



HAL
open science

Incorporating clonality into the plant ecology research agenda

Jitka Klimešová, Gianluigi Ottaviani, Tristan Charles-Dominique, Giandiego Campetella, Roberto Canullo, Stefano Chelli, Zdeněk Janovský, F. Curtis Lubbe, Jana Martínková, Tomáš Herben

► **To cite this version:**

Jitka Klimešová, Gianluigi Ottaviani, Tristan Charles-Dominique, Giandiego Campetella, Roberto Canullo, et al.. Incorporating clonality into the plant ecology research agenda. Trends in Plant Science, 2021, 26 (12), pp.1236-1247. 10.1016/j.tplants.2021.07.019 . hal-03372270

HAL Id: hal-03372270

<https://hal.science/hal-03372270v1>

Submitted on 4 Nov 2022

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Trends in Plant Science

Incorporating clonality into the plant ecology research agenda

--Manuscript Draft--

Manuscript Number:	
Article Type:	Opinion
Keywords:	belowground organs; clonality, plant functions
Corresponding Author:	Jitka Klimesova Institute of Botany, Czech Academy of Sciences Trebno, CZECH REPUBLIC
First Author:	Jitka Klimesova
Order of Authors:	Jitka Klimesova Jitka Klimešová Gianluigi Ottaviani Tristan Charles-Dominique Giandiego Campetella Roberto Canullo Stefano Chelli Zdeněk Janovský Frederic Curtis Lubbe Jana Martínková Tomáš Herben
Abstract:	A longstanding research divide exists in plant ecology: studies either examine specifically plant clonality or are focused on all plants generally but considering them being nonclonal. This gap cascades into a lack of knowledge about the similarities and differences between clonal and nonclonal plants. We aim to bridge this gap by identifying areas that would benefit from incorporating clonal strategies and dynamics into one integrated research platform. These fields are: i) response to habitat productivity, ii) interactions among neighbours, iii) response to disturbance, and iv) population structure and evolution. We are convinced that this would provide valuable insights into the eco-evolutionary dynamics of all plants.
Suggested Reviewers:	Daniel Laughlin daniel.laughlin@uwyo.edu John Morgan J.Morgan@latrobe.edu.au Peter Vesk pvesk@unimelb.edu.au Helge Bruelheide helge.bruehlheide@botanik.uni-halle.de Mark van Kluenen mark.vankleunen@uni-konstanz.de Alessandra Fidelis alessandra.fidelis@unesp.br Markus Fischer Markus.Fischer@ips.unibe.ch Katja Tielborger katja.tielboerger@uni-tuebingen.de

Opposed Reviewers:

Dear Dr. Susanne Brink, Editor of *Trends in Plant Science*,

On behalf of all co-authors, I would like to submit our manuscript of an Opinion piece entitled "*Incorporating clonality into the plant ecology research agenda*" to *Trends in Plant Science*.

Please, accept my apology for delay with submission.

With best regards

Jitka Klimesova

1 **Highlights:**

2 Clonal plants can represent a substantial proportion of species in floras and plant
3 communities. Because they share several functions that are not present in nonclonal plants,
4 the differences in their proportions are likely to scale up as community and ecosystem
5 differences.

6 Clonal reproduction is provided by specialised organs which directly or indirectly affect other
7 plant traits. Clonal growth organs usually serve for the storage of carbohydrates and buildup
8 of the bud bank, both necessary for resprouting in recurrently disturbed habitats. These
9 storage organs (to an unknown but potentially important degree) affect global carbon cycling.

10 Clonal growth leads to increased plant size in the horizontal dimension and to a different
11 degree of ramet aggregation that influences exploration of soil resources, pollination, and
12 biotic interactions.

13 Clonal growth serves as a reproductive insurance mechanism that further affects plant
14 demography and possible evolutionary rates.

15

[Click here to view linked References](#)

1 **Incorporating clonality into the plant ecology research agenda**

2 Authors

3 Jitka Klimesová^{1,2*}, Gianluigi Ottaviani¹, Tristan Charles-Dominique³, Giandiego Campetella⁴,
4 Roberto Canullo⁴, Stefano Chelli⁴, Zdeněk Janovský^{1,2}, F. Curtis Lubbe¹, Jana Martínková¹,
5 Tomáš Herben^{2,5}

6 1 - Institute of Botany, The Czech Academy of Sciences, Dukelská 135, 37901 Třeboň, Czech
7 Republic,

8 2 - Department of Botany, Faculty of Sciences, Charles University, Benátská 2, 12800 Praha,
9 Czech Republic

10 3 - CNRS UMR7618; Sorbonne University; Institute of Ecology and Environmental Sciences
11 Paris; 4, place Jussieu 75005 Paris, France

12 4 - School of Biosciences and Veterinary Medicine, Plant Diversity and Ecosystems Management
13 unit, Camerino University, 62032 Camerino, Italy

14 5 - Institute of Botany, The Czech Academy of Sciences, Zámek 1, 25243 Průhonice, Czech
15 Republic

16 *correspondence: jitka.klimesova@ibot.cas.cz

17 **Keywords:** belowground organs; clonality, plant functions

18

19

20 **Abstract**

21 A longstanding research divide exists in plant ecology: studies either examine specifically plant
22 clonality or are focused on all plants generally but considering them being nonclonal. This gap
23 cascades into a lack of knowledge about the similarities and differences between clonal and
24 nonclonal plants. We aim to bridge this gap by identifying areas that would benefit from
25 incorporating clonal strategies and dynamics into one integrated research platform. These fields
26 are: i) response to habitat productivity, ii) interactions among neighbours, iii) response to
27 disturbance, and iv) population structure and evolution. We are convinced that this would provide
28 valuable insights into the eco-evolutionary dynamics of all plants.

29

30

31 **Introduction**

32

33 An overwhelming portion of plant ecological research focuses solely on aboveground parts.
34 As a result, our understanding of belowground plant functions is lagging behind the aboveground
35 compartment [1, 2]. While the study of root acquisitive functions (and fine roots as **acquisitive**
36 **organs**; see Glossary) has made recent large advances [3], **nonacquisitive organs** such as
37 storage roots, rhizomes, and bulbs, have still been given little attention, despite their almost
38 ubiquitous presence [4]. These organs provide functions that are common to all plants, such as
39 the storage of carbohydrates and buds, but also enable functions limited to only some plants,
40 namely clonal growth. The ability to multiply clonally separates plants into two groups:
41 nonclonal plants that multiply only by seed, and clonal plants that can also multiply by vegetative
42 growth. However, **clonality** does not only affect reproductive mode, it is also involved directly or
43 indirectly in many aspects of plant life and function.

44 There has been a long tradition of research on plant clonality (see [5] for a recent
45 overview), but it has largely remained restricted to a small group of researchers that are focused
46 exclusively on clonals and only use a few model species, otherwise ignoring nonclonal plants.
47 Consequently, we do not have enough comparative information about key differences and
48 similarities between clonal and nonclonal species. Therefore, it is no surprise that functional
49 syntheses incorporating all plants have largely ignored the clonal/nonclonal dichotomy or have
50 pigeonholed clonality as a special, if not exotic feature, important only for a narrow range of
51 ecological functions (but see [6]).

52 Here, we draw a roadmap aimed at bridging this research divide (i.e. clonal-and nonclonal-
53 oriented), by showing that although it is not universal, clonality is a widespread feature of plants
54 (Box 1, 2) and that modifies or determines many essential plant functions and processes, such as
55 i) response to productivity, ii) interactions among neighbours, iii) response to disturbance, and iv)
56 population structure and evolution (Table 1 – Key Table). We provide examples where taking
57 clonality and its absence into account may substantially affect the way we look at plants, making
58 our understanding of plant functional differentiation more complete and finally allowing deeper
59 insights into plant eco-evolutionary dynamics.

60

61 **Clonality along the productivity gradients**

62

63 One key gradient that shapes plant form and function is associated with habitat productivity
64 and the related plant economics spectrum [7, 8]. It was primarily defined for leaves, wherein thin,
65 short-lived, acquisitive leaves with low protection against herbivory and high photosynthetic
66 rates are found at the fast end of the spectrum, whereas leaves with contrasting features are

67 positioned at the slow end [9]. In addition to the leaf economics spectrum, this productivity
68 gradient determines biomass allocation strategies (i.e. biomass investment to organs that acquire
69 different resources), this is typically described as the root/shoot ratio [10, 11]. Plants in highly
70 productive environments (i.e. those characterised by high competition for light) tend to invest
71 preferentially into stems and leaves, while plants growing in nutrient-limited environments
72 primarily increase allocation to fine roots that promote belowground resource acquisition [12].

73 These textbook relationships generally consider all belowground organs to be responsible
74 for resource acquisition. However, the specialised belowground **clonal growth organs** of clonal
75 plants add further functional dimensions. They do not participate in resource acquisition, but in
76 addition to clonal reproduction, these organs store and share resources among the rooting units
77 that they connect [2]. This leads to three major consequences for economics and allocation
78 relationships.

79 First, **storage organs** divert a significant amount of resources which might otherwise be
80 used for growth and competition for light [13]. Although the leaf economics spectrum has been
81 expanded by incorporating other organs (e.g. roots; [7]) and functions into the plant economics
82 spectrum, belowground storage organs have remained sidelined and their role in resource
83 economy along the productivity gradient is essentially unknown [14]. Second, the occurrence of
84 belowground storage organs confounds the use of the simple root/shoot ratio as a proxy of
85 relative investment into nutrient- vs light-acquiring organs [10, 15]. Therefore, the available
86 comparative data [16] cannot be interpreted as evidence for differential roles between light- and
87 nutrient-acquiring investment. In particular, the lifespan of belowground storage organs and its
88 link to the leaf economics spectrum is of key importance and may affect carbon sequestration and
89 cycling globally [17, 18]. Third, in addition to resource storage (also present in many nonclonal

90 perennials), clonal organs are often involved in the horizontal transport of resources, including
91 carbon and soil-borne nutrients [19, 20]. The extent of resource sharing in clonal plants depends
92 on the lifespan of their clonal connections, which has been described in terms of the integrator
93 and splitter strategies. Integrators (i.e. species with clonal connections maintained long after the
94 new **ramet** has become established) are favoured in harsh and low-productivity environments
95 whereas splitters (with short lifespan of connection) in benign conditions [20, 21, 22]. As plants
96 transport limited resources (water, nutrients) or ameliorate stress (e.g. aeration of organs in
97 waterlogged soil), such connections between different parts of a clone may equalise conditions
98 over heterogeneous environments with potentially ecosystem-wide effects [17].

99 Currently, we know very little about the role of storage and clonal growth organs in the
100 plant economics spectrum as well as how traits of these organs vary along productivity gradients.
101 While theoretical models have shown how resource storage may (or may not) contribute to
102 fitness [23], there are almost no comparative studies showing differences in relative size or
103 lifespan of storage organs across a number of species along productivity gradients [24, 25]. While
104 the **splitter-integrator continuum** has been extensively studied, such studies seldom incorporate
105 relevant trends and traits for nonclonal plants [22]. Therefore, we cannot identify general rules
106 about the contribution of clonality (and its types) to fitness via translocation across different
107 micro-environmental patches. Including clonal growth organs, their resource-sharing, and their
108 storage capacities into a broader research on economics spectrum will fill gaps in understanding
109 plant strategies in response to habitat productivity.

110

111 **Interactions among neighbours**

112

113 Plants compete for many resources, especially light, water, and nutrients. Under productive
114 and undisturbed conditions, competition for light along the vertical dimension takes precedence
115 [26]. It is inevitably asymmetric and often leads to competitive exclusion. However, competitive
116 exclusion by overtopping may be delayed or blocked by reducing vertical size differences [27].
117 After disturbance or in harsh environments, the vertical dimension may lose its importance and
118 the success of individual species is then determined by the amount of space they can occupy in
119 the horizontal dimension [28].

120 This space occupation can be attained by generative reproduction or by clonal growth. Both
121 these processes determine aggregation and mobility patterns in communities, but strongly differ
122 in their outcomes. While seed dispersal is not directly controlled by the mother plant and
123 typically follows an exponential or hyperbolic decay curve [29], clonal reproduction is more
124 active and associated with species-specific fixed distances [30]. Distinct morphologies of
125 individual species then determine their aggregation patterns, ranging from loose patches to
126 densely packed aggregations of individuals (often termed the **guerrilla-phalanx growth forms**,
127 another strategy scheme not suited to encompass nonclonal plants; [31]). Aggregation can be a
128 way for plants to respond to environmental harshness ranging from intraspecific (namely clonal)
129 to interspecific (facilitative) aggregations [32, 33].

130 Another important attribute of clonal reproduction can be parental support of offspring,
131 which may last long into their maturity, and can contribute far greater access to resources than
132 afforded by an increase in seed size by nonclonal plants. Resources stored in the seed are
133 necessarily fixed and also lack the feedback mechanism, and thus versatility, present in clonal
134 connections. Clonal growth is also often associated with density regulation of neighbouring

135 ramets from the same genetic individual (i.e. **genet**) [34] which reduces competition among them
136 (Box 3).

137 Finally, an essentially technical yet key detail in plant competition concerns dominance.
138 Dominant species (i.e. those accounting for the highest number of individuals or biomass in a
139 community) are assumed to affect community-level processes more than subordinate species
140 [35]. This assumption is used to scale up plant traits, typically collected at the individual plant
141 level, to ecosystem functions, which is typically based on the proportions of individual species
142 aboveground [36]. However, plant species strongly vary in their investment of biomass into
143 belowground organs, and the clonal-nonclonal divide plays an important role. This implies that
144 scaling up plant traits by aboveground plant dominance is improper, especially when plant
145 belowground functions are the focus of the study and in open, seasonal, or disturbance-prone
146 ecosystems that are characterised by species with large allocation into belowground
147 nonacquisitive organs [4, 37]. This discrepancy may be ameliorated by investing larger effort to
148 collecting information on belowground organs and traits.

149

150 **Clonality and disturbance**

151

152 Disturbance is another essential driver of plant strategies worldwide [38, 39]. It has been
153 hypothesized that clonal plants are particularly common at intermediate disturbance levels [38],
154 but this strongly depends on the disturbance regime in terms of both frequency and severity. The
155 existing literature often distinguishes between **seeders and resprouters**: seeders (regenerating
156 from seeds), and resprouters (i.e. plants that resprout after disturbance from specialised bud-
157 bearing organs) [38]. Clonal plants are typically resprouters, but currently available data do not

158 establish a clear relationship between resprouting and the degree of clonality a plant possesses,
159 hence limiting possible generalisation. While experiments with temperate species have shown
160 that clonal plants accumulate higher belowground biomass (in storage and/or bud-bearing organs)
161 than nonclonals [40, 41], there are no comparative data on storage and **bud banks** in clonal and
162 nonclonal plants available for the majority of disturbance-prone ecosystems.

163 Further, simple quantification of plant investment into bud-bearing organs is not sufficient
164 for assessing the cost of their formation. Loss of a bud is a loss of the outcome of a complex
165 morphogenetic event and cannot be immediately replaced even if the necessary resources are still
166 available. Consequently, such loss must be assessed with respect to the architectural constraints
167 of a given plant. For example, stem-derived clonality provides the plant with a limited number of
168 axillary buds and thus confers only a limited ability to survive disturbance, and therefore it is
169 found mainly in grazed or mown systems. In contrast, root-derived clonality, with a potentially
170 unlimited number of adventive buds, can be very successful under severe disturbance regimes
171 like ploughing or landslides [42].

172 Compared to the other topics, the disturbance-clonality relationship has been addressed
173 more in-depth, making clear that clonality needs to be taken into account when building a
174 comprehensive scheme of response to disturbance. Additional research on the morphological
175 means of clonal and nonclonal bud-bearing organs and their relationship to specific disturbance
176 regimes would be beneficial.

177

178 **Population structure and evolution**

179

180 Plant species have remarkable diversity in their life histories, which partly corresponds to
181 the changes of their bodies in response to productivity and disturbance gradients. This is reflected
182 by the different relative roles of survival, reproduction, and growth in plant life cycles. Again,
183 this simple scheme essentially excludes clonal growth, which can play a decisive role in both the
184 growth and reproduction components of plant life cycles, with consequences on population
185 structure, reproductive strategy, and diversification rates.

186 First, clonal reproduction has a number of short-term ecological effects. It may serve to
187 boost population growth rates under favourable conditions [43] and provide reproductive
188 assurance to genetic individuals. Clonal plants thus have consistently lower investment into
189 reproduction via seed than nonclonal plants [44, 45]. While species differ strongly both in the
190 presence and magnitude of clonal reproduction, we only have limited data for the extent of this
191 differentiation (see also Box 2). In particular, there is almost no demographic information on
192 species with extensive clonal growth because these cannot be studied using simple marking-
193 based demographic techniques [46].

194 Second, the population structure of species that are capable of clonal growth is very
195 different from that of nonclonal species because of the uncoupling of genetic and ecological
196 individuals [43]. Although the overall genetic diversity of clonal species does not necessarily
197 differ from that of nonclonal species, some clonal species may have many genetically identical
198 ramets within a population – a situation that is otherwise found only in apomictic or highly
199 autogamous species [47]. This phenomenon, amplified by the spatial aggregation of genetically
200 identical ramets, may result in restricted pollen dispersal among genets [44, 47, 48] and have
201 important consequences for autogamy, selection of floral traits, and display for pollinators (Fig. 1
202 in [47]). These relationships have never been systematically explored, either by comparing clonal

203 and nonclonal species, or across different types of spatial organisations of clonal plants.
204 Unfortunately, we also do not possess good estimates of genet lifespan in more than a few model
205 plants [49]. Interestingly, both clonal growth and incompatibility are related to whole-genome
206 duplication, and polyploid plants are more likely to be both self-compatible and clonal [50, 51],
207 although links between these two patterns have not been addressed.

208 Finally, clonal reproduction strongly extends generation time and thus may change rates of
209 molecular evolution and speciation, similarly to plant size [52]. While it has rarely been
210 systematically explored, clonality may contribute to explaining latitudinal or environmental
211 gradients in plant diversity as clonal plants tend to prevail in cold and wet environments [25].
212 This may be one of the reasons for the slow evolution of aquatic angiosperms that have extensive
213 clonal growth, although the comparison between resprouters and seeders in fire-prone areas does
214 not support the same conclusion [53].

215 Unfortunately, we do not possess good estimates of genet lifespan in more than a few
216 model plants therefore we are in need of new methodological approaches in studying
217 demographic parameters in clonal plants [44, 49, 54].

218

219 **Conclusions**

220

221 Existing evidence has shown that clonal species differ systematically from nonclonals in a
222 number of features and as their proportion in different communities and biomes varies (Box 2),
223 these functional differences likely scale up to affect community- and ecosystem-level dynamics.
224 It has also become clear that despite its previous treatment in the literature, clonality is not a
225 binary trait (i.e. presence or absence) and there is a large functionally relevant spectrum spanning

226 across different clonal strategies and species. Incorporating clonality into the mainstream plant
227 ecology research platform requires a closer look at the functions that vary across clonal strategies,
228 and identifying comparable functions in nonclonal plants so that all species can be placed into a
229 general strategy scheme.

230 Finally, we point out that one of the key reasons why clonality has been under-developed in
231 plant ecology research at large, lies in the fact that it is inevitably tied to a whole-plant
232 perspective, which is not easily incorporated into current approaches to plant functional
233 differentiation (e.g. [8, 55]). Such approaches (be it trait-based comparative ecology, population
234 ecology, or ecosystem ecology) have their strengths in reducing plant bodies and organs to simple
235 functional traits applicable to any species. This means that dimensions of the plant life that are
236 difficult to reduce to such a scheme are necessarily neglected. We believe that the inclusion of
237 plant clonality into mainstream ecological research would capture better the essential feature of
238 plant life (i.e. the hierarchical and modular structure of any individual; Box 3, [56]) and would
239 add a whole-plant perspective that many of our studies have been lacking.

240

241 **Acknowledgements**

242 The work on the Opinion was supported by Czech Science Foundation (GAČR Project 19-
243 13231S to JK, JM, TH, 19-14394Y to GO), by Ministry of Education, Youth and Sports of the
244 Czech Republic (LTT20003 to ZJ) and by the by long-term research development project of the
245 Czech Academy of Sciences [No. RVO 67985939].

246

247

248 **References**

249 1. Klimešová, J. et al. (2018) Belowground plant functional ecology: towards an integrated
250 perspective. *Funct. Ecol.* 32, 2115-2126

251 2. Klimešová, J. et al. (2019) Handbook of standardized protocols for collecting plant modularity
252 traits. *Perspect. Plant Ecol. Evol. Syst.* 40, 125485

253 3. Freschet, G.T. et al. (2021) Root traits as drivers of plant and ecosystem functioning: current
254 understanding, pitfalls and future research needs. *New Phytol.* doi.org/10.1111/nph.17072

255 4. Ottaviani, G. Et al. (2020) The neglected belowground dimension of dominance. *Trends Ecol.*
256 *Evol.* 35, 763-766

257 5. Franklin, S. et al. (2021) Next-Gen Plant Clonal Ecology. *Perspect. Plant Ecol. Evol. Syst.* 49,
258 125601

259 6. Chytrý, M. et al. (2021) Pladias Database of the Czech Flora and Vegetation. *Preslia* 93, 1–87

260 7. Freschet, G.T. et al. (2010) Evidence of the ‘plant economics spectrum’ in a subarctic flora. *J.*
261 *Ecol.* 98, 362-373

262 8. Reich, P.B. (2014) The world-wide „fast-slow“ plant economics spectrum: a traits manifesto. *J.*
263 *Ecol.* 102, 275–301

- 264 9. Wright, I. et al. (2004) The worldwide leaf economics spectrum. *Nature* 428, 821–827
- 265 10. Mokany, K. et al. (2006). Critical analysis of root: Shoot ratios in terrestrial biomes. *Global*
266 *Change Biol.* 12, 84–96
- 267 11. Poorter, H. et al. (2015) How does biomass distribution change with size and differ among
268 species? An analysis for 1200 plant species from five continents. *New Phytol.* 208, 736-
269 749
- 270 12. Reynolds, H.L. and Antonio, C.D. (1996) The ecological significance of plasticity in root
271 weight ratio in response to nitrogen: Opinion. *Pl. Soil* 185, 75–97
- 272 13. Midgley, J.J. (1996) Why the world's vegetation is not totally dominated by resprouting
273 plants; because resprouters are shorter than reseeders. *Ecography* 19, 92-95
- 274 14. Lubbe, F.C. et al. (2021) Carbohydrate storage in herbs: the forgotten functional dimension of
275 the plant economic spectrum. *Ann. Bot.* doi.org/10.1093/aob/mcab014
- 276 15. Ottaviani, G. et al. (2021) Strong impact of management regimes on rhizome biomass across
277 Central European temperate grasslands. *Ecol. Appl.* doi.org/10.1002/eap.2317
- 278 16. Kobe, R.K. et al. (2010) Optimal partitioning theory revised: non-structural carbohydrates
279 dominate root mass responses to nitrogen. *Ecology* 9, 166–179
- 280 17. Cornelissen, J.H.C. (2004) Plant traits and ecosystem effects of clonality: a new research
281 agenda. *Ann. Bot.* 114, 369–376
- 282 18. Klimešová, J et al. (2021) Are belowground clonal traits good predictors of ecosystem
283 functioning in temperate grasslands? *Funct. Ecol.* 35, 787-795
- 284 19. Stuefer, J.F. (1998) Two types of division of labour in clonal plants: benefits, costs and
285 constraints. *Perspect. Plant Ecol. Evol. Syst.* 1, 47-60
- 286 20. Yu, F.H. (2008) Adaptation of Rhizome Connections in Drylands: Increasing Tolerance of
287 Clones to Wind Erosion. *Ann. Bot.* 102, 571–577

- 288 21. Jónsdóttir, I.S. and Watson, M.A. (1997) Extensive physiological integration: An adaptive
289 trait in resource-poor environments? In *The ecology and evolution of clonal plants* (de
290 Kroon, H. and van Groenendael, J.M., eds.) pp. 109-136, Backhuys Publishers, Leiden
- 291 22. Klimeš, L. (2008) Clonal splitters and integrators in harsh environments of the Trans-
292 Himalaya. *Evol. Ecol.* 22, 351–367
- 293 23. Iwasa, Y. and Kubo, T. (1997) Optimal size of storage for recovery after unpredictable
294 disturbances. *Evol. Ecol.* 11, 41–65
- 295 24. Klimešová, J. et al. (2011) Evolutionary and organismic constraints on the relationship
296 between spacer length and environmental conditions in clonal plants. *Oikos* 120, 1110-
297 1120
- 298 25. Klimešová, J. et al. (2016) Herbs are different: clonal and bud bank traits can matter more
299 than leaf–height–seed traits. *New Phytol.* 210, 13-17
- 300 26. Charles- Dominique, T. et al. (2018) Steal the light: shade vs fire adapted vegetation in
301 forest–savanna mosaics. *New Phytol.* 2018, 1419-1429
- 302 27. Barot, S. and Gignoux, J. (2004) Mechanisms promoting plant coexistence: can all the
303 proposed processes be reconciled? *Oikos* 106, 185–19
- 304 28. Bolker, M. et al. (2003) Spatial Dynamics in Model Plant Communities: What Do We Really
305 Know? *Am. Nat.* 162, 135-148
- 306 29. Bullock, J.M. et al. (2017) A synthesis of empirical plant dispersal kernels. *J. Ecol.* 105, 6–19
- 307 30. Zobel, M. et al. (2010) Clonal mobility and its implications for spatio-temporal patterns of
308 plant communities: what do we need to know next? *Oikos* 119, 802-806
- 309 31. Lovett-Doust, L. (1981) Population dynamics and local specialization in a clonal perennial
310 (*Ranunculus repens*): I. The dynamics of ramets in contrasting habitats. *J. Ecol.* 69, 743-
311 755

- 312 32. Brooker, R.W. (2017) Clonal plants and facilitation research: bridging the gap. *Folia Geobot.*
313 52, 295–302
- 314 33. Lu, H.Z. (2020) When facilitation meets clonal integration in forest canopies. *New Phytol.*
315 225, 135-142
- 316 34. Hara, T. and Šrůtek, M. (1995) Shoot growth and mortality patterns of *Urtica dioica*, a clonal
317 forb. *Ann. Bot.* 76, 235-243
- 318 35. Avolio, M.L. et al. (2019) Demystifying dominant species. *New Phytol.* 223, 1106-1126
- 319 36. Grime, J.P. (1998) Benefits of plant diversity to ecosystems: immediate, filter, and founder
320 effects. *J. Ecol* 86, 902–910
- 321 37. Pausas, J.G. et al. (2018) Unearthing belowground bud banks in fire- prone ecosystems. *New*
322 *Phytol.* 217, 1435-1448
- 323 38. Bellingham, P.J. and Sparrow, A.D. (2000) Resprouting as a life history strategy in woody
324 plant communities. *Oikos* 89, 409-416
- 325 39. Bruelheide, H. et al. (2018) Global trait–environment relationships of plant communities.
326 *Nature Ecol. Evol.* 2, 1906-1917
- 327 40. Martínková, J. et al. (2020) Response of clonal versus non-clonal herbs to disturbance:
328 different strategies revealed. *Perspect. Plant Ecol. Evol. Syst.* 44, 125529
- 329 41. Martínková, J. et al. (2020) Young clonal and non-clonal herbs differ in growth strategy but
330 not in aboveground biomass compensation after disturbance. *Oecologia* 193, 925–935
- 331 42. Guerrero- Campo, J. et al. (2008) Plant traits enabling survival in Mediterranean badlands in
332 northeastern Spain suffering from soil erosion. *J. Veg. Sci.* 19, 457-464
- 333 43. Janovský, Z. and Herben, T. (2020) Reaching similar goals by different means–Differences in
334 life-history strategies of clonal and non-clonal plants. *Perspect. Plant Ecol. Evol. Syst.* 44,
335 125534

- 336 44. Vallejo-Marín, M. et al. (2010) The Ecological and Evolutionary Consequences of Clonality
337 for Plant Mating. *Annu. Rev. Ecol. Evol. Syst.* 41, 193-213
- 338 45. Herben, T. et al. (2015) Clonal growth and sexual reproduction: tradeoffs and environmental
339 constraints. *Oikos* 124, 469–476
- 340 46. Janovský, Z. et al. (2017) Accounting for clonality in comparative plant demography –
341 growth or reproduction? *Folia Geobot.* 52, 433-442
- 342 47. Barrett, S.C.H. (2015) Influences of clonality on plant sexual reproduction. *Proc. Natl. Acad.*
343 *Sci. U. S. A.* 112 (29), 8859-8866
- 344 48. Stephens, S. et al. (2020) Patterns of pollen dispersal and mating in a population of the clonal
345 plant *Sagittaria latifolia*. *J. Ecol.* 108, 1941-1955
- 346 49. Arnaud Haond, S. et al. (2008) Standardizing method to address clonality in population
347 studies. *Molecular Ecol.* 16, 5115-39
- 348 50. Herben, T. et al. (2017) Polyploid species rely on vegetative reproduction more than diploids:
349 a re-examination of the old hypothesis. *Ann. Bot.* 120, 341–349
- 350 51. van Drunen, W.E. and Husband, B.C. (2019) Evolutionary associations between polyploidy,
351 clonal reproduction, and perenniality in the angiosperms. *New Phytol.* 224, 1266-1277
- 352 52. Boucher, F.C. et al. (2017) Plant size: a key determinant of diversification? *New Phytol.*
353 2016, 24-31
- 354 53. J.G. Pausas, et al. (2016) Towards understanding resprouting at the global scale. *New Phytol.*
355 209, 945-954
- 356 54. Eriksson, O. and Jerling, L. (1990) Hierarchical selection and risk spreading in clonal plants.
357 In *Clonal growth in plants: regulation and function* (van Groenendael, J. and de Kroon,
358 H., eds) pp. 79–94, SPB Academic Publishing, The Hague

- 359 55. Salguero-Gómez, R. et al. (2016) Fast–slow continuum and reproductive strategies structure
360 plant life-history variation worldwide. *Proc. Natl. Acad. Sci. U. S. A.* 113 (1), 230-235
- 361 56. Barthelemy, D. and Caraglio, Y. (2007) Plant architecture: a dynamic, multilevel and
362 comprehensive approach to plant form, structure and ontogeny. *Ann. Bot.* 99, 375-407
- 363 57. Howard, C.C. et al. (2018) The monocotyledonous underground: global climatic and
364 phylogenetic patterns of geophyte diversity. *Am. J. Bot.* 106, 850-863
- 365 58. Herben, T. and Klimešová, J. (2020) Evolution of clonal growth forms in angiosperms. *New*
366 *Phytol.* 225, 999-1010
- 367 59. Zhang, H. Et al. (2018) Is the proportion of clonal species higher at higher latitudes in
368 Australia ? *Austral Ecol.* 43, 69-75
- 369 60. Ye, D. Et al. (2014) Clonality-Climate Relationships along Latitudinal Gradient across China:
370 Adaptation of Clonality to Environments. *PloS ONE* 9: e94009
371 [Doi.org/10.1371/journal.pone.0094009](https://doi.org/10.1371/journal.pone.0094009)
- 372 61. Klimešová, J. et al. (2011) Clonal growth forms in eastern Ladakh, Western Himalayas:
373 classification and habitat preferences. *Folia Geobot.* 46, 191-217
- 374 62. Fujinuma, J. et al. (2019) Species- specific clonality in east Asian island flora: Phylogenetic
375 and environmental constraints. *Ecol. Res.* 34, 577-586
- 376 63. Sosnová, M. et al. (2010) Distribution of clonal growth forms in wetlands. *Aquat. Bot.* 92,
377 33-39
- 378 64. Wellstein, K. and Kuss, P. (2011) Diversity and frequency of clonal traits along natural and
379 land-use gradients in grasslands of the Swiss Alps. *Folia Geobot.* 46, 255-270
- 380 65. Klimešová, J. and Doležal, J. (2011) Are clonal plants more frequent in cold environments
381 than elsewhere? *Plant Ecol. Div.* 4, 373-378

- 382 66. Chytrý, M. and Rafajová, M. (2003) Czech National Phytosociological Database: basic
383 statistics of the available vegetation-plot data. *Preslia*. 75, 1-15
- 384 67. Klimešová, J. and Herben, T. (2015) Clonal and bud bank traits: patterns across temperate
385 plant communities. *J. Veg. Sci.* 26, 243-253
- 386 68. Charles- Dominique, T. et al. (2015) Using intra- individual variation in shrub architecture to
387 explain population cover. *Oikos*, 124, 707-716
- 388 69. Charles-Dominique, T. et al. (2009) Architectural strategies of *Cornus sericea*, a native but
389 invasive shrub of Southern Quebec, Canada, under an open or a closed canopy. *Ann. Bot*
390 105, 205-220
- 391 70. Charles-Dominique, T. et al. (2012) Architectural strategies of *Rhamnus cathartica*
392 (*Rhamnaceae*) in relation to canopy openness. *Botany* 90, 976-989

393

394 **Glossary:**

395 **Acquisitive organs:** organs acquiring resources, namely leaves as photosynthesising organs that
396 capture light and produce organic compounds using water, light, and CO₂; and fine roots that
397 capture water and mineral nutrients. They are characterised by high surface to volume ratios.

398 **Bud bank:** a pool of dormant buds borne by belowground storage organs from which the plant
399 regrows after seasonal rest or after the damage of aboveground parts.

400 **Clonality:** the ability of one genetic individual (genet) to produce more than one physically
401 independent rooting unit (ramet) during its lifespan. The degree of clonal multiplication differs
402 among species so that some of them can grow clonally but rarely do so (Potential clonality) while
403 others multiply regularly and clonality is a regular part of their life cycle (Functional clonality).

404 **Clonal growth organs:** organs through which vegetative multiplication takes place. They are
405 usually placed belowground and have different morphologies depending on the participation of
406 stems (e.g. rhizomes), leaves (e.g. bulbs), and roots (e.g. root-sprouting).

407 **Genet:** the term denotes all ramets descending from one zygote irrespectively of their physical
408 connections.

409 **Nonacquisitive organs:** organs providing resource storage, clonal multiplication, lateral spread,
410 and bud bank that often connect aboveground stems and roots, but do not participate directly in
411 acquiring resources. They are characterised by low surface to volume ratios.

412 **Phalanx-guerrilla growth forms:** horizontal distances between ramets of one clone vary
413 strongly, ranging from tightly packed ramets (phalanx) to loose stands intermingled with other
414 species (guerrilla). While intraspecific (and intragenotypic) interactions prevail in the phalanx
415 strategy, interspecific interactions prevail in the guerrilla. The runner vs. clumper continuum
416 denotes the same thing.

417 **Ramet:** the term identifies the smallest potentially independent unit in clonal plants with its own
418 shoot and root system. Ramets may (but do need not to) be linked by clonal growth organs
419 (rhizomes, roots, or stolons); a set of linked ramets is usually called clone or clonal fragment.

420 **Storage organs:** coarse organs of different morphological origin specialised for storing
421 carbohydrates and building bud bank, generally located belowground.

422 **Splitter-integrator continuum:** clonal connections between ramets of one clone strongly vary in
423 their duration, ranging from long-lived (integrators) to short-lived (splitters) connections among
424 ramets. While integrators can share resources over many years (up to tens), the ability to share
425 resources in splitters is lost after a few weeks or months when the offspring ramet becomes
426 mature.

427 **Seeders and resprouters:** strategies described in ecosystems with recurrent severe disturbance,
428 usually fire. Seeders do not survive disturbance as adult plants and thus regenerate after
429 disturbance only from seeds; in contrast, resprouters survive and resprout from bud bearing
430 organs, often located belowground.

431

432 **Box 1.** Phylogenetics of clonality in angiosperms.

433 Clonality is widespread in all major clades of angiosperms. It shows marked phylogenetic
434 clustering, although transitions between clonal and non-clonal forms have been fairly common
435 (Figure I). Specifically, all monocots are at least potentially clonal [57] (although not all of them
436 are clonal functionally, i.e. having clonality as a part of their life cycle). From the morphogenetic
437 point of view, clonal growth is attained by two distinct processes, either by the formation of
438 adventitious roots on stems (stem-derived clonality, i.e. stolons or rhizomes) or by forming
439 adventitious buds on roots (root-derived clonality). Phylogenetic transition rates among stem-
440 derived clonality, root-derived clonality, and nonclonal forms show that both morphological
441 types represent evolutionarily independent pathways for attaining clonal growth (Figure II).

442

443 Figure I - Phylogenetic patterns of **functional clonality** in angiosperms showing large clusters of
444 predominantly clonal species (e.g. monocots, rosids, Asterales) and also clusters where nonclonal
445 forms prevail (fabids, Apiales). Red colour indicates the reconstructed probability of possessing
446 clonal growth at a given node of the phylogenetic tree, covering 2909 angiosperm species of
447 Central European flora. Estimate of Pagel's lambda for clonality here is 0.815 (95.0% confidence
448 interval: 0.774-0.849; lambda = 1 indicates full phylogenetic signal corresponding to the
449 Brownian motion evolution, lambda = 0 indicates complete phylogenetic randomness). Clonality

450 evolution was estimated using stochastic character mapping of 50 simulated character histories
451 based on a continuous-time reversible Markov model of clonality-nonclonality transitions. These
452 histories were used to estimate the expected number of evolutionary transitions over the tree and
453 to reconstruct probabilities of the trait occurrence on individual tree nodes. For the data and
454 further details see [58].

455
456 Figure II - Root-derived clonality and stem-derived clonality represent two evolutionarily
457 independent pathways to attain clonal growth. Transitions between stem-derived clonality and
458 nonclonal forms have been common, as well as between nonclonal plants and root-derived
459 clonality. Root-derived clonality appeared almost exclusively in nonclonal species as an
460 alternative pathway to attain clonal growth [58]. The low frequency of transitions between stem-
461 and root-derived clonality indicates that both types are alternative solutions to the same selective
462 force acting on non-clonal species. Transitions were estimated using simulated stochastic
463 character histories based on the continuous Markov chain rate matrix estimated from 2909
464 angiosperm species of Central European flora. The frequency of transitions is expressed relative
465 to the total number of transitions in the tree. Line width is proportional to the transition
466 probability; transitions with zero relative frequency are indicated by the dotted line. For the data
467 and further details see [58].

468
469 **Box. 2.** Participation of clonal species in floras and communities.
470 Clonal species are more common in wetter, colder and more disturbed environments. This can be
471 shown both by large regional (Figure I) and local (Figure II) differences in the proportion of
472 clonal plants, which reflect structure of species pools as well as of ecological filters.

473
474 Figure I - Proportion of clonal species in different floras of the world, showing a consistent
475 increasing trend in participation of clonal species towards higher latitudes and altitudes. Numbers
476 indicate the total number of species analysed; stars indicate the number of species in a larger
477 region when the number of species analysed was not available. It is to be noted that individual
478 surveys [59-65] do not necessarily work with the identical definition of clonality, potentially
479 adding some noise to the data. To exemplify this, we are showing Central European data using
480 two definitions of clonality: "potential" clonality, i.e. including all species that have the potential
481 to grow clonally, and "functional" clonality, i.e. including only species that regularly have clonal
482 growth as a part of their life cycle.

483
484 Figure II - Participation of clonal species in different vegetation types in one region (Czech
485 Republic), showing a steep increase of clonal species proportion towards community types with
486 higher water availability or moderate disturbance. It is based on a stratified set of 20486
487 vegetation records from the Czech National Phytosociological Database [66], classified into
488 vegetation types using EUNIS habitat classification. Participation is expressed as the proportion
489 of clonal species out of all species in records of the given vegetation types; error bars indicate
490 95% confidence intervals. Numbers indicate the total number of each vegetation type records.
491 Modified from [67].

492
493 **Box 3.** Plant architecture of clonal and nonclonal plants.
494 Analysing architecture (Figure I) helps to elucidate plant response to the environment [68]. For
495 example, plants respond to light limitation by adjusting their architecture and occupation strategy

496 in both clonal and nonclonal plants [69, 70]. As shown for the clonal shrub *Z. americanum*
497 (Figure II), shoots are produced at regular intervals (Fig. 2) on long horizontal roots and they die
498 if growing in the dense shade (Fig. 2 C) or develop into ramets with a complex structure (Fig. 2
499 B). When the clone is developing in the understory, horizontal roots linking ramets senesce and
500 trigger new taproots (Fig. 2 D) emitting radially horizontal roots and resetting soil exploration in
501 all directions. Soil exploration in the horizontal dimension stops when it comes to contact with
502 another clone from the same species (but not from other plant species) suggesting a chemical
503 signaling responsible for this ‘root shyness’ between clones. This example illustrates important
504 parts of clonal strategies: radial colonisation, exploration of suitable patches, and limitation of
505 intraspecific competition.

506
507 Figure I – Plant structure is based on the repetition of different levels of structural units, from
508 phytomeres to growth and architectural units, to branch complexes. The complexity of the
509 structure is shown by highlighting module (yellow), foliage of largest structural unit (purple), and
510 rooting unit (beige) in nonclonal (1, 2, 4) and clonal plants (3, 5, 6, 7). In the simplest case, the
511 genet is also a module and the rooting unit (e.g. annual herb: 2) but the clone can also be
512 composed of several modules (e.g. rhizomatous forb: 6). In more complex structures, the module
513 can be integrated into a larger structural unit that can correspond to a distinct rooting unit,
514 therefore a genet (e.g. tree and shrub: 1, 4) or can be part of an even larger clone (e.g. multi-
515 stemmed shrub: 5, root sprouting shrub: 7). In some plants, modules can include several rooting
516 units, ramets (e.g. creeping herb: 3).

517

518 Figure II – Clone structure of *Zanthoxylum americanum* in open (left) and shaded environments
519 (right). Ramets (B; green arrows or fully drawn) and small shoots (C) are regularly spaced along
520 horizontal roots (light brown) emitted from a single taproot (A; dark brown). When the clone
521 grows in the understory, horizontal roots frequently senesce, triggering development of secondary
522 taproots (D) (4).

523

524

525

1 **Outstanding questions:**

2 What is the role of storage organs in the plant economics spectrum? Are conservative and
3 acquisitive trait strategies related to clonal growth, and to the abundance and mass of bud-
4 bearing and storage organs?

5 How do the main costs and benefits associated with clonality change with key environmental
6 gradients, such as productivity and disturbance?

7 In there a general pattern of how clonal and nonclonal strategies differ in species interactions
8 such as competition or facilitation in plant communities? Do such differences affect species
9 coexistence and persistence in communities?

10 How does clonality affect population dynamics, seed reproduction, and population genetics?

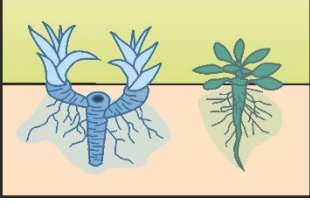
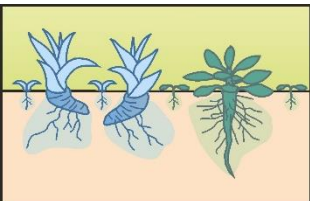
11 Do clonal plants have different evolutionary rates than nonclonal plants?

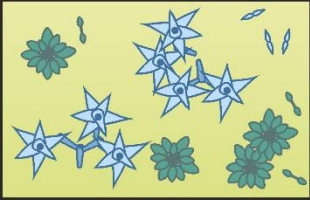
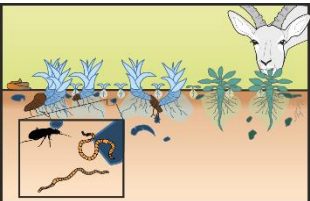
12 What is the role of clonal growth, bud-bearing and storage organs in ecosystem functioning
13 (e.g. soil carbon cycling and storage, protection against erosion)?

14 How can we devise a general plant strategy scheme for all plants and incorporate key
15 functions provided by clonality? How do we account for the different morphological means
16 (such as stem- and root-derived) and strategies (such as phalanx and guerrilla or splitter and
17 integrator) of clonality?

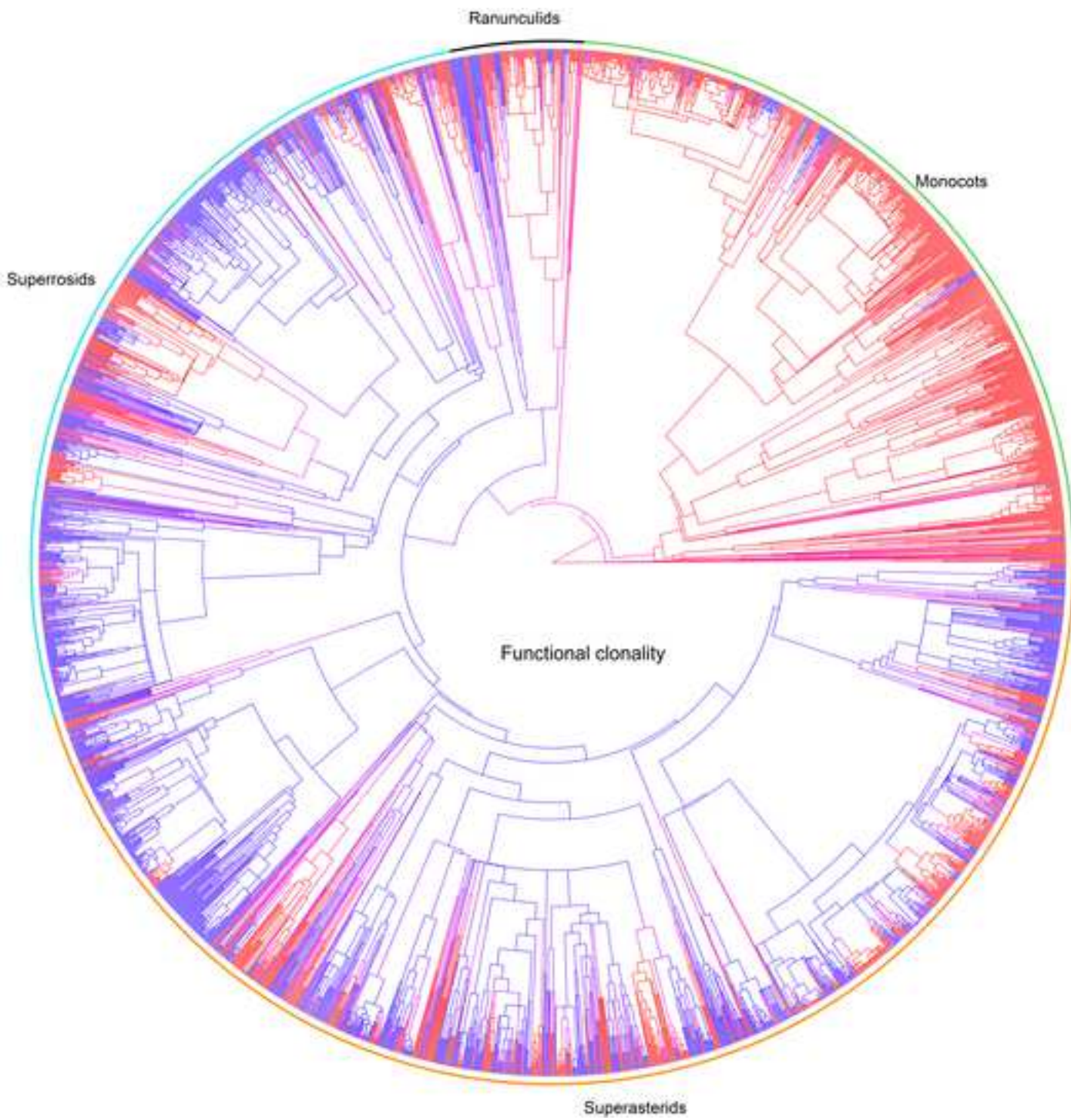
18

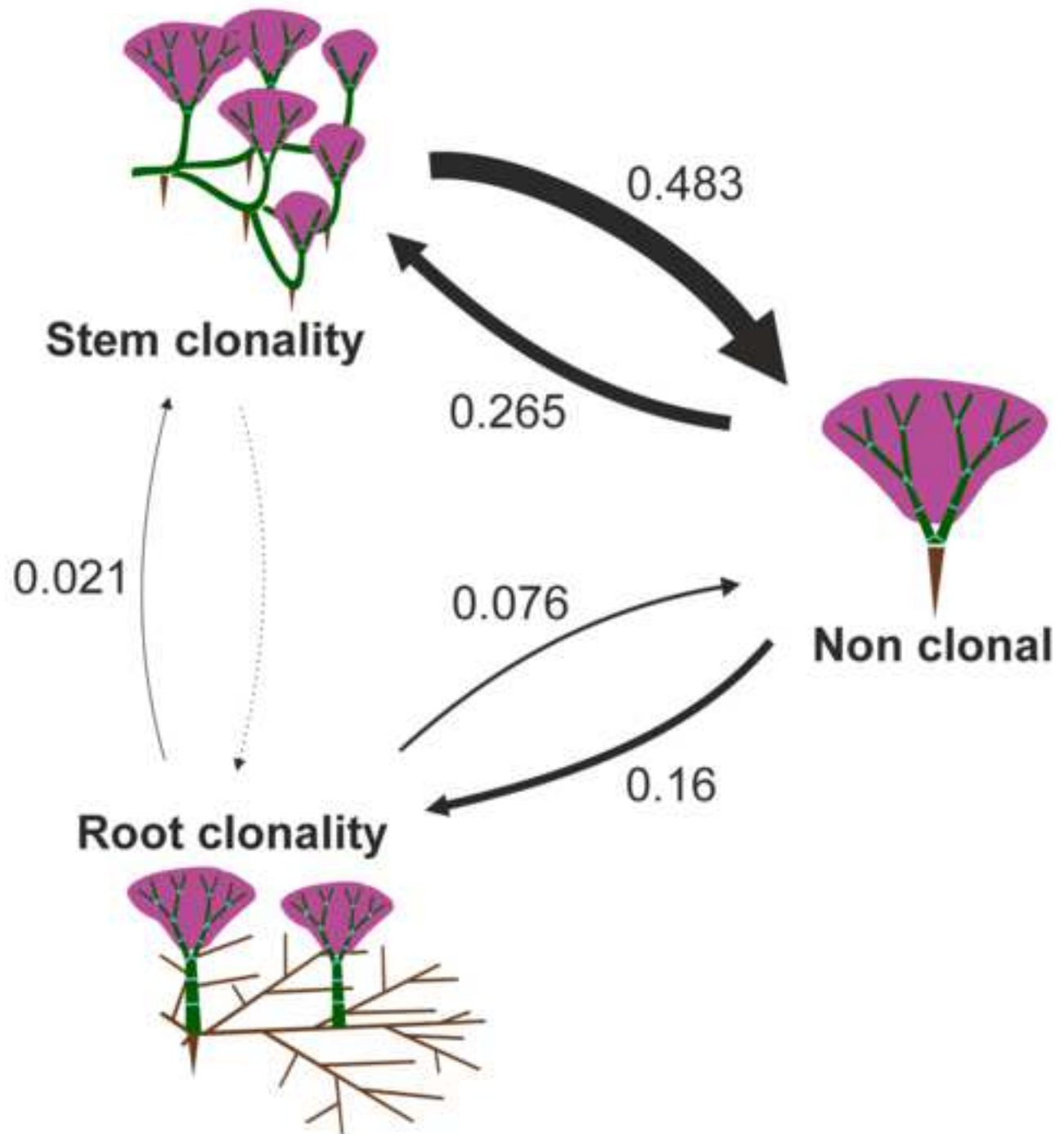
1 **Table 1.** Contrasting features of clonal and nonclonal plants.

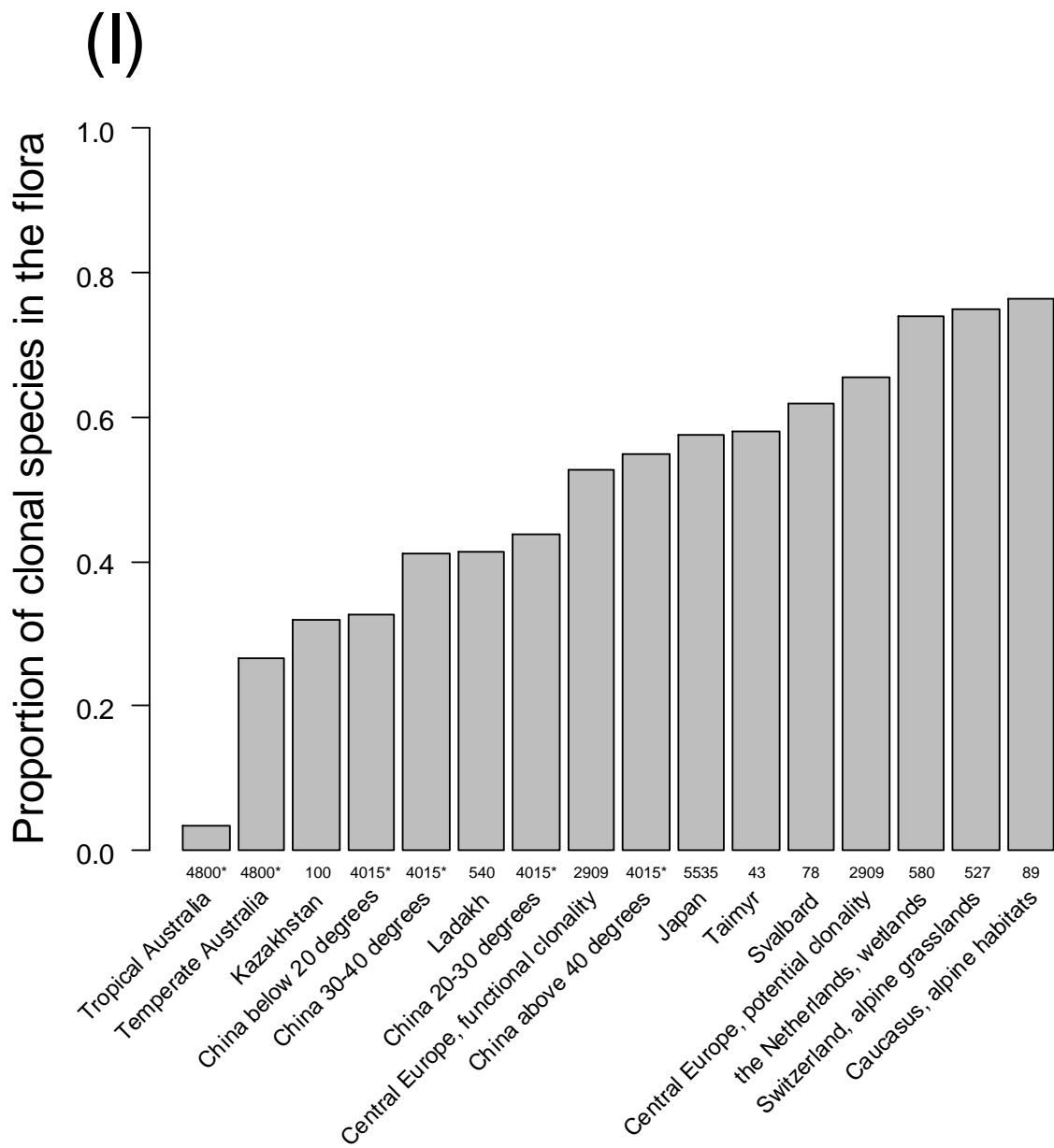
Organisational level	Functions & processes	Clonal plants	Nonclonal plants	Available data
Individual 	Resource allocation	Large biomass investment in storage and/or clonal growth organs	Low biomass investment into storage organs (except resprouting nonclonals)	Rare
	Resource acquisition	Rooting over a large area but with little resource foraging	Deep rooting and with marked resource foraging	Rare
	Resource sharing	Translocation of resources among rooting units forming a clone, specialisation of rooting units for obtaining abundant resources	No possibility of resource sharing (among rooting units) apart from mycorrhizas, no specialisation of rooting units for obtaining limiting resources	Rich (for clonal plants only)
Population 	Reproduction	Vegetative and generative	Only generative	Rich
	Natural selection	Decoupling of genetic and physiological and functional individuals	Genetic corresponds to a physiological individual	Rare
	Regeneration after disturbance	Vegetative and generative	Mostly generative	Rich
	Regeneration niche	Can establish in dense vegetation, large parental provisioning	Dependent on canopy gaps, small parental provisioning	Moderate
	Competition	In vertical and horizontal directions	Mainly vertical direction	Rare

<p>Community</p> 	Coexistence	Alleviation of competition by mobility (guerrilla strategy) or aggregation and clonal integration (phalanx strategy)	Negative density dependence	Rare
	Facilitation	Aggregated rooting units (phalanx) may serve as nurse plants	Only cushion plants	Rare
	Evolvability	Slow	Fast	Unknown
	Dominance	Aboveground abundance is not a good proxy for belowground abundance	Aboveground abundance is a good proxy for abundance belowground (except resprouting nonclonals)	Rare
	Carbon cycling	Continuous litter input belowground thanks to exudation, senescing rhizomes and roots	Continuous carbon input belowground through exudation and senescing roots	Moderate (only for exudates and senescing roots)
<p>Ecosystem</p> 	Environment homogenisation	Translocation of resources in the horizontal direction through ramets forming a clone	Water and nutrients uplift from deep soil layers thanks to deep rooting	Moderate (only for non-clonal)
	Aeration of waterlogged soils	Aeration through rhizomes and roots	Aeration through roots	Unknown

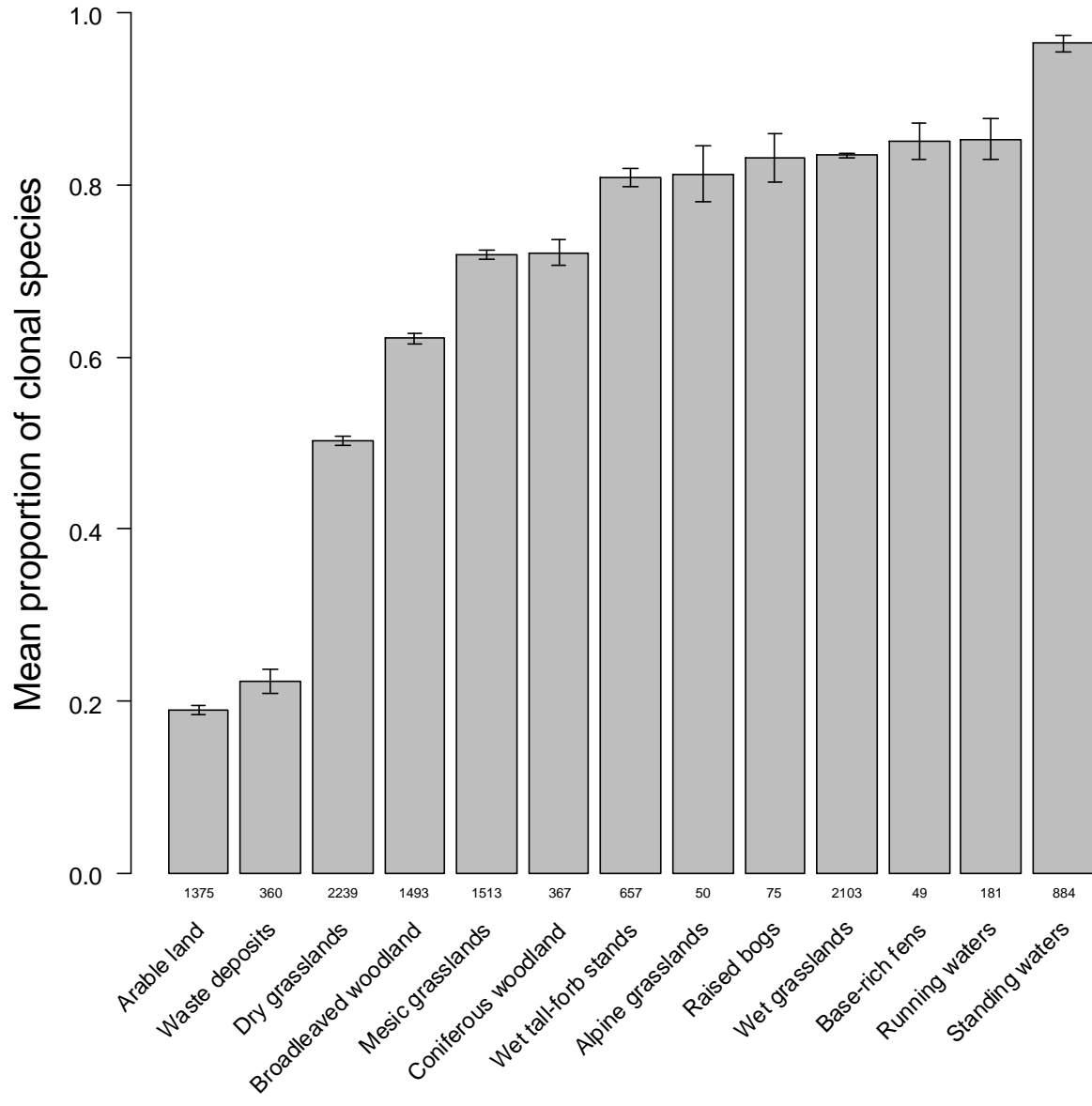
2
3
4







(II)



A Whole plant architecture

B module

