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# POPULATION-LEVEL CONSEQUENCES OF INHERITABLE SOMATIC MUTATIONS AND THE EVOLUTION OF MUTATION RATES IN PLANTS

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**ABSTRACT.** Inbreeding depression, that is the decrease in fitness of inbred relative to outbred individuals, was shown to increase strongly as life expectancy increases in plants. Because plants are thought to not have a separated germline, it was proposed that this pattern could be generated by somatic mutations accumulating during growth, since larger and more long-lived species have more opportunities for mutations to accumulate. A key determinant of the role of somatic mutations is the rate at which they occur, which likely differs between species because mutation rates may evolve differently in species with constrasting life-histories. In this paper, I study the evolution of the mutation rates in plants, and consider the population-level consequences of inheritable somatic mutations given this evolution. I show that despite substantially lower somatic and meiotic mutation rates, more long-lived species still tend to accumulate larger amounts of deleterious mutations because of the increased number of opportunities they have to acquire mutations during growth, leading to higher levels of inbreeding depression in these species. However, the magnitude of this increase depends strongly on how mutagenic meiosis is relative to growth, to the point of being close to nonexistent in some situations.

# 1 Introduction

Plant growth is fueled by cell divisions occurring in meristems. Each shoot is produced by 1 an apical meristem and may bear axillary meristems, which are typically situated in the 2 axils of leaves and grow out to become the apical meristem of a new shoot upon activation 3 (Burian et al., 2016). As meristematic cells generate all the tissues constituting the shoot, 4 any mutation occurring in a meristematic cell will be borne by all the cells it gave rise to, 5 leading to genetic mosaicism within individual plants (Schoen and Schultz, 2019). Fur-6 thermore, because meristems also give rise to reproductive tissues, mutations occurring 7 during growth before the differentiation of the germline, that is somatic mutations, may 8 be present in the gametes and hence be inherited (although how frequently the inheritance 9 of somatic mutations occurs is currently unknown, Lanfear, 2018). All else being equal, it 10 follows that the larger and the older a given plant grows, the more cell divisions it under-11 goes and the more somatic mutations it should accumulate and transmit to its offspring, 12 potentially leading to a higher mutation load in more long-lived and larger species since 13 it is thought that most mutations are deleterious (Eyre-Walker and Keightley, 2007). 14

Inbreeding depression, that is the decrease in fitness of inbred relative to outbred 15 individuals (Charlesworth and Charlesworth, 1987), is thought to be mostly generated 16 by recessive deleterious mutations maintained at mutation-selection balance in popula-17 tions (Charlesworth and Willis, 2009). Hence, Scofield and Schultz (2006) proposed that 18 somatic mutations accumulation could lead to higher inbreeding depression in larger and 19 more long-lived species. Consistent with this view, inbreeding depression was indeed shown 20 to increase strongly as life expectancy increases across plant species (Duminil et al., 2009; 21 Angeloni et al., 2011). Furthermore, Bobiwash et al. (2013) showed that substantial in-22

breeding depression was generated by somatic mutations in a study performed at the 23 phenotypic level in old *Vaccinium angustifolium* clones. This is, however, the only em-24 pirical test of Scofield and Schultz (2006)'s idea. Besides, recent theoretical investigations 25 have shown that variations in inbreeding depression can in principle be generated by dif-26 ferences in the fitness effect of mutations between species with contrastring life-histories 27 (Lesaffre and Billiard, 2020), so that somatic mutations accumulation may not always be 28 needed to explain variations in the magnitude of inbreeding depression across plant species. 29 Moreover, theoretical investigations of the population-level consequences of somatic mu-30 tations accumulation are lacking, so that their role in the maintenance of high inbreeding 31 depression in long-lived species remains poorly understood. Indeed, theoretical studies 32 regarding somatic mutations in plants either focused on the case of favorable mutations, 33 conferring resistance against herbivores (e.g. Antolin and Strobeck, 1985), or studied the 34 fate of deleterious mutations subject to intra-organismal selection (Otto and Orive, 1995; 35 Pineda-Krch and Lehtilä, 2002), but never considered the population-level consequences 36 of recessive deleterious mutations (Schoen and Schultz, 2019). In summary, deleterious 37 somatic mutations accumulation has been proposed as a mechanism to explain the rarity 38 of selfing species among long-lived plants (Scofield and Schultz, 2006), consistent with 39 empirical measures of inbreeding depression, but theoretical support for this idea remains 40 scarce. 41

An important determinant of the consequences of somatic mutations accumulation is the rate at which said mutations accumulate during growth, that is the somatic mutation rate, which is defined here as the number of mutations occurring per unit of vegetative growth. This rate is likely influenced by evolutionary mechanisms similar to those affect-

ing mutation rates in general. For example, Kimura (1967) showed that mutation rates 46 should be shaped by the opposition between the increase in the number of deleterious 47 mutations borne by individuals with higher mutation rates on the one hand, which causes 48 indirect selection against genetic variants increasing mutation rates to increase, and the 49 direct fitness cost there is to increasing the fidelity of DNA replication on the other hand. 50 Besides, Lynch (2011) proposed that selection to decrease the mutation rate should be-51 come weaker than genetic drift at some point in finite populations, thereby favoring the 52 persistence of non-zero mutation rates. Nevertheless, the inheritability of somatic muta-53 tions in plants and their intrinsic link with growth and life expectancy likely contribute 54 to shape the evolution of mutation rates in a specific manner which was never tackled 55 theoretically. Great interest was however taken in empirically detecting somatic muta-56 tions and comparing mutations rates in a variety of plants species ranging from the very 57 short-lived Arabidopsis thaliana to ancient, centuries old trees. In an analysis performed 58 across many plant families, Lanfear et al. (2013) showed that taller species among pairs of 59 sister species have significantly lower rates of molecular evolution, measured as the number 60 of substitutions per site per  $10^6$  years. They argued that contrary to animals, this pattern 61 is not a mere reflection of differences in generation time, which would reflect different rates 62 of genome copying per unit of time, because somatic genome copying events contribute 63 to the inheritable genetic variation in plants. Instead, they proposed that this pattern 64 may be due to slower growth in taller species, which results in a lower number of mitosis 65 (and therefore mutations) per unit of time. Consistent with this view, it was shown at the 66 cellular level that axillary meristems cells are set aside early during the growth of a shoot 67 (Burian et al., 2016), resulting in a number of cell divisions increasing linearly with the 68

number of branching events in trees although the number of terminal branches increases exponentially. Furthermore, multiple studies showed that somatic mutation rates tend to be considerably lower in taller, more long-lived species (Schmid-Siegert et al., 2017; Plomion et al., 2018; Hofmeister et al., 2019; Orr et al., 2020; Wang et al., 2019; Hanlon et al., 2019). For instance, Orr et al. (2020) found the somatic mutation rate per generation to be only ten times higher in *Eucalyptus melliodora* than in *Arabidopsis*, despite being > 100 times larger in size.

Thus, empirical evidence indicates that more long-lived species have acquired mecha-76 nisms to reduce the amount of mutations accumulated during growth on the one hand, but 77 still present high levels of inbreeding depression on the other hand, which suggests that 78 more long-lived species still accumulate more mutations despite above mentioned limiting 79 mechanisms. The aim of the present study is to disentangle the relationship between these 80 two observations. I first study the evolution of the mutation rate in plants, and then con-81 sider the number of mutations and the magnitude of inbreeding depression maintained at 82 mutation-selection balance, given the evolutionarily stable mutation rate reached by the 83 population. To do so, I extend the work of previous authors (Kimura, 1967; Gervais and 84 Roze, 2017) to the case of a perennial population in which individuals grow as they age and 85 accumulate mutations in doing so. I obtain analytical predictions which are then tested 86 against the output of individual-centered simulations. I show that the evolutionarily stable 87 mutation rate should decrease in plants as life expectancy increases, because deleterious 88 mutations have more time to accumulate in more long-lived species. Furthermore, I show 89 that despite substantially lower per year mutation rates, more long-lived species still tend 90 to accumulate larger amounts of deleterious mutations because of higher per generation, 91

leading to higher levels of inbreeding depression in these species. However, the magnitude
of this increase depends strongly on how mutagenic meiosis is relative to growth.

# 2 Methods

Model outline. I consider a large population of hermaphroditic diploids. Individuals 94 survive between mating events with a constant probability S. Juveniles may only settle in 95 replacement of deceased individuals, so that population size is kept constant. Individuals 96 are assumed to be made of a trunk, which grows by one section between each flowering 97 event (FIG. 1). This growth model is neither intended to depict a particular kind of plants 98 nor to be a realistic model of plant growth. It was chosen because it is the simplest growth 99 model incorporating within-individual genetic mosaicism. Besides, as long as mutations 100 do not interfere with the growth process, as it is the case here (see below), more compli-101 cated growth models would only alter the age distribution of sections within individuals, 102 which should not qualitatively alter the results presented in this study provided that older 103 individuals are still made of older sections on average. 104

Mutations at the selected loci occur both during meosis and somatic growth. The 105 meiotic mutation rate of a given individual (u), which includes both mutations occurring 106 during meiosis and during the development of disposable reproductive parts, is determined 107 by its genotype at a single modifier locus. At this locus, I consider the fate of a rare mutant 108 (m) with a weak effect  $(\varepsilon)$  competing with a resident allele (M). This mutant allele is 109 codominant with the resident, so that an individual's meiotic mutation rate is given by 110  $u_{MM} = u_0, u_{Mm} = u_0 + \varepsilon$ , or  $u_{mm} = u_0 + 2\varepsilon$ , depending on its genotype at the modifier. 111 Mutations occur due to the unrepaired misincorporation of nucleotides during DNA 112

replications, or due to DNA lesions occurring between replications which are not repaired 113 in time before the next replication event (*i.e.* cell division), so that they end up being 114 incorporated in the daughter cells' genome (Gao et al., 2016). Because there is, to my 115 knowledge, no reason to expect DNA repair mechanisms to fundamentally differ between 116 meiotic and somatic cell divisions, I hypothesized that meiotic and somatic mutation rates 117 should evolve jointly to some extent. Importantly however, these two rates differ in at 118 least two ways. First, they are not defined on the same scale. Indeed, while the somatic 119 mutation rate is usually defined as a number of mutations per unit of growth, as it is the 120 case in the present model, meiotic mutation rates are defined at the scale of a reproductive 121 event. Thus, they may each cover a very different number of cell divisions, especially 122 since recent empirical evidence has shown that the number of cell divisions separating 123 axillary buds stem cells from those of the apical meristem they emerged from may be 124 much lower than previously thought due to strong quiescence mechanisms (Burian et al., 125 2016). Second, meiotic cell divisions necessarily include recombination, causing additional 126 double-strand DNA breaks and therefore giving the opportunity for more mutations to 127 occur during meiosis than during mitosis (Magni and Von Borstel, 1962). Hence, the 128 relationship between these two mutation rates is not straightforward, because different 129 genetic events may happen and different numbers of cell divisions may occur over the course 130 of a growth season and during a reprodutive event. In the absence of a more mechanistic 131 model, it is hard to give a biologically well-motivated shape to this relationship. Thus, in 132 an effort to keep the model as simple as possible, I will assume that somatic mutations 133 accumulate at rate  $\gamma u$  per unit of growth (that is, per section, FIG. 1), where  $\gamma$  is a 134 positive real number which allows one to tune the intensity of somatic mutation relative 135

to meiotic mutation. In other words, I assume there is a linear relationship between thetwo rates.

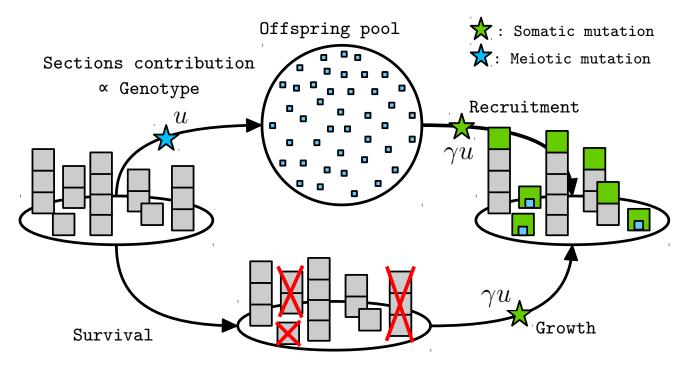


FIGURE 1: Life cycle of the modeled population. Small blue squares depict seeds. Green squares depict the sections grown during the last growing season. Juveniles go through one growing season before reproducing, and are therefore made of a single section as depicted by the green squares wrapping the small blue ones. Stars show the steps at which meiotic (blue) and somatic (green) mutation occurs. The rate at which mutation occurs is indicated beside each star.

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I assume that any section can contribute to reproduction (FIG. 1). Self-fertilisation
occurs at rate \alpha, a fraction \sigma of which imperatively occurs within the same section. The
remaining fraction 1 - \sigma can occur between sections within the individual. Introducing
\sigma into the model enables one to study the effect of within versus between sections selfing
more easily.
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A section's fecundity is determined by its genotype at a very large number of biallelic loci acting multiplicatively. At these loci, allele 0 is an healthy allele, while allele 1 is a mutated allele which diminishes the section's fecundity by a proportion s. In heterozygotes, allele 1 expresses proportionally to its dominance coefficient h. Following previous authors (Gervais and Roze, 2017), I also introduce a DNA replication fidelity cost function, f, which is an increasing function of the meiotic mutation rate u. Gervais and Roze (2017) considered a variety of cost functions and came to qualitatively similar conclusions in every case. Yet, most of their results were obtained using the cost function given in Equation (1),

$$f\left(u\right) = e^{-\frac{c}{u}},\tag{1}$$

where c is the cost of replication fidelity, which is also used in this study. Thus, the fecundity of a section is given by

$$W = f(u) \times (1-s)^{n_{\text{hom}}} (1-sh)^{n_{\text{het}}},$$
(2)

where  $n_{\text{hom}}$  and  $n_{\text{het}}$  are the number of mutations borne in the homozygous and heterozygous states, respectively.

Analytical methods. To study the model, I use the theoretical framework described in Kirkpatrick et al. (2002), which relies on indicator variables to describe individuals' multilocus genotypes. In the analytical work, the effect of the proportion of obligate within-section selfing ( $\sigma$ ) is neglected since it is difficult to incorporate and will prove to have very little impact on the results. For the sake of brevity, derivations of the results presented in the following sections are detailed in Appendices I.1 and I.2 for results regarding the evolution of mutation rate and the mutation-selection equilibrium properties <sup>163</sup> of the population given the evolutionarily stable mutation rate, respectively.

Individual-centered simulations. Individual-centered simulations were run to test the validity of analytical approximations. The simulation program was coded in C++11, is available from GitHub and has been given a DOI using Zenodo (10.5281/zenodo.5166952). In this program, individuals are represented by two chromosomes of length  $\lambda$  (expressed in cM) with the modifier situated at the center and along which mutations can occur at any position, so that infinitely many selected loci are effectively modeled (Roze and Michod, 2010).

Modeled loci. Following the work of Gervais and Roze (2017), it is assumed that 171 infinitely many alleles exist, coding for any value of  $u \in [0, +\infty)$  exist at the modifier. 172 Mutations at the modifier occur at rate  $u_m = 10^{-3}$ , and the value coded by the new 173 allele is sampled from a Gaussian distribution centered on the former allele value with 174 standard deviation  $\sigma_m = 10^{-2}$ , which is truncated at zero to prevent the modifier from 175 going out of range. At selected loci, the number of mutations occurring on a chromosome 176 during a given mutation event is sampled from a Poisson distribution with mean u ( $\gamma u$  for 177 somatic growth), and their position is sampled from a uniform distribution. Recombination 178 is modeled by exchanging segments between homologous chromosomes. The number of 179 crossing-overs is sampled in a Poisson distribution with mean  $\lambda$  and their positions are 180 sampled from a uniform distribution along chromosomes. Every time a mutation occurs, 181 the age of the section at which it occured along the individual is stored, so that the 182 genotype of any section within an individual can be reconstructed at any time from the 183 individual genome. This method allows one to gain substantial computation time because 184

mutations are stored only once per individual instead of being copied once for each new
section.

The population is kept of constant size, N. Between each Sequence of events. 187 mating event, individuals have a constant survival probability S. If they survive, they 188 grow by one section, and mutations occur at rate  $\gamma u$  in this section. If they die, they are 189 replaced by an offspring produced by the population. Any section within any individual 190 can be chosen as a parent, with a probability proportional to its fecundity (Equation 191 2). The offspring is produced by self-fertilisation with probability  $\alpha$ , in which case the 192 chosen section mates with itself with probability  $\sigma$ , and with any section within the same 193 individual with probability  $1-\sigma$ . When selfing occurs between sections, a second parental 194 section is selected within the individual. When the offspring is not produced by self-195 fertilisation, which occurs at rate  $1 - \alpha$ , it is produced by random mating and a second 196 parent is selected from the whole population. Mutation occurs at rate u during meiosis. 197

Measurements. Once the equilibrium was reached, that is when both the muta-198 tion rate and the average number of mutations per chromosome were at equilibrium, the 199 average number of mutations per chromosome in seeds, the average mutation rate and 200 inbreeding depression were measured. Although individuals are chimeric in the model, I 201 stuck with measuring inbreeding depression at the individual level to be in line with its 202 formal definition. To do so, I counted how many times each individual was chosen as 203 a parent before it died (*i.e.* I measured its lifetime reproductive success) and used this 204 quantity as a measure of lifetime fitness. Individuals were marked as being produced by 205 outcrossing (0), selfing within the same section (1), and selfing between sections within 206

the same individual (2), so that I was able to measure fitness differences between these 207 various categories of individuals. Namely, I measured inbreeding depression, that is the 208 decrease in fitness of selfed individuals relative to the outcrossed ( $\delta_{01}$ ), and autogamy de-209 pression (Schultz and Scofield, 2009; Bobiwash et al., 2013), that is the decrease in fitness 210 of within-section selfed individuals relative to between-sections ones ( $\delta_{12}$ ). Ten replicates 211 were run for each parameter set. Simulations were kept running for  $10^6$  and  $2 \times 10^5$  re-212 productive seasons for life expectancies lower and higher than 200 reproductive seasons, 213 respectively. Results were averaged over the last  $10^5$  reproductive cycles and  $2 \times 10^4$  for 214 life expectancies lower and higher than 200 reproductive seasons, respectively, and the 215 95% confidence interval around the mean was also recorded. 216

#### 3 Results

In what follows, life expectancy (E) will be used to discuss results instead of survival probability (S) for the sake of clarity and biological relevance. Given survival probability S, life expectancy can be computed as

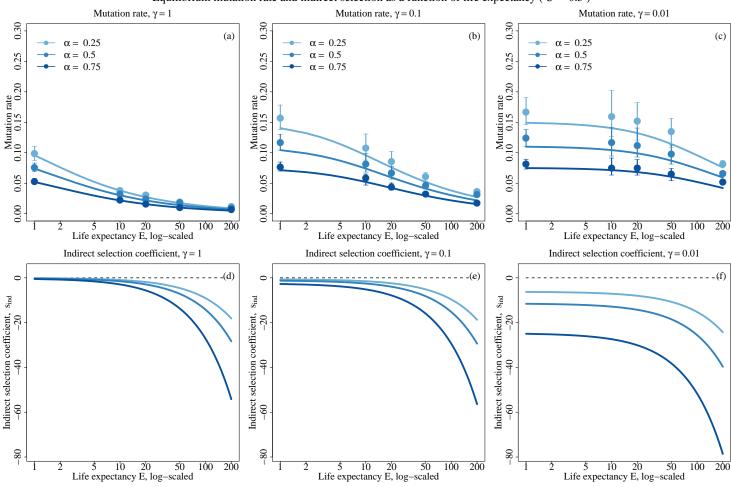
$$E = \frac{1}{1-S}.$$
(3)

#### 3.1 Evolutionarily stable mutation rate

Let us first study the evolution of the mutation rate. It is shown in Appendix I.1 that the evolution of the mutation rate is the result of the opposition between the direct cost of DNA replication fidelity, which is higher when the mutation rate is lower, and the indirect selection caused by deleterious alleles which tend to be more frequently linked with modifier alleles increasing the mutation rate (Equation A23). The resulting evolutionarily stablemutation rate is given by

$$u^* = \sqrt{-\frac{c}{\hat{s}_{ind}}},\tag{4}$$

where  $\hat{s}_{ind}$  encapsulates the intensity of indirect selection acting on the modifier. Its 226 expression is derived in Appendix I.1.5. FIGURE 2 shows the evolutionarily stable mutation 227 rate as a function of life expectancy (top row), along with the intensity of indirect selection 228 (bottom row), for cases where  $\gamma = 1$ ,  $\gamma = 0.1$  and  $\gamma = 0.01$ . I chose to focus on cases 229 where  $\gamma \leq 1$ , that is on cases where more mutations are produced during meiosis (plus the 230 production of disposable reproductive parts) than during the development of a new section, 231 on the basis of three lines of evidence. First, direct observations of plant development at 232 the cellular level indicate that cells destined to form axillary meristems undergo much fewer 233 divisions than other cells from the moment they are produced in the apical meristem, which 234 suggests that the number of cell divisions per branching event, and therefore the number of 235 opportunities for mutations to accumulate, may be lower than previously thought (Burian 236 et al., 2016). Second, estimates of somatic mutation rates per unit of growth tend to 237 be low (Orr et al., 2020). Third, to my knowledge, the only experiment comparing the 238 mutagenicity of meiosis and mitosis was performed by Magni and Von Borstel (1962) 239 in yeast. They found meiosis to be 6 to 20 times more mutagenic than mitosis, which 240 further suggests that  $\gamma$  may tend to be lower than 1. Besides, performing simulations with 241  $\gamma > 1$  proved to be very challenging since the number of mutations accumulated in the 242 population quickly became very high, causing simulations to run very slowly and consume 243 a lot of resources. 244



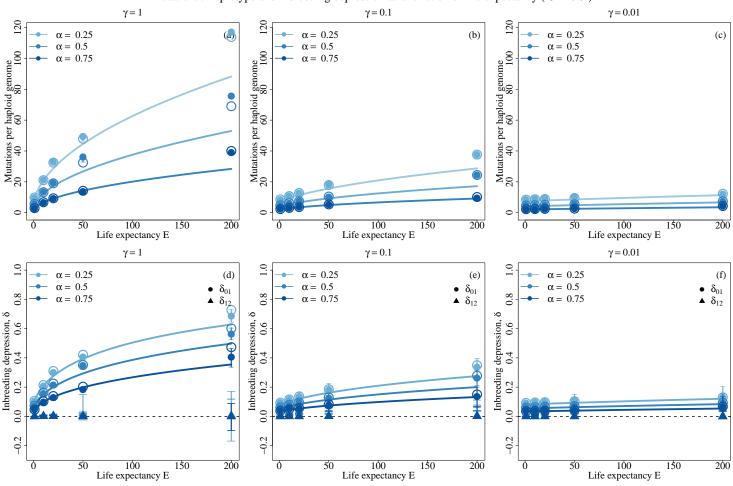
Equilibrium mutation rate and indirect selection as a function of life expectancy (  $\sigma = 0.5$  )

FIGURE 2: Evolutionarily stable mutation rate (top) and intensity of indirect selection (bottom) as a function of life expectancy (log-scaled) for various selfing rates (colors) and for  $\gamma = 1$  (left),  $\gamma = 0.1$  (middle) and  $\gamma = 0.01$  (right). Other parameters values are s = 0.05, h = 0.3, c = 0.0014,  $\lambda = 20$ , and  $\sigma = 0.5$ . Dots depict simulation results and error bars depict the 95% confidence intervals. Lines depict analytical predictions.

The evolutionarily stable mutation rate decreases with life expectancy for all  $\gamma$  values (FIG. 2a-c). In both cases, this is due to the greater number of opportunities to accumulate deleterious mutations in more long-lived species because they go through more growth events, which in turn causes indirect selection to increase against alleles increasing the mutation rate because deleterious mutations become more numerous (FIG. 2d-f).

The mutation rate also decreases as the selfing rate  $(\alpha)$  increases, which may seem

counter-intuitive since selfing tends to reduce the number of deleterious mutations seg-251 regating in the population through purging Roze (2015). However, self-fertilisation also 252 causes genetic associations between selected loci and the modifier to increase, thereby in-253 creasing indirect selection and resulting in a decrease of the evolutionarily stable mutation 254 rate when the selfing rate increases as shown by Gervais and Roze (2017). The results pre-255 sented in FIG. 2 were obtained assuming half of selfing events occured imperatively within 256 the same section ( $\sigma = 0.5$ ). Cases with  $\sigma = 0$  and  $\sigma = 1$  were also investigated and yielded 257 very similar results, which are presented in FIG. S3 and S4, respectively, in Appendix II. 258 The very small effect of  $\sigma$  on the results is due to the relatively low evolutionarily stable 259 mutation rate, which causes few somatic mutations to occur during growth, and to the 260 fact that weak selection was assumed so that mutations have little effect on their bearer's 261 fitness. 262



Mutations / haplotype and inbreeding depression as a function of life expectancy (  $\sigma=~0.5$  )

FIGURE 3: Average number of mutations per haploid genome (top) and inbreeding depression (bottom) as a function of life expectancy (log-scaled) for various selfing rates (colors) and  $\gamma = 1$ (left),  $\gamma = 0.1$  (middle) and  $\gamma = 0.01$  (right). Other parameters values are s = 0.05, h = 0.3, c = 0.0014,  $\lambda = 20$ , and  $\sigma = 0.5$ . Filled dots depict simulation results and error bars depict the 95% confidence intervals. Lines depict analytical predictions. Open circles depict the value predicted by our analytical model when the equilibrium mutation rate from simulations is used instead of Equation 4. On the bottom row, dots indicate inbreeding depression ( $\delta_{01}$ ), while triangles indicate autogamy depression ( $\delta_{12}$ ).

#### 3.2 Mutation-selection balance

<sup>263</sup> Once the mutation rate has reached an equilibrium and the population is at mutation-<sup>264</sup> selection balance, I show in Appendix I.2.1 that a leading order approximation of the average number of mutations per haploid genome in juveniles (n) is given by

$$n \approx \frac{\hat{u}^*}{s \left[h + F(1-h)\right]} - \gamma u^* \frac{S}{1-S},$$
(5)

where  $u^* = \sqrt{-\frac{c}{\hat{s}_{ind}}}$ , and  $\hat{u}^* = \left(1 + \frac{\gamma}{1-S}\right)u^*$  depicts the total mutation rate of the population over the course of one timestep, including both meiotic and somatic mutations. As for inbreeding depression calculated between outcrossed and selfed individuals  $(\delta_{01})$ , it is given by

$$\delta_{01} = 1 - \exp\left[-s(1-2h)\frac{1+F}{2}\left(\frac{\hat{u}^*}{s\left[h+F(1-h)\right]} - \gamma u^*\frac{S}{1-S}\right)\right],\tag{6}$$

where  $F = \frac{\alpha}{2-\alpha}$ , to leading order in s. Again, I do not consider the impact of the 270 proportion of selfing occurring within or between sections ( $\sigma$ ) in the analytical model since 271 it is negligible. FIGURE 3 shows the number of mutations per haploid genome among 272 juveniles (n, top row), and inbreeding and autogamy depression ( $\delta_{01}$  and  $\delta_{12}$ , bottom 273 row) at mutation-selection balance. Deviations between analytical predictions (lines) and 274 simulations results (dots) are observed. They can be explained by the slight differences 275 between the predicted evolutionarily stable mutation rate and the equilibrium mutation 276 rate reached by simulations, which build up large differences in n when life expectancy 277 becomes high. Indeed, when the equilibrium mutation rate from the simulations is used 278 to predict n instead of Equation (4), the agreement between predictions (open circles) and 279 simulation results (dots) is restored. 280

The number of mutations maintained n increases with life expectancy in every case due to the greater amount of opportunities for mutations to accumulate in more long-

lived species. Indeed, the denominator of the first term in Equation (5) shows that the 283 intensity of selection is independent of life expectancy, while the total mutation rate  $\hat{u}^*$  is 284 an increasing function of life expectancy in all investigated cases despite the fact that the 285 equilibrium mutation rate per mutagenic event  $(u^*)$  decreases in more long-lived species 286 (FIG. S1 in Appendix II shows the total mutation rate as a function of life expectancy in 287 said cases). The increase of n with life expectancy becomes much lower when  $\gamma$  decreases 288 to the point of being barely noticeable with  $\gamma = 0.01$ , despite the fact that the equilibrium 289 meiotic mutation rate is slightly higher in that case. This result is generated by the joint 290 effect of  $\gamma$ , which reduces the contribution of somatic mutations as it decreases, and of 291 the evolution of mutation rate which is lower at the evolutionary equilibrium in more 292 long-lived species (as an additionnal illustration, FIG. S2 in Appendix II compares the 293 obtained  $\hat{u}^*$  with the one expected if the evolutionarily stable mutation rate for annuals, 294 that is E = 1, is assumed for all life expectancies for various  $\gamma$  values). As a result of these 295 effects, inbreeding depression gets lower as  $\gamma$  decreases and increases as life expectancy 296 increases, but this increase becomes less and less marked for smaller  $\gamma$  values. Furthermore, 297 consistent with the negligible effect  $\sigma$  had on the evolution of the mutation rate, almost 298 no autogamy depression is generated (triangles in FIG. 2, bottom row). 299

# 4 Discussion

In this paper, I studied the evolution of the mutation rate when mutations accumulating during growth are assumed to be inheritable, and considered the consequences of such mutation accumulation for mutation load and inbreeding depression in species with varying degrees of perenniality.

#### 4.1 Evolution of the mutation rate

I showed that the evolutionarily stable mutation rate decreases as life expectancy increases 304 because of the greater number of opportunities to accumulate mutations during growth 305 in more long-lived species, which makes indirect selection against alleles increasing the 306 mutation rate stronger. However, although the mutation rate per mutagenic event (u), 307 that is per growth season or per meiosis in the present model, decreased in more long-lived 308 species, the total mutation rate  $(\hat{u})$ , that is the rate at which mutations entered the pop-309 ulation through both somatic growth and meiosis, increased. Hence, results indicate that 310 while we should expect more efficient mechanisms reducing the accumulation of deleteri-311 ous mutations during growth to evolve in more long-lived species, so that their per unit of 312 growth and per year mutation rate should be lower, their per generation mutation rates 313 should still be higher. These predictions are in line with empirical evidence, which suggest 314 that mutation rates per generation tend to be higher in more long-lived species although 315 the mutation rates per unit of growth tend to be lower (Hofmeister et al., 2019; Hanlon 316 et al., 2019; Orr et al., 2020). 317

I modeled the evolution of the mutation rate following the work of Kimura (1967), by 318 assuming there is a direct fitness cost to DNA replication fidelity opposing the indirect 319 selection generated by deleterious mutations linked to the modifier, so that the mutation 320 rate was maintained greater than zero in response to a trade-off. An alternative mecha-321 nism, which is not mutually exclusive with the trade-off described above, was put forward 322 by Lynch (2011). They proposed that selection should always act to reduce the mutation 323 rate, down until it becomes so low that the selective advantage brought by any further 324 reduction should be overwhelmed by genetic drift, thus maintaining non-zero mutation 325

rates because alleles further decreasing the mutation rate should at some point become 326 effectively neutral, and thereby creating a lower bound for the evolution of the mutation 327 rate (Lynch, 2011). This lower bound is inevitably influenced by effective population size, 328 as it plays on the relative strength of selection and genetic drift. In the present model, 329 I overlooked Lynch (2011)'s lower bound by assuming a large and fixed population size. 330 Yet, effective population sizes are expected to be higher in more long-lived species in 331 which generations overlap (Felsenstein, 1971; Charlesworth, 1980; Petit and Hampe, 2006; 332 Duminil et al., 2009), which implies the lower bound described by Lynch (2011) should 333 be met for lower mutation rates in said species. Hence, we should expect the decrease 334 in the evolutionarily stable mutation rate described in this study to become sharper in 335 conditions where Lynch (2011)'s lower bound is expected to matter for the evolution of 336 the mutation rate. 337

#### 4.2 Inbreeding depression

The larger total mutation rate in more long-lived species led to the maintenance of more 338 mutations in the population at mutation-selection balance, and therefore to higher in-339 breeding depression in these species, consistent with results from meta-analyses which 340 found inbreeding depression to increase in larger-statured, more long-lived species (Du-341 minil et al., 2009; Angeloni et al., 2011). Importantly however, the magnitude of the 342 increase in the total mutation rate, and therefore in inbreeding depression with life ex-343 pectancy depended strongly on the relative mutagenicity of meiosis and growth, which 344 was controlled by the  $\gamma$  parameter in this model. Indeed, while the increase in inbreeding 345 depression was strong when  $\gamma$  was close to 1, that is when the same amount of mutation 346

was produced during meiosis and during growth between two flowering seasons, it became 347 smaller as  $\gamma$  decreased, to the point of being barely noticeable for  $\gamma = 0.01$ . This was due 348 to both the decrease of the evolutionarily stable total mutation rate  $(\hat{u}^*)$  and to the de-349 crase of  $\gamma$ , which made the contribution of somatic mutations to the mutation load more 350 and more negligible compared with meiotic mutations. Hence, according to the results 351 presented in this paper, for somatic mutations to be the main driver of the empirically 352 observed increase in inbreeding depression in more long-lived species, roughly the same 353 amount of mutations should be produced during growth between two flowering seasons 354 and during reproduction. 355

#### 4.3 Mating system evolution

Inbreeding depression is thought to be one of the main factors preventing the evolution of 356 self-fertilisation (Lande and Schemske, 1985; Barrett and Harder, 2017). In Angiosperms, 357 consistent with the observed increase in inbreeding depression in more long-lived species, 358 there exists a strong correlation between mating systems and life-histories. Indeed, many 359 self-fertilising species are annuals whereas most long-lived species are strictly outcrossing 360 (Barrett and Harder, 1996; Munoz et al., 2016). Thus, somatic mutations accumulation 361 was proposed as an explanation for this correlation (Scofield and Schultz, 2006). While the 362 results presented in this study indicate that inbreeding depression increases with respect 363 to life expectancy due to somatic mutations accumulation, particularly when  $\gamma$  is large, 364 this increase is tempered by the decrease of the evolutionarily stable mutation rate with 365 life expectancy. Furthermore, in agreement with results obtained by Gervais and Roze 366 (2017), I showed that the evolutionarily stable mutation rate decreases as the selfing rate 367

increases because the modifier becomes more strongly associated with selected loci. These 368 decreases of the mutation rate with respect to mating system and life expectancy, together 369 with the purging effect of self-fertilisation (Roze, 2015), result in a substantial drop in the 370 magnitude of inbreeding depression as the selfing rate increases in more long-lived species, 371 potentially opening the way for the evolution of self-fertilisation. Hence, whether somatic 372 mutations accumulation is sufficient to explain the correlation between life-history and 373 mating system in Angiosperms when the mutation rate is allowed to evolve jointly with 374 the mating system remains an open question. 375

#### 4.4 Autogamy depression

In order to empirically estimate the contribution of somatic mutations accumulation to 376 inbreeding depression using phenotypic data, a method was developed by Schultz and 377 Scofield (2009). This method, called the autogamy depression test, relies on the compar-378 ison of the fitnesses of individuals produced by selfing within an inflorescence with those 379 of individuals produced by selfing between distant inflorescences on the plant's crown 380 (Schultz and Scofield, 2009; Bobiwash et al., 2013). In this paper, I performed such test 381 by measuring autogamy depression ( $\delta_{12}$ ). Contrary to inbreeding depression, I found auto-382 gamy depression to be almost null in every case, even in situations where the contribution 383 of somatic mutations accumulation to inbreeding depression was high. This result can be 384 explained by the low evolutionarily stable mutation rates, and by the fact that we only 385 considered mutations with a weak fitness effect. It suggests that the autogamy depression 386 test should only be able to detect mutations with a large fitness effect in large individ-387 uals, where mutations have had time to accumulate. Thus, it implies that detecting no 388

autogamy depression in a given population cannot be taken as evidence of a negligible
contribution of somatic mutations accumulation to the population's mutation load.

# 4.5 Germline segregation and relative mutagenicity of growth and meiosis

The results presented above suggest that valuable insights into the evolutionary relevance of somatic mutations and the evolution of the mutation rate in plants could be gained by further investigating the  $\gamma$  parameter in this model, which depicts the relative mutagenicity of meiosis and growth between two flowering seasons, and is likely influenced by at least three important factors that were either overlooked or only partially accounted for in this study.

#### 4.5.1 Relative mutagenicity of meiosis and mitosis

First, it is necessarily influenced by how mutagenic meiotic divisions are in comparison with mitotic divisions, about which little is known although one may expect meiotic divisions to generate more mutations, as they generate many more double strand DNA breaks which are required for recombination and are known to be particularly mutagenic events (Magni and Von Borstel, 1962; Arbel-Eden and Simchen, 2019).

#### 4.5.2 Number of cell divisions separating meristems

Second, it is influenced by the number of mitoses occurring between flowering buds. This number depends on the growth habit of the considered species, because fast growing species undergo more mitoses per unit of time than slow growing species, and because the rate at which mitoses occur, and thus the growth rate, may interact with the evolution of the <sup>406</sup> mutation rate. For instance, investing in a higher fidelity of DNA replication may tend to
<sup>407</sup> slow down individual growth.

The number of mitoses separating two meristems also depends on patterns of meris-408 tematic stem cell divisions that were recently brought to light (Burian, 2021). Indeed, 409 although it has long been thought that the germline remains unsegregated up until a 410 meristem switches to the floral state in plants, Burian et al. (2016) showed that within 411 the apical meristem, the stem cells give rise to a specific cell lineage which will serve as 412 the axillary meristems' stem cells and spend most of their time in a quiescent, almost 413 non-dividing state, contrary to surrounding cell lineages which divide vigorously to effect 414 plant growth. Thanks to this mechanism, the number of mitoses separating two meristems 415 is greatly reduced and so is the number of cell divisions separating the seed from the ga-416 metes, as the germline directly emerges from these stem cells. Hence, although it is clear 417 that the plant germline remains undifferentiated up until reproduction is triggered, it may 418 be considered segregated prior to differentiation, because the cells giving rise to it do not 419 suffer the same fate as surrounding somatic cell lineages, thus behaving as a functional 420 germline (Romberger et al., 1993; Burian, 2021). The exact timing of such segregation 421 during plant development is, however, not known (Lanfear, 2018). Therefore, it is impor-422 tant to point out that the results presented in this study not only hold if the germline 423 segregates late in development, but that they would also hold if the germline was actually 424 segregated as early as the first embryonic cell division (as it is the case in animals) and 425 remained sheltered within meristems. Indeed, such segregated germline would still have 426 to go through a non-zero number of mitotic cell divisions to be passed from one meristem 427 to the next due to developmental constraints (Burian, 2021), so that the number of cell 428

divisions it goes through before reproduction would still be affected by individual growth and be higher in more long-lived, larger species. In summary, the validity of the results presented in this paper does not depend on the degree to which the germline is actually segregated in plants, but the existence of a functional germline as described above, irrespective of when germline segregation occurs, supports the idea that plants acquired physiological mechanisms favoring lower values of  $\gamma$ .

#### 4.5.3 Intra-organismal selection

Finally, apart from mechanisms reducing the amount of mutations produced during growth, 435 deleterious mutations may also be affected by intra-organismal selection, which may not 436 only reduce the growth rate by eliminating mutated cells, but also efficiently purge delete-437 rious mutations from the organism, so that little to no somatic mutation may be present 438 in the gamete, which could make  $\gamma$  smaller among the mutations effectively transmit-439 ted to offspring. This could in turn affect the evolution of the mutation rate. Little is 440 known, however, about the actual efficacy of intra-organismal selection in removing dele-441 terious mutations since it was seldom investigated theoretical (Otto and Orive, 1995), and 442 mostly empirically demonstrated to occur in the case of strongly beneficial mutations (e.g. 443 Edwards et al., 1990; Simberloff and Leppanen, 2019). 444

The various elements discussed above show that  $\gamma$  is an emerging property of the interaction between a variety of physiological mechanisms rather than a fixed quantity, which advocates for the development of theoretical models treating it as such rather than as a fixed parameter, by incorporating growth, mutation and selection at the cellular level.

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