS). A recent survey of certified CF centers indicates difficulty in achieving a QNS rate of <10% as stipulated by CFF guidelines for infants in the first three months of life. Some reported QNS rates reach values of 40%, with poor mechanical sealing of the MSCS to the skin and cumbersome extraction protocol being primary contributing factors (2, 5, 20, 22, 25, 26).

Despite systematic quality improvement efforts and modest center-specific decreases in QNS instances, insufficient sweat collection remains the most common issue in cystic fibrosis

diagnostics(22). Promising diagnostics use alternative analysis approaches(27) and form factors(28) to reduce the required volume of sweat. Nevertheless, high failure rates(29) and poor performance in clinical trials(30) prevent widespread adoption. The additional requirement of laboratory-scale analytics tools and controlled environments for evaluation further limit their utility in remote settings or in the home(27, 31–33). A system for simple and rapid sweat collection and chloride analysis, preferably within the collection device itself, in a manner that bypasses these drawbacks represents a foundational unmet need for the CF community.

Emerging classes of bio-integrated sensors(34), a type of wearable technology characterized by the noninvasive, intimate integration with the curved surfaces of the body, offer the potential to surmount these limitations in sweat testing. A recent demonstration(35) of a wearable electrochemical sensor provided localized sweat stimulation and electrochemical analysis via a single battery-powered platform for real-time, in situ measurement of sweat chloride. Other examples rely on commercial iontophoresis systems with instrumented collection platforms that minimize analytical sweat volume requirements through alternative quantification methods such as those based on conductivity(36), potentiometry(37, 38) and/or fluorimetry(39, 40). Although successful in minimizing sweat collection requirements, these platforms rely on complex analytical techniques not accepted by established clinical guidelines(6). Challenges that arise from various calibration procedures, extended sensor stabilization times, individual sensor variability, and need for complex battery-powered electronic circuitry(33, 35, 38) represent fundamental constraints. Recent work offers a path to circumvent these limitations in devices that leverage networks of embedded microfluidic channels in soft, elastomeric substrates for the passive capture, storage, and quantitative analysis of sweat(41–46). Demonstrations of these epidermal microfluidic devices (“epifluidic” devices) utilize the natural pressure generated by the sweat glands(47) to guide the flow of sweat through sophisticated channel geometries(48–50),

colorimetric reagents(49, 51), and wireless (e.g. battery-free) integrated sensors for real-time(48, 52) and time-sequenced analysis(51) of sweat biomarkers in a variety of environments without risk of evaporation(53, 54).

Here we introduce a soft, wearable device that collects and analyzes sweat via an intimate skin-compatible microfluidic construct that operates via pilocarpine iontophoresis. This epifluidic device (“sweat sticker”) uses a newborn-safe adhesive as a skin interface and an optimized multi-layer geometry to gently but tightly seal to the epidermis, supporting near-perfect efficiency in sweat collection without leakage. Real-time image analysis for colorimetric chloride-based chemical reagents allows for quantitative readout of chloride concentrations directly on the sticker, using digital image capture with a smartphone. Validation studies involved patients with CF and healthy control subjects of varying ages, with direct comparisons to sweat collection performed using the MSCS method and to chloride analysis using coulometric chloridometers. Our results support clinical equivalency in accuracy and demonstrate substantial reductions in rates of QNS. These technologies establish the foundations for a low-cost wearable platform for diagnosis of CF in patients.

Results

A representative device consists of a sealed network of microfluidic channels and reservoirs for sweat collection and chemical analysis in an overall circular geometry, designed to facilitate highly efficient capture of sweat over a region of the skin stimulated by pilocarpine iontophoresis (Fig. 1A). The soft mechanical properties, the thin (~1 mm), flexible layout and the small size of the device facilitate attachment onto various body locations (e.g. forearm, thigh) for both infants and adults. The construction involves three layers of low modulus (1-2 MPa) silicone elastomers

(polydimethylsiloxane, PDMS) that define the microchannel structures. The physical properties enable a zero-pressure, fluid-tight interface to the skin via a medical-grade, irritation-free, FDA-cleared gentle skin adhesive. Laser-patterned openings define inlets in the skin-facing side of the device to allow sweat to enter the system, driven by the natural pressures created by the stimulated glands (~ 3 kPa). To optimize sweat collection, the device incorporates patterned adhesives that allow sweat capture in the collection area while maintaining robust adhesion. The collection surface area is identical to the stimulation area (28 mm diameter region, defined by the dimensions of the iontophoresis electrodes) and corresponds to ~ 640 sweat glands at 104 glands per cm^2 (55). As it enters the device, the sweat passes into three independent chambers defined by serpentine channels (Fig. 1B). Each chamber has a capacity of 47 μL . An embedded chloride-free, food-safe dye (located near the inlet) aids in visualization of sweat ingress during collection. The “sweat sticker” nomenclature and inclusion of child-friendly thin-film graphical overlay (250- μm thickness, Fig. 1, A and B) represent design choices intended to minimize anxiety in pediatric patients. Motivated by the CARE principles (Choices, Agenda, Resilience, Emotional support) (56), this user-friendly strategy mitigates uncertainty and offers comfort during sweat testing by reframing a “sweat collection device” into a familiar and safe concept (*i.e.*, a sticker). The MSCS serves as a comparative reference device for sweat collection (Fig. 1C), where application to the skin involves a band or disposable tourniquet to apply sufficient pressure to form an adequate seal between the skin and the rigid, plastic housing that encloses the coiled collection tube. Figure 1D illustrates the operation typically used with MSCS devices during study protocols. Examples of MSCS failure modes appear in fig. S1.

A key design consideration for sweat-monitoring devices is the efficacy and ease of sweat extraction to facilitate measurement by the guideline-specified process. Sweat must be fully extracted from the device and the extraction mechanics should promote ease of operation while

eliminating sources of contamination. At the conclusion of a traditional sweat test, the technician must perform a multistage, high dexterity procedure (Fig. 1E) to extract collected sweat from the MSCS while the device remains attached to the subject and transfer the sample to a container for chloride quantification. In contrast, the sweat sticker can be removed from the subject's arm to facilitate extraction (Fig. 1F). The central location of the collection inlets (i.e. openings to skin) are easy to access upon sticker removal, to minimize sweat leakage. A standard pipette (1 mL) enables sweat extraction from the device. The device-layer inlet size (1.2-mm diameter) is smaller than the pipette opening (1.5 ± 0.1 mm diameter), thereby allowing for a strong, temporary water-tight and air-tight seal for application of negative pressure sufficient to fully extract sweat from each chamber, independent of filled volume. The sweat extraction rate depends linearly on the magnitude of negative pressure applied (set volume) and rate of draw on the pipette. Extracted sweat from all three chambers yields a total sweat volume approaching 140 μ L.

Initial sweat sticker design optimizations yielded a multi-layer construct (Fig. 1, A, B, and D) to: (i) increase sweat storage volume, (ii) minimize volume changes during mechanical deformation, and (iii) eliminate puncture risk during sweat extraction. A potential sweat test failure mode could occur in overfilling the collection device, resulting in sample contamination. Clinical guidelines specify the collection area, which restricts the overall size and collection volume of a single-layer device. An increase in device thickness accommodates greater collection of volume (proportional to channel depth). The addition of a tiered, multi-layer design preserves collection volume and the conformal skin interface (Fig. 2). This design reinforces the sticker, and thereby reduces risk of overfilling or ejecting collected sweat from the inlets during removal from skin (movie S1) and puncture of the device with a pipette tip during sweat extraction.

The soft, flexible mechanical properties of the device (Fig. 2A) provide gentle, conformal coupling to the skin of neonatal and pediatric patients. Phantom arm assemblies (Fig. 2B) similar in size to

the arms of average adults and infants demonstrated the dynamics of sweat ingress as a function of bending radius of the device. Microbore tubes (1.2-mm internal diameter, ID) integrated within the phantoms connect to pumps that simulate sweat production. Measurements of sweat rate are important to ensure adherence to established testing protocols (minimum of 1 g/m²/min; equivalent to 0.5 μL/min for MSCS)(57). The geometries and layouts of the microchannels must not restrict fluid transport phenomena across physiological ranges of sweat release. Replicate data (N = 3) for fluid volume versus time (normalized by time to completion) for devices at three different radii of curvature are shown in Fig. 2C. Measurements of flow at a constant pressure (~2 kPa, comparable to that produced by eccrine glands)(47) maintained by a pressure displacement pump exhibited negligible variation associated with microchannel geometries explored here (Fig. 2D), consistent with constant flow rates and absence of fluidic resistance. This observation applies both for flows induced by pressure displacement (Fig. 2C) and syringe-based pumps (fig. S2).

Clinical sweat testing with the sweat sticker

A clinical pilot study in patients with CF and healthy controls conducted at the Cystic Fibrosis Center at the Ann & Robert H. Lurie's Children's Hospital (Chicago, IL) demonstrated the clinical utility of this platform as a sweat collection and analytics system. Table 1 summarizes the participant demographics and sample characteristics of the study cohort (N = 51). Bilateral sweat testing enabled comparison of collected sweat volume by the sweat sticker and the clinically established MSCS. In both cases, a pilocarpine iontophoresis process initiated the sweat response. Figure 3A shows the average sweat volume and QNS rate for the sweat sticker and the MSCS, with additional data from four recent large cohort studies. The mean sweat volume collected by the sweat sticker was 74 ± 35 μL, whereas the mean MSCS sweat volume was 58 ± 24 μL, consistent with previous reports(16, 20, 58, 59), with age-stratified individual volumes shown in fig. S3. The improvement in sweat volume collection with the sweat sticker as compared to the

MSCS was statistically significant (Wilcoxon test, $W = 1603.5$, $P = 0.02$), with the matched-pair average approaching 33% for sweat volume ($P = 0.05$). Three instances of QNS occurred with the MSCS tests (5.8%); in contrast, no such instances appeared with the sweat sticker (0%) in this study. Furthermore, the MSCS QNS rate is below estimates reported in previous studies (16, 20, 58, 60) and comparable to the routine QNS rate of this clinical site (5%) (21). Figure S4 reports sweat volume from ten consecutive bilateral sweat tests (MSCS on both arms) performed by trained staff during routine clinical CF screening, as another point of comparison. Four collection events resulted in QNS.

A calibrated chloridometer determined the chloride concentrations of the collected samples in both cases. The equivalence chart in Fig. 3B shows excellent correlation (Pearson's $r = 0.99$; $N=51$) between the paired bilateral tests for both population groups. A Bland-Altman plot in Fig. 3C supports this result, indicating strong correlation for sweat chloride values < 80 mmol/L. The increase in variability above 80 mmol/L likely follows from the increase in the coefficient of variation of the commercial chloridometer at these elevated chloride concentrations (1.02% at 100 mM).

For age stratification analysis, the study cohort was classified into three groups: (i) infants (0 – 6m), (ii) pediatric (7 m – 18 y), and (iii) adult (+18 y). The sweat sticker showed enhanced performance in collected sweat volume across all age groups as compared to the MSCS (Fig. 3D). Wilcoxon analysis of the adult age group showed a significant increase in collected volume by the sweat sticker ($W = 151$, $P = 0.03$). QNS events occurred in both the pediatric and adult groups for the MSCS, but not the sweat sticker. Age-stratified Bland-Altman plots further support the superior performance of the sweat sticker (fig. S5).

Figure 4 offers representative examples of the sweat stickers and MSCS devices worn (observed at the conclusion of testing) by pediatric (Fig. 4A) and infant (Fig. 4D) participants. Fundamental

design improvements evident in the sweat sticker as compared to the MSCS include method for device attachment (adhesive Fig. 4B; tourniquet, Fig. 4C), sweat visualization, patient comfort (fig. S6), and child-friendly aesthetics. Fig. 4D illustrates the conformal sweat sticker interface.

Multimodal sweat sticker with integrated chloride analysis

A configuration of the sweat sticker (similar in form to Fig. 1A) that includes an integrated colorimetric chloride assay (Fig. 5A) allows for in situ analysis, without need for sweat extraction or measurement with external instruments. In contrast to conventional electrochemical methods, use of a colorimetric approach enables in situ sweat chloride analysis with a completely passive, battery-free form factor. The reagent silver chloranilate, inserted into the microchannel structure of the device, reacts with the chloride in sweat to produce a color response that is proportional to chloride concentration (Fig. 5B). The device in this case includes a thin (100 μm) transparent film of a specified color to enhance the signal-to-noise ratio of colorimetric readout of chloride concentration. The film contains three 1-mm diameter regions, each with an opaque PANTONE color swatch pattern as a color balance reference. Assay imaging with a digital single-lens reflex camera (dSLR) camera enables facile conversion from standard Red Green Blue color space (sRGB) to CIE (International Commission on Illumination) LAB color space for color extraction and image processing, as shown in fig. S7. CIELAB color space expresses color in three components: lightness (L) from black (0) to white (100); the amount of green to red (a); and the amount of blue to yellow (b).

Investigations of blue, red, and green colors helped to identify an effective color for use as an overlay. A clear overlay served as a reference. Fig. 5C displays the color shift that results from the addition of a color overlay as a function of CIELAB a^* and b^* color values. Each value corresponds to the average of the evolved color of a reaction of silver chloranilate with solutions of known sodium chloride concentrations ($N = 3$ independent solutions). Fig. S8 shows the color

variation for one solution at the individual pixel level within the region of interest (ROI) that comprises one of the three measurements. Converting a^* and b^* color values to chroma, defined as $C^* = (a^{*2} + b^{*2})^{1/2}$, yields color-overlay specific calibration curves by a power-law relation (Fig. 5D). Sampling colors in a device-independent color space, such as CIELAB color space, provides a facile method for color comparison. Fig. S9 further highlights this device independence as analysis of iPhone and dSLR images yielded equivalent measurement values.

Three commercial clinical sweat chloride standards (ELITech Group) corresponding to the prescribed CF diagnostic ranges served as analytical benchmarks (low standard: 23 mM; intermediate standard: 49 mM, high standard: 100 mM). Difference plots for each standard (Fig. 5E) show the analytical performance for each color overlay and the chloridometer with the externally defined valid measurement range (red dashed lines). Results also compare the performance of the color-analysis proposed here (calibration fit using C^*) with a method used in previous work (calibration fit using a^*). Many colorimetric assays (such as silver chloranilate) contain a “white point measurement” to indicate the absence of an analyte (i.e. 0 mM chloride). The utilization of white in a colorimetric assay is problematic when attempting to distinguish between subtle variations in color and thus analyte concentration as white is defined as $L^* = 100$, $a^* = 0$, $b^* = 0$. This effect propagates uncertainty into colorimetric assay analysis at low concentrations. As Fig. 5E highlights, the results show that the green overlay minimizes the difference in measured chloride values with respect to the chloridometer. All methods lack agreement with the expected value for the low standard reference solution; however, the green overlay curves and a^* clear overlay curve are within accepted values of the chloridometer. The C^* fit offers the best agreement to the reported chloridometer values across the range of measured chloride concentrations.

A human subject pilot study comprising patients with CF (N = 2) and healthy subjects (N = 3) explored the clinical relevance of this approach to in situ sweat chloride measurements. Each chamber (clear overlay, green overlay, chloridometer control) of the sweat sticker captured an independent sweat sample for analysis upon completion of sweat collection (Fig. 6A). C^* -based calibration curves converted color values to chloride concentrations. The values determined with the green overlay agreed closely (within 1 SD) with chloridometer measurements for all members of the study cohort (Fig. 6B). The values measured in the absence of a color overlay were similar for study member P5 but deviated from chloridometer values for subjects with elevated sweat chloride concentrations. These findings suggest that the color analysis approach introduced here can enhance the measurement accuracy of a colorimetric chloride assay to yield values comparable to established clinical approaches.

Discussion

The results presented here demonstrate that sweat sticker devices (N = 51) and associated approaches for colorimetric chloride analysis (N = 5) can address clinical requirements, with advantages over the current standard of care by potentially reducing the QNS rate, improving the comfort and experience for the patient, and enabling accurate measurements outside of hospital or clinical laboratory settings. These outcomes rely on optimized microfluidic designs, soft device mechanics, gentle adhesive interfaces, and advanced image processing methods. By establishing conformal, water-tight interfaces to the skin, the device offers: (i) an increase in collection volume compared to MSCS, for otherwise similar conditions; (ii) improved resistance to motion-induced collection failures; and (iii) facile processes for mounting the devices on the skin and extracting samples of sweat from them. Our clinical studies demonstrated that these engineering features led to improved outcomes, compared to those possible with the MSCS platform. For example, the

sweat sticker successfully addresses insufficient sweat by enhancing collected sweat volume via the certified stimulation methods used in CF diagnostic testing centers. In all testing, both for patients with CF and healthy subjects, the sweat sticker adhered robustly and maintained conformal attachment without sweat leakage. Furthermore, the zero-pressure skin adhesive interface avoided transient deep epidermal indentations that occur with the high-pressure interface utilized by the MSCS. The additional ease of use and handling for the clinical staff further minimizes common pitfalls of contamination and insufficient volumetric extraction observed in post-test sample processing. This clinical demonstration of sufficient sweat collection establishes the foundation for an integrated collection and chloride analysis platform.

Integrated colorimetric analysis

The ability to measure chloride concentrations in situ during and immediately after collection represents a powerful feature of the technology that has the potential to allow diagnostic CF screening outside of approved CF centers. The consequences are wide ranging, especially in the context of the recent emergence of *CFTR* modulator therapeutics for which sweat chloride could serve as a biomarker for assessing and monitoring treatment efficacy(57, 61–63). This potential appears here as study members with CF and sweat chloride concentrations below 100 mM were on *CFTR* modulators at the time of the tests. Sweat stickers offer facile, passive methods for assessing sweat chloride values on a regular, routine basis, thereby advancing the utility of sweat chloride measurements from a clinical diagnostic to a mechanism for frequent monitoring of disease progression.

The capture and analysis of sweat chloride concentrations via a smartphone, with sufficiently high accuracy, represents a key advance over previous work(41, 43, 44, 46, 49, 51–53). When coupled with enhanced color analysis methods reported here, colorimetric chemical reagents offer performance comparable to that of commercial chloridometers.

While the analytical range of the chloride bioassay is suitable across the key diagnostic windows for CF (< 30 mM, CF unlikely; 30 – 60 mM, intermediate; 60 mM, CF likely), previous work(49) lacked the accuracy and precision at low concentrations necessary for clinical utility. The evolution of the dark violet color of the assay (with increasing chloride concentration) is stark at high sweat chloride concentrations; however, resolving slight variations, especially at lower concentrations, presents difficulties via optical imaging. In the context of CF, eliminating this deficiency is critically important to ensure accurate diagnosis and eliminate the profound consequences of a false positive or negative test. Our image processing strategy improves colorimetric assay accuracy by eliminating the white point by hue shift through a prescribed color overlay with a contrasting color. The absence of a color overlay introduces measurement error into the calibration chart yielding magnified inaccuracies in reported sweat chloride values, typically at either low or high concentrations, as observed for the three standard controls. Optimization of the overlay color elongates the relevant detection range of the assay to increase the range of distinguishable color measurements.

Although several methods exist to relate measured differences in color in CIELAB space to assay concentration(49), C^* offers a single value based upon both measured a^* and b^* color values suitable for use in a calibration curve. In combination with a robust color correction process and a hue-shift analytical approach, this method achieves an enhanced degree of accuracy in quantifying chloride concentrations across the physiological range of interest, with performance comparable to established clinical methods (chloridometer).

Study limitations

Although this study demonstrates substantial improvements in volume of sweat collected by a statistically significant margin in adult populations, the sample sizes for younger age groups could be expanded in follow-up investigations. These studies could also increase the range of CF

conditions, birth weights and environmental factors (e.g. hydration state). These latter effects may contribute to high inter-subject variability in the amount of sweat volume captured, as demonstrated by the large standard deviations (MSCS SD = 24.15 μ L, sweat sticker SD = 35.19 μ L) and consistent with expectation based previous studies (16, 58, 64–68). The variability in sweat volume likely contributes to the lack of differences in the age-stratified analysis for the childhood and adolescent groups, thereby further emphasizing the need for larger subsequent studies.

Pathway to clinical validity for colorimetric analysis with smartphone

The CLSI guidelines specify coulometric titration with a chloridometer as the only recommended chloride measurement method for CF diagnostics(69). Specific concerns regarding the absence of systematic validation of ion-selective performance and decreased sensitivity at lower chloride concentrations could compromise measurement accuracy and precision. An alternative sweat chloride and collection quantification method using the sweat sticker and digital image analysis thus requires rigorous verification and validation, and approval by a regulatory agency.

This feasibility study and the clinical equivalence of the colorimetric sweat sticker supports further investigation to establish full clinical validity. Although the accuracy of the sweat sticker matches the laboratory-grade diagnostic technique in this study, additional studies in uncontrolled environments will be needed for real-world deployment and data capture. Physical variations in device color overlay or non-uniform lighting (shadows) can adversely affect calculated fits, which in turn, could affect accuracy. Risk mitigation strategies include inclusion of color reference markers, utilization of a custom smartphone application for imaging and analysis, and employing rigorous manufacturing quality control protocols.

Although this sweat chloride assay has been validated in athletic performance applications through comparisons to high pressure liquid chromatography (34, 43, 49, 53), careful validation in a large clinical trial could provide statistical evidence in support of analytical equivalence for patients with CF. Specifically, further studies must demonstrate (i) assay accuracy, (ii) assay precision, (iii) measurement interval, and (iv) limit of quantitation per the CLSI evaluation protocol(30).

The emergence of *CFTR* modulator therapeutics represents an inflection point for sweat chloride measurements in CF patient care. Sweat chloride tracking has transitioned from a biomarker exclusive to initial screening and diagnostics to a long-term reference in a manner similar to blood glucose for diabetic patients. This raises the critical need for low-cost, quantitative sweat chloride wearable sensors suitable for real-time and daily assessment of CF biomarkers.

Utilization for diagnostics and disease management

The requirements imposed by calibrated analytical equipment, access to centralized laboratory facilities, and need for trained technicians restrict the use of conventional MSCS diagnostic testing to CFF accredited clinics. This can result in considerable hardship as many rural locations and some U.S. states lack access to such centers. In contrast, the results presented here favor rapid and decentralized modes of deployment that could increase clinical diagnostic access and expand the breadth of sweat testing utility. As fig. S10 illustrates, these modes can be classified according to three categories: (i) diagnostics, (ii) screening, and (iii) daily management of CF. The ease of use of the microfluidic platform supports deployment in both clinical and free-living (home) environments. For clinical applications, the device could support collection alone (equivalent function to MSCS) as well as integrated in situ chloride quantification (equivalent function to MSCS and chloridometer), using traditional techniques for sweat stimulation, pending clinical trials to validate the efficacy and utility compared to these predicate devices (30). Extending these applications to at-home sweat chloride monitoring without requiring pilocarpine iontophoresis

could provide insights in CF drug efficacy and patient quality of life. As shown in recent work, thermal exposure in a warm bath or shower is sufficient to induce eccrine sweat for purposes of monitoring biomarkers for chronic kidney disease(51). A similar sweat induction mechanism has the potential to eliminate the need for pilocarpine iontophoresis. A key consideration is establishing sweat composition and rate equivalence between cholinergic and thermal stimulation. Initial work(51) suggests a warm shower supports sufficient sweat rates (10-min shower: 2 $\mu\text{L}/\text{min}$ sweat rate) exceeding the ~ 0.5 $\mu\text{L}/\text{min}$ minimum sweat rate specified for the clinical CF sweat test. If validated, such an approach could enable the broad use of the sweat sticker and smartphone application as a daily or weekly management tool for clinical trials for drugs and disease management strategies. Estimates suggest per device costs, when produced at scale, of less than \$5 USD, thereby allowing for single-use strategies with potential for considerable savings relative to costs of clinical sweat tests [ranging from \$200 to \$250 USD at most CF centers(70)].

Implications for diagnosis and management of CF

High QNS rates in neonates with positive CF newborn screens motivates the work reported here. Minimizing the QNS rate while enabling remote tracking of sweat chloride will have strong positive effects on the management of CF, as a delay in CF diagnosis and management may have profound negative consequences on the pulmonary function and the quality of life of patients(23). The sweat sticker and image processing technologies introduced here have the potential to reduce QNS rates while enabling CF management at home, with implications across neonatal, pediatric, and adult patients.

Materials and Methods

Study design

The purpose of this study was to develop and demonstrate a soft, epidermal microfluidic device for use in collecting and analyze sweat for the diagnosis of cystic fibrosis. Patients were recruited from CF populations and from adult volunteers (control). The study was IRB-approved through the Ann & Robert H. Lurie's Children's Hospital (IRB #2018-1440). Briefly, patients with CF and healthy subjects were recruited (at random) from the Cystic Fibrosis Center at Lurie Children's Hospital and through Northwestern University, respectively (time frame:1 year, number of subjects: 51, study power: 0.8078). Informed consent was obtained after the nature and possible consequences of the study was explained. The sweat tests were administered by trained personnel at Northwestern University and Lurie Children's Hospital. Patient sweat tests followed established clinical protocols for sweat capture and analysis(71) using the Wescor Sweat Stimulator and Macroduct (MSCS, ELITech Group) as detailed in the materials and methods section of the Supplemental Materials. The pilot study (N=5) was conducted as part of the primary study whereby an additional cohort of adults were recruited (at random) from the same population to evaluate the performance of a sweat sticker with an integrated colorimetric chloride assay. Data collected in this pilot study is independently reported from the primary study (N = 51).

Sweat stimulation and analysis

A single measurement consisted of sweat stimulation via pilocarpine iontophoresis (~5 min, forearm) followed by sweat collection (~30 min), using either MSCS or the microfluidic sweat sticker. An alignment guide allowed for consistent placement of iontophoresis electrodes across different patients. A patient evaluation session involved two sweat measurements performed sequentially, one for each arm (left, then right), with the device and arm designation determined *a priori*. Patients were instructed to refrain from arm movement during MSCS sweat collection, as prescribed by clinic protocol, and instructed to move freely during the sweat sticker collection period. Sweat was extracted from the sweat sticker and MSCS at the conclusion of the collection

period. Chloride measurements were then performed using a clinically certified ChloroCheck Chloridometer (ELITech Group, with a coefficient of variation of 1.02% at 100 mM). Details of the study population are provided in table S1 with individual patient data in table S2. Performance of the pilot study followed the same procedure. A photograph of the sweat sticker was recorded at the conclusion of the collection period prior to removal from the arm to enable quantitative analysis of the colorimetric chloride assay.

Microfabrication of microfluidic molds for layer-specific microchannel networks

Ultraviolet light exposure (420 mJ cm^{-2} , Heidelberg MLA150) of a $10 \text{ }\mu\text{m}$ -thick layer of negative resist (KMPR 1010, spin cast at 3000 RPM for 30 s, soft baked at $110 \text{ }^\circ\text{C}$ for 5 min, cooled to ambient at $15 \text{ }^\circ\text{C min}^{-1}$) on a silicon wafer (1.5 mm thickness, 100 mm diameter) followed by a soft bake ($110 \text{ }^\circ\text{C}$ for 5 min, cooled to ambient at $15 \text{ }^\circ\text{C min}^{-1}$) and development (AZ 300MIF, 7 min) process formed the patterns for each mold. A deep reactive ion etching (DRIE) process (Plasma-Therm Versaline DSE, Plasma-Therm, Inc.) removed the exposed regions of the silicon wafer to a controlled depth (layer 1 mold: $400 \text{ }\mu\text{m}$, layer 2 mold: $300 \text{ }\mu\text{m}$). A spin cast thin film of polymethyl methacrylate (495 PMMA C6, MicroChem Corp., 4000 RPM, 5 min bake at $110 \text{ }^\circ\text{C}$) served as a release layer to facilitate production of channel replicas in silicone via soft lithography.

Microfabrication of sweat stickers

The sweat sticker platform comprises three layers of micro-molded silicone (polydimethylsiloxane, PDMS, Dow, Inc.). Spin coating a layer of 20:1 PDMS (base to curing agent, Sylgard 184, Dow, Inc.), mixed in a planetary mixer (Thinky ARE-310; 30 s mix, 30 s defoam), on molds for layers 1 (200 RPM, 30 s) and 2 (350 RPM, 30 s) followed by flash curing for 5 min at $120 \text{ }^\circ\text{C}$ formed solid channel replicas. The addition of 3 wt% white silicone pigment (Silc Pig, Smooth-on) prior to mixing layer 1 PDMS yielded a uniform white background for sweat

visualization and chloride concentration measurements. A similar process created interconnect layer blanks comprising a 20:1 PDMS layer (150 μm thick at 500 RPM, 30 s) on a laser-defined PMMA sheet (100 mm circle, 3 mil thickness). A CO₂ laser cutting system (Universal Laser, VLS3.50) defined the outer diameter of the layer and the necessary openings to permit fluid transport between layers 1 and 2. A biopsy punch (1.2 mm diameter) formed the openings to the epidermis in layer 1 and a circular steel punch (20 mm diameter) defined the outer diameter of layer 2 and channel exits. Device assembly followed a logical progression. Adding 5 μL of chloride-free gel food dye (AmeriColor) at reservoirs near the ingress point of each chamber prior to assembly permitted visualization of sweat during device operation. Exposure of layer 1 and the interconnect layer to plasma from a corona wand (ElectroTechnic BD-20AC, 30 s) activated the surface to facilitate bonding upon lamination with applied pressure. Baking at 70 °C for 1 hr ensured complete interfacial bonding. Upon removal of the PMMA sheet, the same process enabled integration of layer 3 to the device with a circular steel punch (30 mm diameter) that defined the outer diameter of the completed assembly. Subsequent corona treatment exposure to the backside of the device ensured a permanent bond to a laser-patterned skin adhesive (3M 2475P, skin-facing; 3M 91022, device facing; 3M, Inc.).

Characterization of fill rate as a function of bend radius

Phantom arm assemblies for studies of flow used tubing [Tygon, ID = 1.5 mm, outer diameter (OD) = 3 mm] embedded in a 3D-printed model of an arm (Black Resin, Formlabs Form 2). A calibrated pressure displacement pump (Fluigent, France) controlled the pressure of water introduced into the system. Each measurement consisted of a “fill test” in which water at a fixed pressure (2 kPa) entered the device until full. Simultaneously, the progression of the fluid front was recorded using a camera (Canon 90D, Canon EF 100mm f/2.8 L USM lens) at 30 frames per

second. Three separate phantoms reflective of different arms (infant, $r = 15$ mm; adult, $r = 30$ mm; flat/control) were used for the testing described above.

Colorimetric chloride measurements

Systematic reactions of silver chloranilate with solutions of artificial sweat (5, 10, 15, 20, 30, 50, 60, 80, 90, 100, 125, 135, 160 mM of NaCl) yielded reference solutions for establishing a colorimetric calibration curve. Imaging a sweat sticker filled with a reference solution with a DSLR camera (Canon 90D) under uniform illumination for each concentration formed a set of reference images. Four image sets were recorded corresponding to the color and clear overlays. A 24-color card (SpyderCHECKR Color Chart, Datacolor) enabled color correction of RAW images prior to image processing via a custom MATLAB script. Procedures for extracting and processing LAB color values appear in detail in the materials and methods section of the Supplemental Materials.

Preparation of sweat stickers for integrated chloride analysis required replacement of the sweat visualization dye with silver chloranilate and the addition of overlays (one clear, one green) to two chambers. The overlays comprised 1 mil PET (polyethylene terephthalate) transfer films with solid colors added by a laser printer (color calibrated, Canon ImageClass MX722). Human subject testing followed the previously described sweat testing procedure with replacement of the MSCS with a sweat sticker. Devices were removed upon the completion of the test prior to extraction of the sample and imaged as previously described. The concentration of the sweat chloride collected by the control chamber was measured using a ChloroChek Chloridometer (ELITech Group).

Statistical analysis

Data are presented with mean values and SD, unless noted in the figure caption. All statistical analysis of study data was performed using the R statistical language. Wilcoxon Rank Sum test

(Mann-Whitney test) was used instead of Student's t-test as it is valid for data from any distribution. Power-law curve fitting of measured colorimetric data was used to generate calibration curves for colorimetric sensing using MATLAB (v. 2020a, MathWorks, Inc.).

Supplementary Materials

Materials and methods

Fig. S1. Examples of MSCS failure modes observed during clinical testing.

Fig. S2. Linear fill characterization.

Fig. S3. Sweat volume collected by sweat sticker and MSCS.

Fig. S4. MSCS volume comparison for left versus right arms.

Fig. S5. Bland-Altman analysis of sub populations.

Fig. S6. Dermatological impact of collection methods on infants.

Fig. S7. Colorimetric image analysis procedure.

Fig. S8. Example of individual pixel measurements within a region of interest (ROI).

Fig. S9. Comparison of image capture methodology on chloride measurement.

Fig. S10. Modes of device operation in clinical practice and disease management.

Table S1. Description of the population study cohort grouped by age.

Table S2. Individual data from participants in the study.

Movie S1. Demonstration of sweat sticker removal.

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Author Contributions:

T.R.R., S.H. and J.A.R. led the development of the concepts, designed the experiments, interpreted the results. T.R.R., R.G., and J.A.R. wrote the paper. T.R.R. led the experimental work supported throughout by M.I.; P.M.C., D.F., W.J.J., J.C., H.G., G.Y., S.R., J.T.R., A.H-F, A.J.B., J.C. contributed to device fabrication and testing; J.C. to fabrication process development; D.F., K.G., J.B.M., S.P.L., J.C., and A.J.A. to colorimetric analysis. T.R.R., S.A.M., and S.H. led the design

and development of the patient trials and were assisted by M.I. All authors contributed to proofreading the manuscript.

Competing interests:

T.R.R. and J.A.R. are inventors on patents and patent applications related to epidermal microfluidics, including applications in cystic fibrosis diagnostics (Patent “Thin, soft, skin-mounted microfluidic networks for detection and analysis of targets of interest in sweat,” # US 2020/0093416 A1 and Patent “Microfluidic systems for epidermal sampling and sensing,” # US 2020/0155047 A1). R.G., S.P.L., A.J.A., J.B.M., and J.A.R. are cofounders of Epicore Biosystems, a company that develops epidermal microfluidic devices. T.R.R. has a consulting and advisory relationship with Epicore Biosystems. The remaining authors are not aware of any affiliations, memberships, funding, or financial holdings that might be perceived as affecting the objectivity of this research.

Data and materials availability:

All data associated with this study are present in the paper or the Supplementary Materials. All information and materials can be requested from one of the corresponding authors.

Figures:

Fig. 1. Sweat sticker design and function. (A) Exploded view illustration of the device, highlighting key layers. (B) Microfluidic channel geometry. (C) Optical images of the clinical standard device for sweat collection (MSCS) on the arm after pilocarpine iontophoresis. The guideline-specified collection time for sweat (light blue) is 30 minutes. (D) Images of the epidermal microfluidic device (sweat sticker) on the contra-lateral arm after pilocarpine iontophoresis. (E) Optical images of removal of the protective cover of the MSCS, exposing the microbore tubing to permit insertion of a blunt-tip needle. Positive pressure from the needle (and syringe) ejects sweat from the tubing into a microcentrifuge tube (not shown). (F) Removal of sweat sticker from the skin after the collection period. Full extraction of sweat occurs via application of negative pressure using a 1-mL laboratory pipetter. Transfer of the sweat sample into a microcentrifuge tube (not shown) follows this extraction step. Scale bars, 5 mm.

Fig. 2. Device characterization and benchtop testing. (A) Images of a device (left) while bent (center) and twisted (right). (B) Characterization of bending effects on performance in sweat collection. Images of the sticker attached to a flat plate (top) and phantoms of an adult (middle) and infant (bottom) arm. (C) Graph of repeat filling tests ($N = 3$) at fixed physiological pressures (2 kPa) via a pressure displacement pump for a flat surface and phantoms of an adult and infant arm. The results show consistent, linear filling behavior. Volumes measured by image processing indicate a standard deviation of 0.1 μL . (D) Magnified region of the graph from Fig. 2C. Time error bars represent standard deviation of $N = 3$ measurements. Volume error bars omitted for clarity. Scale bar, 5 mm.

Fig. 3. Clinical study inclusive of patients with CF. (A) Results of collected sweat volume from a clinical pilot study using the microfluidics sticker (blue data, $N = 51$) relative to the

MSCS in both this work (difference is statistically significant, Wilcoxon test, $W = 1603.5$, $P = 0.02$; matched-pair average difference $\sim 33\%$, $P = 0.05$) and published studies. Reported QNS rates (%) appear as red bars (red bars). Reported volumes of collected sweat appear as boxes color-coded to this study (blue) or published studies (black). Specific studies referenced are: LeGrys 2017(58), LeGrys 2018(16), De Boeck 2017(59). **(B)** Equivalence plot for sweat chloride estimates from the sweat sticker versus the MSCS. **(C)** Bland-Altman plot of sweat data shown in (B). Blue line indicates the mean, red dashed lines indicate 1.96 SD. **(D)** Histograms of sweat volume from collection methods across three age groups (infants: 0 – 6m, $N = 3$; pediatrics: 7m – 18y, $N = 11$; and adults: +18y, $N = 37$). Difference in mean volume collected for adult age group is statistically significant (Wilcoxon test, $W = 151$, $P = 0.03$).

Fig. 4. Photographs of device operation. **(A)** Pediatric subject at the conclusion of a sweat test with magnified insets that show volumes collected by the **(B)** sticker and **(C)** the MSCS prior to sweat extraction. **(D)** Sweat sticker on an infant subject at the conclusion of a sweat test. This evaluation involved a nearly complete filling of the device. Scale bars: (A) 10 mm, (B)-(D) 5 mm.

Fig. 5. Colorimetric chloride assay analysis. **(A)** Exploded view illustration of the sweat sticker with an integrated colorimetric chloride assay. A single-layer device comprises three independent microfluidic collection chambers and an overlay (either colored or clear; shown in green in illustration). **(B)** An example set of optical images show the progressive color shift of the chloride assay in response to increasing chloride concentration (in water). Clinical ranges for CF diagnosis indicated for reference. **(C)** Color measurements of the assay images in (B) without a color overlay (clear) shown in CIELAB color space (A and B components). Color overlays (green, blue, red) shift the measured color response. **(D)** Calibration chart of chloride

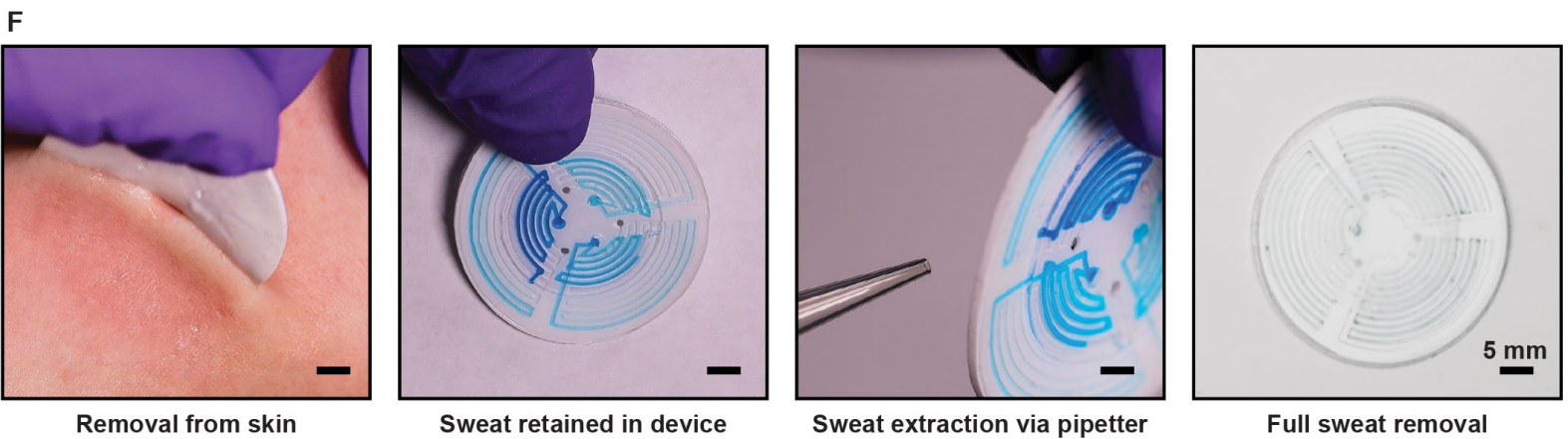
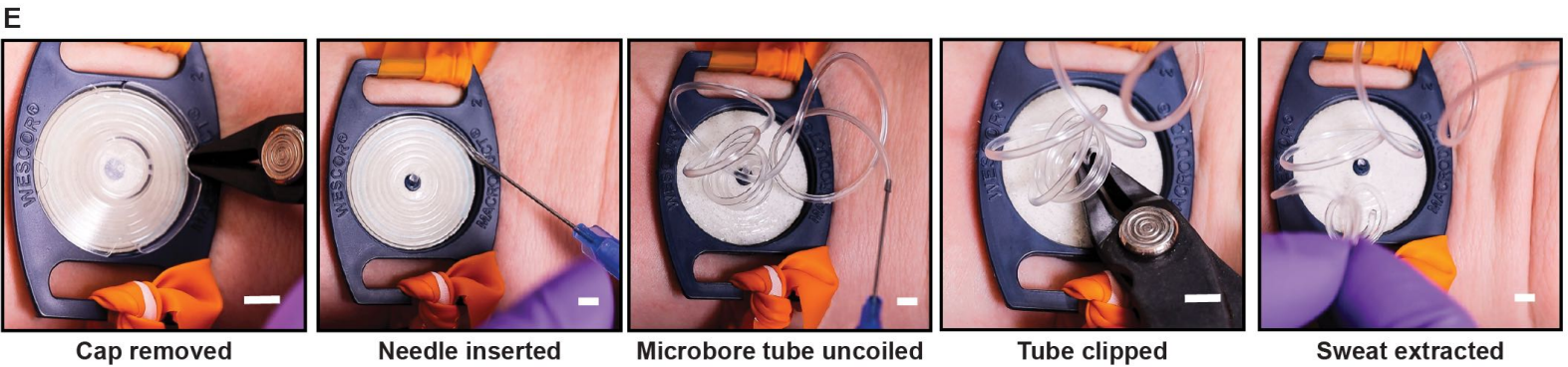
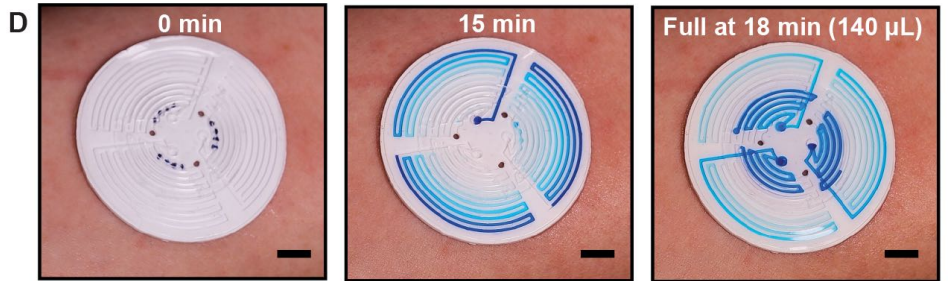
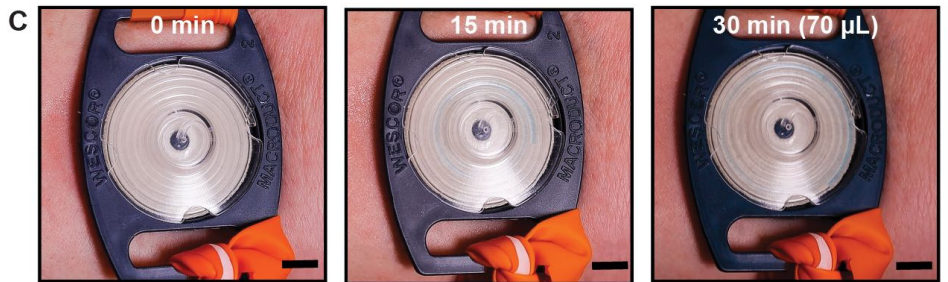
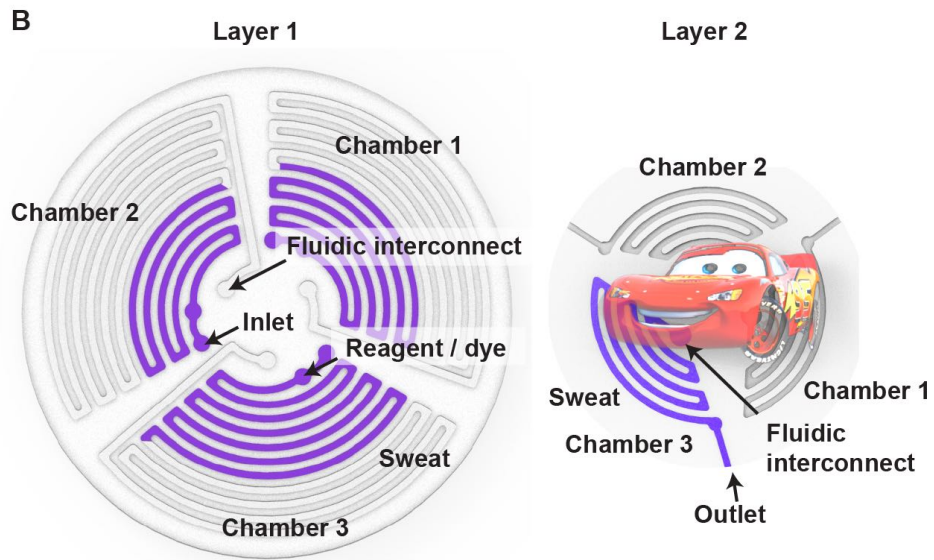
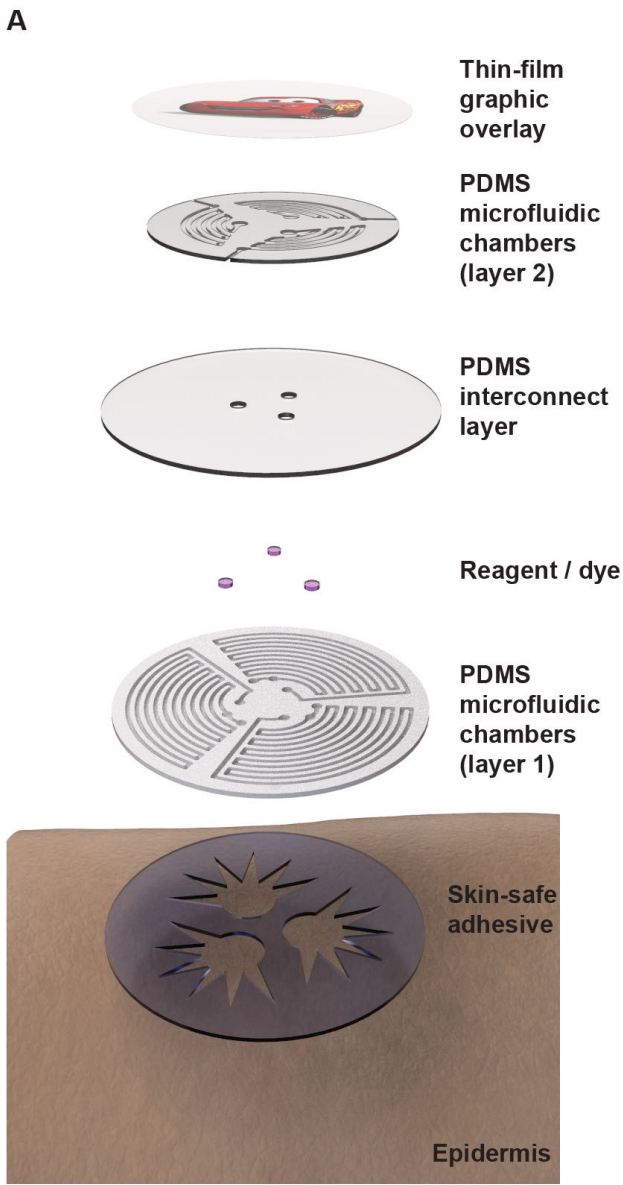
concentration vs. chroma (C^*) for the color and clear overlays with corresponding power-law fits. **(E)** Comparison of colorimetric assay performance (with and without overlays) to chloridometer measurements for three clinical chloride standards. Assay measurements are from calibration fits using chroma (C^* , solid) or a^* values (checked).

Fig. 6. Pilot study of integrated sweat sticker. **(A)** Optical image of a body-worn sweat sticker after a 30 min collection period. Violet chambers contain the colorimetric assay and the blue chamber is a control sample for analysis using a commercial chloridometer. Scale bar, 5 mm. **(B)** Comparison of sticker performance in human subject pilot study ($N = 5$).

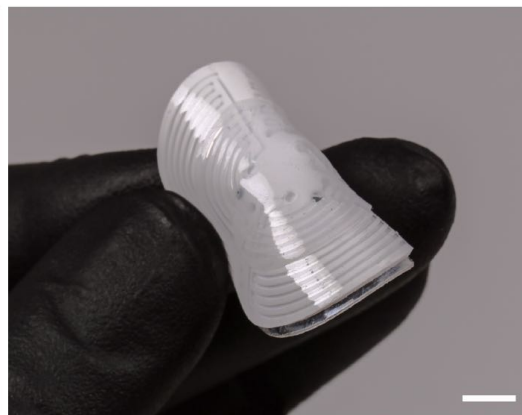
Tables:

Table 1. Study population demographics.

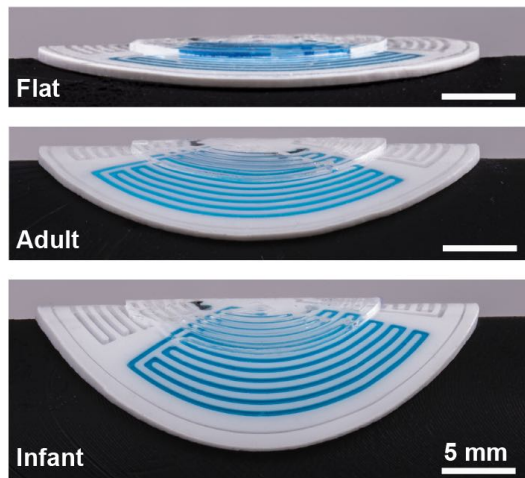
		This study	LeGrys Study 1(58)	LeGrys Study 2 (58)
N		51	28	100
Age				
	Mean	23	29	24
	Range	2m – 51y	18y – 44y	8y – 54y
	0 to 6m	3	–	–
	7m – 18y	11	–	–
	+18y	37	–	–
Gender				
	Female	25	18	48
	Male	26	10	52
Condition				
	CF	33	28	100
	Non-CF	18	–	–
Number of samples				
	Total	102	292	687
	Left arm	51	146	346
	Right arm	51	146	341
Collected volume (Mean ± SD)				
	MSCS	58.4 ± 24.1 µL	57 mg	53 mg
	Sweat sticker	73.6 ± 35.2 µL	–	–
Range				
	MSCS	7.2 – 94.5 µL	17 – 105 mg	15 – 110 mg
	Sweat sticker	17 – 133 µL	–	–
QNS occurrences				
	MSCS	3 (5.8%)	14.3% (4/28)	13.0% (100)
	Sweat sticker	0	–	–



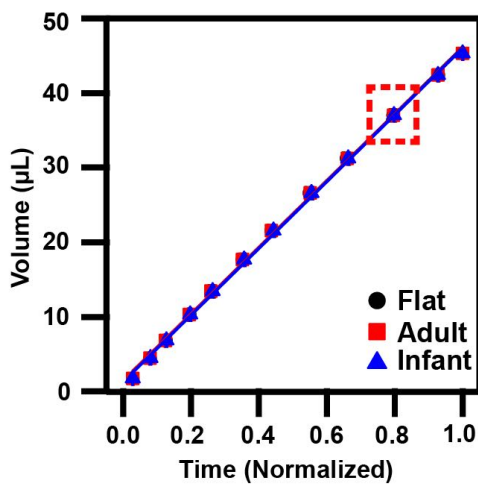
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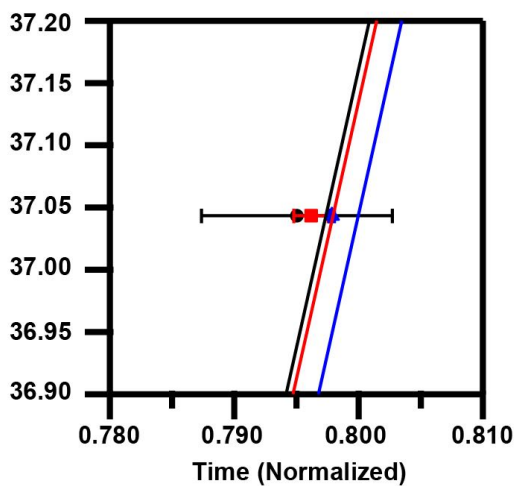
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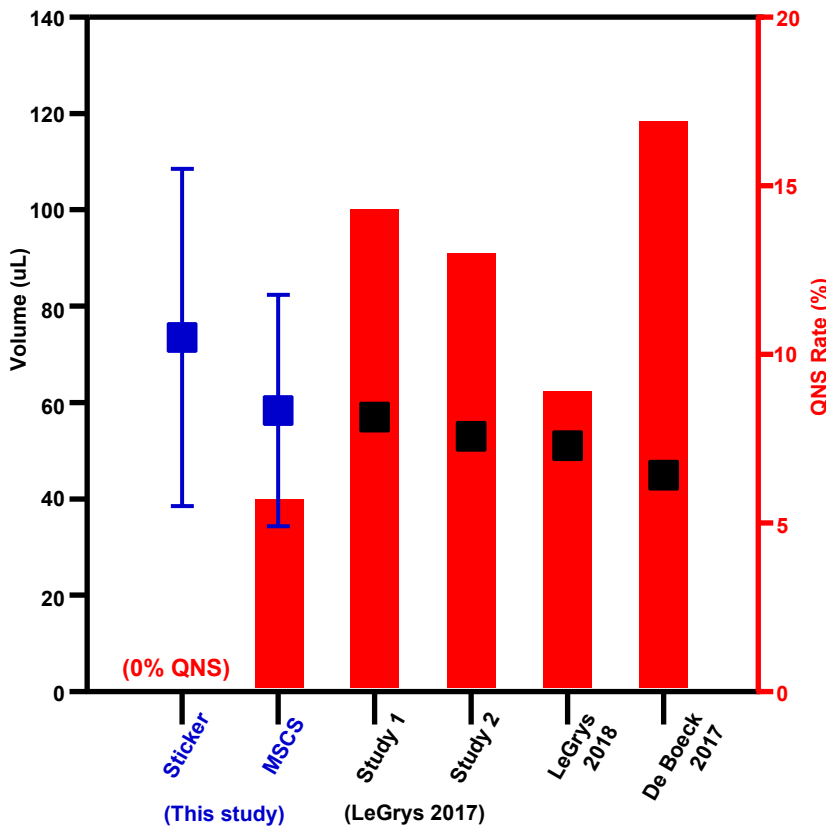
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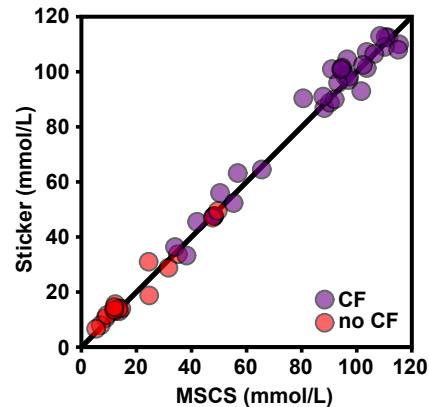
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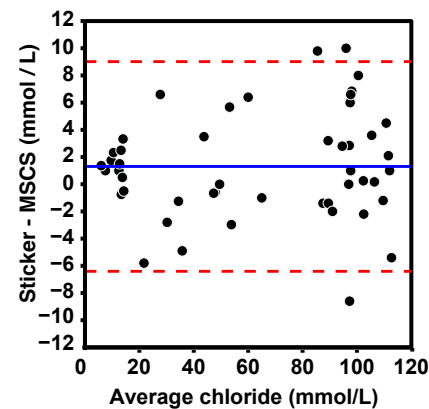
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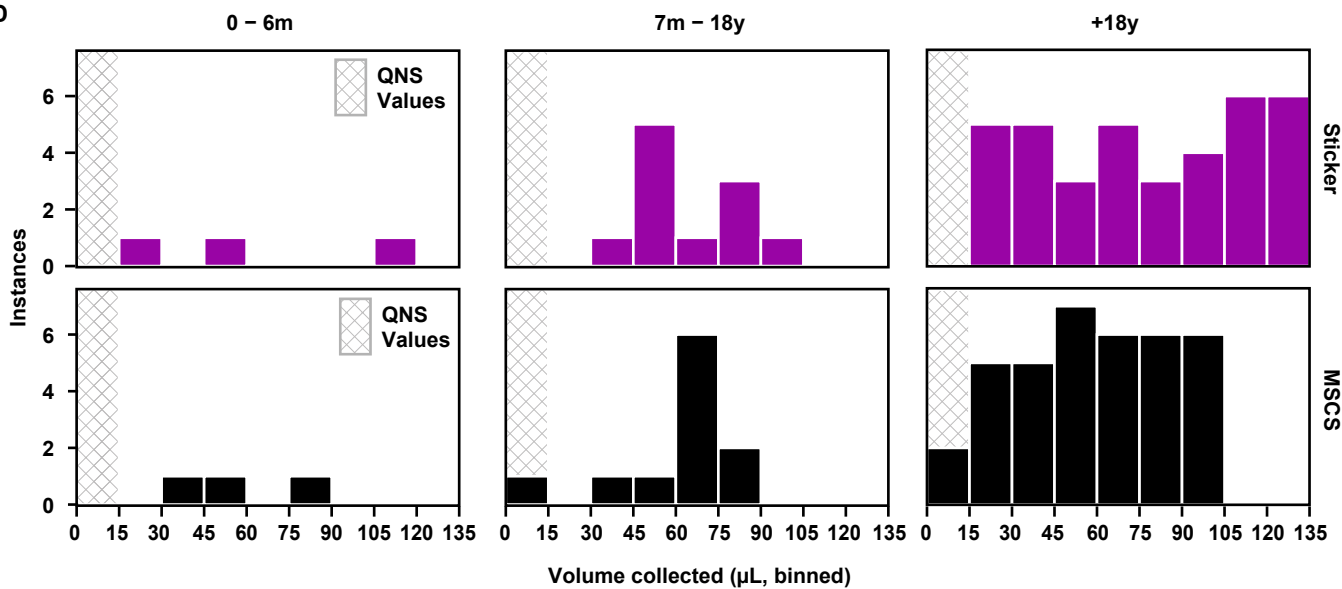
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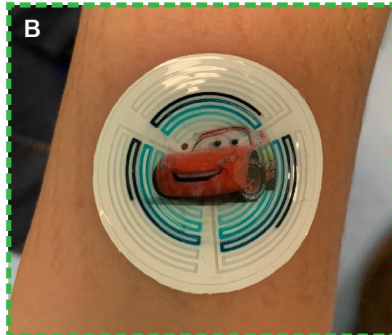


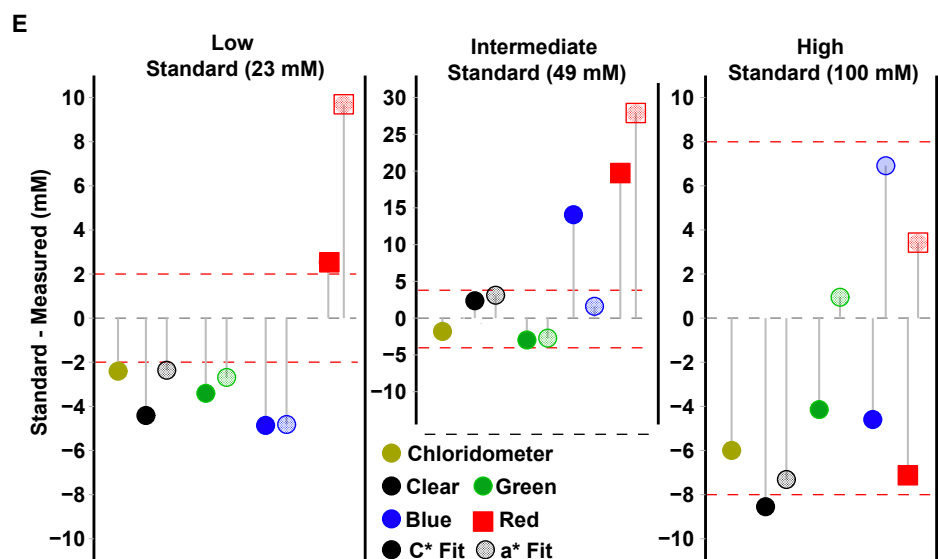
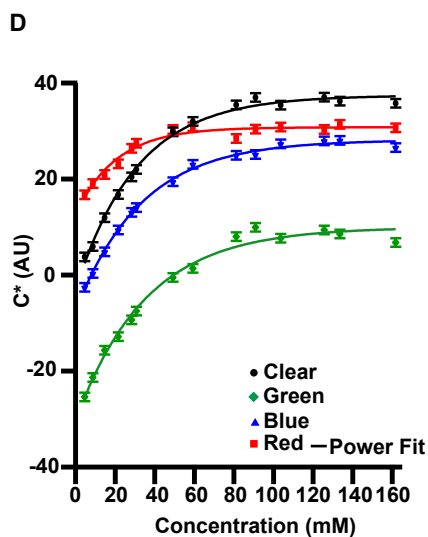
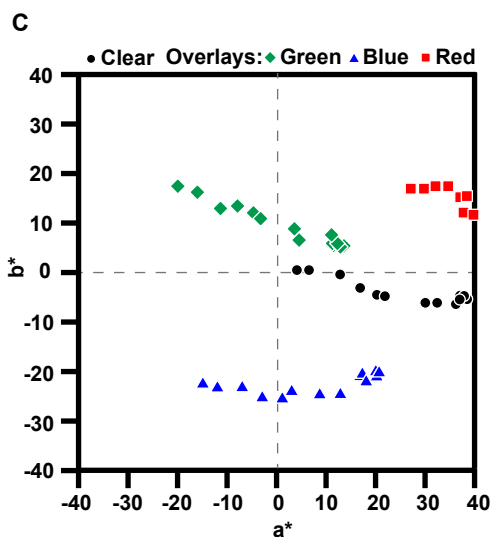
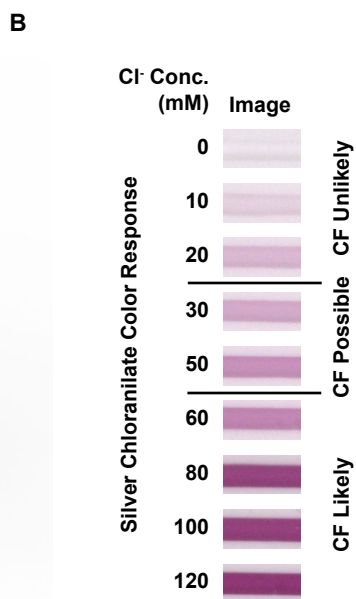
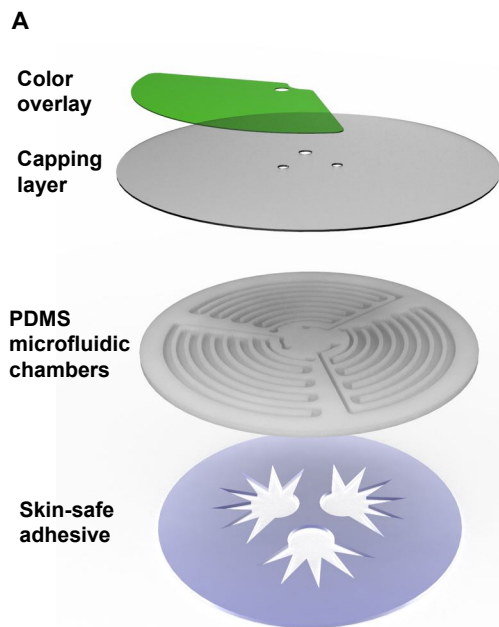
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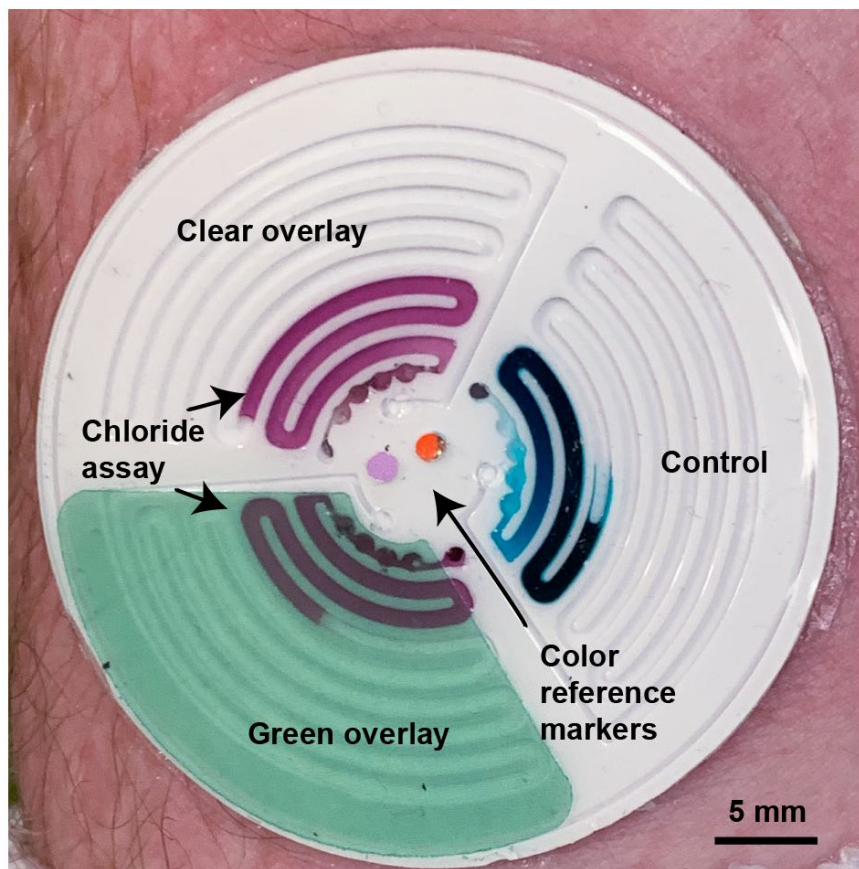
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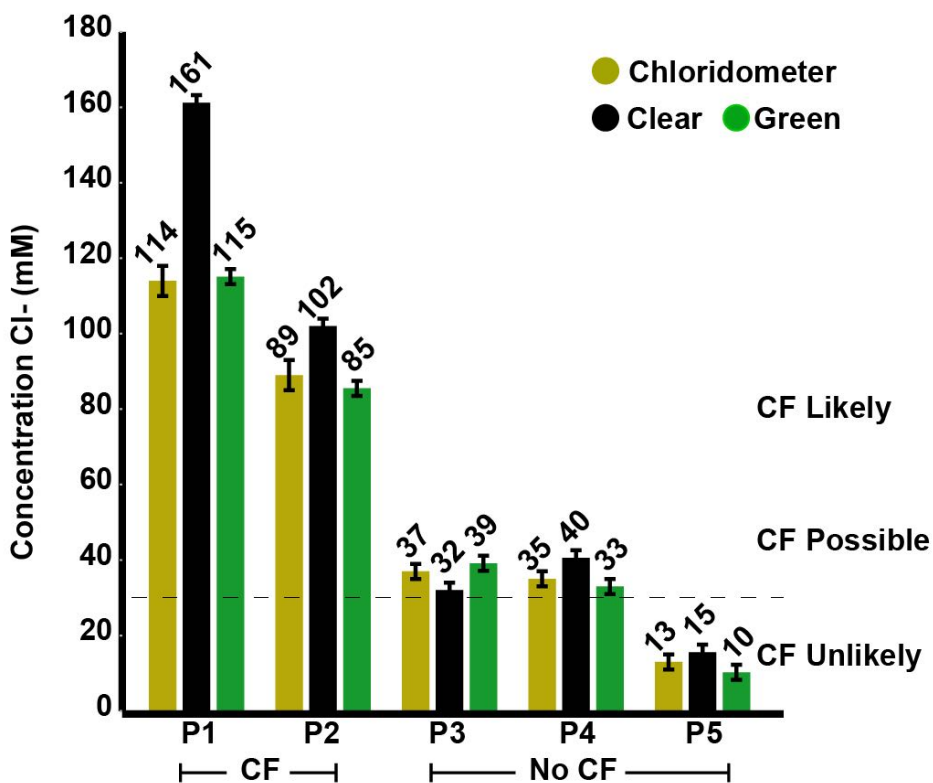




A



B



Supplementary Materials for

Soft, skin-interfaced sweat stickers for cystic fibrosis diagnostics and management

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This PDF file includes:

Materials and Methods
Fig. S1 to S10
Table S1 to S2
Caption for movie S1

Materials and methods

(i) Study protocol

Study objectives involve sweat data collection and exploratory clinical endpoints:

1. Sweat collection devices meet performance specifications for collection volume and fill time compared to conventional technique.
2. Sweat collection devices meet robustness criteria required for use in clinical laboratory setting compared to conventional technique.
3. Sweat collection using these devices enables accurate and precise measurement of chloride compared to current clinical laboratory-based chloride analysis methods.

Inclusion / exclusion criteria:

Adult and children CF patients and healthy volunteers were recruited to conduct various parts of this study.

Inclusion criteria:

- (1) Healthy adult volunteers
- (2) Adults with CF or known carriers of cystic fibrosis gene mutations
- (3) Healthy children volunteers (7 days to 17 years old)
- (4) Children undergoing sweat chloride testing for clinical purposes (7 days to 17 years old)
- (5) Children with known diagnosis or carrier status for CF (7 days to 17 years old)

Exclusions criteria:

- (1) Patients receiving oxygen by an open delivery system.
- (2) Patients with insufficient access for sweat collection (i.e., inadequate areas on the arms or legs that are free from diffuse inflammation or rash.)

Summary of procedures to induce and extract sweat:

The performances of the sweat sticker and chloride analysis were evaluated using both benchtop analysis and camera-based image processing, and compared to a conventional method (MSCS). Sweat extraction and chloride measurement using benchtop analyzers include the following steps: i) pilocarpine application and iontophoresis to stimulate sweating; ii) sweat collection following iontophoresis; and iii) quantitative sweat chloride analysis.

Sweat glands are locally activated by iontophoretic delivery of pilocarpine. Briefly, an electric current is applied to transport the pharmacologically active agents into the skin. Once the sweat glands have been stimulated for ~5 minutes, the electrodes are then removed, the skin is cleaned with an alcohol wipe, and the sweat collection device is placed over the local region where the pilocarpine was applied. Sweat is collected for up to ~30 minutes. A visual indicator aids staff in recognizing when sufficient sweat volume has been collected. Clinical staff document the iontophoresis steps and sweat collection times.

The captured sweat is then extracted from the device and stored in a pre-weighed microfuge tube and analyzed using a chloridometer (Wescor ChloroChek, ELITech Group). A minimum of 15 μL of sweat is required to generate results. If available, sweat samples are measured in duplicate or triplicate. Quality control samples are processed and pass acceptance criteria prior to the analysis of study participant samples.

In parallel, sweat is also induced iontophoretically and collected using the control MSCS and chloride is measured with Wescor ChloroChek instrument. Paired experimental (sweat sticker) and control (MSCS) data were analyzed across 51 subjects as part of this comparative study. In addition to the sweat extraction and external sweat chloride analysis, a subset of CF patients and healthy volunteers (n= 2 CF patients; n = 3 healthy volunteers) had sweat stickers with onboard chloride assay for image-based colorimetric analysis. These results were compared to the MSCS and chloridometer results.

Best practices for iontophoresis and sweat collection were followed (69,71):

- a. Collection of sweat should be carried out at a time when the patient is clinically stable, well hydrated, free of acute illness, and not receiving mineralocorticoids.
- b. Sweat can be collected from the arm or leg, using a site free from diffuse inflammation or rash. When using the arm, the positive electrode is placed on the inner flexor surface of the forearm. For the leg, the positive electrode is placed on the thigh. Under no circumstance should the electrodes be placed so that the current crosses the trunk.
- c. Iontophoresis is withheld from patients receiving oxygen by an open delivery system.
- d. Attachment of the MSCS to the inner flexor surface of the forearm is by a disposable tourniquet. The assembly is then overwrapped by gauze to improve contact with the epidermis as described in (71)
- e. Hands should not touch any portion of the collection materials, collection device, collection site, or weighing vials. Powder-free gloves should be used.

Informed consent:

Prior to study entry, a written informed consent was obtained from the parental guardians of the subjects or from the adult subjects participating in the study. This trial was conducted in accordance with Good Clinical Practices and the Declaration of Helsinki.

Note regarding MSCS sweat collection process:

The manufacturer's instructions state that the tubing should be held tight; however, in practice, this is difficult for phlebotomists to maintain, especially with moving patients. Failures of trained technicians to maintain strict adherence to CLSI testing procedures under all circumstances is a common issue determined by Quality Improvement programs at CF testing centers. This is a broadly-identified issue over the past two decades with sweat testing in general and the MSCS in particular (72-74).

As referenced above, this study utilizes a disposable tourniquet to attach the MSCS to the patient, which is not provided by the manufacturer. The use of a disposable tourniquet (and a gauze overwrap) as implemented by the study location (Lurie Children's Hospital, Chicago, IL) serves as a means to reduce instances of QNS rates. It also has the added advantage for infection prevention over the manufacturer-provided straps in that they do not need to be cleaned between patients, as they are disposable. The use of disposable tourniquets vs. manufacturer provided straps reduces rates of QNS from a QNS rate of 15% for all ages to a QNS rate of 5% for all ages ($p < 0.003$) (71). We utilized this method to ensure a best-case comparison to the MSCS.

(ii) Colorimetric-based image analysis procedure

Figure S5 shows the steps involved in colorimetric analysis of the sweat sticker. Images were captured with a DSLR camera in RAW format, under controlled lighting, at a fixed distance from the device. A calibrated color card (SpyderCHECKR Color Chart, Datacolor) served as a color reference target in each photograph to correct for ambient lighting and color-balance. RAW images were converted to color-corrected lossless tiff format images by a software program (RAWdigger, LibRaw, LLC.) to facilitate image processing in MATLAB. A custom MATLAB script processes each tiff formatted image by first converting from the native sRGB color space (camera default) to CIELAB color space via a native function (`rgb2lab`) in the Image Processing Toolbox. Color values in regions of interest (ROI) corresponding to (i) three PANTONE reference colors, (ii) a white color reference, (iii) regions of the device containing samples of sweat and colorimetric assay mixture, and (iv) background reference regions immediately adjacent to the sweat assays were extracted at the individual pixel level.

An example plot of uncorrected colorimetric measurements for a sample ROI is shown in Fig. S6. The ROI regions for reference colors and the white point were defined as 100x100 pixel square areas. The median values for each region were used to compute a color calibration transformation matrix using the measured CIELAB values. This calibration matrix enables quantitative correction to known colors for the sample and background reference regions at the single pixel level. After color correction, each median value of the background reference region is subtracted from the median value for the corresponding colorimetric assay region to eliminate slight variations in background illumination. The corrected a^* and b^* components were then used to calculate chroma (C^*) for each reference solution. From this, a power-law fit was derived as the basis of a calibration curve used to calculate sweat chloride concentrations.

We note that assessment and interpretation of colorimetric assays can be affected by ambient light if the analytical approach does not account for such variations or effects. Our previous work⁽⁴⁹⁾ provides a characterization study of these effects in the absence of a color overlay. As described in that work and in described process, incorporation of several reference colors enables utilization of industry-standard color correction processes to mitigate or eliminates this concern.

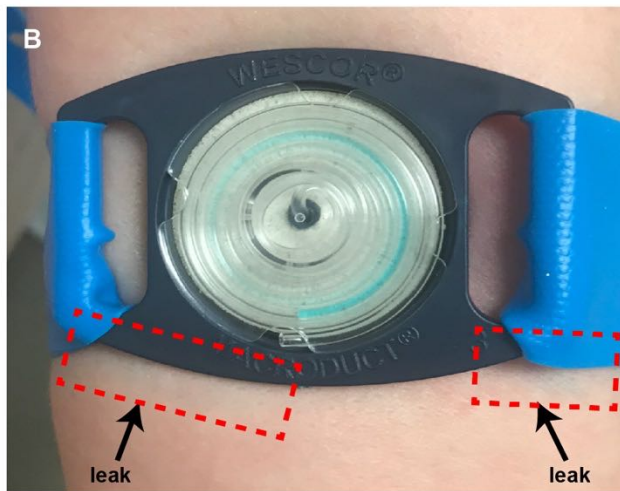
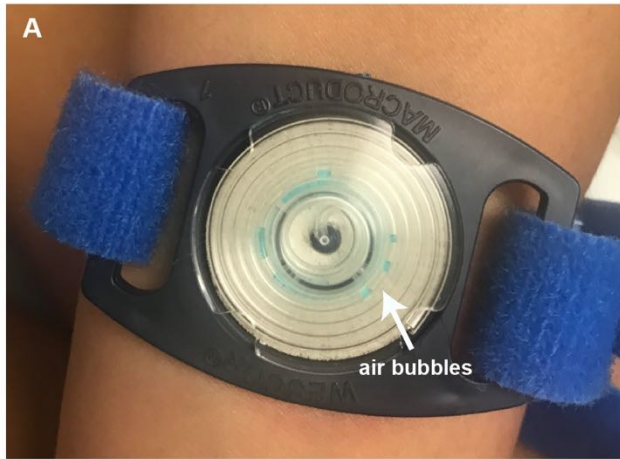


Fig. S1. Examples of MSCS failure modes observed during clinical testing.

(A) Photograph highlights motion-induced failure mode of MSCS by which air bubbles in the sweat collection tube result in insufficient sweat capture. **(B)** Photograph shows the motion-induced failure mode of MSCS whereby sweat leaked from the device caused staining of the skin, (71). **(C)** Photograph of the skin showing the effects of motion-induced leaks (B) after removal of MSCS.

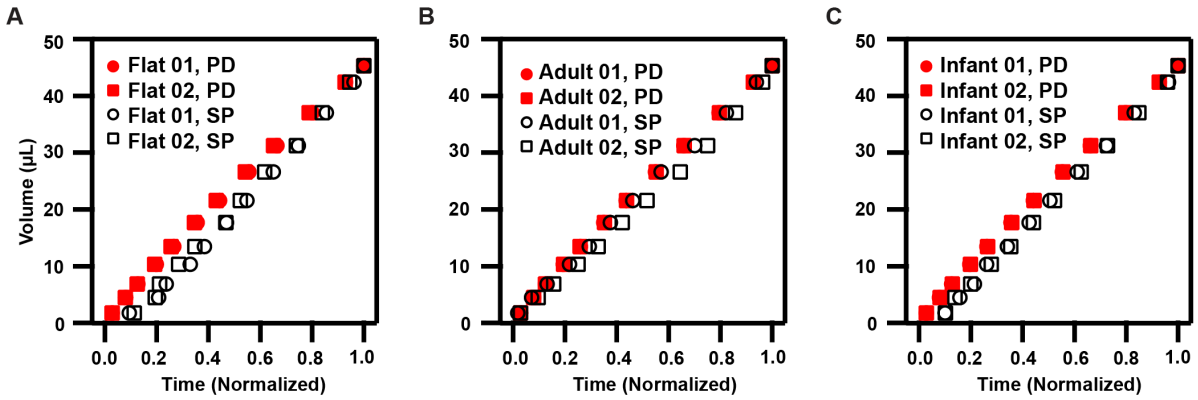


Fig. S2. Linear fill characterization.

Evaluation of fluid fill using a pressure displacement (PD) method delivered with a syringe pump (SP). Two sweat stickers were tested on a (A) flat surface, on an (B) arm phantoms representing an adult arm and on an (C) infant arm.

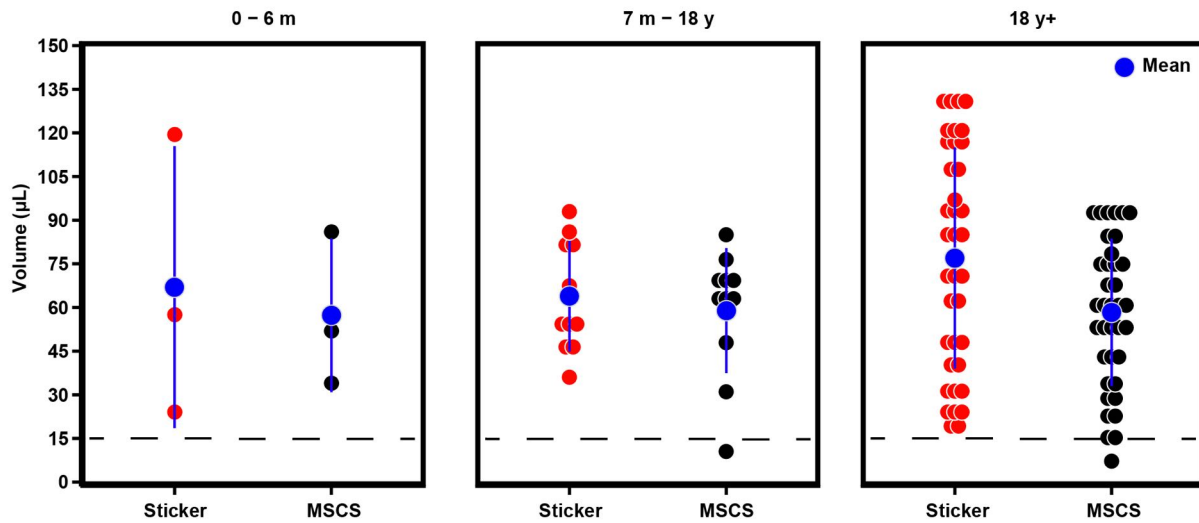


Fig. S3. Sweat volume collected by sweat sticker and MSCS.

Sweat volume results for sweat sticker and MSCS across three age groups (infant: 0-6m, pediatric: 7m-18y, and adult: +18y). Means (blue) and standard deviation are included for each age group. Dashed lines at 15 µL indicate the volume cut-off for a QNS (quantity not sufficient) event.

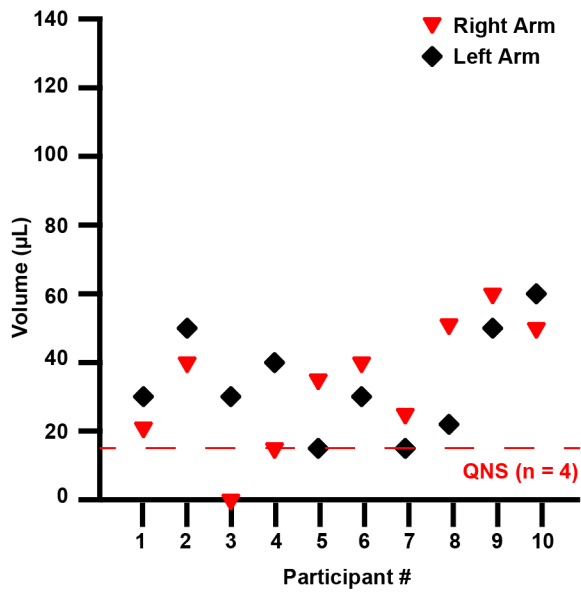


Fig. S4. MSCS volume comparison for left versus right arms.

A snapshot of sweat testing by phlebotomy staff as part of routine clinical operation. Data shown for 10 sweat tests conducted during a 3-day period to validate the routine clinical QNS rate for this study site. Four collection events resulted in QNS. The dashed line at 15 µL indicates the volume cut-off for a QNS (quantity not sufficient) event.

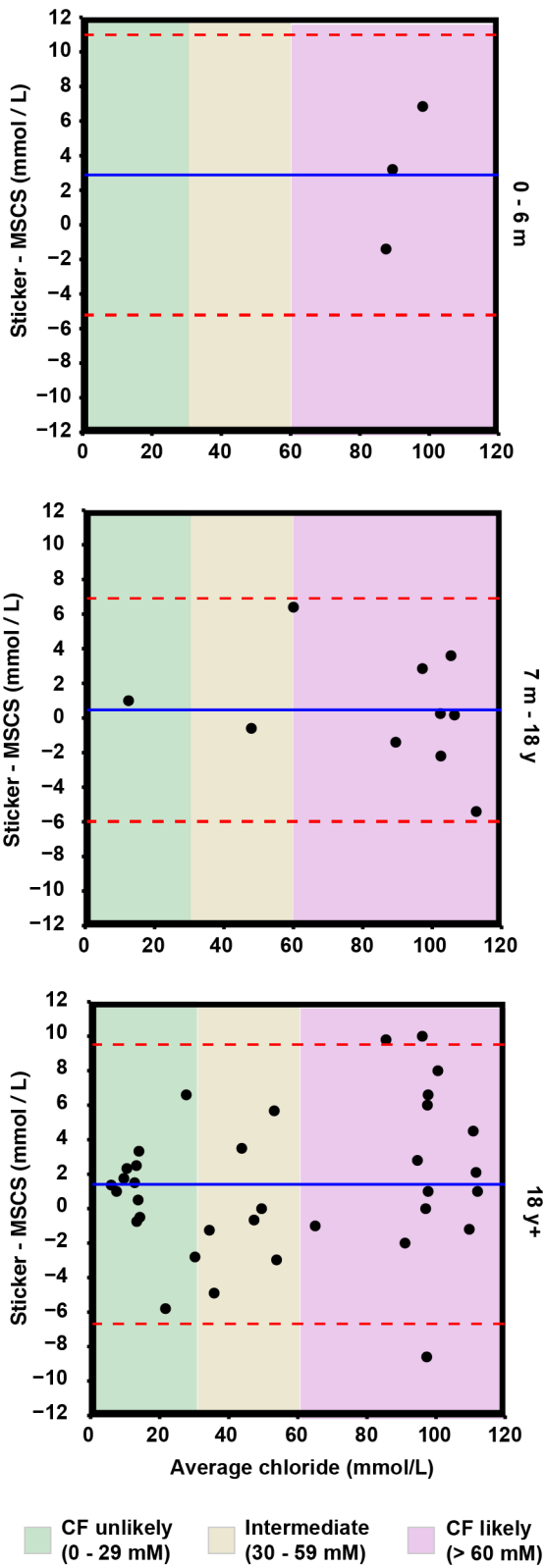


Fig. S5. Bland-Altman analysis of sub populations.

Sweat chloride analysis of the sweat sticker vs MSCS across infant (top), pediatric (middle), and adult (bottom) populations.

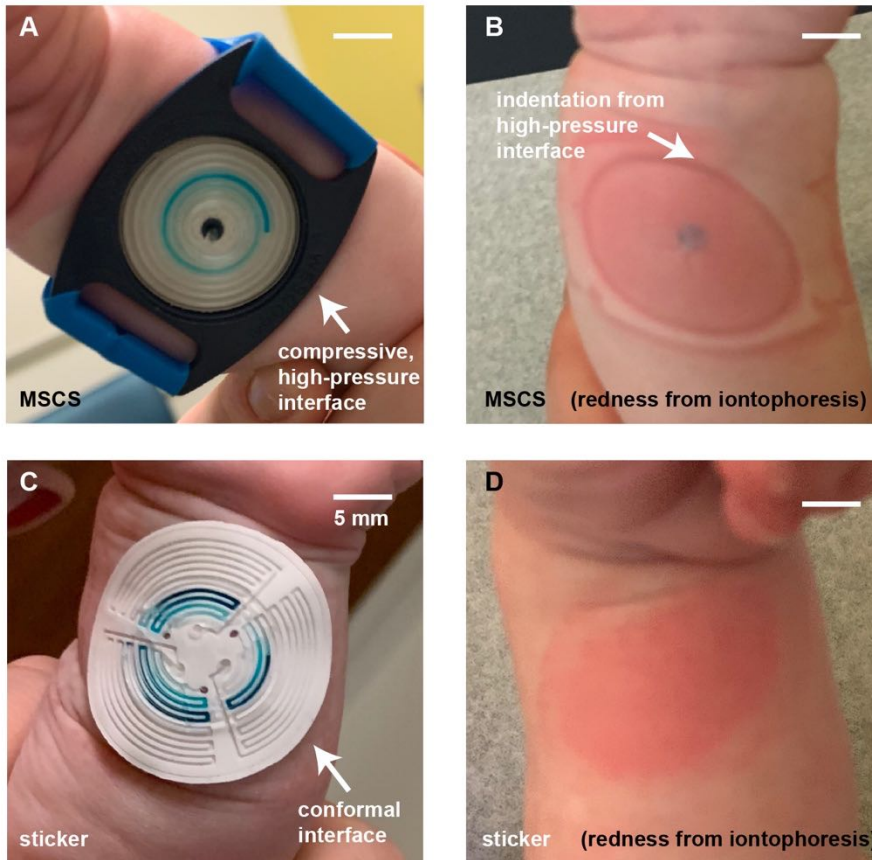


Fig. S6. Dermatological impact of collection methods on infants.

Photographs of collection devices [(A) – MSCS, (C) – sweat sticker] captured at the conclusion of the sweat test. (B) and (D) show the skin region upon removal of the devices. As a result of the high-pressure interface required for successful sweat collection events, the MSCS typically causes a transient indentation in the skin (~1-2 mm in depth). The sweat sticker does not leave a tactile mark on the skin. Skin reddening results from the pilocarpine iontophoresis prior to application of the sweat collection device.

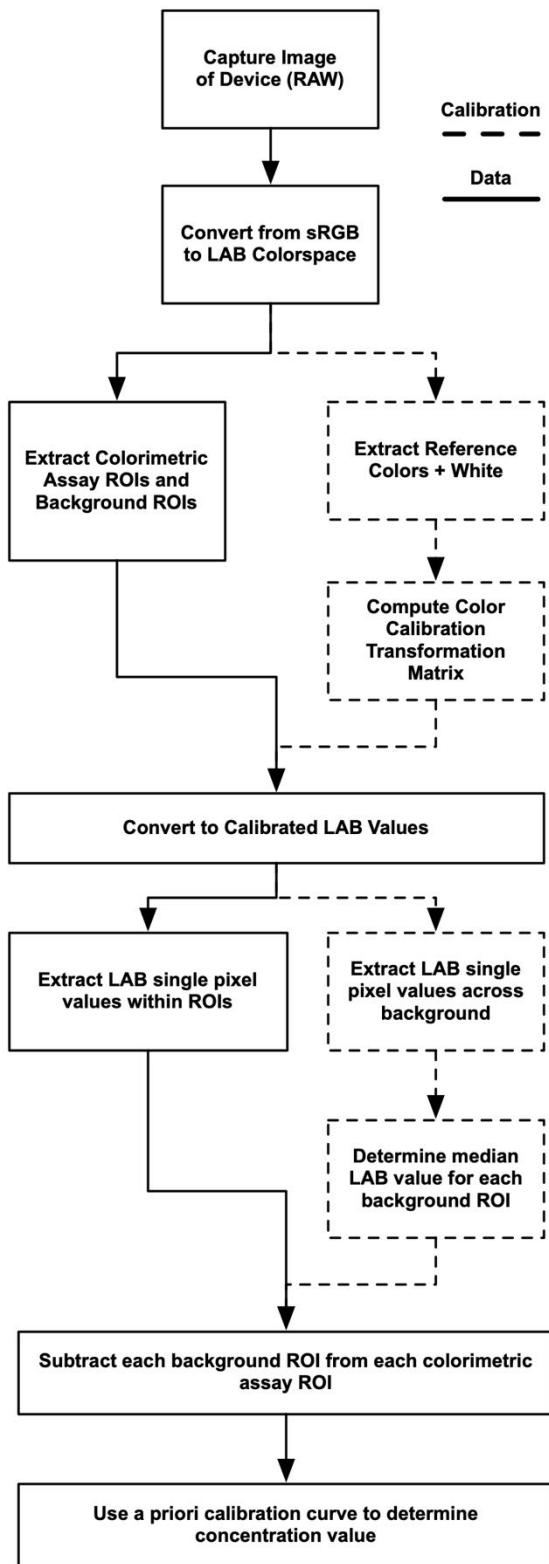


Fig. S7. Colorimetric image analysis procedure.

RAW images from camera (Canon 90D) are converted into LAB color space. Subsequent processing steps generate values suitable for quantitative analysis of chloride via a calibration curve generated *a priori*.

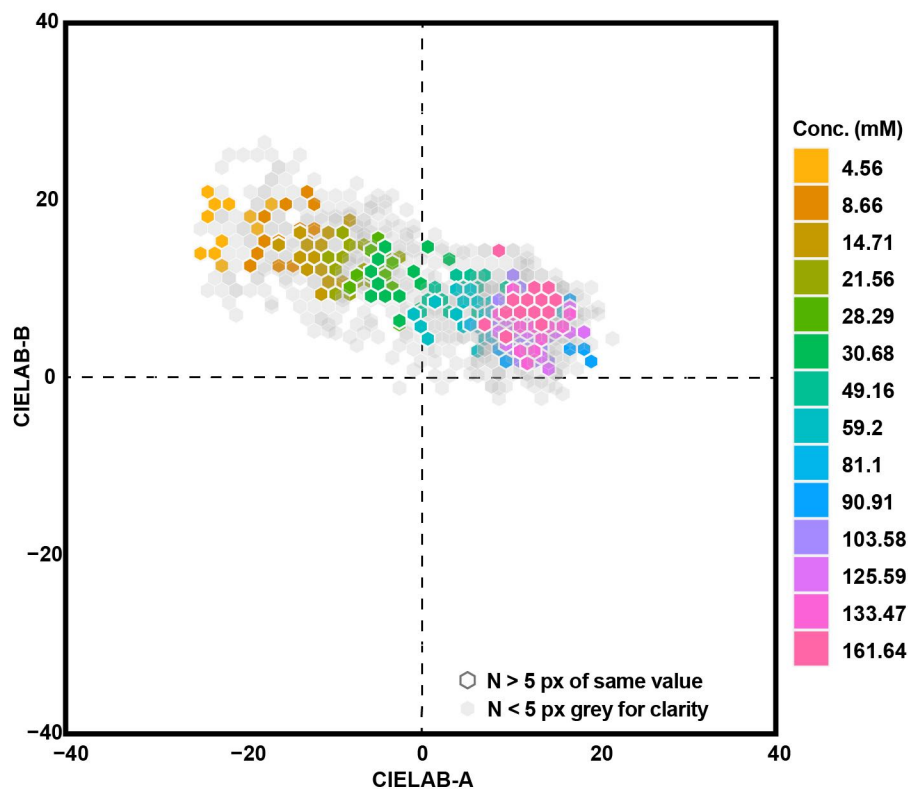


Fig. S8. Example of individual pixel measurements within a region of interest (ROI). Measurements are from a ROI with color overlay. CIELAB values were converted directly from RAW values.

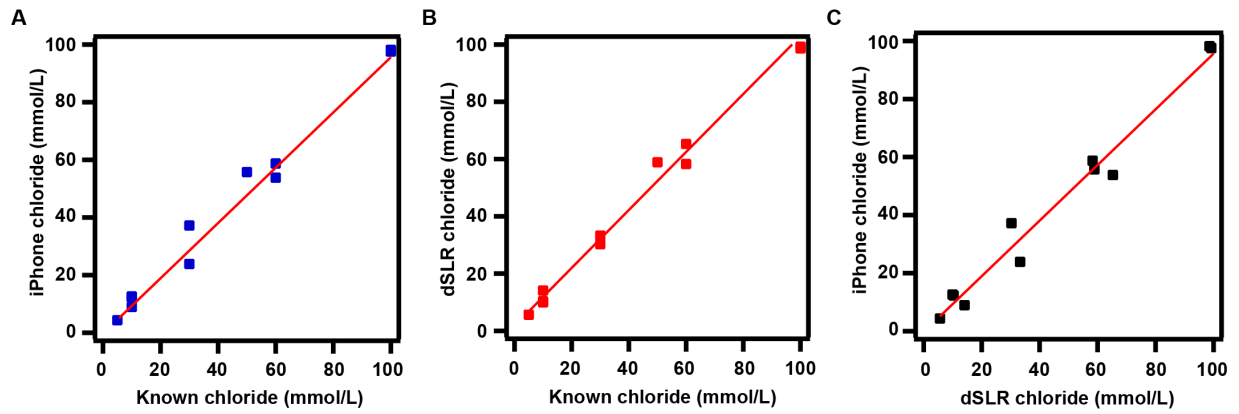


Fig. S9. Comparison of image capture methodology for chloride measurements.

Measurements of calibration techniques using known chloride concentrations from images captured with (A) iPhone 8 and (B) dSLR camera. (C) iPhone 8 versus dSLR camera equivalence plot.

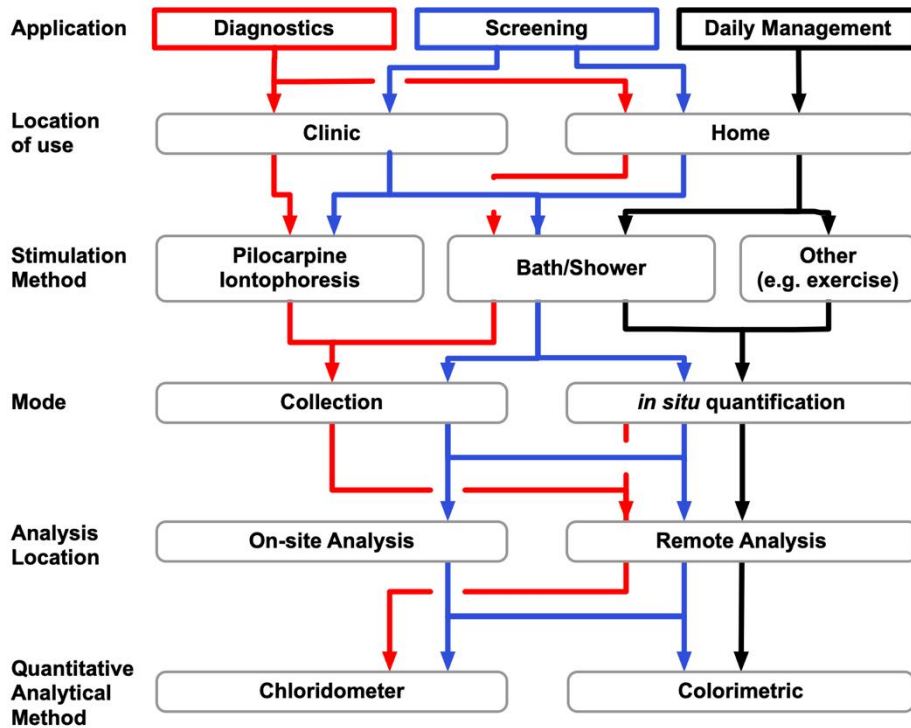


Fig. S10. Modes of device operation in clinical practice and disease management.

Table S1. Description of the population study cohort grouped by age.

(A) Divisions as specified by Medical Subject Headings. (B) Divisions as specified by the Age-Phenome Knowledge-base. (C) Divisions as specified as newborn, pediatric, or adult.

A. Population statistics

Age range	Age classification	Male (#)	Female (#)	CF (#)	Control (#)	Total (#)
0-23 months	Infant	1	3	3	1	4
2-5 years	Child, preschool	1	1	1	1	2
6-12 years	Child	4	2	6	0	6
13-18 years	Adolescent	1	1	2	0	2
19-44 years	Adult	18	16	19	15	34
45-64 years	Middle aged	1	2	2	1	3

B. Population statistics

Age range	Male (#)	Female (#)	CF (#)	Control (#)	Total (#)
birth - 4 years	2	4	4	2	6
5 - 9 years	2	1	3	-	3
10 - 14 years	2	1	3	-	3
15 - 19 years	3	2	4	1	5
20 - 29 years	10	12	12	10	22
30 - 39 years	5	2	5	2	7
40 - 49 years	2	1	2	1	3
50+ years	-	2	1	1	2

C. Population statistics

Age range	Male (#)	Female (#)	CF (#)	Control (#)	Total (#)
birth - 6 months	-	3	3	-	3
7 months - 18 years	7	4	9	2	11
18+ years	19	18	21	16	37

Table S2. Individual data from participants in the study. For reference, red denotes instances of QNS (quantity not sufficient). Chloride values are reported averages of N = 3 measurements. Chloride values reported for QNS instances (where collected sweat exceeded 10 μ L) are N = 1.

ID	Age (y)	Age (m)	Gender	CF	Modulator	MSCS Arm	MSCS Volume (μ L)	MSCS Cl (mM)	Sticker Arm	Sticker Volume (μ L)	Sticker Cl (mM)
1	51	0	F	No	No	Right	60.5	35	Left	46	33.75
2	12	0	F	Yes	Yes	Left	10.5	34	Right	36	36.33
3	24	0	F	Yes	No	Left	51.3	94.5	Right	69.5	100.5
4	19	0	F	Yes	Yes	Right	7.2	NA	Left	25.5	66.5
5	22	0	M	Yes	Yes	Right	94.1	96.5	Left	133.1	104.5
6	29	0	M	Yes	Yes	Left	77.4	111.5	Right	120	112.5
7	10	0	M	Yes	No	Right	62	103.6	Left	82.2	107.2
8	9	0	F	Yes	Yes	Right	68.1	115.4	Left	56.5	110
9	20	0	M	Yes	Yes	Left	25	110.5	Right	71.3	112.6
10	7	0	M	Yes	Yes	Right	47.9	95.75	Left	55.7	98.6
11	31	0	F	Yes	Yes	Right	34	42	Left	21.5	45.5
12	29	0	M	Yes	No	Left	18	97	Right	72	97
13	1	6	M	No	No	Right	63	12	Left	86	13
14	27	0	F	No	No	Left	93	24.6	Right	130.5	18.8
15	30	0	M	No	No	Right	78.5	31.6	Left	97	28.8
16	27	0	F	No	No	Left	85.5	8.75	Right	49.5	10.5
17	24	0	F	Yes	No	Right	29	65.5	Left	25.5	64.5
18	28	0	M	Yes	No	Left	55	38.2	Right	115	33.3
19	26	0	F	Yes	No	Right	77	110.2	Left	91	109
20	4	0	F	Yes	No	Right	31	106.33	Left	47	106.5
21	4	0	M	No	No	Left	85	48.2	Right	93	47.6
22	7	0	M	Yes	No	Right	64	103.6	Left	52	101.4
23	15	0	M	Yes	No	Right	69.5	102.25	Left	46	102.5
24	40	0	M	Yes	Yes	Right	76	101.6	Left	30.5	93
25	22	0	F	Yes	No	Left	60	97.2	Right	83	98.2
26	50	0	F	Yes	Yes	Right	45	55.3	Left	31.5	52.33
27	30	0	F	Yes	No	Right	28.5	91	Left	17	101
28	27	0	F	Yes	No	Left	20.5	108.5	Right	50	113
29	5	0	F	No	No	Right	30	31	Left	63.5	34.3
30	0	2	F	Yes	No	Left	86	88.2	Right	119.5	86.8
31	0	6	F	Yes	No	Right	34	94.66	Left	24	101.5
32	12	0	M	Yes	No	Right	76.5	90.2	Left	81	88.8
33	20	0	M	Yes	Yes	Left	67	94.4	Right	106.5	101
34	19	0	M	Yes	Yes	Right	90.5	80.6	Left	128.5	90.4
35	32	0	M	Yes	Yes	Left	33.5	50.33	Right	22.5	56
36	29	0	F	No	No	Right	72.5	7	Left	95.5	8
37	43	0	F	No	No	Left	41	13.75	Right	32	13
38	33	0	M	No	No	Right	94	24.4	Left	116	31
39	21	0	F	No	No	Left	54.5	5.3	Right	87	6.67
40	26	0	M	No	No	Left	52	47.66	Right	108.5	47
41	30	0	M	No	No	Left	63	12.33	Right	129.5	15.66
42	24	0	F	No	No	Right	94.5	9.33	Left	120.5	11.66
43	26	0	M	No	No	Right	94.5	49.5	Left	64	49.5
44	23	0	M	No	No	Left	55	14.5	Right	60.5	14
45	28	0	F	No	No	Right	68.5	13.5	Left	41.5	14
46	29	0	M	No	No	Right	61	12	Left	86.5	13.5
47	19	0	M	No	No	Left	58.5	12	Right	39	14.5
48	49	0	M	Yes	No	Left	43.7	92	Right	118.8	90
49	38	0	M	Yes	No	Right	14.2	115	Left	121.6	108
50	0	5	F	Yes	No	Right	52	87.8	Left	57.5	91
51	18	0	F	Yes	Yes	Left	70.5	56.8	Right	67.5	63.2

Movie S1. Demonstration of sweat sticker removal.

Demonstration of sweat sticker removal from the arm at the conclusion of sweat test. The adhesive is robust in the presence of sweat, but facile to remove. Removal does not result in loss of sweat from the collection chambers. The adhesive occludes the subjacent sweat glands resulting in compensatory sweating(42). Although compensatory sweating occurs in the regions of skin covered by the adhesive, this excretion of sweat from surrounding skin pores does not affect sweat collection by the device.