

# Harbor porpoise losing its edges: genetic time series suggests a rapid population decline in Iberian waters over the last 30 years

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1	Harbor porpoise losing its edge: genetic time series suggests a rapid
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#### Abstract

39 Impact of climate change is expected to be especially noticeable at the edges of a species' 40 distribution, where they meet sub-optimal habitat conditions. In Mauritania and Iberia, two 41 genetically differentiated populations of harbor porpoises (Phocoena phocoena) form an 42 ecotype adapted to local upwelling conditions and distinct from other ecotypes further north 43 on the NE Atlantic continental shelf and in the Black Sea. By analyzing the evolution of 44 mitochondrial genetic variation in the Iberian population between two temporal cohorts (1990-45 2002 vs. 2012-2015), we report a substantial decrease in genetic diversity. Phylogenetic 46 analyses including neighboring populations identified two porpoises in southern Iberia 47 carrying a divergent haplotype closely related to those from the Mauritanian population, yet 48 forming a distinct lineage. This suggests that Iberian porpoises may not be as isolated as 49 previously thought, indicating possible dispersion from Mauritania or an unknown population 50 in between, but none from the northern ecotype. Demo-genetic scenario testing by approximate 51 Bayesian computation showed that the rapid decline in the Iberian mitochondrial diversity was 52 not simply due to the genetic drift of a small population, but models support instead a 53 substantial decline in effective population size, possibly resulting from environmental 54 stochasticity, prey depletion, or acute fishery bycatches. These results illustrate the value of 55 genetics time series to inform demographic trends and emphasize the urgent need for 56 conservation measures to ensure the viability of this small harbor porpoise population in 57 Iberian waters.

58

59 **KEYWORDS**: cetacean conservation genetics, genetic time series, genetic diversity,

60 by catch, approximate Bayesian computation, population genetic modelling

#### 61 **1. Introduction**

The impact of climate change on species is expected to be especially noticeable at their distribution edges, where species meet the limits of their ecological tolerance. Marginal populations of temperate species at the rear (warm) distribution edge are often considered more susceptible to warming than central populations because of the warmer ambient temperatures they experience, but this overlooks the potential for local adaptation (Bennett et al., 2015; Vilà-Cabrera et al., 2019).

68 Cetaceans are particularly vulnerable to climate change (MacLeod, 2009) due to their crucial function as apex predators in marine ecosystems (Bowen, 1997). Climate change 69 70 influences the availability of their prey and modifies the habitat, thereby impacting several (if 71 not all) cetacean species and may even put some of them at a high risk of extinction (Learmonth 72 et al., 2006). As a response, several cetacean species are shifting their distribution ranges to 73 track suitable habitats or adapt locally by switching to different food resources (Lambert et al., 74 2011; Learmonth et al., 2006; MacLeod, 2009; Williamson et al., 2021). For example, 75 temperate and subpolar small cetaceans like the harbor porpoises (Phocoena phocoena) are 76 likely to show a poleward shift (Heide-Jørgensen et al., 2011; MacLeod, 2009). Due to an 77 elevated metabolic rate and relatively short generation time, this small cetacean is heavily 78 dependent on a continuous food supply (Hoekendijk et al., 2018; Wisniewska et al., 2016). 79 This species is thus expected to be particularly sensitive to climate changes (Heide-Jørgensen 80 et al., 2011). In addition, due to their coastal distribution, thousands of harbor porpoises are 81 also killed accidentally each year by commercial fisheries. This by-catch mortality is 82 threatening local populations to a level that is still hard to quantify, but it is likely unsustainable, 83 especially with a highly fluctuating environment (Carlén et al., 2021; IMR-NAMMCO, 2019; 84 Vingada & Eira, 2018).

85 Harbor porpoises are officially subdivided into three distinct subspecies: P. p. vomerina 86 in the Pacific, P. p. phocoena in the North Atlantic, and P. p. relicta in the Black Sea (Fontaine, 87 2016; Read, 1999; Rosel et al., 1995). A fourth lineage or Evolutionary Significant Unit (ESU) 88 has been suggested for the Afro-Iberian harbor porpoises (IBMA; currently unnamed sub-89 species, but possibly P. p. meridionalis following Fontaine et al. 2014) owing to their 90 distinctive ecology, morphology (Donovan & Bjørge, 1995; Smeenk et al., 1992), and genetic 91 divergence (Fontaine et al., 2014) (Figure 1). However, no formal description or subspecies 92 taxonomic assignment has been made for this distinct lineage so far (Pierce et al., 2020). IBMA 93 porpoises are found around the relatively cold and productive upwelling waters off the coasts 94 of Iberia and on the NW Atlantic coasts of Africa from southern Morocco, to Mauritania, and 95 Senegal (Fontaine, 2016; Gaskin, 1984; IMR-NAMMCO, 2019; Read, 1999). They are 96 significantly larger in term of skull morphology, growth curve, and adult body size (typically 97  $\geq$ 2m long) compared to the two other ecotypes (or subspecies) (~1.5m long) (Galatius & 98 Gol'din, 2011; Read, 1999; Sequeira, 1996; Smeenk et al., 1992). Beside their distinctive 99 morphological features, the IBMA porpoises belong to the Eastern Canary Coastal province 100 with equatorward surface circulation in the form of the Canary current, rich in eddies and large 101 upwellings that are permanent along the NW African coast and seasonal along the Iberian coast 102 (Álvarez-Salgado et al., 2003; Fraga, 1981; Moustahfid et al., 2021; Pierce et al., in revision). 103 While P. p. phocoena in the northern Bay of Biscay and further north in the North Atlantic are 104 generally considered opportunistic feeder relying primarily on bottom dwelling on the shallow 105 continental shelf (i.e. in waters of <200 m depth), the IBMA porpoise feeding ecology is still 106 poorly known. It is hypothesized that they exhibit similar foraging behaviors as their 107 counterparts on the continental shelf to the north, but with a greater emphasis on pelagic fish (Pierce et al., in revision; Pinela et al., 2010; Santos & Pierce, 2003). From a genetic 108 109 perspective, the Iberian and Mauritanian porpoises are closely related but they were found genetically differentiated from each other and form two distinct populations separated by at
least 2500 km (Ben Chehida et al., 2021; Fontaine, 2016; Fontaine et al., 2014).

112 The upwelling habitat of the Iberian porpoise (IB) population is at the border of a 113 biogeographical transition zone between temperate and subtropical waters where species with 114 different affinities co-occur and reach their respective ecological limits (Bowen, 1997; 115 MacLeod, 2009). Because species' habitat suitability at the margins between biogeographic 116 zones are often suboptimal, populations like the Iberian harbor porpoises are expected to be 117 among the first to show the impacts of environmental changes. This is particularly true with 118 the small-scale Iberian upwelling system, which is known to fluctuate in response to ongoing 119 climate change (Casabella et al., 2014; Pires et al., 2013). Such environmental fluctuation may 120 impact the Iberian harbor porpoises in various ways, possibly leading to dramatic impacts on 121 the population demography and genetic diversity. Porpoises may adapt locally, for example, 122 by adapting to various prey species (Heide-Jørgensen et al., 2011), by migrating to more 123 suitable environments, or the population may also become extirpated if environmental changes 124 outpace their ability to adapt. Previous population genetic studies (Ben Chehida et al., 2021; 125 Fontaine et al., 2014; Fontaine et al., 2017; Fontaine et al., 2010), based on samples collected 126 between 1990 and 2002, showed that the Iberian population was a source population with highly asymmetric geneflow, sending more migrants to the North into the P. p. phocoena 127 128 populations on the northern side of the Bay of Biscay and to the South into the Mauritanian 129 population than in the other direction (Figure 1). In fact, a tension zone was detected in the 130 northern side of the Bay of Biscay with Iberian genetic ancestry decreasing sharply with the 131 increasing latitude (Fontaine et al., 2007; Fontaine et al., 2014; Fontaine et al., 2017). If 132 recruitment does not keep up with the mortality and migration rate, the census size of the 133 Iberian population may decrease, leading to a gradual reduction in number of reproducing 134 individuals, in genetic diversity, and thus in effective population size over time.

135 Unlike previous studies, which inferred population structure between the Iberian 136 population and other populations (Ben Chehida et al., 2021; Fontaine et al., 2014; Fontaine et 137 al., 2017; Fontaine et al., 2010), the goal of the present study is to assess how mtDNA variation 138 evolved over time in the Iberian population, comparing sampling cohorts spanning about 3 139 decades and how this information can guide conservation of the Iberian population. More 140 precisely, using genetic diversity as a proxy for effective population size variation, we tested 141 if mitochondrial genetic diversity decreased through time by comparing the genetic 142 composition of two cohorts of porpoises, one sampled in the 1990's (1990-2002), and the other 143 one from the 2010's (2012-2015).

- 144
- 145 **2. Material and methods**

146 Sampling and molecular analyses. Tissue samples from 52 by-caught or stranded 147 individuals collected along the Portuguese coast between 2012 and 2015 were obtained from 148 the Portuguese Marine Animal Tissue Bank (MATB). These samples form the 2010's cohort 149 of the Iberian population (also called the 'new' cohort or IB N in this study) (Figure 1, Table 150 1 and S1). Tissues samples were stored in 70% ethanol and kept at -20°C until molecular 151 analyses. Genomic DNA was extracted with the DNeasy Blood and Tissue kit (QIAGEN Inc.) 152 following the manufacturer's protocol. Following Fontaine et al. (2014), we screened five mitochondrial coding regions (ATP-6, ATP-8, Cyt-b, ND5, and COI) for genetic 153 154 polymorphisms using Sanger sequencing. Target regions were PCR amplified, quality 155 controlled, and purified following the protocol described in Fontaine et al. (2014). Sanger 156 sequencing was outsourced to GATC Biotech ltd. The resulting forward and reverse sequences 157 were visually inspected, cleaned, and assembled with Geneious 10.0.9 (Kearse et al., 2012). 158 We augmented the dataset with the 82 previously published sequences from Fontaine et al. 159 (2014) encompassing the same five mitochondrial genes. These sequences were obtained for 160 82 harbor porpoises collected between 1990 and 2002 (Figure 1, Table 1 and S1). These 161 included: 14 individuals collected along the French coast of the Bay of Biscay (BB), 23 further 162 north in the North Atlantic (NAT) from the Channel to the Nordic waters of Norway and 163 Iceland; 19 from the Iberian coast (a.k.a. the `old` Iberian cohort or IB O), 14 from Mauritania 164 (MA); and 12 from the Black Sea (BS). The 'old' Iberian cohort from Fontaine et al. (2014) 165 obtained between 1990 and 2002 included 19 stranded and bycaught porpoises from the 166 Galician coasts of Spain and Portugal (Figure 1 and Table S1). Previous studies based on 167 mtDNA data and nuclear microsatellite data demonstrated that the Spanish and Portuguese 168 samples were part of a same panmictic genetic group (Ben Chehida et al., 2021; Fontaine et 169 al., 2007; Fontaine et al., 2014; Fontaine et al., 2017). The main focus of the present study thus 170 consists in comparing the temporal evolution of the mtDNA variation between the two Iberian 171 cohorts of porpoises: *i.e.* the 'new' [2010's: 2012-2015, n=52] versus the 'old' [1990's:1990-172 2002, n=19]). The temporal coverage of this serial sampling of the Iberian population is thus 173 at most 25 years, which corresponds to 2.5 generations, assuming a generation time of 10 years 174 (Read, 1999). The mtDNA sequences from the other populations of Fontaine et al. (2014) 175 neighboring the Iberian population were included here to provide a broader perspective to 176 interpret the phylogenetic analyses.

177 One additional sequence from the closest outgroup species, the Dall's porpoise 178 (*Phocoenoides dalli*) also generated in Fontaine et al. (2014), was included in the present 179 dataset to root the mitochondrial phylogeny. All mtDNA sequences (n=135) were aligned with 180 *MUSCLE* (Edgar, 2004).

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182 Phylogenetic and population genetic analyses. In order to place the Iberian porpoises into 183 a global phylogeographic context, we first reconstructed a maximum-likelihood (ML) 184 phylogeny with all the 135 mtDNA sequences (including the outgroup). Unique mtDNA 185 haplotypes were identified using DnaSP v.5.10.1 (Librado & Rozas, 2009). Then, we used 186 *PhyML* (Guindon et al., 2010) implemented in *Geneious* to estimate the ML tree based on all 187 available unique haplotypes identified in the dataset using a nearest neighbor interchange 188 topology search. We used the Tamura and Nei (1993)'s (or TN93) model of nucleotide 189 evolution as identified as the best-fitting model for our sequence alignment using the Akaike 190 Information Criterion (AIC) in jModelTest 2.1.10 (Darriba et al., 2012). Each node's support 191 was evaluated by conducting 5,000 iterations of bootstrap resampling. The phylogenetic tree 192 was rooted with the sequence from a Dall's porpoise. In addition to the phylogenetic trees, we 193 also reconstructed a median-joining haplotype network (Bandelt et al. 1999) using PopART 194 (http://popart.otago.ac.nz).

195 Various statistics informative on the mtDNA genetic variation of the two cohorts of 196 Iberian porpoises ("old" vs. "new") were estimated using the C library libdiversity 197 (https://bioinfo.mnhn.fr/abi/people/achaz/cgi-bin/neutralitytst.c) and the R package pegas 198 v.1.2 (Paradis, 2010) (Table 1). Genetic diversity was estimated using the number of 199 haplotypes (H), haplotype diversity (Hd), nucleotide diversity ( $\pi$ ), and Watterson's theta ( $\theta_W$ ). 200 Differences in sample size can significantly impact the values of some genetic diversity 201 estimators (Goodall-Copestake et al., 2012). Therefore, a rarefaction approach was applied to 202 estimate measures of genetic diversity while correcting for difference in sample size (Sanders, 203 1968) using a custom python script. More precisely, for the two cohorts of Iberian porpoises, 204 for each statistic and each sample size increment k, we randomly subsampled 5,000 times with 205 replacement a number of k sequences available in each group, with k ranging between 2 and the  $k_{max}$  available sequences (up to a maximum of 50). At each sample size increment k, we 206 207 estimated the mean and standard error values per statistic and per group. Subsequently, genetic 208 differentiation between the two cohorts was assessed using the Hudson's estimator of  $F_{ST}$ (Hudson et al., 1992) and  $\varphi_{ST}$  (Excoffier et al., 1992) using respectively the libdiversity C-209

210 *library* and the R-package *haplotypes v.1.1.3.1*. The significance levels of  $F_{ST}$  and  $\varphi_{ST}$  were 211 assessed by randomization tests (1,000 permutations). Finally, we investigated evidence of 212 changes in mtDNA effective population size (*Ne*) between the two cohorts (old vs. new) by 213 calculating values of Tajima's *D* (Tajima, 1989) and Achaz's *Y* (Achaz, 2008) accounting for 214 differences in sample sizes by implementing the same rarefaction approach as described for 215 the diversity statistics. The Achaz's *Y* statistic is analogous to the *Tajima's D*, but accounting 216 for potential sequencing errors (see details in Achaz, 2008).

217

218 Approximate Bayesian Computation (ABC). In order to evaluate the support for different 219 demographic scenarios and assess whether there is a signal of a recent population decline in 220 the Iberian population based on the mtDNA data, we applied an ABC approach relying on 221 coalescent simulations (Lopes & Beaumont, 2010). More precisely, we compared six 222 demographic scenarios: (1) a population with a large constant effective population size, (2) a 223 small constant effective population size, (3) an old expansion ( $\geq 150$  generations, gen.), (4) an 224 old decline (>150 gen.), (5) a recent decline (<3 gen.), and (6) an old expansion (>150 gen.) 225 followed by a recent decline scenario ( $\leq 3$  gen.) (see Figure 2 and Table S2 for details). In the 226 coalescent simulations, we leverage the time series data information available and specified explicitly at which time the samplings of the two cohorts were performed. 227

For each of the six scenarios, we ran 10,000 coalescent simulations using *fastsimcoal2 v2.6* (Excoffier & Foll, 2011). Details on the coalescent simulations and the mutation model used are provided in the supplementary **Table S2**. We used the program *Arlsumstat v3.5.2* (Excoffier & Lischer, 2010) to compute a set of 55 summary statistics listed in **Table S3**, augmented with the principal axes obtained from a linear discriminant analysis (LDA) on the summary statistics (Pudlo et al., 2016), to describe and summarize the genetic variation of each simulation. 235 We employed a random forest classification (RF) procedure for the model selection (Pudlo 236 et al., 2016). RF is a machine-learning algorithm that uses bootstrapped decision trees to 237 classify and select the best scenario using the summary statistics as a set of predictor variables 238 (Breiman, 2001). RF was therefore applied to compare the likelihood of the six competing 239 scenarios (Figure 2). This procedure provides classification votes that accounts for the number 240 of times a scenario is selected as the best one among the trees used to constructed the random 241 forest. Based on a forest of a 1,000 decision trees, the scenario that gathered the highest number 242 of votes was selected as the most likely.

The *prior* error rate and the type-II error rate were estimated using the out-of-bag simulations (i.e. simulations not used to build the decision trees). In addition, we computed the confusion matrix, which provides a more global assessment of the performance of our RF procedure (Schrider & Kern, 2018). The convergence of the results was evaluated over 10 independent RF runs, as recommended by Fraimout et al. (2017).

248 Finally, to check whether the formulated scenarios and associated priors were compatible 249 with the observed dataset, we plotted both the simulated datasets from the training set and the 250 observed dataset on the two first principal axes of a Linear Discriminant Analysis (LDA) and 251 of a Principal Component Analysis (PCA), following the recommendation of Pudlo et al. (2016). We visually checked whether the summary statistics from the observed data was 252 253 located within the clouds formed by those from the simulated data for at least one demographic 254 scenario. In addition, we assessed the goodness-of-fit between the simulated and the observed 255 data following Gelman et al. (2004): the summary statistics for which the cumulative simulated 256 distribution was larger or smaller than 95% CI of the distribution compared to the observed 257 value were considered as significantly different from the observed data. All ABC-RF analyses 258 were performed using the R package *abcrf* v.1.8.1 (Pudlo et al., 2016) in R 4.1.2 (R Core Team, 259 2016).

260

### 261 **3. Results and Discussion**

In total, we successfully sequenced 52 Iberian harbor porpoises from the new cohort 262 263 (2012-2015) for fragments of five mitochondrial genes including Cytochrome b (Cyt-b: 1,111 264 base-pairs, bp), and ATP-6 and -8 (842bp), NADH dehydrogenase 5 (ND5: 707bp), and the 265 Cytochrome c oxidase subunit I (COI: 1,515 bp), When aligned with the 82 previous sequences 266 of harbor porpoises from Fontaine et al. (2014) and an additional one of the Dall's porpoise 267 used here as outgroup, the final alignment of 4,175 base-pairs (pb) and included 135 sequences 268 and contained 384 segregating sites with 267 singletons and 117 parsimony informative sites 269 defining 61 distinct haplotypes. Without the outgroup, the 134 mtDNA sequences of harbor 270 porpoise contained 170 segregating sites with 65 singletons and 105 parsimony informative 271 sites defining 60 distinct haplotypes (Table 1).

272 Consistent with previous studies, the phylogenetic relationships among mtDNA 273 haplotypes depicted by the ML tree (Figure 3 and S1) and by the haplotype network (Figure 274 S2) revealed the three main monophyletic lineages corresponding to the three previously 275 identified subspecies or ESU's (Fontaine et al., 2014): (i) P. p. relicta in the Black Sea; (ii) P. 276 p. phocoena composed of the porpoises from the North Sea to the waters of Norway; and (iii) 277 the Afro-Iberian (IBMA) porpoises (unnamed ssp. possibly P. p. meridionalis), which 278 comprises two distinct monophyletic sub-lineages in the upwelling waters of Iberia and 279 Mauritania, respectively.

Evidence of migration from the Iberian to the Mauritanian populations has been previously documented (Ben Chehida et al., 2021; Fontaine et al., 2014) and is also observed here with two haplotypes (Hap\_55 and Hap\_59) found in the Mauritian population, but clustering with the Iberian lineage (Figure 3, S1 and S2). However, we also identified one haplotype (Hap\_10 or Haplotype 10, see Figure 3, S1 and S2) present in two porpoises from the new Iberian 285 cohort, but clustering with the Mauritanian lineage. This haplotype was carried by two 286 individuals in the southernmost part of the Iberian sampling (red arrow in Figure 1). This result 287 provides the first evidence of possible dispersal (or migration) from the Mauritanian population 288 into Iberian waters, suggesting that these two populations may not be as isolated as previously 289 thought. Haplotype 10 could also belong to a distinct unknown population as it is relatively 290 divergent from the other Mauritanian haplotypes (Figure 3, S1 and S2). Indeed, the ML tree 291 (Figure 3 and S1) shows that this haplotype forms a strongly supported (bootstrap support 292 >70%) distinct lineage from the other Mauritanian porpoises. The haplotype network also 293 illustrates this divergence (Figure S2): while the Mauritanian haplotypes are separated from 294 each other with at most two mutational steps, at least ten mutational steps separate this 295 haplotype 10 from the other Mauritanian haplotypes (Figure S2). Notably, both porpoises 296 carrying this haplotype 10 were found on the southern extremity of the Portuguese coasts 297 (8.78W-37.07N; Praia do Burgau, Portugal), at least 400km south from all other Iberian 298 sampling locations (Figure 1). Stranding records of harbor porpoises were previously reported 299 in Morocco, on the east coasts of Gibraltar, and in Cadiz (Spain; Rojo-Nieto et al., 2011). If 300 unsampled populations occur between the Iberian Peninsula and the Mauritanian waters, they 301 are most likely confined to a narrow and seasonal upwelling area. This relatively divergent haplotype 10 closely related to the Mauritanian rather than the Iberian porpoises could be 302 303 evidence of such unsampled populations. This hypothesis of such unsampled population could 304 be plausible since harbor porpoise sightings, stranding, and bycatches have been reported from 305 the North of Morocco to the Gambia (Boisseau et al., 2010; IMR-NAMMCO, 2019). Haplotype 10 was excluded from subsequent analyses of genetic diversity and coalescent 306 307 simulations because it could be part of a distinct population and therefore would artificially 308 inflate measures of genetic diversity (Table 1 and S1). Future studies with additional samples 309 from these relatively unsampled regions (southern Iberian coast and Morocco) and data from 310 the nuclear genome are needed to confirm whether these two porpoises are genetically distinct 311 from the other Iberian porpoises, possibly forming a distinct population, and whether or not 312 they are admixing with the other Iberian porpoises.

313 Aside from the two porpoises carrying the haplotype 10, our results indicate that both 314 cohorts sampled at different times along the Iberian coasts belong to the same mtDNA gene 315 pool. We detected no genetic differentiation between the two Iberian cohorts suggesting no 316 significant differences in haplotype frequencies between them ( $F_{ST}$  and  $\varphi_{ST}$  values = 0; *p*-value 317 > 0.54, Table S4). This result reflects the high prevalence of haplotype 4 which dominates in 318 both cohorts (Figure 3, S1 and S2). However, once we accounted for differences in sample 319 size between the two cohorts using a rarefaction method, we observed a significant reduction 320 in genetic diversity in the new cohort compared to the old one as shown by the lower number 321 of haplotypes (*H*; Figure 4a), and in the two measures of nucleotide diversity ( $\pi$  and  $\theta_W$ ; Figure 322 4b and 4c, respectively). Rarefaction curves provided striking evidence of this reduction in 323 genetic diversity (Figure 4). The non-overlapping standard errors from a standardized sample 324 size beyond eight sequences suggest that the decline of genetic variation is likely highly 325 significant. For example, the average number of haplotypes (H) observed from 19 sampled 326 sequences in each cohort decreases from 10 to 6 when comparing the old vs. new cohorts. This suggests that the proportion of rare haplotypes has dropped substantially over the 25-years 327 328 sampling period of this study. Consistent with this observation is the rapid loss in nucleotide 329 genetic diversity as measured by  $\pi$  and  $\theta_W$  (Figure 4b and 4c). Such a substantial loss of genetic 330 variation over 2.5 generations is often attributed to an enhanced effect of genetic drift rather 331 than other evolutionary forces like selection (Hartl, 2020). This could suggest a dramatic 332 decline of the Iberian population in term of effective population size, possibly reflecting a 333 substantial decline in the number of reproducing individuals.

334 Tajima's D (Tajima, 1989) and Achaz's Y (Achaz, 2008) statistics estimated for the two 335 cohorts sampled at different times along the Iberian coast also displayed negative values 336 (Figure 5). At first glance, such negative values would be indicative of a population that 337 underwent a "recent" expansion at evolutionary time-scales (Epps & Keyghobadi, 2015; Smith 338 et al., 2011), most likely due to the foundation of the Iberian population during the period that 339 followed the Last Glacial Maximum (19kyr ago; Fontaine et al. 2014; Fontaine 2016). 340 However, the values estimated for the new cohort for both statistics were significantly less 341 negative (based on non-overlapped standard error estimates) than those from the old cohort. 342 For example, the Tajima's D values obtained from 19 randomly sampled sequences increased 343 from -1.5 for the old cohort to -1.25 for the new (Figure 5a). The change in Achaz's Y values, 344 which account for possible sequencing errors (Achaz, 2008), show an even stronger increase 345 from -1.75 to -0.95 between the old and new cohorts (Figure 5b). It is remarkable to see how 346 both values have become dramatically less negative in the time lapse of only 2.5 generations 347 between the two cohorts. This increase in D and Y values also provides a strong indication of 348 a population losing rare haplotypes as it would be expected for a small and isolated population 349 strongly subject to genetic drift that could be declining.

350 Such a loss of genetic diversity may reflect different demographic scenarios, for example 351 a small population that did not experience any population size changes, but still experiencing 352 strong genetic drift. Alternatively, a scenario of ancient expansion and a very recent decline 353 could be also possible. We used a simulation-based Approximate Bayesian computation 354 analysis relying on the Random-Forest classifier (ABC-RF) to explored statistically six 355 plausible demographic scenarios (Figure 2) describing the evolutionary history of the Iberian 356 population since it diverged from the Mauritanian population within the last 150 generations 357 (Fontaine et al., 2014). Out of the six scenarios compared, the analysis strongly supported the 358 SC6 scenario assuming an "old" expansion (>150 generations) followed by recent decline with 359 the past 3 generations (SC6; Figure 2, S3 to S5). Ten replicates of the ABC-RF analysis 360 supported strongly and unanimously this result (Figure S3). Scenario SC6 gathered in average 361  $(\pm$  SD) over the ten runs 56.6  $\pm$  1.3% of the RF votes (Figure 2 and S3), with a posterior 362 probability of 66.3  $\pm$  2.5%, a *prior* error rate of 9.5  $\pm$  0.0%, and type II errors  $\leq$  3.1%  $\pm$  0.0%. 363 Furthermore, the confusion matrix (Figure S5) depicting the overall performance of the RF 364 classification analysis indicated high power to distinguish between the competing scenarios 365 (>90%). Such a high-resolution power to discriminate among the different demographic scenarios likely comes from the multiple sampling time points included in the ABC analysis. 366 367 The goodness-of-fit analysis provided additional support to the demographic model identified 368 by showing that scenario SC6 was the only scenario that produced simulated values that closely 369 matched the observed values for each summary statistics used in this analysis (Table S5, 370 Figure S4 and S6). This scenario SC6 (Figure 2) suggests that the effective size (*Ne*) of the 371 Iberian population expanded at least ~150 generations ago, thus ~1500 years ago, assuming a rough generation time of 10 years (Fontaine et al., 2014). Then, under this scenario, Ne values 372 373 of the Iberian population would have declined by a factor > 10 within the last three generations 374 (thus 30 years). Thus, this ABC analysis, together with the rarefaction results, adds an 375 additional line of evidence suggesting a rapid loss of genetic variation over the last three generations due to a very recent decline in effective population size. The reader should note 376 377 however that the effective population size (Ne) is a population genetic concept that differs from 378 and is usually smaller than the *census* population size. It does not represent the number of 379 individuals actually alive at any given time point. Following Frankham (2019), Ne is the 380 number of individuals that would result in the same loss of genetic diversity, inbreeding, 381 genetic drift, etc., if they behaved in the manner of an idealized population. Such idealized 382 population would assume non-overlapping generations, random mating, and no migration, 383 mutation or selection. Ne would thus roughly represent the number of breeding individuals in this idealized population, or more precisely the number of breeding females since we analyzed here maternally inherited fragments of the mitogenome that behave as a single nonrecombining haploid unit. Therefore, our results are not intended to estimate exactly how many Iberian porpoises there were at different points in history, but rather they suggest that the number of reproducing females may have been substantially declining in the Iberian population over the last 2.5 generations. However, such interpretation must be taken with caution considering all the assumptions related to the *Ne* concept.

391 In this study, we also confirmed previous evidence of gene flow from the Iberian 392 population to more northern populations with Iberian haplotypes present in the Bay of Biscay 393 and in the English Channel where admixture between the two lineages, IBMA (or P. p. 394 meridionalis) and P. p. phocoena, has been reported (see Figure 3, S1 and S2; Table S4, and 395 also Ben Chehida et al., 2021; Fontaine et al., 2014; Fontaine et al., 2017). Additionally, we 396 did not find any evidence of gene flow from the populations located further North into the 397 Iberian population, as previously reported (Ben Chehida et al., 2021; Fontaine et al., 2007; 398 Fontaine et al., 2014; Fontaine et al., 2017; Fontaine et al., 2010). Indeed, Fontaine et al. (2014) 399 estimated very asymmetric values of gene flow using the coalescent approach of MIGRATE 400 (Beerli, 2006; Beerli, 2009) based on microsatellite data. While no evidence of immigration 401 was detected toward the Iberian porpoises from the neighboring populations, ca. 3 individuals 402 per generation were estimated migrating from the Iberian to the Mauritanian populations, and 70 per generation from the Iberian to the northern Bay of Biscay populations (see table 2 in 403 404 Fontaine et al., 2014). In the northern Bay of Biscay and adjacent waters, a geographically 405 restricted admixture zone was reported forming a well-delimited tension zone (Fontaine et al., 406 2017). Because the population size of *P. p. phocoena* in the North Sea and adjacent waters is 407 much larger than that of the Iberian population (census N estimates from the SCANS-III survey 408 (Hammond et al., 2017): ~400,000 vs.  $\leq$  3,000 individuals respectively), the observed 409 northward migration may possibly lead to the assimilation of the Iberian gene pool by P. p. 410 phocoena. In the long term, this process could be an additional cause of concern as it could 411 result in the disappearance of the unique Iberian genetic pool. This scenario is advanced by 412 Macleod et al. (2009), which used habitat modeling to show that species with a similar range 413 as the Iberian harbor porpoises (Figure 1C in MacLeod, 2009) may shift their range northward 414 in response to a 5°C increase in water temperature due to global warming (Figure 1D in 415 MacLeod, 2009). This model suggests that the future range of IBMA could completely overlap 416 with the current range of *P. p. phocoena* (Figure 1D in MacLeod, 2009). It is worth mentioning 417 that Ben Chehida et al. (2021) recently modeled the future distribution of the harbor porpoises 418 in the North Atlantic under the most aggressive scenario of the Intergovernmental Panel on 419 Climate Change (IPCC) in 2050 (RCP8.5; Schwalm et al., 2020) and did not capture this strong 420 northward range shift. The discrepancy between the two models may be related to differences 421 in assumed temperature increase, 2°C and 5°C, in Ben Chehida et al. (2021) and MacLeod 422 (2009), respectively. In addition, the environmental niche modeling in Ben Chehida et al. 423 (2021) did not account for any biotic factors in the simulations. However, environmental niche 424 modelling such as those conducted by MacLeod (2009) and Ben Chehida et al. (2021) needs 425 to be taken with some caution since they do not account for local population adaptation. In the 426 case of IBMA porpoises, their morphological and genetic distinctiveness together with the 427 peculiar upwelling habitat they inhabit suggest that they are adapted to the local conditions. 428 Future genomic studies could assess whether selective processes have impacted the genome of 429 IBMA porpoises and whether there is genetic evidence of such local adaptation process 430 compared to other populations.

431 The results of this study support the hypothesis that the Iberian population is both small 432 and isolated. Furthermore, both the sharp decline in mtDNA genetic diversity over the past 25 433 years and northward migration may represent reasons for concern for the long-term persistence 434 of the Iberian porpoises. One plausible explanation is that the decay of mtDNA genetic 435 variation we reported here could simply reflect the fact that Iberian porpoises constitute a stable 436 population with a very low effective population size (Ne) (SC2 in Figure 2). This scenario SC2 437 was not the best model according to our simulation-based ABC-RF analysis, but the third best 438 one, receiving only 16% of the total RF classification votes, but it remains plausible (Figure 439 2, S3 to S6, Table S5). In such a condition, genetic drift would have a disproportionate 440 influence on the genetic make-up of this population and could lead to the observed loss of 441 genetic variation over a few generations. Several previous studies showed that the Iberian 442 population has indeed a very low Ne at nuclear microsatellite loci (Ne < 100 individuals; Ben 443 Chehida et al., 2021; Fontaine et al., 2014; Fontaine et al., 2010). The mtDNA employed in 444 this study has an effective size four times smaller compared with nuclear autosomal loci. This 445 means that the stochastic loss of genetic variation due to genetic drift would be drastically 446 increased for mtDNA compared to nuclear loci. Thus, the Iberian population might be at a 447 stable census size losing rare haplotypes through the enhanced effect of genetic drift at the 448 mtDNA. Another more plausible explanation receiving the highest support in our ABC-RF 449 analysis (scenario SC6, Figure 2, S2 to S6, Table S5) is that the decay of mtDNA genetic 450 variation might be associated with a rapid and recent decline of the number of reproductive females in addition to the overall low Ne (Lande, 1988), as depicted by the scenarios SC6 in 451 452 Figure 2. This scenario SC6 support a severe population decline in the last three generations, 453 leaving at most 10% of the population pre-decline Ne. This hypothesis is even more plausible 454 since there is accumulating evidence that the Iberian population is sending more migrants to 455 adjacent populations than the reverse (Ben Chehida et al., 2021; Fontaine et al., 2014; Fontaine 456 et al., 2010). Thus, in addition to being small and isolated, the recent and strong population 457 decline envisioned under SC6 of the ABC-RF analysis would imply that the Iberian porpoises 458 may become increasingly subject to demographic stochasticity (Allendorf et al., 2012).

Because this population is also known to be subject to strong genetic drift and environmental stochasticity (Casabella et al., 2014; Pires et al., 2013), all these effects can accumulate and reinforce each other, potentially precipitating the decline of the Iberian porpoises. This socalled extinction vortex (Allendorf, et al., 2012) can be particularly severe for a long-lived Kstrategist species, inhabiting a marginal habitat highly subject to environmental variation like the Iberian harbor porpoises (Casabella et al., 2014; Pires et al., 2013) and could put them at a high risk of extinction.

466 The present study relies on a single molecular marker (mtDNA). Therefore, future studies 467 with additional samples and data from the nuclear genome are highly needed to confirm the 468 trends highlighted here. Indeed, genome-wide data can provide further insights into the 469 temporal dynamics of the Iberian population. For example, the reconstruction of historical 470 demography based on genome-wide data (Liu & Fu, 2020; Schiffels & Wang, 2020; Terhorst 471 et al., 2017) can be used to test whether these harbor porpoises maintained a long-term low Ne 472 (Morin et al., 2020; Robinson et al., 2022) since their post-glacial emergence (Fontaine, 2016; 473 Fontaine et al., 2014; Fontaine et al., 2010), or if on the contrary, the population declined 474 recently (Hu et al., 2020). A simulation framework, similar to the one used by Robinson et al. 475 (2022), could be applied to assess whether these porpoises are already facing deleterious effects of the mutation load, despite their current low Ne, or if purging processes have been ongoing 476 477 similar to those recently reported for the vaquita (Phocoena sinus) (Morin et al., 2020; 478 Robinson et al., 2022).

Additional data sources also point to large fluctuations in population size in the Iberian porpoises and require careful considerations: the SCANS-III survey in summer 2016 estimated a census size as low as 2,898 animals (CV=0.32) in Iberian waters and Bay of Biscay (Hammond et al., 2017). Annual aerial surveys in Portuguese continental waters between 2011 and 2015 estimated an overall average abundance of 2,254 porpoises and corresponding

484 density of 0.09 ind/km<sup>2</sup> (CV=21.99%) with large interannual fluctuations. The highest annual 485 abundance estimated for that period was 3,207 individuals (CV=38.14%) in 2013, followed by 486 a sharp decrease in 2014, when only 1,653 individuals (CV=43.27%) were estimated (Torres-487 Pereira et al., 2022). López et al. (2013) estimated an abundance of 683 (CV=0.63, 95%CI: 488 345-951) animals for the Spanish part of the IBMA range (Galicia and Bay of Biscay). 489 Globally, these low census sizes were associated with particularly high bycatches rate (always 490  $\geq$ 8%) with the best estimates varying from 90-197 deaths per year (Pierce et al., 2020) to 130-491 330 deaths per year (Read et al., 2020). However, it is important to mention that these estimates 492 are mainly based on "on-board observer" and aerial surveys data that deliver wide confidence 493 limits. This is due to low observer coverage of fishing activities in the region, low density of 494 porpoises, and estimates from various sources (observer data, strandings, aerial survey, and 495 interviews). Furthermore, Iberian harbor porpoises are regularly found stranded, often with 496 signs of bycatch mortality, potentially indicating unsustainable mortality rates (IMR-497 NAMMCO, 2019; López et al., 2002; Pierce et al., 2020; Read et al., 2020; Torres-Pereira et 498 al., 2022; Torres-Pereira et al., 2023a; Vingada & Eira, 2018). In fact, based on Portuguese 499 stranding data, an average of 9.19% (CI=5.25-16.10%) of the porpoise population was 500 estimated to be removed annually by fisheries between 2011 and 2015, corresponding to an 501 average of 207 porpoises removed per year, which largely surpasses the Potential Biological 502 Removal rate conservatively estimated for the same period (22 porpoises, CI: 12-43) (Torres-503 Pereira et al., 2023a). An additional cause for concern is related to potential overexploitation 504 of the feeding resources (Méndez-Fernandez et al., 2013; Santos & Pierce, 2003). A scarcity 505 in prey availability can be harmful because it enhances interspecific competition, as already 506 reported between harbor porpoises and other marine mammals, for example with the bottlenose 507 dolphin *Tursiops truncatus* whose ecological niche largely overlaps with the harbor porpoise 508 in some portions of Iberian waters (Méndez-Fernandez et al., 2013; Spitz et al., 2006). Hostile interactions between the two species have been documented in Iberian waters and hypothesized
to cause harbor porpoises to disperse to less productive narrow shelf areas (Alonso et al., 2000;
Spitz et al., 2006).

512

#### 513 Conclusions

514 The Afro-Iberian (IBMA) lineage of harbour porpoises was previously recognized as a 515 unique Evolutionarily Significant Unit (Crandall et al., 2000; Moritz, 2002) distinct from the 516 other sub-species devribed in the N. Atlantic and in the Black Sea. IBMA display unique 517 features in terms of genetics, morphology, ecology and evolutionary history following Fontaine 518 et al. (2014) (see also the review of Fontaine, 2016). Nevertheless, they are currently not 519 recognized as a distinct subspecies (Pierce et al., 2022; Pierce et al., 2020). There is also 520 currently no mention of these porpoises in the IUCN Red List of Threatened Species despite 521 the multiple threats faced by this vulnerable lineage. Given the results of this study, the 522 previous findings on their genetic distinctiveness from other populations at both mtDNA and 523 nuclear markers (Ben Chehida et al., 2021; Fontaine, 2016; Fontaine et al., 2007; Fontaine et 524 al., 2014; Fontaine et al., 2017; Fontaine et al., 2010), and the previous concerns raised about 525 their viability (IMR-NAMMCO, 2019; Pierce et al., 2022; Pierce et al., 2020), we recommend immediate actions and further research from the community in the following areas: (1) a formal 526 527 taxonomic description is needed for these IBMA porpoises to be officially recognized as a 528 distinct subspecies, following the guidelines of the International Commission on Zoological 529 Nomenclature as recommended by the Taxonomic Committee of the Society for Marine 530 Mammalogy; and (2) a better assessment for the Iberian porpoises in term of their life history 531 traits, ecology, behavior, genetics and genomics, and the threats to the viability for this unique This knowledge will help designing better conservation measures and 532 group. 533 recommendations. It is indeed paramount to ensure that all measures are taken to gain 534 additional knowledge and to apply effective conservation efforts to protect the Iberian 535 population, as recently underline by the OSPAR Marine Mammal Expert Group (OMMEG) 536 (ICES-WGMM, 2021; Pierce et al., 2022). Recently, both countries (Portugal and Spain) 537 updated the conservation status of the harbor porpoises inhabiting Iberian waters: from "Vulnerable" to "Endangered" in Spain (Boletin oficial del estado, 2020) and to "Critically 538 539 Endangered" in Portugal (Torres-Pereira et al., 2023b). In Portugal, a new Marine Protected 540 Area (PTCON0063) in the northwestern coast was recently designated whereas another one 541 was enlarged in the southwestern coast (PTCON00012) in order to incur higher protection for 542 harbor porpoise hotspots in the country (Vingada & Eira, 2018). Nonetheless, none of the 543 conservation and mitigation measures within the areas' management plan (Portaria 201/2019) 544 is currently in place and bycatch mortality remains effectively unaddressed. In the future, 545 improving our understanding of the demographic trends and the threats these porpoises are 546 facing is required to devise suitable and tailored larger scale conservation plans for the whole 547 Iberian population. Such measures are timely needed to mitigate the decline of this unique 548 lineage potentially on the brink of extinction.

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### 570 **Conflict of interest**

571 The authors declare that they have no conflict of interest.

573

#### 574 Data Availability

575 The data sets and scripts supporting this study are available in the IRD Porpoises Genetics and 576 Genomics Dataverse Repository (https://dataverse.ird.fr/dataverse/porpoise\_genomics) at 577 https://doi.org/10.23708/GOPRGV. Mitochondrial haplotypes were also deposited on the 578 NCBI–GenBank under the Accession Numbers (Accession numbers are listed in Table S1).

579

### 580 Author Contributions

- 581 Conceptualization: MCF. Data curation: MF, ATP, LN, CE, MCF. Formal analysis: YBC,
- 582 TS, MCF. Funding acquisition: MCF. Investigation: JT, TS, JPAH, MCF. Methodology: JT,
- 583 TS, JPAH, MCF. Project administration: MCF. Resources: JT, TS, JPAH, MCF. Software:
- 584 YBC. Supervision: MCF. Validation: MCF, YBC, JT. Visualization: TS, YBC, MCF.
- 585 Writing original draft: YBC, TS, MCF. Writing review & editing: YBC, MCF with inputs
  586 from all the co-authors.

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### **Tables and Figures**

**Table 1**: Mitochondrial genetic diversity. The statistics display per groups and overall include the mtDNA sample sizes (*N*), the number of segregating sites (*S*), singleton mutations (*Singl.*); sites informative in parsimony (*Parsim.*), number of haplotypes (*# Hap*), haplotype diversity (*Hd*), the nucleotide diversity ( $\pi$ ), Watterson's theta ( $\theta_W$ ), Tajima's *D*, *Y*, Achaz's *Y*. Population codes are as follow: BS=Black Sea; NAT=North Atlantic; BB=Bay of Biscay; MA=Mauritania; IB\_O=Iberian Old 1990's cohort; IB\_N= Iberian new 2010's cohort. Superscript next to each population code indicate the origin of the mtDNA sequence : <sup>1</sup>.Fontaine et al. (2014); <sup>2</sup>. Fontaine et al. (2014) + this study; 3. This study. See Table S1 for more details on the samples.

	Ν	S	Singl.	Parsim.	# Hap	Hd	π (%)	θw (%)	D	Y
BS <sup>1</sup>	12	23	21	2	10	0.96	0.10	0.18	-2.04	-1.56
NAT <sup>1</sup>	23	99	57	42	18	0.97	0.41	0.64	-1.46	-0.83
BB <sup>1</sup>	14	66	19	47	10	0.95	0.52	0.50	0.22	0.05
MA <sup>1</sup>	14	11	1	10	7	0.90	0.07	0.08	-0.66	-1.93
All IB <sup>2</sup>	71	27	7	20	17	0.72	0.05	0.13	-1.85	-2.12
IB_O <sup>1</sup>	19	12	6	6	10	0.74	0.05	0.08	-1.53	-1.77
IB_N <sup>3</sup>	52	24	5	19	11	0.70	0.06	0.13	-1.77	-2.19
IB_N* <sup>3</sup>	50	14	7	7	10	0.73	0.04	0.08	-1.58	-1.26
All	134	170	65	105	60	0.91	0.40	0.74	-	-

<sup>\*</sup>Excluding haplotype 10



**Figure 1.** Map showing the sampling locations and sample sizes. Geographic locations are based on approximate GPS coordinates or reported discovery locations. The red arrow indicates the geographical location of the two porpoises carrying Haplotype 10. (\* Fontaine et al. (2014); \*\* This study).



**Figure 2.** Alternative scenarios tested in the ABC-RF procedure. See **Table S2** for further details on the model parameters. Below each scenario are provided the main results including the number of Random Forest (RF) votes (in percent), the posterior probability of the best scenario, the prior-error rate, the type-II error rate, and the number of outlying statistics in the goodness-of-fit testing (**see Table S5**). The colored scenario, SC6, was the scenario gathering the highest number of Random Forest votes. The time scale is indicated by the arrow on the left, in generations before present (gen.).



**Figure 3.** Cladogram derived from the maximum-likelihood mitochondrial phylogeny among unique mitochondrial haplotypes. A branch length transformation proportional to the number of tips under the node (as implemented in *FigTree*) was applied to facilitate the visualization of the branching patterns (see Figure S1 for the actual phylogenetic tree with branch lengths proportional to sequence divergence). The color-coded labels show the geographic origin of the haplotype. The numbers within the boxes refer to the number of individuals carrying the haplotype. No number in a box means that the haplotype was observed only once. Population code are as follow: BS=Black Sea (N=12)\*; NAT=North Atlantic (N=23)\*; BB=Bay of Biscay (N=14)\*; MA=Mauritania (N=14)\*; IB\_O=Iberian Old (N=19)\*; IB\_N= Iberian new (N=52)\*\*. [\* Fontaine et al. 2014; \*\* This study].



**Figure 4.** Rarefaction curves showing three statistics describing mitochondrial genetic diversity including (a) number of haplotypes (*H*), (b) the nucleotide diversity ( $\pi$ ), and (c) Watterson's theta ( $\theta_W$ ). The red arrow in each panel shows the direction of change between the old and new cohorts. The X-axis shows the number of sequences randomly resampled in the rarefaction procedure to estimate the statistical values. The mean and standard error are provided at each sample size increment (*k*) for each statistic and each group, calculated from 5000 sampling with replacement of the *k* sequences available in each group, with *k* ranging from 3 to  $k_{max}$  up to 50.



**Figure 5.** Rarefaction curves showing two neutrality indexes indicative of demographic changes including (a) Tajima's D and (b) Achaz's Y. The red arrow inside each panel shows the direction of the change between the old and new cohorts. The bidirectional arrow shows the interpretation of D and Y in terms of demography. The X-axis shows the number of sequences randomly resampled in the rarefaction procedure to estimate the statistical values.