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Monodispersed MOF-808 nanocrystals synthesized via scalable room-temperature approach for efficient heterogeneous peptide bond hydrolysis

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Abstract:

Zr(IV)-based metal-organic frameworks (MOFs) such as the Zr(IV) trimesate MOF-808 are promising materials for catalytic applications. In this work, we report an aqueous solution-based room temperature strategy to produce well-defined monodispersed MOF-808 nanocrystals down to 35 nm with a high space-time yield, up to 2516 g/ m³/ day, and an excellent crystallinity and porosity. The resulting nanocrystals show remarkable colloidal dispersion during one day in a wide range of nanoparticles concentrations. As a result, 35 nm MOF-808 colloidal-level nanocrystals exhibit the highest rate of selective peptide bond and protein hydrolysis among reported Zr(IV)-based MOFs. This result may open new opportunities for highly efficient peptide or protein hydrolysis using scalable nano-catalyst.

1. Introduction

In the past decades, nanotechnology has generated interest in many fields, including bio-applications,¹ sensing² and catalysis.³ Metal-Organic Frameworks (MOFs) are ordered hybrid porous materials that are promising for a wide scope of potential applications,⁴⁻⁷ including new properties upon downsizing.⁸⁻¹⁰ Nano-MOFs are particularly appealing in catalysis for their highly tunable porosity and reactivity (presence of acid/basic sites, redox active cations...) that can even sometimes surpass previously studied nanomaterials.¹¹ However, formulation of MOFs as high quality nanomaterials has been relatively underdeveloped despite a few well-developed strategies (e.g., microwave,¹² modulating chemical,¹³ spray-drying¹⁴). Often, the produced nano-MOFs are lacking in uniformity, crystallinity/porosity and even more frequently, are obtained without size control.^{15,16} Thus, the development of new high quality MOF nanomaterials with controllable size may open some new perspectives for catalysis. For example, one emerging application for MOFs is their potential use as artificial nanozymes for middle down proteomics. For accurate protein fragment identification via mass spectrometry, proteins require partial breakdown into smaller fragments, which is achieved through hydrolysis of the highly stable peptide bond ($t_{1/2} \sim 400$ years).¹⁷ It has been shown that peptide bond hydrolysis can be catalyzed by some metal salts that can however be prone to hydro-gel formation when used in physiological conditions.¹⁸⁻²² This problem can be overcome through incorporation of the active metal salts into soluble complexes or MOF materials. A few MOFs have already been proven to be active towards hydrolysis of peptide bonds in a range of dipeptides and proteins.²³⁻²⁵ In addition, they are more cost effective than traditional enzymatic methods due to their stability

and reusability. Therefore, nanoMOFs are highly promising for the development of extremely efficient catalysts to produce peptides suitable for middle down proteomics.

Zr(IV)-carboxylate based MOFs are known as a subclass of MOFs that usually provide high chemical robustness.²⁶ A prominent example, UiO-66 (University of Oslo), was first reported in 2008 by Lillerud and co-workers and later studied for many potential applications.²⁷⁻²⁹ MOF-808 is another benchmark solid that is promising for catalytic applications, where the trimesate linker is used to bridge Zr_6 oxoclusters with a molar-ratio of 2:1 to give a cubic structure with potentially unsaturated coordination sites, high BET surface area (2060 m²/g) and a large pore size (18.4 Å).³⁰⁻³² Synthesizing Zr(IV)-based MOFs mostly relies on solvothermal strategies using toxic solvents (e.g. DMF), which is not an environmentally-friendly method and is not suitable for some applications such as the synthesis of host-guest architectures using the bottle-around-ship approach.^{33,34} Nonetheless, a few pioneering works have reported the room temperature (RT) synthesis of Zr(IV)-carboxylate based MOFs but most of these attempts were dedicated to 12-connected (each metal cluster coordinated to 12 organic linkers) UiO-type MOFs and the use of DMF was often inevitable.³⁵⁻⁴¹ Considering the appealing properties of MOF-808 and its use in several well-documented advanced applications,³¹ developing an approach to produce MOF-808 nanocrystals at room temperature using environmentally friendly conditions appears really promising.

Herein, we report a two-step, room temperature approach for the synthesis of MOF-808 in environmentally friendly conditions. The MOF nucleation-growth kinetics was investigated by in-situ powder X-ray diffraction (PXRD), which facilitated the understanding of the MOF synthesis. Tuning the synthesis conditions allowed us to obtain monodisperse MOF crystals from sub-micron scale (850 nm) down to the nanoscale (35 nm) via a concentration-induced method,

which simultaneously enabled a space-time yield (STY) reaching 2516 kg/m³/day for the smallest nano-MOF-808. To the best of our knowledge, this is the first study tuning the crystal size of MOF-808 with such a high yield at room temperature. Finally, we demonstrated not only the high long-term colloidal stability of the synthesized nano-MOF-808 in a large concentration range but also the effect of particle's size on the efficiency of the highly selective peptide bond hydrolysis in a Hen Egg White Lysozyme (HEWL) protein by MOF-808. Notably, the remarkable catalytic efficiency of nano-MOF-808 allows us to obtain protein fragments of suitable size, which are in the range for the analysis in the emerging middle-down proteomic applications.

2. Experimental

All chemicals were purchased from commercial suppliers and used as received without further purification. Trimesic acid, 98%, Alfa Aesar. ZrCl₄ anhydrous, 98%, Acros. Zirconyl chloride octahydrate, 98%, Acros. Isopropyl alcohol, 99+%, Sigma. Formic acid, 98%, Acros. Acetic acid, 99+%, Fisher. Acetone, 99%, Acros. Ethanol absolute, >=99%, Acros. Distilled water, Millipore system.

Synthesis of Zr₆ oxoclusters:

ZrCl₄ (200g, 0.84 mol) was added in a mixture of 300 mL of acetic acid and 500 mL of isopropanol under stirring at 500 rpm and heated at 120 °C for 60 min. The product was collected either through suction filtration or centrifugation at 10000 rpm. The collected white solid was subsequently washed with acetone twice and dried under vacuum at room temperature. The dry powder mass was 175 g, corresponding to 98% of product yield (calculation based on the Zr salt) and to a space-time yield of 5340 kg/m³/day.

Synthesis of MOF-808:

MOF-808-DMF: MOF-808 was obtained via solvothermal synthesis in N,N-dimethylformamide (DMF) at 110 °C for 48 hours using a method adapted from literature.³⁰ Typically, 1,3,5-benzenetricarboxylic acid (1.49 g, 7 mmol) was added to N,N-dimethylformamide (128 mL) in a thick glass jar while stirring at 600 rpm. ZrOCl₂·8H₂O (2.16 g, 6.7 mmol) and formic acid (133 mL) were subsequently added and the reaction mixture was stirred 20 minutes at room temperature. The jar was then heated at 110 °C for 48 hours. A white solid was collected by centrifugation at 14500 rpm for 5 minutes and washed in N,N-dimethylformamide (250 mL) at 100 °C overnight under stirring at 350 rpm. The solid was collected again by centrifugation and further washed in acetone (250 mL) at 60 °C overnight under stirring at 600 rpm. The solid was finally collected by filtration and dried in air.

850 nm MOF-808: Zr₆ oxoclusters (0.6 g) was dispersed in formic acid (3 mL) under stirring at 600 rpm. H₂O (5 mL) was subsequently added and the reaction mixture was stirred until being completely colorless. 1,3,5-Benzenetricarboxylic acid (150 mg, 0.7 mmol) was then added and the reaction was stirred overnight. The solid was collected by centrifugation at 14500 rpm for 5 minutes and then simply washed with 40 mL of H₂O and 40 mL of EtOH once, respectively. The collected solid was dried in vacuum for 3 hours affording a white powder.

210 nm MOF-808: A similar procedure as for 850 nm MOF-808 was used with solvent volumes divided by 2 (1.5 mL of formic acid and 2.5 mL of H₂O).

60 nm MOF-808: A similar procedure as for 850 nm MOF-808 was used with solvent volumes divided by 4 (0.75 mL of formic acid and 1.25 mL of H₂O).

35 nm MOF-808: Zr₆ oxoclusters (1.2 g) was dispersed in formic acid (1 mL) under stirring at 600 rpm. H₂O (1.67 mL) was subsequently added and the reaction mixture was stirred until

being completely colorless. 1,3,5-Benzenetricarboxylic acid (300 mg, 1.4 mmol) was then added and the reaction was stirred for 5 h. The solid was collected by centrifugation at 14500 rpm for 60 minutes and then simply washed with 40 mL of H₂O and 40 mL of EtOH once, respectively. The collected solid was dried in vacuum for 3 hours affording a white powder.

MOF activation for hydrolysis reaction: Prior to being used as catalyst for peptide and protein hydrolysis, MOFs were activated in batches of 50 mg in 0.5 M HCl - acetone solution for 72 h (3 x 2 mL) and methanol for 24 h (2 x 2 mL). After that, the MOFs were dried at 70°C for 2 h and activated at 160°C for 24 h at atmospheric pressure.

Glycylglycine hydrolysis: For each experiment, 2 μmol of MOF and 1 mL of glycylglycine (GG) in D₂O (2 mM) were mixed and pD was adjusted with NaOD and DCl to pD 7.6. Each sample was incubated at 60 °C for 20 h. 600 μL aliquots were taken at different time points, centrifuged and the resulting supernatant, to which internal standard TMSP-d₄ was added, was analyzed with ¹H NMR (Bruker 300 MHz, 32 scans, Topspin 4.0.8 software).

Protein hydrolysis: 0.1 mM protein solution (1 mL) was mixed with 2 μmol MOF and incubated at 60 °C at pH 7.2 for 72 h. 100 μL aliquots were taken for each time point, and analyzed with sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE).

Electrophoresis: SDS-PAGE was performed with a 0.5 M Tris - HCl buffer (pH 6.8) 4% (w/v) polyacrylamide stacking gel and a 1.5 M Tris - HCl buffer (pH 8.8) 16% (w/v) polyacrylamide resolving gel. 15 μL of reaction mixture and 5 μL of buffer were combined and incubated at 95 °C for 5 min prior to the loading of 10 μL into gel wells. Page Ruler unstained low range protein ladder was used as the molecular weight standard, and gels were run at 200 V for 2 h. Subsequently, the gels were stained with Coomassie Blue and imaged with a Gel Doc EZ Imager and Bio-Rad Image Lab software.

3. Results and discussion

In the MOF-808 framework, Zr_6 -nodes are at the vertices and trimesate linkers at the faces of a tetrahedral cage with internal pore size of 4.8 Å. Ten contiguous tetrahedral cages are assembled to form a large adamantane cage with an inner diameter of 18.4 Å (**Figure 1a**) assembled in a *spn* topology. The room temperature synthesis of MOF-808 was performed using preformed octahedral Zr_6 oxoclusters, which is the most common SBU (secondary building unit) in the family of Zr(IV)-carboxylate based MOFs. The synthesis of the Zr_6 oxoclusters was carried out by using a scalable reflux method in a mixture of isopropyl alcohol and acetic acid at 100 °C during 1 h. In order to confirm the composition of the obtained species, ESI-MS (electrospray ionization mass spectroscopy) was applied. As shown in Figure S3, the spectrum pattern observed from solution evidenced the presence of Zr_6 oxoclusters, with an average formula of $Zr_6O_4(OH)_4(C_2H_3O_2)_8(H_2O)_2Cl_3$ that is consistent with the atomic ratio (67% /33%) between Zr/Cl obtained from EDX (Energy-dispersive X-ray spectroscopy, see Figure S1). The efficient preparation of Zr_6 oxoclusters was performed with a high space-time yield (~5340 kg/ m³/ day) compatible with up-scaled synthesis (>100 g). From a lab-scale point of view, we demonstrated (Figure S2) that these Zr_6 acetate oxoclusters can be synthesized either on a small or large scale (100 g) without any drop in yield and quality. This new environmentally friendly preparation of Zr_6 oxoclusters with high STY surpassed the previously reported ones that were either performed in toxic solvents or present limited scalability.^{36, 40} MOF-808 synthesis was then performed by simply mixing Zr_6 oxoclusters with formic acid, water and BTC (trimesic acid) under stirring at 600 rpm. The alternative use of acetic acid led to completely clear solution without any product after 7 days at room temperature and therefore, formic acid was used for all the synthesis in this

work. After 18 h of stirring, the appearance of a white slurry indicated the formation of MOF-808-RT (RT: Room Temperature). The synthesis yield was calculated based on the amount of ligand, reaching almost 100 %. After washing with water and ethanol once respectively, powder X-ray diffraction (PXRD) (**Figure 1b**) demonstrated the successful synthesis of highly crystalline MOF-808 by comparison to the simulated diffraction pattern. Taking into account the ease of the washing step, this water-based green synthesis has a great advantage in comparison to the DMF-based one, where 10 days are needed to fully activate the MOF.^{30, 42}

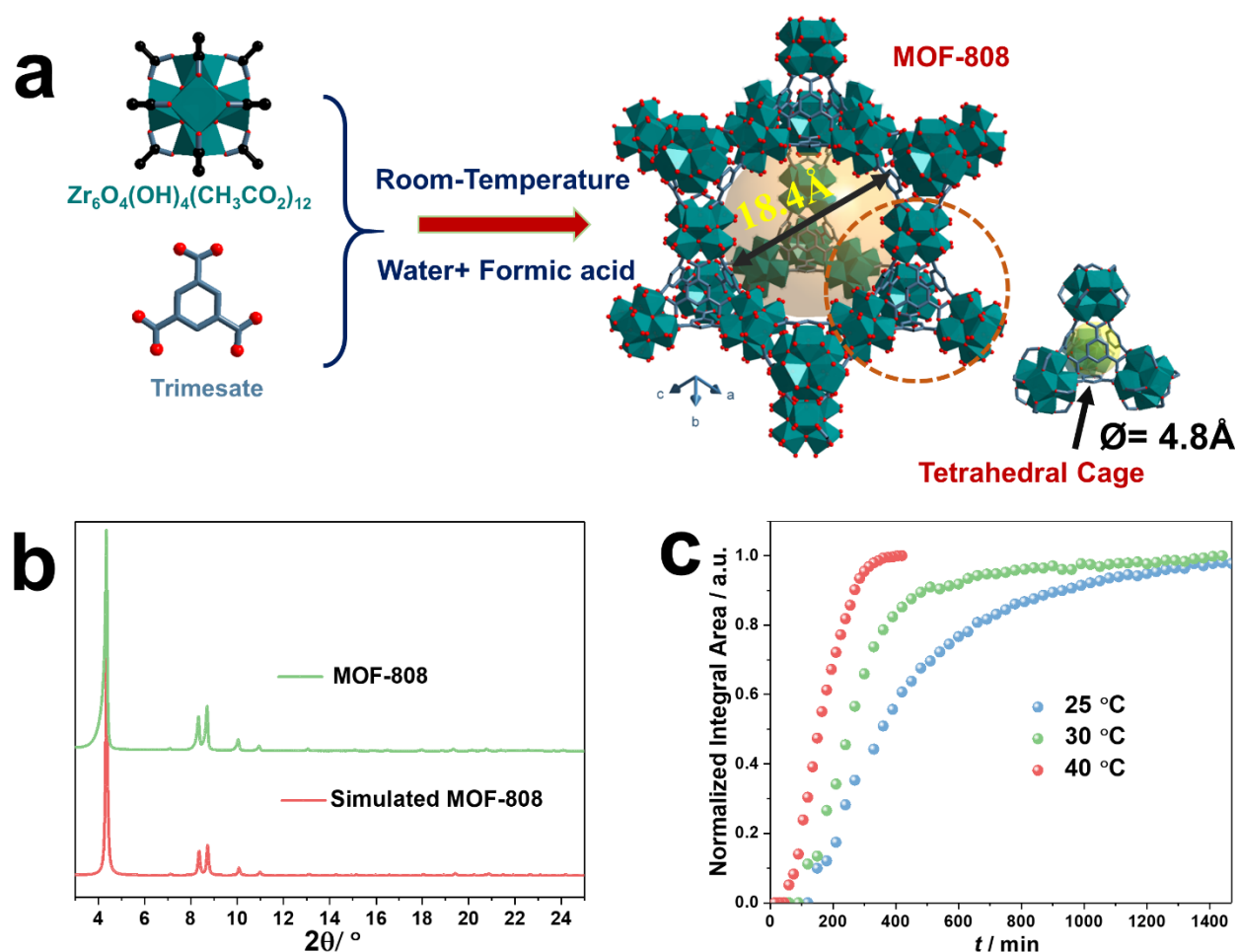


Figure 1. (a) Structures of $Zr_6O_4(OH)_4(CH_3CO_2)_{12}$ nodes, trimesate linker, and the MOF-808 (tetrahedral and adamantane-shaped cages); (b) PXRD ($\lambda_{Cu} = 1.5406 \text{ \AA}$) pattern of MOF-808 and the corresponding simulated PXRD pattern; (c) Normalized integral intensities of (3 1 1) Bragg reflection as a function of time (t) for different temperatures.

MOF-808-RT displayed a typical N₂ sorption isotherm at 77 K (Figure S4) with a high N₂ adsorption capacity of 512 cm³/g along with a high Brunauer-Emmett-Teller (BET) surface area (2090±17 m²/g), which matches very well with the reported value (2060 m²/g).³⁰ The pore size of MOF-808-DMF, which was calculated using density functional theory (DFT) on a sample prepared using the reported synthesis in DMF, showed two types of apertures at ca. 6.5 Å and 15.5 Å, corresponding to the windows of the tetrahedral and adamantane-shaped cages respectively (Figure S5). In the case of MOF-808-RT, the peak at 6.5 Å was not observed, whereas the large pore shifted to 18.6 Å and an additional new peak at 12.5 Å was detected, suggesting very likely the presence of low temperature-induced defects (LTID).⁴¹ Infrared spectra (IR) (Figure S6) evidenced the absence of a C=O stretching band at around 1700 cm⁻¹ - corresponding to uncoordinated COOH groups- in the washed MOF-808-RT solid, suggesting that the compound does not present missing nodes. Thermogravimetric analysis (TGA) (Figure S7) proved that each Zr₆ node of MOF-808-RT is connected to 1.82 trimesate ligands, compared to 2 for the defectless structure, suggesting that 9% of linkers are missing, which might explain the slightly increased pore size compared to the ideal structure.

In order to shed light on the crystallization process, we performed in-situ diffraction measurements using a laboratory X-ray powder diffractometer in order to get a quantitative insight into the nucleation kinetics (e.g., activation energy, reaction rate).^{43, 44} The reaction kinetics has been studied at 25 °C, 30 °C and 40 °C, respectively. Notably, such temperature-dependent in-situ kinetic studies were usually restricted to hydro/ solvo-thermal synthesis and therefore relying on the use of synchrotron pressure-resistant systems. Here, the ambient conditions and relatively fast kinetics allowed us to investigate the thermal kinetics of a MOF material using a laboratory diffractometer. Figure S8 evidenced the direct formation of MOF-808

without any other phase or impurity. The integrated intensity of the (3 1 1) reflection was monitored as a function of crystallization time (**Figure 1c**), evidencing that even a slight increase in temperature from 25 °C to 40 °C induces a much faster reaction. One could therefore imagine multiplying by 5 the space-time yield by a temperature increase of 15 °C (Figure S9 and Table S2). The estimated Arrhenius activation energy of 34.5 kJ·mol⁻¹ is in good agreement with the activation energies measured for the crystallization of other MOFs using mild temperature conditions.^{45, 46}

As low temperature synthesis can be effective to produce nano-MOFs,^{47, 48} our protocol with low activation energy is appealing to prepare size-controlled MOF-808 nanocrystals. Indeed, homogeneously tuning the crystal size while maintaining high product yield and quality is an interesting but still challenging topic in MOF chemistry. For example, one very recent effort with MOF-808 produced nanoparticles with low BET surface area (622 m²/g) and poor crystallinity.⁴⁹ In our case, we control the crystal size via a concentration-induced method (illustrated in **Figure 2a**), where the volume of water and formic acid are kept constant and the concentration of Zr₆ oxoclusters and BTC linker simultaneously tuned. The MOF-808-RT synthesized with BTC concentration of 18.75 mg/ mL exhibits a uniform crystal size of 850 (±10) nm (labeled “850 nm MOF-808” in the following) according to scanning electron microscopy (SEM) images (Figure 2b). By increasing the concentration of reactants by 6, the nucleation-growth kinetics of MOF-808 was significantly accelerated, which shortened the synthesis time to 5 h in comparison to the 14 h for 850 nm MOF-808. As shown in Figure S10, PXRD patterns evidenced the high crystallinity and purity of all the materials with a progressive peak broadening (FWHM from 0.126° to 0.26° in 2θ) that is consistent with the formation of nanocrystals. SEM and High-resolution transmission electron microscope (HRTEM) measurements in **Figure 2b-e** further

revealed that this concentration-induced method produced well-defined cubic crystals from sub-micron (850 nm) to nanoscale (35 nm). These samples display very narrow size distribution with a low coefficient of variation (CV) (Figure S13). The statistical analysis also confirms the strong dependence of crystal sizes on the concentration of precursor (**Figure 2f**). The monodispersity of the synthesized MOF-808 could be tentatively attributed to the use of Zr_6 oxoclusters as the metal source that bypasses the formation of Zr_6 from Zr(IV) salts, therefore facilitating the MOFs nucleation. In addition, the syntheses were performed under stirring, which allows the reactants to be mixed in a homogeneous way. Indeed, the absence of stirring in the synthesis of 850 nm MOF-808 resulted not only in prolonged synthesis duration (> 5 days) but also in a more polydispersed sample (see Figure S14).

Additional N_2 porosimetry experiments were carried out to evaluate the impact of the particle size on the surface areas and pore sizes of MOF-808. All values were consistent with the literature (**Figure 2g**), with only slight differences in surface areas following the relationship: 850 nm MOF-808 ($2090 \pm 17 \text{ m}^2/\text{g}$) > 210 nm MOF-808 ($2010 \pm 20 \text{ m}^2/\text{g}$) > 60 nm MOF-808 ($1920 \pm 20 \text{ m}^2/\text{g}$) \approx 35 nm MOF-808 ($1920 \pm 20 \text{ m}^2/\text{g}$). The pore size distributions followed the same trend (Figure S15), 850 nm MOF-808 (18.6 Å) > 210 nm MOF-808 (17.4 Å) > 60 nm MOF-808 (16.0 Å) \approx 35 nm MOF-808 (16.2 Å), which may be explained by small variation of the defect content in the structure according to the TGA (Figure S16 and Table 1). Indeed, the drop in size is accompanied by a decrease of the modulator (formic acid) to substrate ratio, which can lead to a lower number of defects, as commonly observed in Zr(IV)-based MOF chemistry. Owing to the very low amount of solvents used in the synthesis of 35 nm MOF-808 (1.5g MOF precursors vs 2.67 mL solvent) and the high reaction yield (98 %), the compound showed a gel-like feature (Figure S17) and was obtained with a high STY at $2516 \text{ kg/ m}^3/\text{ day}$,

suggesting possible applications in aerogels^{50,51} and possible up-scaling (Figure S18, synthesis of 10g of 35nm MOF-808 particles) of the green synthesis. To the best of our knowledge, this value is comparable to a few commercialized MOF examples, which are commonly synthesized through conventional solvothermal approaches. It should be noted that the reaction cannot be performed when the concentration of the BTC is above 112.5 mg/ mL due to the very limited solvent volume that does not allow reactants be fully dispersed.

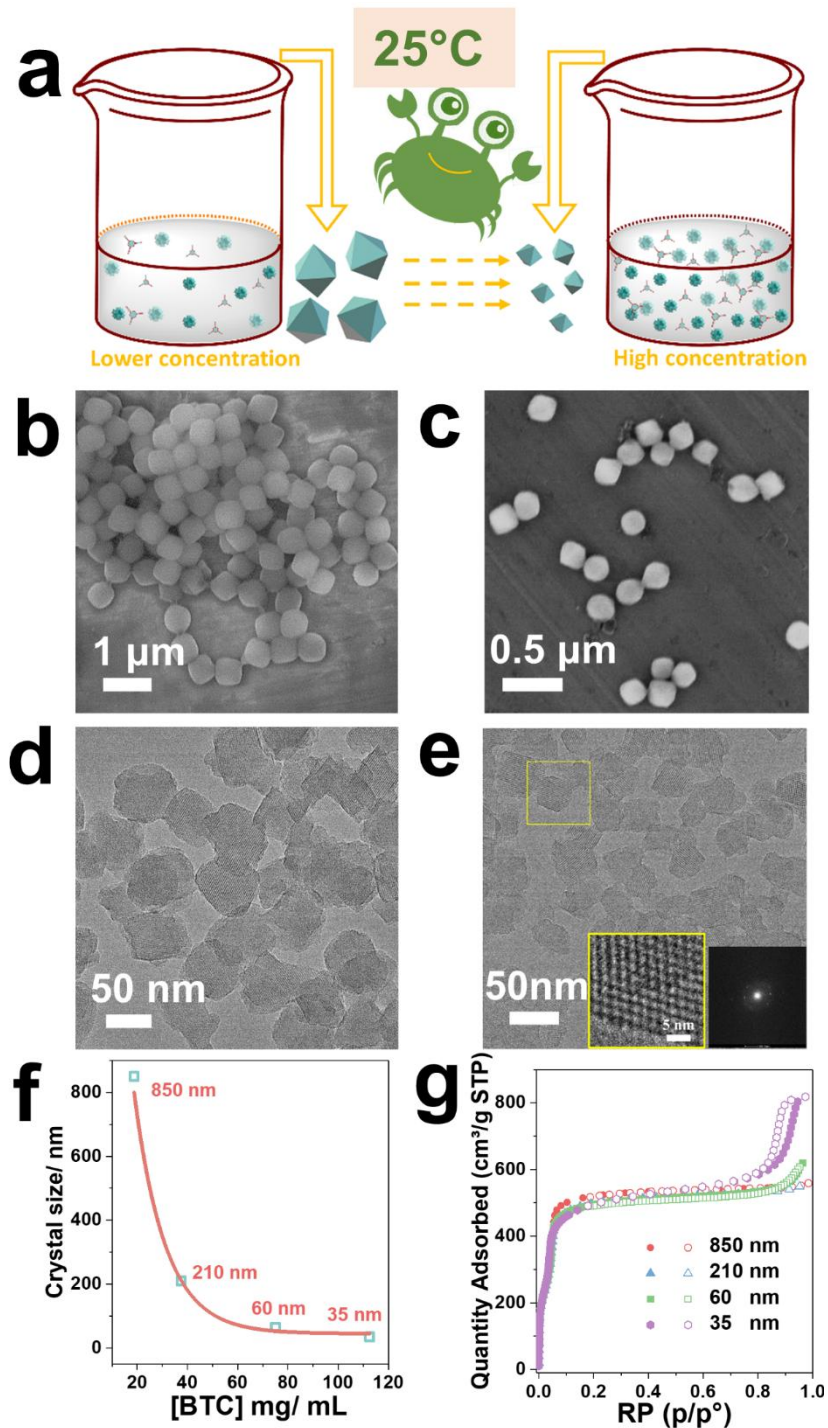


Figure 2. (a) Schematic representation of the concentration-induced method to tune the MOF-808 size; SEM images of (b) 850 nm MOF-808 and (c) 210 nm MOF-808; HRTEM image of (d) 60 nm MOF-808, (e) 35 nm MOF-808, Insets: Fast Fourier transform (FFT), cropped at the predominant lattice fringes and enlarged image inside the yellow square; (f) Statistical mean size of the synthesized MOF-808 in different reactants concentrations; (g) 77 K N_2 adsorption-desorption of 850 nm MOF-808, 210 nm MOF-808, 60 nm MOF-808, and 35 nm MOF-808, RP= Relative pressure.

The size reduction might be explained by the faster reaction kinetics associated with the lower formic acid to MOF precursor ratio. It is common in MOF carboxylate chemistry to use monocarboxylic acids -called modulators- to competitively coordinate with metal sites. In such cases, the more modulator used in the synthesis, the slower the synthesis is, which usually results in larger crystals. In order to confirm this hypothesis, additional experiments were carried out. As the increase in the concentration of reactants leads to a decrease of the modulator/ reactant ratio, we changed the relative ratio between the volume of formic acid and the mass of Zr_6 oxoclusters while keeping constant the concentration of reactants as in the 60 nm MOF-808 synthesis. For example, when this volume to mass ratio (VTMR) was 1.93 (1.93 mL formic acid per gram of Zr_6 clusters) instead of the value 1.25 for 60 nm MOF-808, we observed a clear deceleration of the kinetics, confirming that the amount of modulator influences the reaction kinetics. The obtained well-crystalline product was collected and analyzed with SEM (Figure S19), evidencing the presence of 170 nm MOF-808 crystals. This experiment shows that increasing the amount of modulator leads to an increase in particle size and that therefore, the concentration-induced size tuning effect is related to modulator-induced methods. As further proof, we used the parameters of the 850 nm MOF-808 synthesis but decreased the VTMR to 1.93. The SEM image in Figure S20 indicated that the obtained MOF-808 exhibited a smaller mean size of 175 nm, very close to the MOF-808 particle size synthesized under the same VTMR but with much higher reactant concentrations. However, when the VTMR was further decreased in order to produce nano-MOF-808, a significant drop in reaction yield (12% after 5 days with VTMR= 1.1) and quality (amorphous gel) was observed. We hypothesized this might be due to solubility issues because a lower formic acid content decreases the quantity of BTC dissolved in solution as BTC has a very low solubility in pure H_2O at room-temperature,

therefore requiring a much longer synthesis duration associated with a dramatically lower yield. Notably, this can be circumvented by a slight heating (to 60 °C, see Figure S21) or addition of another organic solvent (e.g., Methanol, see Figure S22) that enhance the solubility of BTC to form smaller crystals with reasonable product yield (92%).

Table 1. Main properties of the MOF-808 nanocrystals.

MOF-808 particle size [nm]	BET Surface area [m ² /g]	External Surface area [m ² /g] ^a	Total Pore volume [cm ³ /g]	Pore size [Å]	Connectivity ^b	Space-time yield [kg/m ³ /day]
850	2090	47	0.85	18.6	1.82	102
210	2008	51	0.84	17.4	1.92	420
60	1920	452	0.83	16.0	1.98	1491
35	1920	775	0.83	16.2	1.98	2518

^{a)} External surface area was determined through t-plot method; ^{b)} Connectivity refers to the bridging ligands per Zr₆ oxoclusters.

NanoMOFs are suitable candidates for heterogeneous catalysis, sensing or bio-applications,⁵²⁻⁵⁴ mainly due to their easy processability and fast diffusion kinetics. However, when using heterogeneous catalysts in liquid phase, a high colloidal stability is required. Hence, Dynamic Light Scattering (DLS) was performed on MOF-808 in deionized water. As shown in **Figure 3a** (distribution in number) and Figure S23 (distribution in intensity), 35 nm MOF-808 displayed a constant hydrodynamic size at around 53 (±13) nm and 91 (±27) nm, respectively, in a wide range of particle concentrations (from 0.17 mg/ mL to 7 mg/ mL, pH from 3.63-3.82), which provided a flexible set of conditions to investigate their catalytic activity. Subsequently, time-dependent DLS measurements were carried out (**Figure 3b**) and demonstrated the excellent dispersion stability of this nano-MOF during 24h with an extremely low polydispersity index (0.05-0.1), indicating that no additive or treatment is needed to stabilize the dispersion. The

satisfying colloidal stability and dispersion can be attributed to 1) the highly positive surface charge of the MOF-808 nanoparticles (+32 mV, Figure S24), and 2) the well-defined monodispersed nanoparticles.

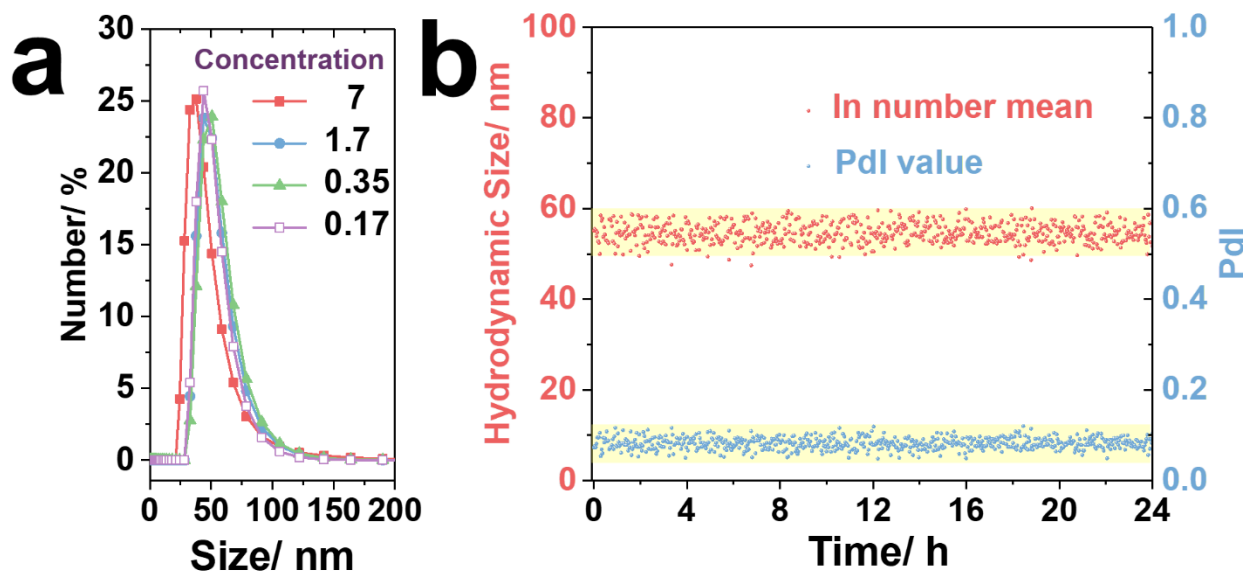


Figure 3. (a) Hydrodynamic sizes of the 35 nm large MOF-808 nanocrystals in aqueous solution with varying concentrations of 7, 1.7, 0.35, 0.17 mg/ mL. (b) Time-resolved hydrodynamic size and polydispersity index (PDI) of the 35 nm MOF-808 in aqueous solution.

In order to investigate the effect of particle size on the catalytic activity of MOF-808, the rate of peptide bond hydrolysis at physiological pH was followed with both the model dipeptide glycylglycine (GG) and with the protein Hen Egg White Lysozyme (HEWL) which consists of 129 amino acids. Previous studies have shown that several Zr_6O_8 -based MOFs, including MOF-808, were highly efficient catalysts for peptide bond hydrolysis.²³ Conversion of GG to glycine (G) was followed by quantitative 1H -NMR spectroscopy over 20 h (Figure S25). Noteworthy, after 20 h hydrolysis the MOFs did not suffer from any significant loss of crystallinity under these reaction conditions (Figure S28). **Figure 4a** shows the conversion of GG with time, in

addition to the formation of cyclic glycyglycine (cGG), consistent with previous reports involving GG and Zr_6O_8 -carboxylate based MOFs.^{24, 25} Hydrolysis followed first order rate kinetics (Figure S26), from which, k_{obs} was calculated for each MOF used in the study (Figure 4b). After 20 h, 35 nm MOF-808 hydrolyzed 94.0% of the GG substrate, compared with 91.3%, 80.0% and 70.6% respectively for the 60, 210 and 850 nm MOF-808 crystals. This resulted in k_{obs} that ranged from $3.87 \times 10^{-5} \text{ s}^{-1}$ to $1.73 \times 10^{-5} \text{ s}^{-1}$ depending on the particle size (**Figure 4b**), showing that smaller particles are the most efficient. Such differences might be due to the external surface area of the MOF. 850 nm MOF-808, with external surface area of $47 \text{ m}^2/\text{g}$ is significantly less efficient at peptide bond hydrolysis than 35 nm MOF-808, which has an external surface area of $775 \text{ m}^2/\text{g}$, while both compounds present relatively similar BET surface areas (Figure S27). This supports the notion that the hydrolysis is likely to occur on the external surface of the MOFs, as external surface area has the most significant influence on the efficiency of peptide bond hydrolysis.

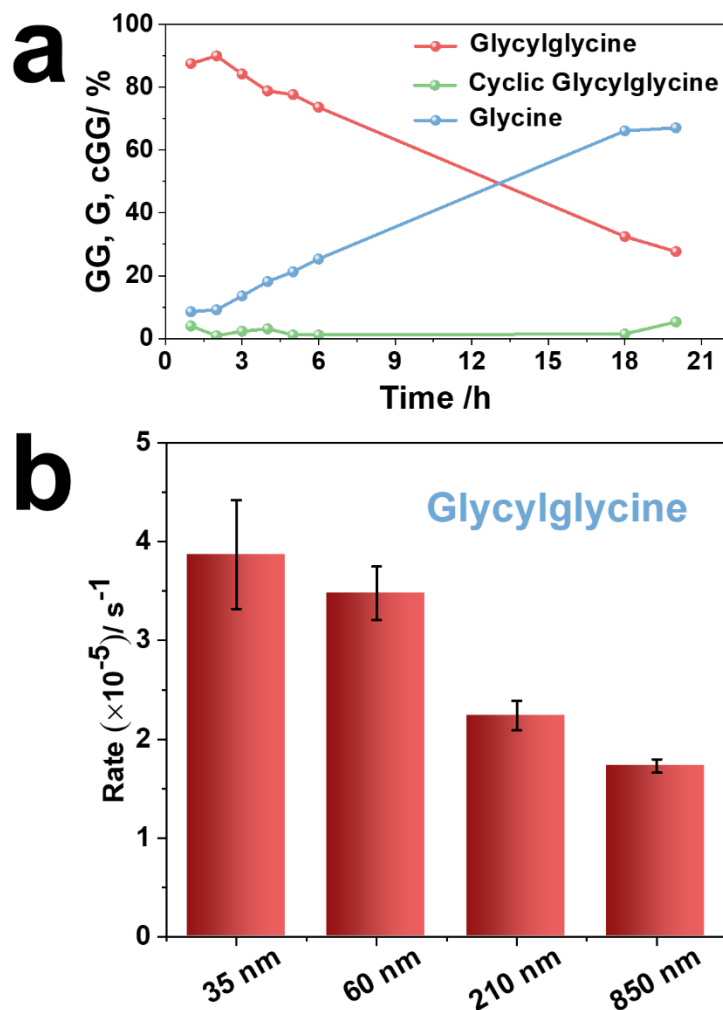


Figure 4. (a) Conversion of glycylglycine (GG) to glycine (G) and the side product cGG with time in the presence of 850 nm MOF-808. (b) k_{obs} of GG hydrolysis with different MOF-808 particle sizes.

Hydrolysis of HEWL was analyzed with Coomassie stained 18 % SDS-PAGE gels (**Figure 5a** and S29). After 72 h of HEWL incubation with 850 nm MOF-808, new fragments appear at 12.7, 11.3, 8.3, 5.8 and 4.7 kDa in addition to the intact protein at 14.3 kDa. The four fragments with the largest molecular weight (MW) were previously observed in HEWL hydrolysis with MOF-80823 and NU-1000,²⁵ suggesting that the new MOF synthesis protocol described here does not affect the selectivity of the MOF catalysts. The MW of the fragments observed in SDS PAGE indicates that they are most likely generated through hydrolysis of the peptide bonds at Asp18

(12.7 kDa), Asp52 (8.3 kDa), and Asp66 (5.8 kDa), as well as sequential hydrolysis of the 12.7 kDa fragment at Asp 119, to give the 11.3 kDa fragment, in analogy with the previously reported HEWL hydrolysis by metal complexes.⁵¹ The lightest fragment (4.7 kDa) was not observed before in MOF catalyzed hydrolysis of HEWL but it corresponds well to the MW of a fragment previously observed in HEWL hydrolysis by an Hf(IV)-Polyoxometalate,⁵⁵ resulting from hydrolysis at the Asp101 site. In addition to these 6 Asp residues, there are two additional Asp residues in HEWL (Asp48 and Asp87) that are located on short stretches of random coils in between larger regions of secondary structure (Figure S30). They most likely remain buried in the tertiary structure of the protein and as such are inaccessible to the active metal sites of the MOF even after partial hydrolysis of the intact protein. The rate of peptide bond hydrolysis in HEWL showed the same trend as with GG hydrolysis, with an increase in the rate observed as particle size decreases (**Figure 5b**).

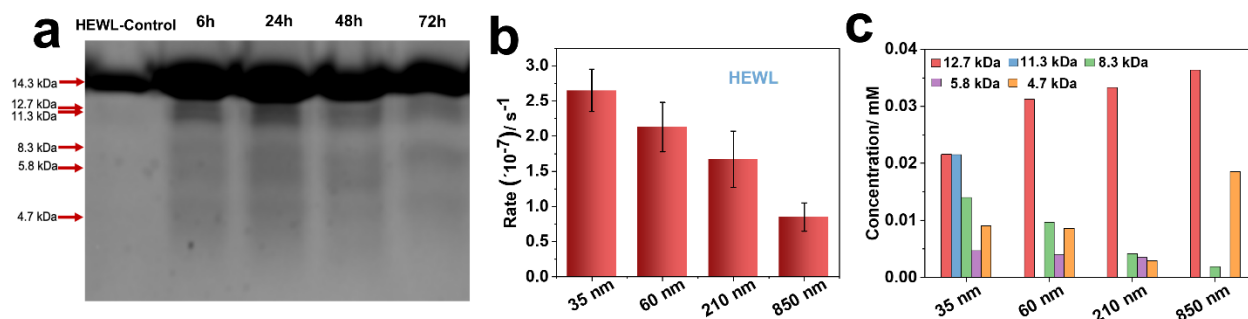


Figure 5. (a) Hydrolysis of HEWL by 850 nm MOF-808 followed by Coomassie blue stained SDS-PAGE. Native weight of protein – 14.3 kDa (b) k_{obs} of HEWL hydrolysis with all MOFs. (c) Concentration of HEWL hydrolysis fragments after 72 h incubation with all MOFs, from SDS-PAGE band analysis.

In addition, as demonstrated in **Figure 5c**, the number of resulting polypeptide fragments and their concentration can be tuned by changing the MOF particle size. While MOFs with smaller particle size produce more fragments within comparable range of concentrations, the larger particle sizes appear to be more efficient at hydrolyzing the Asp18 site, and show limited

hydrolysis at Asp52 and Asp66 residues. Furthermore, these MOFs exhibited higher activity towards the secondary hydrolysis at the Asp119 site, which resulted in higher concentration of the lightest fragment at 4.7 kDa. With 35 nm MOF-808, the fragment coming from the hydrolysis at Asp18 bond is likely further hydrolyzed at Asp119 to give the 11.3 kDa fragment, which has not been detected for other MOFs. Taking into account the dimensions of HEWL (35 x 25 x 45 Å), it is unlikely that the protein is able to diffuse into the MOFs pores without some degree of unfolding as the maximum pore size is 18.6 Å. Such unfolding has been reported previously for the incorporation of the smaller cytochrome c protein into NU-1000 and Tb-meso MOF.^{56, 57} Therefore, we propose that the initial hydrolysis occurs on the surface exposed Zr-nodes present within the half-spherical pockets on the surface of particles that are of similar size to HEWL, after which, subsequent hydrolysis of smaller fragments may also occur to a limited extent, within the pores, as the parent fragments are small enough to diffuse through the pore windows.²³ As was proposed for GG hydrolysis, the external surface area of the MOFs contributes significantly to the observed hydrolysis patterns. For 210 nm MOF-808 and 850 nm MOF-808, which have similar external surface areas (51 and 47 m²/g respectively) the hydrolysis patterns are similar, with the major HEWL fragment being 12.7 kDa. However, as the external surface area increases to 775 m²/g for 35 nm MOF-808, the efficiency of HEWL hydrolysis and number of resulting fragments increase in line with the increase in number of active metal sites present on the external surface of the MOF. The more efficient protein hydrolysis could also be due to the more effective MOF-protein interaction on the structure of the protein as smaller size nanoMOFs with a significantly larger external surface area may induce a substantial structural change in HEWL, causing a greater degree of protein unfolding over the surface of the

nanoMOF. This in turn may expose more Asp residues to come into contact with the active metal sites, promoting hydrolysis at more sites on the protein surface.

4. Conclusion

This article reports for the first time an efficient room-temperature synthesis of the benchmark 6-connected MOF-808 under environmentally-friendly conditions. The novel synthesis conditions allowed us to achieve a size tuning from sub-micron range (850 nm) to the nanoscale (35 nm) while maintaining high crystallinity, porosity, homogeneity and a high space-time yield. In addition, we showed the high quality of the colloidal dispersion of the nano-MOF-808, reaching a nearly monodispersed suspension both at various concentrations (from 7 mg/ mL to 0.17 mg/ mL) and long-time-scale (24h). Such ambient conditions are particularly appealing for the incorporation of temperature sensitive species (e.g. small nanoparticles, enzymes, etc.) inside MOFs. Subsequently, the nano-MOF-808 exhibited a remarkable size-dependent catalytic efficiency towards a challenging peptide bond hydrolysis, with the smallest nanoMOF showing the highest catalytic potential. The trend is directly proportional to the external surface area of the MOF particles, thus suggesting that the catalytic reaction is likely to occur on the MOF external surface. The nanoMOF-808 was also able to selectively hydrolyze HEWL protein, efficiently producing fragments with MW that are suitable for the analysis in the emerging middle-down proteomics applications. Additionally, varying the particle size gives control over the specific protein fragments produced. This shows that MOFs can be synthesized with a specific particle size to produce desired fragments, proving these MOFs can act as superior artificial protease nanozymes for use in the field of proteomics. Finally, we hope this work

can pave the way for efficient heterogeneous catalysis applications but also for applications that require colloidal-level nano-MOF-808.

Supporting Information

Additional synthetic details, PXRD, MS, EDX, TGA, *In-situ* PXRD, and SEM data (PDF)

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