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DESIGN OF GPR54 (KISS1R) CROSSLINKER PEPTIDES: NEW TOOLS TO STUDY G PROTEIN-COUPLED RECEPTORS (GPCR) LOCALIZATION AND FUNCTION





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**Belgium Introduction**. The neuropeptide kisspeptin and its receptor, GPR54 (KISS1R) play a central stimulatory role in mammalian reproduction. However, GPR54 localization is still incompletely mapped and its pharmacology only partially known. Receptor localization and receptor deorphanization remain important issues in drug discovery. To approach these problems, we designed new tools and used kisspeptin and its receptor to begin testing them.

**Materials and Methods**. Four molecules (TMV-33-2-A, TMV-34, TMV-160A and TMV160B) were created based on the scaffold of the kisspeptin analog C6, that shows robust *in vivo* activity. These molecules bear modifications to improve the pharmacodynamics, to have tag (biotin insertion) and to bind covalently the receptor (furan insertion). *In vitro* activity of the new molecules was evaluated by a calcium mobilization assay in HEK-293 cells transfected with hGPR54. *In vivo* activity was assessed in adult C57BL6 male mice and in ovariectomized and estrogen replaced (implant) female mice. Molecules were injected ip with of test compounds (0,4 nmol/mouse) and

Sequence of human kisspeptin 10 and of modified ssay. H-Tyr-Asn- Trp -Asn-Ser-Phe- Gly -Leu-Arg-Kp10 Phe-NH<sub>2</sub> peptides. Palm- $\gamma$ -Glu-Tyr-Asn- Trp -Asn-Ser-Phe- Gly $\psi$ [Tz]-Leu-Arg(Me)-Tyr-NH<sub>2</sub> C6 Palm-γ-Glu-Tyr-Asn-FurAla-Asn-Ser-Phe- Glyψ[Tz]-Leu-Arg(Me)-Tyr-NH<sub>2</sub> TMV-33-2A **Biotin-linker-Tyr-Asn-FurAla-Asn-Ser-Phe- Glyψ[Tz]-Leu-Arg(Me)-Tyr-NH**<sub>2</sub> TMV-34 Biotin-linker Palm-y-Glu Lys-Tyr-Asn- Trp -Asn-Ser-Phe- Glyψ[Tz]-Leu-Arg(Me)-Tyr-NH<sub>2</sub> TMV-160A Biotin-linker Palm-γ-Glu Lys-Tyr-Asn-FurAla-Asn-Ser-Phe- Glyψ[Tz]-Leu-Arg(Me)-Tyr-NH<sub>2</sub> TMV-160B The modifications introduced into the human kp10 are indicated in color in the amino acid sequence. These modifications allow: i) protection against enzymatic degradation, ii), reduced renal excretion, iii) to tag the molecule and iv) to bind covalently the

receptor. Lysine is present as a chemical moiety to anchor both biotin and palmitoyl.



EC 50

Scheme showing the structure of FurAla and the general principle of furan cross



Kinetic of LH concentration for differents compound in females (A, D); males (B, E). Area under induceds bylc@latedHowteveren theirhoAksCorisfe

**Conclusions.** These preliminary results suggest that it is possible to develop highly potent and selective crosslinker molecules as new tools to study the kisspeptin system. More importantly, this strategy holds the promise to be applicable to study other targets. Structural optimization is in progress to evaluate various combinations of chemical modifications and different tags to allow *ex vivo* receptor localization by crosslinking the receptor with

## the tagged analog.



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