

Strong links between plant traits and microbial activities but different abiotic drivers in mountain grasslands

Sarah-sophie Weil, Camille Martinez-almoyna, Gabin Piton, Julien Renaud, Louise Boulangeat, Arnaud Foulquier, Amélie Saillard, Philippe Choler, Jérôme Poulenard, Tamara Münkemüller, et al.

▶ To cite this version:

Sarah-sophie Weil, Camille Martinez-almoyna, Gabin Piton, Julien Renaud, Louise Boulangeat, et al.. Strong links between plant traits and microbial activities but different abiotic drivers in mountain grasslands. Journal of Biogeography, 2021, 48 (11), pp.2755-2770. 10.1111/jbi.14235. hal-03409587

HAL Id: hal-03409587

https://hal.science/hal-03409587

Submitted on 28 Nov 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Strong links between plant traits and

microbial activities but different abiotic

drivers in mountain grasslands

Short running title: Plant-soil relationships in mountains

Names and affiliations of authors:

Sarah-Sophie Weil¹, Camille Martinez-Almoyna¹, Gabin Piton¹, Julien Renaud¹, Louise

Boulangeat¹, Arnaud Foulquier¹, Amélie Saillard¹, Philippe Choler¹, Jérôme Poulenard², The

ORCHAMP consortium, Tamara Münkemüller¹ and Wilfried Thuiller¹

¹Univ. Grenoble Alpes, Univ. Savoie Mont Blanc, CNRS, LECA, F-38000, Grenoble, France

²Univ. Savoie Mont Blanc, Univ. Grenoble Alpes, CNRS, EDYTEM, F-73000, Chambéry,

France

Correspondence: Sarah-Sophie Weil, Univ. Grenoble Alpes, Univ. Savoie Mont Blanc,

CNRS, LECA, F-38000, Grenoble, France; mail: sarah-sophie.weil@gmx.de

1

Acknowledgments

The research presented here belongs to the ORCHAMP monitoring programme. We thank the few dozens of interns, field assistants, park managers and rangers for their help conducting the field observations and the lab measurements. We thank Cindy Arnoldi for her help with the laboratory work on the AEEM technical platform. The S2M data were provided by Météo-France - CNRS, CNRM Centre d'Etudes de la Neige, through AERIS. We thank Samuel Morin (Centre d'Etudes de la Neige) for interactions with respect to meteorological data and indicators. We also thank Tristan Ubaldi for his help in the field and for his outreach work for ORCHAMP. Part of the funding came from the French Agence Nationale de la Recherche (ANR) through the GlobNets (ANR-16- CE02-0009) and (ANR-18-CE02-0010) projects, and from 'Investissement d'Avenir' grants (Trajectories: ANR-15-IDEX-02; Montane: OSUG@2020: ANR-10-LAB-56). We also received support from the METRO Grenoble, Conseil Départemental de l'Isère and the Jardin du Lautaret UAR 3370 (Univ. Grenoble Alpes, CNRS, Lautaret, 38000 Grenoble, France), member of AnaEE-France (ANR-11-INBS-0001AnaEE-Services, Investissements d'Avenir frame) and of the LTSER Zone Atelier Alpes, a member of the eLTER-Europe network. We thank the National Alpine Botanical Conservatory (CBNA) that carried out part of the vegetation surveys and that took part in the implementation of the flora protocols.

Statement of Significance

The tight links between plant and soil communities are drivers of biodiversity responses to environmental change. However, it is unclear how those links vary over biogeographic scales. Here, we used partial correlation networks to investigate plant-soil linkages along fourteen elevational gradients in the French Alps. We found tight linkages of plant traits with microbial activities, the former being driven by climate and the latter by soil properties. Climate did not affect soil properties directly, but through plant functional traits. Overall, we demonstrate how soil functioning can be integrated in studies of ecosystem shifts under environmental change at large spatial scales.

Abstract

Aim: Plant-soil interactions can be major driving forces of community responses to

environmental changes in terrestrial ecosystems. These interactions can leave signals in

aboveground plant functional traits and belowground microbial activities and these signals

can manifest in observed covariations. However, we know little about how these plant-soil

linkages vary in response to environmental conditions at biogeographic scales for which

experiments are impossible. Here, we investigate patterns of direct and indirect linkages

between plant functional traits, soil microbial activities and environmental conditions in

mountain grasslands along elevational gradients.

Location: The French Alps.

Taxon: Vascular plants and soil microbiota.

Methods: We analysed observational grassland data sampled along 14 elevational gradients

across the entire French Alps (between 1500 and 2800 m of elevation). Using Graphical

Lasso, we inferred a partial correlation network to tease apart direct and indirect plant-soil

linkages without defining the direction of interactions a priori.

Results: We found tight spatial associations of plant traits with microbial activities, climate

driving the former and soil properties the latter. In these plant-soil linkages, the dominance of

specific plant traits was more important than their diversity. We then showed that in sites with

conservative plant traits and reduced organic matter quality, soil microbes invested strongly in

nutrient acquisition.

Main conclusions: By investigating plant-soil linkages along elevational gradients in the

French Alps, we showed that plant functional traits and belowground microbial activity are

tightly linked and how they depend on environmental conditions. Overall, we demonstrated

4

how soil functioning can be integrated in studies of ecosystem shifts under environmental change at large spatial scales.

Keywords: elevational gradient, graphical lasso, ORCHAMP, partial correlation network, plant functional traits, plant-soil linkages, soil enzymatic activities

Introduction

Plant-soil interactions are fundamental elements of ecosystem functioning and structure (Wardle et al., 2004; Bardgett & Wardle, 2010; de Vries et al., 2013; Martinez-Almoyna et al., 2019). Plants rely on soil microbial communities that drive decomposition of detrital organic matter and the recycling of nutrients through their extracellular enzymatic activities (Burns & Dick, 2002; Schimel & Bennet, 2004; Burns et al., 2013). Conversely, the resource-acquisition strategy of soil microbes depends on the quantity and nutritional quality of plant litter inputs, and is reflected in the relative investment in extracellular enzymes that target C-, N- or P-rich compounds (Bowman, Steltzer, Rosenstiel, Cleveland & Meier, 2004; Bardgett & Wardle, 2010; Fanin, Hättenschwiler & Fromin, 2014; Legay et al., 2014; Piton, Legay, et al., 2020).

Observed correlations between components of plant communities and soil microbial communities, called plant-soil linkages in the following, can be the result of plants affecting soil, of soil affecting plants, or of both processes simultaneously (Fig. 1). Concerning the effect of plants on soil ecosystem properties, there are two conflicting visions in the literature. Grime (1998; see also García-Palacios, Shaw, Wall & Hättenschwiler, 2017) proposed that plant species in a community control ecosystem functions, such as primary production, resistance and resilience to perturbations and nutrient cycling and storage, proportionally to their biomass. In other words, the traits of dominant plant species determine ecosystem properties more strongly than the traits of rare species ("mass-ratio-hypothesis"). On the other hand, it was suggested that functional diversity affects ecosystem functioning through complementary use of resources ("diversity-function-hypothesis"; Tilman et al., 1997; Hooper et al., 2005). For example, diverse plant communities may increase rates of nitrogen (N) cycling by providing variable litter qualities that can be broken down at different rates, thus

providing a more consistent supply of organic N (Laughlin, 2011). Since rare species can have keystone effects and strongly influence pathways of energy and material flows (see also Violle et al., 2017), the relative abundance of species is not always a good predictor of ecosystem-level importance of species. These hypotheses have mostly been tested at regional scales for individual ecosystem processes, and most studies supported the mass-ratio-hypothesis (Díaz et al., 2007; Grigulis et al., 2013; Lavorel, 2013; García-Palacios et al., 2017). However, we still lack knowledge about whether functional diversity or traits of dominant species are of greater importance in plant-soil linkages at biogeographic scales.

Soil functioning also impacts the plant community. On fertile soils with high nutrient availability, plant communities are often composed of exploitative plants (high leaf nitrogen content and low leaf dry matter content) and are associated with bacteria-dominated belowground communities that quickly decompose the easily degradable litter (Wardle et al., 2004; Quested, Eriksson, Fortunel & Garnier, 2007; Fortunel et al., 2009). On infertile soils with low nutrient availability, conservative plant communities with slow growth rates are often linked to fungi-dominated belowground communities with slower cycling rates but with the ability to decompose more recalcitrant material (Aerts, 1999; Bardgett & Wardle, 2010). Although these associations are well-known locally, at biogeographic scales it is rarely studied how the changes in nutrient and energy availability in soils influence the traits of plant communities along the exploitative to conservative plant spectrum (Piton, Legay, et al., 2020). Extracellular enzymatic activities (EEAs) of microbial communities are related to both nutrient availability and microbial growth, acting at the interface between aboveground and belowground communities. EEAs can be either directed towards nutrient (e.g. N or P) or energy (i.e. C) acquisition. Ratios of EEAs therefore reflect the equilibria between elemental composition of microbial biomass and soil organic matter (Sinsabaugh, Hill & Shah, 2009), and the shift between nutrient and energy flow associated with microbial community metabolism. Although EEAs might underlie plant-soil linkages through organic matter recycling, their relationship with plant traits have rarely been investigated.

On top of plant-soil interactions, environmental conditions, i.e. soil physicochemical properties and climate, also influence the plant and microbial communities differentially, and therefore modulate their observed linkages (Bonito et al., 2014; Van der Putten, Bradford, Brinkman, van de Voorde & Veen, 2016) (Fig. 1). In mountain systems, for instance, temperature and snow cover determine growing season length, solar radiation that plants are exposed to, soil moisture regime and soil pH. These environmental conditions might affect nutrient and energy availability which impacts soil microbial communities, EEAs, plant community composition, and associated litter quality, and in turn, nutrient availability. In particular, freezing cycles constitute a physiological stress which alters microbial demand and allocation of C and nutrients (Freppaz, Williams, Edwards, Scalenghe & Zanini, 2007; Schimel, Balser & Wallenstein, 2007). However, it is difficult to distinguish between direct and cascading effects of the environment on plant-soil linkages because of the multitude of interactions between all components of the system. This is why all components implicated in the complex system of plant-soil linkages have rarely been studied together in the past (e.g. Wardle et al., 2004; Kulmatiski, Beard, Stevens & Cobbold, 2008; Delgado-Baquerizo et al., 2019).

At biogeographic scales, it is almost impossible to uncover and study interactions through experiments. But plant-soil interactions leave correlative signals in spatial patterns of aboveground functional traits and belowground activity that we can study with observational data (de Vries et al., 2012; Delgado-Baquerizo et al., 2018). We know, for instance, that at large scales aboveground plant properties (e.g. exploitative-conservative spectrum) are drivers of soil microbial community diversity and composition (de Vries et al., 2012; Delgado-Baquerizo et al., 2018; Boeddinghaus et al., 2019). However, such studies have

focused on abundance weighted trait values and have rarely assessed plant functional diversity. Moreover, most models used in previous studies to infer patterns in aboveground-belowground linkages, such as structural equation models, assume causality and directionality. In particular, microbial community composition was mostly assumed to be the response variable (de Vries et al., 2012; Delgado-Baquerizo et al., 2018; Boeddinghaus et al., 2019; but see Martinez-Almoyna et al., 2020). However, aboveground-belowground linkages result from interactions and feedbacks. From observations alone we can neither determine how the observed patterns were created nor the most prominent directions of the effects. They may result from plant traits influencing the soil community through litter quality, or from the soil community influencing the plant community through nutrient availability, or they may result from a combination of both. Also, we do not know where abiotic factors come into play and it is difficult to disentangle direct and indirect effects.

For these reasons, here, we used a graphical model approach to tease apart direct and indirect influences in plant-soil linkages without defining the direction of interactions *a priori*. Using a graphical lasso to infer a partial correlation network, we analyse interdependencies between aboveground plant functional traits, belowground enzymatic activities and abiotic conditions. We apply this method to grassland communities sampled along 14 elevational gradients from a large-scale observatory representative of the entire French Alps (ORCHAMP). Elevational gradients are associated with drastic abiotic gradients over short distances and are excellent systems to study ecosystem processes for a multitude of climatic and edaphic conditions, and to test how ecosystems respond to environmental variation (Sundqvist, Sanders & Wardle, 2013; Martinez-Almoyna et al., 2019). We focus on mountain grasslands because little is known about large scale patterns in aboveground-belowground linkages in these vulnerable ecosystems. In the context of unprecedented environmental changes, understanding the relevance of plant-soil linkages for biogeographic distributions of plant diversity and

microbial activity, and measuring the effects of single components on overall plant-soil functioning is of tremendous importance (Hagedorn, Gavazov & Alexander, 2019).

To better characterize plant-soil linkages along climatic and edaphic gradients in the French Alps, we ask three specific questions:

- (1) How do abiotic and biotic interdependencies between climate, abiotic soil conditions, and above- and belowground communities vary over biogeographic scales (focus on between-compartment correlations)?
- (2) Are plant-soil linkages better explained by plant functional diversity or by dominant trait values?
- (3) Which specific associations exist between extracellular enzyme activities and other specific variables involved in plant-soil linkages (focus on within-compartment correlations)?

Materials and Methods

Study area and the observatory ORCHAMP

ORCHAMP (Spatio-temporal observatory of biodiversity and ecosystem functioning of mountain socio-ecosystems, Supp. Mat.) is a long-term observation network covering the French Alps (Fig. 2, Fig. S1). It consists of multiple elevational gradients across the entire study area. For each gradient, 30 m * 30 m permanent plots were established every ~200 m of altitude. Plant community relevees (presence/absence and abundances), microbial activities, climatic conditions and soil properties were measured between 2016 and 2018 (Fig. S2). For this study, we selected 41 plots in open habitat (including grasslands and few shrublands) along 14 elevational gradients on different metamorphic and sedimentary bedrock, such as schist, gneiss and limestone (Fig. 2, complete sampling protocols and descriptions of the plots available on orchamp.osug.fr).

Biotic plant-soil linkages – Aboveground community

In each plot, we recorded plant species abundances using a pin-point sampling along a 30 m transect parallel to the slope in the middle of the plot. Two measurements were taken every 20 cm: one measurement 25 cm upslope, the other 25 cm downslope from the transect. All individuals touching the pole that marked the two measure points per 20 cm increment were identified and counted. Furthermore, an additional dataset of presence/absences of all species along the same 30 m transect but of 3 m width was sampled to obtain a value of species richness representative of the whole plot. All plant species in the subplot were identified by professional botanists.

To test the mass-ratio-hypothesis (Grime, 1998), we calculated community-weighted means (CWMs) of plant traits (Garnier et al., 2004; Lavorel et al., 2008, Eq. 1).

(1)
$$CWM = \sum_{i=1}^{n} (p_i \times trait_i)$$

where p_i is the relative abundance of species i in a plot, $trait_i$ the trait value of species i and n the number of species

We calculated the CWMs for the following traits: Leaf nitrogen content (LNC), leaf carbon to nitrogen ratio (C/N), leaf dry mass per area (LMA), leaf dry matter content (LDMC), plant height and root depth (RD). These traits were not measured on individuals on the Orchamp plots but in multiple populations across the whole French Alps to account for intra-specific variability (Albert, Thuiller, Yoccoz, Soudant, et al., 2010) between 2008 and 2018. More specifically, at least 20 individuals were measured for traits related to height (vegetative and reproductive plant height) and 10 for leaf traits (LNC, C/N, LMA and LDMC). The measurements were carried out according to the protocol developed by Cornelissen et al. (2003). We used Flora indicativa (Landolt et al., 2010) to complement missing data on vegetative and reproductive plant height, and to extract data on root depth (RD). Leaf carbon and nitrogen contents were measured with an elemental analyzer (Flash EA1112; Thermo Scientific). Mean trait values per species were then calculated.

In the 41 selected Orchamp plots, 484 different plant species were identified in total. On average, trait data was available for 82 % of all individuals present in a given plot. They were transformed to approximate a normal distribution: a natural logarithm was applied to LNC and plant height, LDMC was square-root transformed and LMA, C/N and RD were log-transformed (basis 10).

To measure the overall functional diversity of the plot, we calculated three independent measures: Functional richness (FRic), functional divergence (FDiv) and functional evenness (FEve) (Villéger, Mason & Mouillot, 2008). FRic represents the multidimensional volume of traits (trait space) filled by the species present in the community. For a single trait, this is simply the range of trait values spanned by the species present in the community. FDiv describes the overall functional divergence between species in the community. Low FDiv values mean that species within the community have similar combinations of traits, thought to be indicative of strong environmental filtering (Mouchet, Villéger, Mason & Mouillot, 2010) but also asymmetric competition (Mayfield & Levine 2010). Conversely, high FDiv values indicate that species in the community have a wide range of trait values, leading to a high degree of complementarity and resource use. FEve measures the evenness in the distribution of abundances in trait space. In other words, whether species' abundances tend to be more abundant on one side (low complementarity) or to be evenly distributed in the trait space indicating a high degree of complementarity or niche partitioning among coexisting species. We included all six traits presented above to calculate the diversity measures using the R package "FD" (Laliberté et al., 2014). The traits were standardised (mean $\mu = 0$, standard deviation $\sigma = 1$) before calculating FRic, FDiv and FEve. Since FRic and species richness were correlated (Villéger et al., 2008; Pearson correlation between FRic and species richness > 0.98), we used a randomisation process and calculated the standard effect size to remove the effect of richness (FRic ses; Supp. Mat., Pearson correlation between FRic ses and species richness < 0.5).

The normalized difference vegetation index (NDVI) has been shown to be related to chlorophyll abundance and leaf area index (Myneni, Hall, Sellers & Marshak, 1995). We used the mean yearly sum of NDVI greater than 0.2 of the period 2000-2018 (NDVI_{int}) as a proxy of plant primary productivity. Estimates of the surface spectral reflectance at a resolution of

250 m were derived from MODIS (Moderate Resolution Imaging Spectroradiometer) bands 1 and 2; MOD09Q1: MODIS/Terra Surface Reflectance 8-Day L3 Global 250 m SIN Grid V006 satellite MODIS (Terra), available online: https://lpdaac.usgs.gov/products/mod09q1v006/. The preprocessing of NDVI time series followed the method described by Choler (2015).

Biotic plant-soil linkages—Belowground community

At 5 m downside of the vegetation transect in each plot, soil samples were taken in three 2 m * 2 m sub-plots. In each subplot, an average of ten soil cores were taken from the top-soil (0-10 cm depth) and mixed to get around 1 kg of soil.

The collected composite samples were sieved at 5.6 mm, and 40-50 ml per sample were extracted and frozen at -20 °C for subsequent analysis of microbial activities. The potential activities of seven different extracellular enzymes involved in carbon (α -Glucosidase (AG), β -Glucosidase (BG), β -D-cellobiosidase (CB), β -Xylosidase (XYL)), nitrogen (Leucine aminopeptidase (LAP), N-acteyl- β -Glucosaminidase (NAG)) and phosphorus (Phosphatase (PHOS)) acquisition, were measured according to an adapted protocol of Bell et al. (2013) (see Supp. Mat.). The sum of all measured potential enzymatic activities (total EEA) was used as an overall indicator for the potential of the soil to depolymerize and recycle organic compounds and is therefore an important component of plant-soil linkages (Piton, Legay, et al., 2020). Soil microbial community is the primary driver of this potential activity as microbes produce most of these enzymes. Then, abiotic factors (e.g. temperature) and processes (e.g. substrate diffusion and enzyme stabilisation in the soil matrix) modulate potential and realized enzymatic activity *in situ* (Wallenstein and Weintraub, 2008; Nannipieri, Trasar-Cepeda & Dick, 2018; Piton, Foulquier, et al., 2020).

Enzyme activity vectors have recently been shown to quantify simultaneous resource demands of the belowground community and the belowground community's relative investment in C, N and P acquisition (Fanin, Moorhead & Bertrand, 2016; Moorhead, Sinsabaugh, Hill & Weintraub, 2016; Chen et al., 2018). The proportional activity of enzymes targeting carbon (AG, BG, CB and XYL) and nitrogen (LAP and NAG) - i.e. (AG+BG+CB+XYL)/[(AG+BG+CB+XYL)+(LAP+NAG)] - was plotted against C vs. P acquiring enzymes (PHOS) - i.e. (AG+BG+CB+XYL)/[(AG+BG+CB+XYL)+(PHOS)]. In general, a relative increase in the activity of enzymes targeting a specific element indicates that this element is limited in the environment (Olander & Vitousek, 2000). The length of the vector created by each point in the plot and the origin quantifies therefore relative C vs. nutrient limitation, with increasing vector length indicating increasing C limitation. The angle formed by the x-axis and the vector on the other hand quantifies the relative P vs. N limitation (Supp. Mat., Fig. S3). The three replicates of each plot were averaged, and a natural logarithm was applied to total EEA.

Abiotic factors modulating plant-soil linkages – Soil physicochemical properties

Soil dry weight was measured by weighing soil samples after drying at 70 °C for 48 h, and soil organic matter (SOM) was determined by loss on ignition using the previously dried samples (4 h at 550 °C).

The rest of the composite samples was then dried, sieved at 2 mm and homogenised to obtain representative subsamples for the following analyses. We measured pH following the ISO 10390:2005 norm using a pH-meter (pH7110, inoLab) in a 1:5 soil/distilled water-solution. We used an Ultra Centrifugal Mill ZM 200 (Retsch ZM200) to grind part of the dried and sieved samples below 250 µm. Carbon and nitrogen contents were then determined using an

elemental analyser (Flash EA1112, Thermo Scientific). The three replicates per plot were averaged and SOM and C/N log-transformed.

Abiotic factors modulating plant-soil linkages – Climate variables

We characterized climate conditions at each plot using: Mean annual soil temperature (MAT), mean total annual precipitation (MAP), growing degree days (GDD), freezing degree days (FDD), climate water stress (CWS) and solar radiation (SR). To do this, we extracted for each plot meteorological time series of surface conditions from the SAFRAN-Crocus (S2M) reanalyses that are available at hourly resolution for the time period 1988-2018 (Durand, Giraud et al., 2009; Durand, Laternser et al., 2009; Vionnet et al., 2012; Vernay et al., 2019). S2M is a regional-scale reanalysis that takes topographical effects into account. Data are produced for 23 massifs of the French Alps as a function of elevation in 300 m increments, slope aspects and angles. The S2M reanalysis has been used for many real-time and climatological applications in the French mountain areas (e.g. Corona, Morin & Choler, 2018; Verfaillie et al., 2018; see also Fig. S6). Rainfall and incoming shortwave radiation (direct+scattered) from the SAFRAN reanalysis were aggregated at a daily time resolution. The soil temperature values (temperature in the first centimetre of the soil column, variable TG1 in S2M datasets) were computed by the ISBA-Crocus model fed by meteorological information from SAFRAN.

Growing degree days correspond to daily average soil temperature sums (using data for the first centimetre of the soil column) above 0 °C and were calculated based on the assumption that plant growth is more affected by cumulated heat than by mean temperature (Körner & Hiltbrunner, 2018). To capture frost intensity and length of frost episodes, FDD was calculated analogously to GDD (daily average ground temperature sums below 0 °C)

following Choler (2018). GDD was calculated over the growing season, the onset of which was defined by four consecutive days without snow and a soil temperature of at least 4 °C in the simulations, and the offset of which was defined by the first day of the winter season with snow on at least four consecutive days. FDD was calculated over the whole year. Both variables were calculated on an annual timescale and then averaged over the whole time period (1988-2018). We log-transformed absolute values of yearly averaged FDD. We calculated the climatic variables over a 30-year time period. This was done as a compromise since plant communities in alpine systems are known to respond to medium-term climatic conditions rather than annual variations (Körner, 2003). Soil communities might instead be more affected by shorter climatic conditions but it has been suggested that soil communities might actually be very tolerant to climatic variations (Thakur & Geisen, 2019; Thakur, 2020; Thakur et al. 2020).

To quantify water stress and severity of summer drought, we used an approximation of climate water deficit (CWD). CWD is the difference between potential evapotranspiration and water availability in the soil (Stephenson 1998) and usually accounts for actual evapotranspiration and local soil conditions. Since local soil conditions, and especially available water capacities, are poorly understood in the Alps, we approximated CWD by calculating climate water stress (CWS): daily CWS corresponds to the difference of daily potential evapotranspiration (calculated after Vannier & Braud (2012) using the Penman-Monteith Equation (Allen, Pereira, Raes & Smith, 1998)) and daily precipitation. Water stress increases as values of CWS increase. In the case of precipitation exceeding evapotranspiration there is no water limitation in the soil compartment, therefore CWS values were set to zero (see e.g. Aragão et al. 2007). We calculated CWS sums over the growing season for each year between 1988 and 2018, and then averaged these sums to characterize mean water stress and severity of summer drought of the soil compartment.

We calculated cumulative solar radiations over the growing season, and then averaged the cumulative values over the whole period 1988 - 2018.

The graphical lasso

Given that plant-soil linkages are by definition bi-directional and some of the potential relationships between variables are not clearly identified, we used Graphical Lasso (glasso, Friedman, Hastie & Tibshirani, 2007; Mazumder & Hastie, 2012). Glasso is a type of model that is able to decipher the partial correlations between all the variables across our 41 plots without imposing a directional structure, like it would be the case with a structural equation model. In glasso, the partial correlations between all variables are inferred from the inverse of the variance-covariance matrix. Similarly to the lasso regression (Least Absolute Shrinkage and Selection Operator, Tibshirani, 1996), glasso uses a regularization penalty to set all spurious partial correlations to zero. To do so, partial networks are estimated for different values of λ (the penalty coefficient). We used the Extended Bayesian Information Criterion to select the optimal λ , as implemented in the package qgraph (Epskamp, Cramer, Waldorp, Schmittmann & Borsboom, 2012; see also Ohlmann et al., 2018). The optimal λ depends on the number of variables and the strength of the partial correlations in the network; different sets of variables will therefore lead to different λ being selected and different correlations being considered as spurious. The obtained regularized partial correlation network was then visualised using Gephi (Bastian, Heymann & Jacomy, 2009). The network consisted of nodes representing the variables and edges which connect the nodes. Two variables were connected if they were conditionally dependent on each other, i.e. they were partially correlated given all other variables in the graph. Reciprocally, two unconnected variables are said to be conditionally independent (i.e. they cannot causally influence each other, Murphy, 2012).

Within the glasso, we included several variables representing the different compartments implicated in plant-soil linkages: Plant community, belowground microbial community, physicochemical soil properties and climatic conditions (Fig. 1; Table 1). To assess the importance of a variable within its own group and its importance for other groups, we extracted the degree of each variable (i.e. number of edges that are incident to a given node). We calculated two different measures: the probability of a node to form a connection within its own group (number of realised edges within group/number of possible edges within group), and the probability of a node to form an edge with a node not belonging to its own group (number of realised edges across groups/number of possible edges across groups).

Moreover, we analysed the weights of the edges to estimate the strength of a node (i.e. how important a variable was to the overall plant-soil linkages). The weight of the edges incident to a node corresponds to the strength of its partial correlations. We calculated the sum of absolute partial correlations incident to a node within its own group and the sum of absolute partial correlations incident to a node across groups. The sum of all absolute partial correlations of a node was divided by the number of realised edges to calculate the mean strength of partial correlations.

To assess the importance of interactions between groups, we aggregated the results at the group level, i.e. the probability to form connections as well as the mean partial correlations were calculated for all variables of one group (plant community, EEA, abiotic soil and climate) towards all variables of another group.

Variable selection

From the initial pool of variables (Table 1), a pre-selection was made to avoid redundant information. To select those variables that explained most of the environmental variation

while being weakly correlated with each other, we used a principal component analysis (PCA) on plant traits and on climatic variables, respectively. We identified MAT, MAP, SR, plant height and leaf C/N as variables with little additional information and thus removed them from further analysis.

The glasso was run two times. First, all variables in Table 1 except those eliminated in the PCA step were included (Fig. S4). Then, nodes were excluded that were deemed of minor importance because they were completely unconnected or only connected to other variables in their own group. The network presented in this paper is the result of this process, and only includes five plant community variables (the CWMs LDMC, LMA, RD, LNC, and the canopy variable NDVI_{int}), three abiotic soil variables (C/N, SOM, pH), two climatic variables (GDD, CWS) and two EEA variables (total EEA, vector angle (P/N)). To test the robustness of the result, we ran a regression between the partial correlations from the network containing all variables and the network containing the selection of variables (Fig. S5).

Results

General linkages between biotic plant-soil components and abiotic factors

The probability to observe linkages (Fig. 3a) between the plant community and abiotic components was twice as high for climate as for soil (0.40 and 0.20, respectively). The same pattern was observed for the mean strength of those links (0.44 and 0.26, respectively, Fig. 3b). Enzymatic activity of the soil microbial community was strongly linked to abiotic soil variables (probability 0.67 and mean strength 0.42), but not to climate. There was neither a direct connection between climate and soil properties, nor between the plant community and the soil microbial community (Fig. 3).

Role of the different facets of the plant community in plant-soil linkages

Neither CWMs nor functional diversity indices were linked to microbial activities (Fig. 4, Fig. S4). However, functional diversity measures were completely isolated (spnm, FDiv, FRic_{ses} and FEve, Fig. S4) and thus removed from the final graph, whereas CWMs of several traits showed clear and strong links to the soil C/N ratio which was a focal variable linking the plant and microbial communities. We observed a partial correlation between RD and soil C/N of -0.24 and of 0.26 between LMA and soil C/N (Fig. 4). NDVI_{int} and soil C/N were positively correlated (0.27).

Variables linked to soil enzymatic activities

We observed no direct linkages between plant traits and ratios of enzymatic activities. However, soil C/N was a focal variable linking plant traits to microbial activity, suggesting that the interaction between plant traits and enzymatic activities passes by litter quality. Soil

C/N showed the highest probability to form edges across groups overall (0.44), albeit of medium strength (0.29, Table 2). Soil C/N showed a strong linkage to the conservative-exploitative plant continuum. Low soil C/N was associated with low LMA, which in turn was linked to high LNC, which was linked to low LDMC, thus representing a trait assemblage of an exploitative strategy with high investment in photosynthetic leaf area. This spectrum of conservative-exploitative species was indirectly related to soil enzymatic activities via soil C/N. Conservative species (high LMA) and high soil C/N were associated with a high vector angle, indicating that sites with conservative species were less nitrogen and more phosphorus limited than sites with more exploitative species. Conservative species (high LDMC) were also associated with higher water stress (CWS, 0.21).

In addition to plant functional traits, primary productivity played an important role since $NDVI_{int}$ was the link between climate variables and soil C/N. It was positively correlated to GDD (0.7) and negatively to CWS (-0.58), indicating that higher temperatures increased biomass production but associated water stress limited it. $NDVI_{int}$ and soil C/N were positively correlated (0.27) suggesting an increased soil C/N in productive sites.

Soil abiotic characteristics were strongly linked to variables of enzymatic activities. SOM and total EEA showed a strong positive correlation indicating high enzymatic activity in organic matter rich sites, and pH was negatively correlated to both vector angle and total EEA (-0.31 and -0.38, respectively). This suggests strong phosphorus limitation and increased enzymatic activity in acidic soils. Vector length was not related to any other variables (Fig. S4), suggesting that P vs. N limitation is of more importance than carbon vs. nutrient limitation in alpine open habitats.

Discussion

Plant soil linkages and environmental gradients

Studying for the first time the spatial interdependencies between plant functional composition and soil microbial enzymatic activities across grasslands in the French Alps, we found that plant functional composition was mostly associated with climatic variables, while microbial enzymatic activities were primarily related to soil properties. Abiotic factors thus dominate direct biotic linkages in our study system which confirms what has been proposed before: that abiotic factors are of primary importance in mountain systems (Neuwinger, 1970; Körner, 2003). Our results further revealed that an interdependency between plant traits and soil C/N links the aboveground community to the belowground community. This is congruent with the hypothesis that plant traits play a central role in plant-soil linkages through control of litter quality (Schweitzer et al., 2008; Lau & Lennon, 2011; van der Putten et al., 2013). Moreover, our results showed that in alpine open habitats effects of climate propagate to the soil community through plant traits. This is in accordance with previous studies that demonstrated links between climate and plant traits (Mayor et al., 2017), as well as indirect effects of climate on the soil community via plant traits (Delgado-Baquerizo et al., 2018).

However, in contrast to previous studies (de Vries et al., 2012; Delagado-Baquerizo et al., 2018), we neither found a direct link between soil microbial community activity and plant functional CWMs, nor between soil microbial community activity and climatic variables. This might be due to the fact that the focus of our study was assessing the functioning of the microbial community through extracellular enzymatic activity (e.g. recycling of organic matter) rather than the composition of the microbial community itself. The link between microbial community composition and function is not necessarily straightforward. Potential changes in microbial community composition, resulting from changes in plant functional

traits for example, that could occur here might not have had an impact if there is some level of functional redundancy in the microbial community for the extracellular enzymatic activities we measured (Louca et al., 2018). Future studies considering diversity or composition of microbial communities along with their function could reveal more direct links with plant traits as observed in de Vries et al. (2012) and Delgado-Baquerizo et al. (2018), and help understand spatial interdependencies between below- and aboveground communities along large environmental gradients. An explanation for the missing link between soil microbial and climatic variables may be the 30-year time period over which the climatic variables were calculated. This period was chosen because plant communities in alpine systems are known to respond to medium-term climatic conditions rather than annual variations (Körner, 2003). While soil communities might instead be more affected by shorter climatic conditions it has also been suggested that soil communities might actually be very tolerant to climatic variations (Thakur & Geisen, 2019; Thakur, 2020; Thakur et al. 2020). The choice of the time period over which climatic variables are calculated is, in a large-scale ecosystem study, necessarily a compromise between the studied compartments (see also last section of the discussion).

Role of the different facets of the plant community in plant-soil linkages

Contrasting the mass-ratio-hypothesis (Grime, 1998) with the diversity-function-hypothesis (Tilman et al., 1997; Hooper et al., 2005), we found that CWMs of plant traits were more important than diversity metrics, supporting the mass-ratio-hypothesis. CWMs showed clear and strong linkages to soil C/N, propagating on to microbial activities and SOM content, indicating a strong effect on organic matter decomposition and the recycling of nutrients (Garnier et al., 2004; Quested et al., 2007; Fortunel et al., 2009; Grigulis et al., 2013; Lavorel,

2013). These results are consistent with the growing body of evidence for the mass-ratio-hypothesis as a central mechanism controlling ecosystem functioning and services (Lavorel, 2013; Li et al., 2017). They also confirm that it is the linking mechanism between plants and the functional properties of microbial communities (Grigulis et al., 2013; Piton, Legay, et al., 2020).

The isolation of functional diversity indices in our analyses may be in part due to the mountain context of our study. Hooper et al. (2005) suggested that ecosystems might be less defined by functional diversity in challenging abiotic conditions. They also argued that species with different response traits to environmental changes are needed under increasing temporal and spatial variability in order to ensure a stable supply of ecosystem functions. Furthermore, it has been shown that mass ratio effects better explain individual functions, but that multifunctionality of ecosystems is more driven by diversity effects (Le Bagousse-Pinguet et al., 2019). Therefore, considering today's climate and land-use changes and biological invasions, even a system where the diversity-function-hypothesis does not play an important role under current conditions may depend on trait diversity if certain thresholds are crossed in the future. Thus, although our results confirm the importance of CWMs for ecosystem functioning (e.g. decomposition and nutrient recycling) their importance for ecosystem stability still needs further exploration (e.g. Piton, Legay, et al., 2020), especially in alpine systems where species' trait values may vary a lot depending on altitude due to high plasticity (Albert, Thuiller, Yoccoz, Douzet, et al., 2010; Albert, Thuiller, Yoccoz, Soudan, et al., 2010).

Variables linked to soil enzymatic activities

It is well understood that plant traits are linked to microbial community composition and functioning, directly and via plant litter quality (Grigulis et al., 2013; Fanin et al., 2014; Legay et al., 2014; Martinez-Almoyna et al., 2020). In our study, plant traits associated with the plant conservative-exploitative continuum (LMA, LNC and LDMC) were indirectly connected to SOM, as well as soil enzymatic activities, via a central link between plant traits and soil C/N. This supports at a biogeographic scale the hypothesis that litter quality rather quantity role in microbial functions involved than plays central aboveground-belowground linkages in grassland ecosystems, as has been shown previously also for soil microbial community composition and diversity in studies of temperate grasslands in England and Germany (de Vries et al., 2012; Delgado-Baquerizo et al., 2018). Interestingly, our partial correlation network is consistent with the expected ecosystem functioning along a conservative-exploitative continuum (Moore, 1988; Bardgett & Wardle, 2010; Mulder et al., 2013). On the conservative side, reduced organic matter quality (high soil C/N) associated with conservative plant traits favours SOM accumulation, increasing total enzymatic activity. High soil C/N was also associated with an increased investment of microbial communities in P-acquisition relative to N-acquisition, suggesting that conservative plant traits and high soil C/N are also associated with a high N/P ratio of resources available for microbes. The link between soil C/N and microbial nutrient acquisition strategies might be central in the feedback from soil to plants. In infertile soils, microbial communities immobilize nutrients in their biomass, releasing less nutrients to the soil (Mooshammer et al., 2014; Zechmeister-Boltenstern et al., 2015). This creates a negative feedback-loop since a nutrient poor soil will result in a more conservative plant community which will then produce more recalcitrant litter increasing further SOM accumulation and the nutrient limitation of microbial communities (Wardle et al., 2004; de Vries & Bargett, 2012).

Our network also showed soil C/N to be linked with plant community productivity (NDVI_{int}) and root depth. High plant productivity was associated with a warm growing season, low water deficit and exploitative plant traits (low LMA). It was also positively associated with soil C/N, likely explained by higher plant derived C input in the soil with higher productivity. On the contrary, high root depth associated with high water deficit (stress resistance strategy) decreased soil C/N, possibly because a higher fraction of the plant derived C was released deeper into the soil and thus could not be captured in our soil data from the upper horizon. Our results also suggest stronger microbial phosphorus than nitrogen limitation (high vector angle) and increased enzymatic activity in acidic soils, in accordance with Piton, Legay, et al. (2020). In acidic soils (mean pH of our sites was 5.5), leaching of acids towards the bedrock releases (Fe, Al or Mn) cations. These cations form compounds with bioavailable forms of phosphorus, such as H₂PO₄, and hence decrease P availability (Wild, 1950; Iqbal, 2012).

Critical aspects and perspectives

Our study gives insights about linkages between plant and microbial soil communities in alpine habitats at large scales, and we show how abiotic climatic conditions and soil properties influence these linkages. However, our study is not without flaws. First, there may be a conflict of scales in our analysis. Plant and microbial soil communities have a very different temporal turnover, i.e. composition and abundances may change faster in microbial than in plant communities, and microbial communities may hence respond faster to changes in environmental conditions (Bardgett and Wardle, 2010; but see Thakur & Geisen, 2019; Thakur, 2020; Thakur et al. 2020). They may thus be more determined by short-term fluctuations in abiotic conditions while plant communities may respond more to long-term changes. This may create a mismatch in temporal scales when analysing both components at

the same time. For example, we calculated climatic variables like growing degree days (GDD) over a period that could be considered more of a determinant of the plant community. To determine whether this temporal mismatch was truly an issue, we compared GDD calculated over a 30-year time period (as used here), with the GDD calculated over 10 years and over the year before sampling for all our 41 plots. All correlations were very high, demonstrating that the differences between plots remain relatively constant whatever the time-scale used to calculate growing degree days (Fig. S7, S8). We are thus confident that this should not influence the results and the conclusion of our paper. In addition to a temporal mismatch, there may also be a spatial one. The quantitative prediction of precipitation in mountainous areas can be biased because precipitation is a very local phenomenon that changes from one mountain massif to another (Roe, 2005; Quintana-Segui et al., 2008; Quintana-Segui et al., 2017). Spatial variability in precipitation may therefore be difficult to capture with climate models if few observation stations are available (Hofstra, New & McSweeney, 2010). However, while SAFRAN is known to overestimate the number of precipitation days and underestimate high precipitation events (Quintana-Segui, Turco, Herrera & Miguez-Macho, 2017), it is very likely that plot differences (i.e. the ranking between the plots) are relatively robust to these uncertainties and should thus not influence the Graphical lasso estimation.

Second, plant-soil linkages are complex and they may be affected by factors that we did not include in this study, such as herbivores and soil meso- and macrofauna. Furthermore, we did not include direct measures of diversity or composition of the microbial community but only variables related to their activities. Specifically, we did not use environmental DNA metabarcoding approaches because they do not provide reliable information on the biomass of different organisms (Taberlet, Bonin, Zinger & Coissac, 2018; Calderón-Sanou, Münkemüller, Boyer, Zinger & Thuiller, 2020). It would thus neither have been possible to calculate ratios of fungal and bacterial biomass, nor would we have had information about

abundances of oligotrophic and copiotrophic groups. While we showed that functional diversity of the plant community did not play a role in our study system, we cannot exclude the possibility that diversity of the microbial community did. In addition, we showed in our study that litter quality may be a key determinant of microbial functions in alpine grassland ecosystems. We deduced this from measures of topsoil C/N content which we interpreted as a proxy of litter quality. In future studies it may hence be of interest to include direct measures of litter quality.

Third, it is important to note that we quantified EEA *in vitro* and therefore used measures of potential enzymatic activity and not *in situ* activity. It has been argued that *in vitro* assays may be a limited proxy for EEA in real conditions since substrate diffusion and abundance, as well as soil type and temperature are not reflected in vitro assays (Wallenstein and Weintraub, 2008; Bell et al., 2013). However, *in situ* measurements provide only a snapshot of the belowground community's activity and may therefore not adequately represent longer-term linkages between the plant and the soil microbial communities.

Last, we chose to study plant-soil-linkages with the graphical lasso because it is difficult or even impossible to uncover interactions through experiments at large scales. More importantly, we do not know if observed patterns result from the plant community influencing the soil community, or the other way around, or both influencing each other reciprocally. Hence, an undirected partial correlations network is a very well-suited method to disentangle direct and indirect effects between the plant and the soil microbial communities, and to look at biogeographic patterns. Nevertheless, experimental validation is necessary to determine causal pathways, which is especially important considering the potential effects of rising temperatures or changes in precipitation on plant-soil relationships.

Conclusions

Studying the spatial interdependencies between the plant functional composition, soil microbial enzymatic activities and abiotic conditions across the French Alps, we found abiotic drivers to be key elements. Plant traits were more strongly controlled by climate, whereas enzymatic activities were more associated with soil abiotic properties. The connections between climate and soil biotic and abiotic properties were only indirect through plant trait association with soil C/N. At the same time, the links between microbial enzymatic activities and plant traits through soil C/N highlighted the role of plant-soil interdependencies in ecosystem responses to abiotic changes. The observed plant-soil linkages are well in accordance with the conservative-exploitative continuum with soil C/N as an important player. Our study is one of the first to identify a signal of the conservative-exploitative plant continuum and microbial nutrient acquisition strategies in patterns of co-distributions of plant functional traits and microbial activity on a biogeographic scale (the French Alps) in alpine ecosystems. It thus demonstrates how the soil functioning can be integrated in studies of ecosystem shifts under environmental change at large spatial scales.

Tables

Table 1: Overview of variable pool from which a selection was made to use in the graphical lasso.

Variable	Acronym	Unit	Data source	Mean (median)
				deviation
Extracellular enzymatic activity				
Total EEA	Total EEA	nmol/(h*g dry	Orchamp	12677 (12174) ±
		mass)		
Vector length (proportion	EEC/EEN	0	Orchamp	$0.79(0.78) \pm 0.0$
Carbon/Nutrients)				
Vector angle (proportion P/N)	EEN/EEP	-	Orchamp	$66.1 (65.7) \pm 6.0$
Physico-chemical soil properties				
Soil organic matter	SOM	%	Orchamp	$25.1(23.3) \pm 11.$
Carbon to nitrogen ratio	Soil C/N	-	Orchamp	14.68 (14.42) ± 2
рН	рН	-	Orchamp	$5.45(5.43) \pm 0.8$
Vegetation variables				
Leaf nitrogen content (CWM)	LNC	mg/g		$21.4(20.9) \pm 3.3$
Leaf carbon to nitrogen ratio	C/N	-		$22.8(22.4) \pm 4.1$
(CWM)				
Leaf dry mass per area (CWM)	LMA	mg/mm ²		$0.068(0.067) \pm$
Leaf dry matter content (CWM)	LDMC	mg/g		$296(291) \pm 41$

Plant height (CWM)		cm		$26(25) \pm 11$
Root depth (CWM)	RD	-	Landolt et al. 2010	$2.02(2.00) \pm 0.4$
Species richness	spnm	Number of species	Orchamp	$55(55) \pm 13$
Standard effect size of functional richness	FRic _{ses}	-		$-0.08 (-0.08) \pm 0$
Functional evenness	FEve	-		$0.57 (0.57) \pm 0.0$
Functional divergence	FDiv	-		$0.94(0.95) \pm 0.0$
Integrated normalized difference vegetation index (Proxy for biomass production)	NDVI _{int}	-	Terra MODIS 2000-2018	80 (86) ± 38
Climatic variables				
Mean annual temperature	MAT	°C	Safran-Crocus 1988-2018	$3.10(2.97) \pm 0.7$
Mean total annual precipitation	MAP	mm/year	Safran-Crocus 1988-2018	$673 (650) \pm 223$
Growing degree days (growing season)	GDD	°C/year	Safran-Crocus 1988-2018	$1164 (1087) \pm 28$
Freezing degree days (whole year)	FDD	°C/year	Safran-Crocus 1988-2018	42 (38) ± 21
Mean climate water stress (growing season)	CWS	mm/year	Safran-Crocus 1988-2018	$276(263) \pm 46$
Mean solar radiation (growing season)	SR	kJ/(m²*year)	Safran-Crocus 1988-2018	282823 (280069 40191

Table 2: Degree analysis of the partial correlations network presented in Fig. 4. "Edges within" corresponds to the probability to observe partial correlations between a given variable and other variables that belong to its own group. "Mean strength within" corresponds to the mean sum of all absolute partial correlations that are realised within the group. "Edges across" and "Mean strength across" are analogous metrics to the ones cited before concerning partial correlations *across* groups. Abbreviations: LNC: leaf nitrogen content, RD: root depth, LMA: leaf mass per area, LDMC: leaf dry matter content, NDVI_{int}: integrated normalized difference vegetation index, soil C/N: soil carbon to nitrogen ratio, SOM: soil organic matter content, GDD: growing degree days, CWS: climate water deficit, total EEA: total enzymatic activity.

Variable	Group	Edges	Mean	Edges	Mean
		within	strength	across	strength
			within		across
LNC	Vegetation	0.50	0.49	0	0
RD	Vegetation	0.25	0.22	0.29	0.26
LMA	Vegetation	0.50	0.38	0.14	0.26
LDMC	Vegetation	0.25	0.50	0.14	0.21
NDVI _{int}	Vegetation	0.50	0.25	0.43	0.52
soil C/N	Soil	0.50	0.23	0.44	0.29
SOM	Soil	0.50	0.23	0.11	0.57
pН	Soil	0	0	0.22	0.35
GDD	Climate	1	0.81	0.10	0.70
CWS	Climate	1	0.81	0.30	0.36

Vector angle	Enzymatic	0.00	0.00	0.20	0.36
(proportion P/N)	activity				
Total EEA	Enzymatic	0.00	0.00	0.20	0.48
	activity				

Figures

Figure 1: Linkages between aboveground plant communities and belowground microbial communities and abiotic factors affecting them. Variables defining the compartments plant community, soil properties and climate are written next to them: LNC – leaf nitrogen content, LDMC – leaf dry matter content, LMA – leaf mass per area, SOM – organic matter, C/N – soil carbon nitrogen ratio. References for the specific linkages are indicated by numbers: 1: Bowman et al., 2004; 2: de Vries et al., 2012; 3: Freppaz et al., 2007; 4: Grigulis et al., 2013; 5: Legay et al., 2014; 6: Martinez-Almoyna et al., 2020; 7: Mayor et al., 2017; 8: Piton, Legay, et al., 2020; 9: Quested et al., 2007; 10: Sundqvist, Sanders & Wardle, 2013, and references therein; 11: van der Putten et al., 2016, and references therein; 12: Wardle et al., 2004, and references therein.

Figure 2: The ORCHAMP observatory network in the French Alps (with years of sampling) and measurements that were taken on the plots of the individual elevational gradients.

Figure 3: Probability of observing links between compartments (a) and mean strength of links between compartments (b).

Figure 4: Graph of a partial correlations network in alpine communities, including plant community weighted means, soil microbial enzymatic activities, climatic variables and physico-chemical soil properties. Positive partial correlations are represented by dotted edges, negative ones by continuous ones. The weight of the edges (thickness) corresponds to the absolute strength of their partial correlations. Abbreviations: RD: root depth, LMA: leaf mass per area, LNC: leaf nitrogen content, LDMC: leaf dry matter content, NDVI_{int}: integrated normalized difference vegetation index, GDD: growing degree days, CWS: climate water

deficit, soil C/N: soil carbon to nitrogen ratio, SOM: soil organic matter content, total EEA: total enzymatic activity.

Figure 1

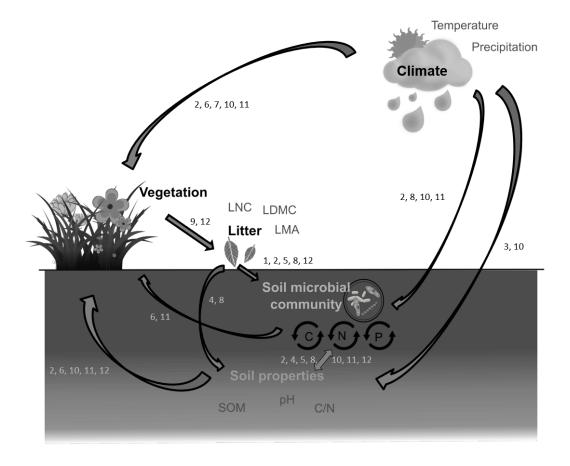


Figure 2

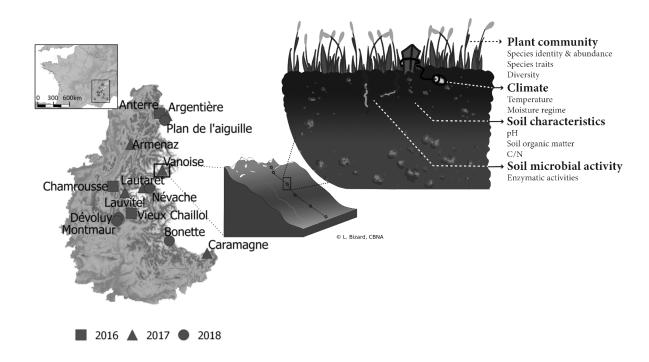


Figure 3

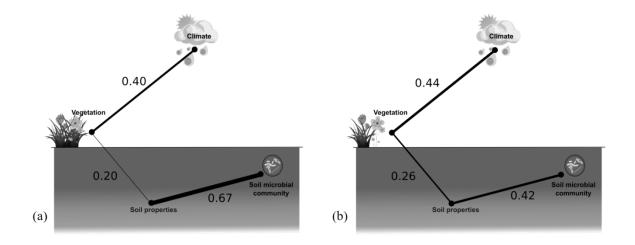
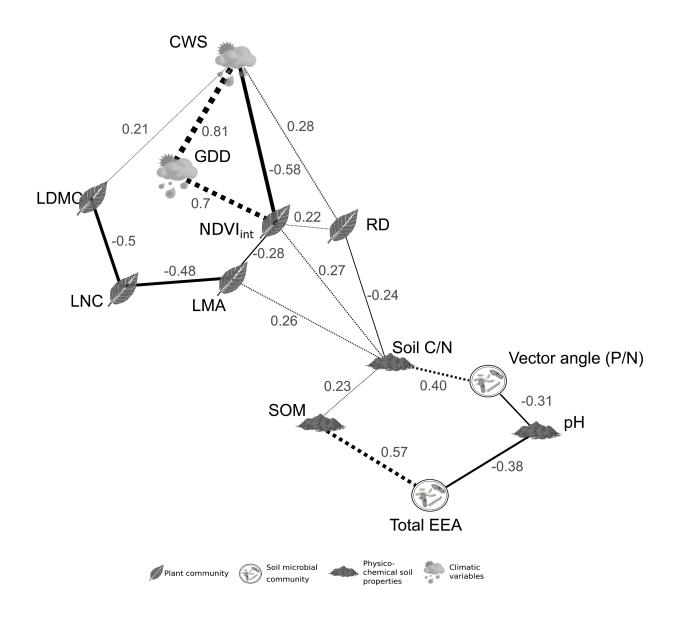


Figure 4



Data Availability Statement: Most of the raw data used in the manuscript are directly available on the ORCHAMP website (https://orchamp.osug.fr/home). Should the manuscript be accepted, the data supporting the results will be archived on Dryad and the data DOI will be included at the end of the article.

References

Aerts, R. (1999). Interspecific competition in natural plant communities: mechanisms, trade-offs and plant-soil feedbacks. *Journal of Experimental Botany*, 50(330), 29–37.

Albert, C.H., Thuiller, W., Yoccoz, N.G., Douzet, R., Aubert, S., & Lavorel, S. (2010). A multi-trait approach reveals the structure and the relative importance of intra-versus interspecific variability. *Functional Ecology*, 24(6), 1192–1201.

Albert, C.H., Thuiller, W., Yoccoz, N.G., Soudant, A., Boucher, F., Saccone, P., & Lavorel, S. (2010). Intraspecific functional variability: extent, structure and sources of variation within an alpine valley. *Journal of Ecology*, 98(3), 604–613.

Allen, R. G., Pereira, L. S., Raes, D., & Smith, M. (1998). Crop evapotranspiration - guidelines for computing crop water requirements-FAO irrigation and drainage paper 56. *FAO, Rome,* 300(9), D05109.

Aragão, L. E. O., Malhi, Y., Roman-Cuesta, R. M., Saatchi, S., Anderson, L. O., & Shimabukuro, Y. E. (2007). Spatial patterns and fire response of recent Amazonian droughts. *Geophysical Research Letters*, 34(7), L07701, doi:10.1029/2006GL028946.

Bardgett, R. D., & Wardle, D. A. (2010). *Aboveground-belowground linkages: biotic interactions, ecosystem processes, and global change*. Oxford University Press, Oxford, United Kingdom.

Bastian, M., Heymann, S., & Jacomy, M. (2009). Gephi: an open source software for exploring and manipulating networks. *International AAAI Conference on Weblogs and Social Media*. http://www.aaai.org/ocs/index.php/ICWSM/09/paper/view/154.

Bell, C. W., Fricks, B. E., Rocca, J. D., Steinweg, J. M., McMahon, S. K., & Wallenstein, M. D. (2013). High-throughput fluorometric measurement of potential soil extracellular enzyme activities. *JoVE (Journal of Visualized Experiments)*, 81, e50961.

Boeddinghaus, R. S., Marhan, S., Berner, D., Boch, S., Fischer, M., Hölzel, N., ... & Prati, D. (2019). Plant functional trait shifts explain concurrent changes in the structure and function of grassland soil microbial communities. *Journal of Ecology*, 107(5), 2197–2210.

Bonito, G., Reynolds, H., Robeson, M. S., Nelson, J., Hodkinson, B. P., Tuskan, G., ... & Vilgalys, R. (2014). Plant host and soil origin influence fungal and bacterial assemblages in the roots of woody plants. *Molecular Ecology*, 23(13), 3356–3370.

Bowman, W. D., Steltzer, H., Rosenstiel, T. N., Cleveland, C. C., & Meier, C. L. (2004). Litter effects of two co-occurring alpine species on plant growth, microbial activity and immobilization of nitrogen. *Oikos*, 104(2), 336–344.

Burns, R. G., DeForest, J. L., Marxsen, J., Sinsabaugh, R. L., Stromberger, M. E., Wallenstein, M. D., ... & Zoppini, A. (2013). Soil enzymes in a changing environment: current knowledge and future directions. *Soil Biology & Biochemistry*, 58, 216–234.

Burns, R. G., & Dick, R. P. (Eds). (2002). *Enzymes in the environment: activity, ecology, and applications*. CRC Press.

Calderón-Sanou, I., Münkemüller, T., Boyer, F., Zinger, L., & Thuiller, W. (2020). From environmental DNA sequences to ecological conclusions: how strong is the influence of methodological choices? *Journal of Biogeography*, 47(1), 193–206.

Chen, H., Li, D., Zhao, J., Zhang, W., Xiao, K., & Wang, K. (2018). Nitrogen addition aggravates microbial carbon limitation: evidence from ecoenzymatic stoichiometry. *Geoderma*, 329, 61–64.

Choler, P. (2015). Growth response of temperate mountain grasslands to inter-annual variations in snow cover duration. *Biogeosciences*, 12(12), 3885–3897.

Choler, P. (2018). Winter soil temperature dependence of alpine plant distribution: implications for anticipating vegetation changes under a warming climate. *Perspectives in Plant Ecology, Evolution and Systematics*, 30, 6–15.

Cornelissen, J. H. C. A., Lavorel, S. B., Garnier, E. B., Díaz, S. C., Buchmann, N. D., Gurvich, D. E. C., ... & Pausas, J. G. (2003). Handbook of protocols for standardised and easy measurement of functional traits worldwide. *Australian Journal of Botany*, 51(4), 335–380.

Corona-Lozada, M. C., Morin, S., & Choler, P. (2019). Drought offsets the positive effect of summer heat waves on the canopy greenness of mountain grasslands. *Agricultural and Forest Meteorology*, 276, 107617.

De Vries, F. T., & Bardgett, R. D. (2012). Plant-microbial linkages and ecosystem nitrogen retention: lessons for sustainable agriculture. *Frontiers in Ecology and the Environment*, 10(8), 425–432.

De Vries, F. T., Manning, P., Tallowin, J. R., Mortimer, S. R., Pilgrim, E. S., Harrison, K. A., ... & Kattge, J. (2012). Abiotic drivers and plant traits explain landscape-scale patterns in soil microbial communities. *Ecology Letters*, 15(11), 1230–1239.

De Vries, F. T., Thébault, E., Liiri, M., Birkhofer, K., Tsiafouli, M. A., Bjørnlund, L., ... & d'Hertefeldt, T. (2013). Soil food web properties explain ecosystem services across European land use systems. *Proceedings of the National Academy of Science of the USA*, 110(35), 14296–14301.

Delgado-Baquerizo, M., Fry, E. L., Eldridge, D. J., de Vries, F. T., Manning, P., Hamonts, K., ... & Bardgett, R. D. (2018). Plant attributes explain the distribution of soil microbial communities in two contrasting regions of the globe. *New Phytologist*, 219(2), 574–587.

Delgado-Baquerizo, M., Bardgett, R. D., Vitousek, P. M., Maestre, F. T., Williams, M. A., Eldridge, D. J., ... & Sala, O. E. (2019). Changes in belowground biodiversity during ecosystem development. *Proceedings of the National Academy of Science of the USA*, 116(14), 6891–6896.

Díaz, S., Lavorel, S., de Bello, F., Quétier, F., Grigulis, K., & Robson, T. M. (2007). Incorporating plant functional diversity effects in ecosystem service assessments. *Proceedings of the National Academy of Science of the USA*, 104(52), 20684–20689.

Durand, Y., Giraud, G., Laternser, M., Etchevers, P., Mérindol, L., & Lesaffre, B. (2009). Reanalysis of 47 years of climate in the French Alps (1958–2005): climatology and trends for snow cover. *Journal of Applied Meteorology and Climatology*, 48(12), 2487–2512.

Durand, Y., Laternser, M., Giraud, G., Etchevers, P., Lesaffre, B., & Mérindol, L. (2009). Reanalysis of 44 years of climate in the French Alps (1958–2002): methodology, model validation, climatology, and trends for air temperature and precipitation. *Journal of Applied Meteorology and Climatology*, 48(3), 429–449.

Epskamp, S., Cramer, A. O. J., Waldorp, L. J., Schmittmann, V. D., & Borsboom, D. (2012). qgraph: network visualizations of relationships in psychometric data. *Journal of Statistical Software*, 48(4), 1–18.

Fanin, N., Hättenschwiler, S., & Fromin, N. (2014). Litter fingerprint on microbial biomass, activity, and community structure in the underlying soil. *Plant and soil*, 379(1), 79–91.

Fanin, N., Moorhead, D., & Bertrand, I. (2016). Eco-enzymatic stoichiometry and enzymatic vectors reveal differential C, N, P dynamics in decaying litter along a land-use gradient. *Biogeochemistry*, 129(1), 21–36.

Fortunel, C., Garnier, E., Joffre, R., Kazakou, E., Quested, H., Grigulis, K., ... & Doležal, J. (2009). Leaf traits capture the effects of land use changes and climate on litter decomposability of grasslands across Europe. *Ecology*, 90(3), 598–611.

Freppaz, M., Williams, B. L., Edwards, A. C., Scalenghe, R., & Zanini, E. (2007). Simulating soil freeze/thaw cycles typical of winter alpine conditions: implications for N and P availability. *Applied Soil Ecology*, 35(1), 247–255.

Friedman, J., Hastie, T., & Tibshirani, R. (2007). Sparse covariance estimation with the graphical lasso. *Biostatistics*, 9(3), 432–441.

García-Palacios, P., Shaw, E. A., Wall, D. H., & Hättenschwiler, S. (2017). Contrasting mass-ratio vs. niche complementarity effects on litter C and N loss during decomposition along a regional climatic gradient. *Journal of Ecology*, 105(4), 968–978.

Garnier, E., Cortez, J., Billès, G., Navas, M., Roumet, C., Debussche, M., ... & Neill, C. (2004). Plant functional markers capture ecosystem properties during secondary succession. *Ecology*, 85(9), 2630–2637.

Grigulis, K., Lavorel, S., Krainer, U., Legay, N., Baxendale, C., Dumont, M., ... & Pommier, T. (2013). Relative contributions of plant traits and soil microbial properties to mountain grassland ecosystem services. *Journal of Ecology*, 101(1), 47–57.

Grime, J. P. (1998). Benefits of plant diversity to ecosystems: immediate, filter and founder effects. *Journal of Ecology*, 86(6), 902–910.

Hagedorn, F., Gavazov, K., & Alexander, J. M. (2019). Above- and belowground linkages shape responses of mountain vegetation to climate change. *Science*, 365(6458), 1119–1123.

Hofstra, N., New, M., & McSweeney, C. (2010). The influence of interpolation and station network density on the distributions and trends of climate variables in gridded daily data. *Climate dynamics*, 35(5), 841–858.

Hooper, D. U., Chapin III, F. S., Ewel, J. J., Hector, A., Inchausti, P., Lavorel, S., ... & Schmid, B. (2005). Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. *Ecological Monographs*, 75(1), 3–35.

Iqbal, M. T. (2012). Acid tolerance mechanisms in soil grown plants. *Malaysian Journal of Soil Science*, 16(1), 1–21.

Körner, C. (2003). *Alpine plant life: functional plant ecology of high mountain ecosystems*, 2nd edition. Springer, Germany.

Körner, C., & Hiltbrunner, E. (2018). The 90 ways to describe plant temperature. *Perspectives in Plant Ecology, Evolution and Systematics*, 30, 16–21.

Kulmatiski, A., Beard, K. H., Stevens, J. R., & Cobbold, S. M. (2008). Plant–soil feedbacks: a meta-analytical review. *Ecology Letters*, 11(9), 980–992.

Laliberté, E., Legendre, P., & Shipley, B. (2014). FD: Measuring functional diversity from multiple traits, and other tools for functional ecology. R package version. 1. 0-12.

Landolt, E., Bäumler, B., Erhardt, A., Hegg, O., Klötzli, F., Lämmler, W., ... & Urmi, E. (2010). Flora indicativa: ökologische Zeigerwerte und biologische Kennzeichen zur Flora der Schweiz und der Alpen. Haupt. Bern.

Lau, J. A., & Lennon, J. T. (2011). Evolutionary ecology of plant–microbe interactions: soil microbial structure alters selection on plant traits. *New Phytologist*, 192(1), 215–224.

Laughlin, D. C. 2011. Nitrification is linked to dominant leaf traits rather than functional diversity. *Journal of Ecology*, 99(5), 1091–1099.

Lavorel, S. (2013). Plant functional effects on ecosystem services. *Journal of Ecology*, 101(1), 4-8.

Lavorel, S., Grigulis, K., McIntyre, S., Williams, N. S. G., Garden, D., Dorrough, J., ... & Bonis, A. (2008). Assessing functional diversity in the field – methodology matters! *Functional Ecology*, 22(1), 134–147.

Le Bagousse-Pinguet, Y., Soliveres, S., Gross, N., Torices, R., Berdugo, M., & Maestre, F. T. (2019). Phylogenetic, functional, and taxonomic richness have both positive and negative effects on ecosystem multifunctionality. *Proceedings of the National Academy of Science of the USA*, 116(17), 8419–8424.

Legay, N., Baxendale, C., Grigulis, K., Krainer, U., Kastl, E., Schloter, M., ... & Poly, F. (2014). Contribution of above-and below-ground plant traits to the structure and function of grassland soil microbial communities. *Annals of Botany*, 114(5), 1011–1021.

Li, W., Epstein, H. E., Wen, Z., Zhao, J., Jin, J., Jing, G., ... & Du, G. (2017). Community-weighted mean traits but not functional diversity determine the changes in soil properties during wetland drying on the Tibetan Plateau. *Solid Earth*, 8(1), 137–147.

Louca, S., Polz, M. F., Mazel, F., Albright, M. B., Huber, J. A., O'Connor, M. I., ... & Wegener Parfrey, L. (2018). Function and functional redundancy in microbial systems. *Nature ecology & evolution*, 2(6), 936–943.

Martinez-Almoyna, C., Piton, G., Abdulhak, S., Boulangeat, L., Choler, P., Delahaye, T., ... & Münkemüller, T. (2020). Climate, soil resources and microbial activity shape the distributions of mountain plants based on their functional traits. *Ecography*, 43(10), 1550–1559.

Martinez-Almoyna, C., Thuiller, W., Chalmandrier, L., Ohlmann, M., Foulquier, A., Clément, J. C., ... & Münkemüller, T. (2019). Multi-trophic β-diversity mediates the effect of environmental gradients on the turnover of multiple ecosystem functions. *Functional Ecology*, 33(10), 2053–2064.

Mayfield, M. M., & Levine, J. M. (2010). Opposing effects of competitive exclusion on the phylogenetic structure of communities. *Ecology letters*, 13(9), 1085–1093.

Mayor, J. R., Sanders, N. J., Classen, A. T., Bardgett, R. D., Clément, J. C., Fajardo, A., & Cieraad, E. (2017). Elevation alters ecosystem properties across temperate treelines globally. *Nature*, 542(7639), 91–95.

Mazumder, R., & Hastie, T. (2012). The graphical lasso - new insights and alternatives. *Electronic Journal of Statistics*, 6, 2123–2149.

Moore, J. (1988). Arthropod regulation of microbiota and mesobiota in below-ground detrital food webs. *Annual Review of Entomology*, 33(1), 419–439.

Moorhead, D. L., Sinsabaugh, R. L., Hill, B. H., & Weintraub, M. N. (2016). Vector analysis of ecoenzyme activities reveal constraints on coupled C, N and P dynamics. *Soil Biology & Biochemistry*, 93, 1–7.

Mooshammer, M., Wanek, W., Hämmerle, I., Fuchslueger, L., Hofhansl, F., Knoltsch, A., ... & Keiblinger, K. M. (2014). Adjustment of microbial nitrogen use efficiency to carbon: nitrogen imbalances regulates soil nitrogen cycling. *Nature communications*, 5, 3694.

Mouchet, M. A., Villéger, S., Mason, N. W., & Mouillot, D. (2010). Functional diversity measures: an overview of their redundancy and their ability to discriminate community assembly rules. *Functional Ecology*, 24(4), 867–876.

Mulder, C., Ahrestani, F. S., Bahn, M., Bohan, D. A., Bonkowski, M., Griffiths, B. S., ... & Lewis, O. T. (2013). Connecting the green and brown worlds: allometric and stoichiometric predictability of above-and below-ground networks. *Advances in Ecological Research*, 49, 69–175.

Murphy, K.P. (2012). *Machine learning: a probabilistic perspective*. The MIT Press, Cambridge, USA.

Myneni, R. B., Hall, F. G., Sellers, P. J., & Marshak, A. L. (1995). The interpretation of spectral vegetation indexes. *IEEE Transactions on Geoscience and Remote Sensing*, 33(2), 481–486.

Nannipieri, P., Trasar-Cepeda, C., & Dick, R. P. (2018). Soil enzyme activity: a brief history and biochemistry as a basis for appropriate interpretations and meta-analysis. *Biology and Fertility of Soils*, 54(1), 11–19

Neuwinger, I. (1970). Böden der subalpinen und alpinen Stufen in den Tiroler Alpen. Mitteilungen der ostalpin-dinarischen Gesellschaft für Vegetationskunde, 11, 135–150.

Ohlmann, M., Mazel, F., Chalmandrier, L., Bec, S., Coissac, E., Gielly, L., ... & Chave, J. (2018). Mapping the imprint of biotic interactions on b-diversity. *Ecology Letters*, 21(11), 1660–1669.

Olander, L. P., & Vitousek, P. M. (2000). Regulation of soil phosphatase and chitinase activity by N and P availability. *Biogeochemistry*, 49(2), 175–190.

Piton G., Foulquier A., Martinez L., Legay N., Brussaard L., Hedlund K., ... & Clement, J. C. (2020). Disentangling drivers of soil microbial potential enzyme activity across rain regimes: an approach based on the functional trait framework. *Soil Biology & Biochemistry*, 148, 107881.

Piton, G., Legay, N., Arnoldi, C., Lavorel, S., Clément, J. C., & Foulquier, A. (2020). Using proxies of microbial community-weighted means traits to explain cascading effect of management intensity, soil and plant traits on ecosystem resilience in mountain grasslands. *Journal of Ecology*, 108(3), 876–893. doi 10.1111/1365-2745.13327

Quested, H., Eriksson, O., Fortunel, C., & Garnier, E. (2007). Plant traits relate to whole-community litter quality and decomposition following land use change. *Functional Ecology*, 21(6), 1016–1026.

Quintana-Segui, P., Le Moigne, P., Durand, Y., Martin, E., Habets, F., Baillon, M., ... & Morel, S. (2008). Analysis of near-surface atmospheric variables: validation of the SAFRAN analysis over France. *Journal of applied meteorology and climatology*, 47(1), 92–107.

Quintana-Seguí, P., Turco, M., Herrera, S., & Miguez-Macho, G. (2017). Validation of a new SAFRAN-based gridded precipitation product for Spain and comparisons to Spain02 and ERA-Interim. *Hydrology and Earth System Sciences*, 21(4), 2187–2201.

Roe, G. H. (2005). Orographic precipitation. *Annual Review of earth and planetary sciences*, 33(1), 645–671.

Schimel, J., Balser, T. C., & Wallenstein, M. (2007). Microbial stress-response physiology and its implications for ecosystem function. *Ecology*, 88(6), 1386–1394.

Schimel, J. P., & Bennett, J. (2004). Nitrogen mineralization: challenges of a changing paradigm. *Ecology*, 85(3), 591–602.

Schweitzer, J. A., Bailey, J. K., Fischer, D. G., LeRoy, C. J., Lonsdorf, E. V., Whitham, T. G., & Hart, S. C. (2008). Plant-soil-microorganism interactions: heritable relationship between plant genotype and associated soil microorganisms. *Ecology*, 89(3), 773–781.

Sinsabaugh, R. L., Hill, B. H., & Shah, J. J. F. (2009). Ecoenzymatic stoichiometry of microbial organic nutrient acquisition in soil and sediment. *Nature*, 462(7274), 795–798.

Stephenson, N. (1998). Actual evapotranspiration and deficit: biologically meaningful correlates of vegetation distribution across spatial scales. *Journal of biogeography*, 25(5), 855–870.

Sundqvist, M. K., Sanders, N. J., & Wardle, D. A. (2013). Community and ecosystem responses to elevational gradients: processes, mechanisms, and insights for global change. *Annual Review of Ecology, Evolution, and Systematics*, 44, 261–280.

Taberlet, P., Bonin, A., Zinger, L., & Coissac, E. (2018). *Environmental DNA: for biodiversity research and monitoring*. Oxford University Press, Oxford, United Kingdom.

Thakur, M.P. (2020). Climate warming and trophic mismatches in terrestrial ecosystems: the green–brown imbalance hypothesis. *Biology Letters*, 16(2), 20190770.

Thakur, M.P., & Geisen, S. (2019). Trophic regulations of the soil microbiome. *Trends in Microbiology*, 27(9), 771–780.

Thakur, M.P., Phillips, H.R.P., Brose, U., De Vries, F.T., Lavelle, P., Loreau, M., ... & Cameron, E.K. (2020). Towards an integrative understanding of soil biodiversity. *Biological Reviews*, 95(2), 350–364.

Tibshirani, R. (1996). Regression shrinkage and selection via the Lasso. *Journal of the Royal Statistical Society: Series B (Methodological)*, 58(1), 267–288.

Tilman, D., Knops, J., Wedin, D., Reich, P., Ritchie, M., & Siemann, E. (1997). The influence of functional diversity and composition on ecosystem processes. *Science*, 277(5330), 1300–1302.

Van der, Putten, W. H., Bardgett, R. D., Bever, J. D., Bezemer, T. M., Casper, B. B., Fukami, T., ... & Suding, K. N. (2013). Plant–soil feedbacks: the past, the present and future challenges. *Journal of Ecology*, 101(2), 265–276.

Van der Putten, W. H., Bradford, M. A., Pernilla Brinkman, E., Voorde, T. F. van de, & Veen, G. F. (2016). Where, when and how plant–soil feedback matters in a changing world. *Functional Ecology*, 30(7), 1109–1121.

Vannier, O., & Braud, I. (2012). Calcul d'une évapotranspiration de référence spatialisée pour la modélisation hydrologique à partir des données de la ré-analyse SAFRAN de Météo-France. SOMME (Synergie Observation Modélisation en Modélisation de l'Environnement)-Envirhônalp.

Vernay, M., Lafaysse, M., Hagenmuller, P., Nheili, R., Verfaillie, D., & Morin, S. (2019). The S2M meteorological and snow cover reanalysis in the French mountainous areas (1958 - present) [Data set]. AERIS. https://doi.org/10.25326/37.

Verfaillie, D., Lafaysse, M., Deque, M., Eckert, N., Lejeune, Y., & Morin, S. (2018). Multi-component ensembles of future meteorological and natural snow conditions for 1500m altitude in the Chartreuse mountain range, Northern French Alps. *Cryosphere*, 12(4), 1249–1271.

Villéger, S., Mason, N.W., & Mouillot, D. (2008). New multidimensional functional diversity indices for a multifaceted framework in functional ecology. *Ecology*, 89(8), 2290–2301.

Violle, C., Thuiller, W., Mouquet, N., Munoz, F., Kraft, N.J.B., Cadotte, M.W., ... & Mouillot, D. (2017). Functional rarity: the ecology of outliers. *Trends in Ecology & Evolution*, 32(5), 356–367.

Vionnet, V., Brun, E., Morin, S., Boone, A., Faroux, S., Le Moigne, P., ... & Willemet, J. M. (2012). The detailed snowpack scheme Crocus and its implementation in SURFEX v7.2. *Geoscientific Model Development*, 5(3), 773–791.

Wallenstein, M. D., & Weintraub, M. N. (2008). Emerging tools for measuring and modelling the in situ activity of soil extracellular enzymes. *Soil Biology & Biochemistry*, 40(9), 2098–2106.

Wardle, D. A., Bardgett, R. D., Klironomos, J. N., Setälä, H., Van Der Putten, W. H., & Wall, D. H. (2004). Ecological linkages between aboveground and belowground biota. *Science*, 304(5677), 1629–1633.

Wild, A. (1950). The retention of phosphate by soil. A review. *Journal of Soil Science*, 1(2), 221–238.

Zechmeister-Boltenstern, S., Keiblinger, K. M., Mooshammer, M., Peñuelas, J., Richter, A., Sardans, J., & Wanek, W. (2015). The application of ecological stoichiometry to plant–microbial–soil organic matter transformations. *Ecological Monographs*, 85(2), 133–155.

Biosketch

Sarah-Sophie Weil is interested in large scale biogeographic patterns and the underlying mechanisms and processes. She is currently working on the role of traits in long-distance dispersal in the context of her joint PhD at the Laboratoire d'Ecologie Alpine (LECA, France) and the Biosciences Department in Swansea (Wales, UK). This work was part of her Master's thesis in the LECA.

Author contributions: WT and TM conceived the ideas, field observations and lab measurements were carried out by the ORCHAMP consortium. WT, CMA and AS collected soil samples, GP, AS and AF analysed the enzymatic activities and LB performed the trait measurements. JR managed and maintained databases of traits, climatic variables and ORCHAMP field measurements, PC provided NDVI data, CMA calculated the climatic indices. SSW performed the modelling work and all subsequent analyses. GP, AF and JP gave additional perspectives to the paper. SSW wrote the first draft of the manuscript and all authors contributed substantially to revisions and gave final approval for publication.