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Jean Michel Brunel, Sophie Négrel. Synthesis and Biological Activities of Naturally Functionalized Polyamines: An Overview. *Current Medicinal Chemistry*, 2021, 28 (17), pp.3406-3448. 10.2174/0929867327666201102114544 . hal-03367292

HAL Id: hal-03367292

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Submitted on 11 Oct 2021

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Synthesis and biological activities of natural **functionalized** polyamines: An overview

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Abstract

Recently, extensive researches have emphasized the fact that polyamines conjugates are becoming important in all the biological and medicinal fields. In this review we will focus our attention on natural polyamines and highlight recent progress in both fundamental mechanism studies and interest for the development and application for a therapeutic human use of polyamine derivatives.

Keywords: natural polyamines, antimicrobial activities, toxins, marine drugs

I-Introduction

The discovery of new compounds from nature is still one of the most efficient methods for finding lead molecules for the development of pharmaceuticals. Thus, following the discovery of penicillin and among all the families of organic compounds encountered in nature, polyamines have long been neglected as potent molecules of biological interest. On a structural point of view, polyamines are polycationic molecules at physiological pH possessing a hydrocarbon backbone and multiple amino groups (Figure 1). [1] The first depicted polyamines were isolated from prokaryotic and eukaryotic organisms and possessing at least two amino groups as part of an aliphatic chain. [2, 3]

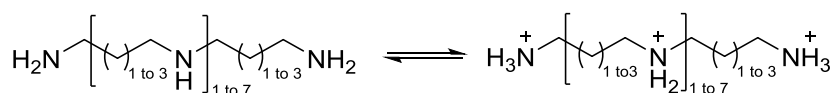


Figure 1. General structure of polyamines.

Therefore, due to the proximity of at least two N-atoms in their structures, polyamines show chemical reactivities significantly different from monoamines as well as peculiar biological activities. [2] In this review, we will only focus our attention on natural polyamines and highlight the potent interest for the development and application for a therapeutic human use.

II-Natural aliphatic polyamines

This class of polyamines constitutes the simplest class of polyamines naturally encountered and presenting NH₂ groups separated by a small or large aliphatic chain. The most known polyamines are the aliphatic low-molecular weight putrescine **1**, cadaverine **2**, spermidine **3** and spermine **4** and, found in all eukaryotic and prokaryotic cells and involved in a wide range of biological phenomena from bacteria to plants and animals (

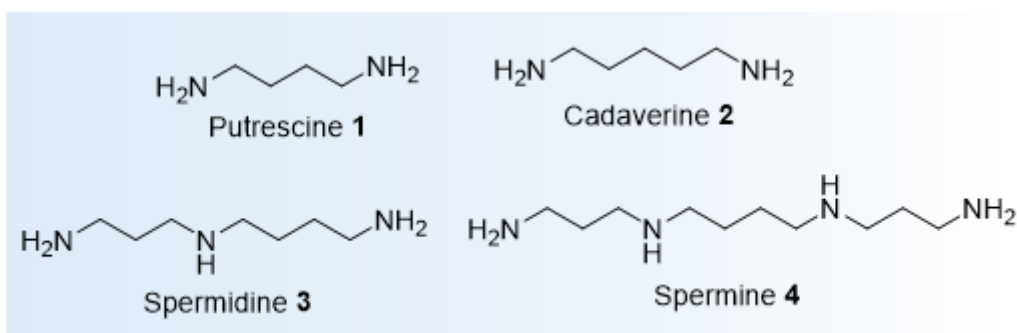


Figure 2).

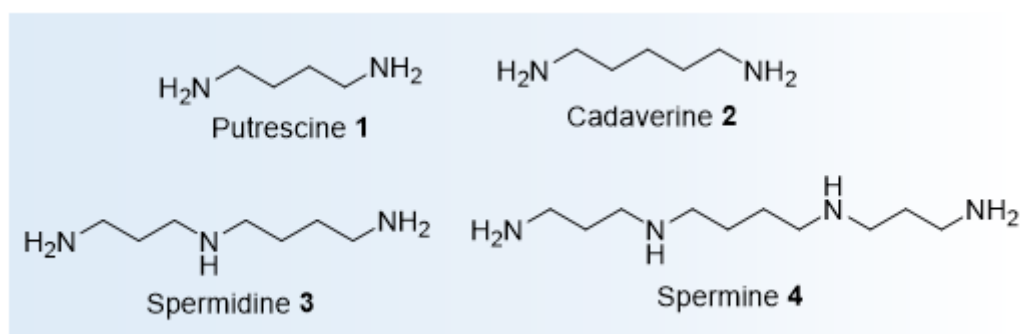


Figure 2. Structure of usual polyamines.

Thus, these polyamines are essential for cells growth due to their polybasic character conferring them a much higher affinity for acidic constituents and allowing reversible ionic interactions leading to the stabilization of macromolecules such as DNA, RNA, and proteins. [3-7] They can also act as second messengers in protein phosphorylation and hormonal signal transduction. [3, 8] Polyamines **1**, **3** and **4** are synthesized in a highly regulated biochemical pathway as described in Figure 3 from ornithine **5** through a biomechanism involving ornithine decarboxylase, spermidine and spermine synthetases, spermidine/spermine N¹-acetyltransferase and FAD-dependent polyamine oxidase. [3] On the other hand, cadaverine **2** came from lysine and its biosynthesis is catalyzed by a lysine decarboxylase via a pyridoxal phosphate-dependent decarboxylation. [8]

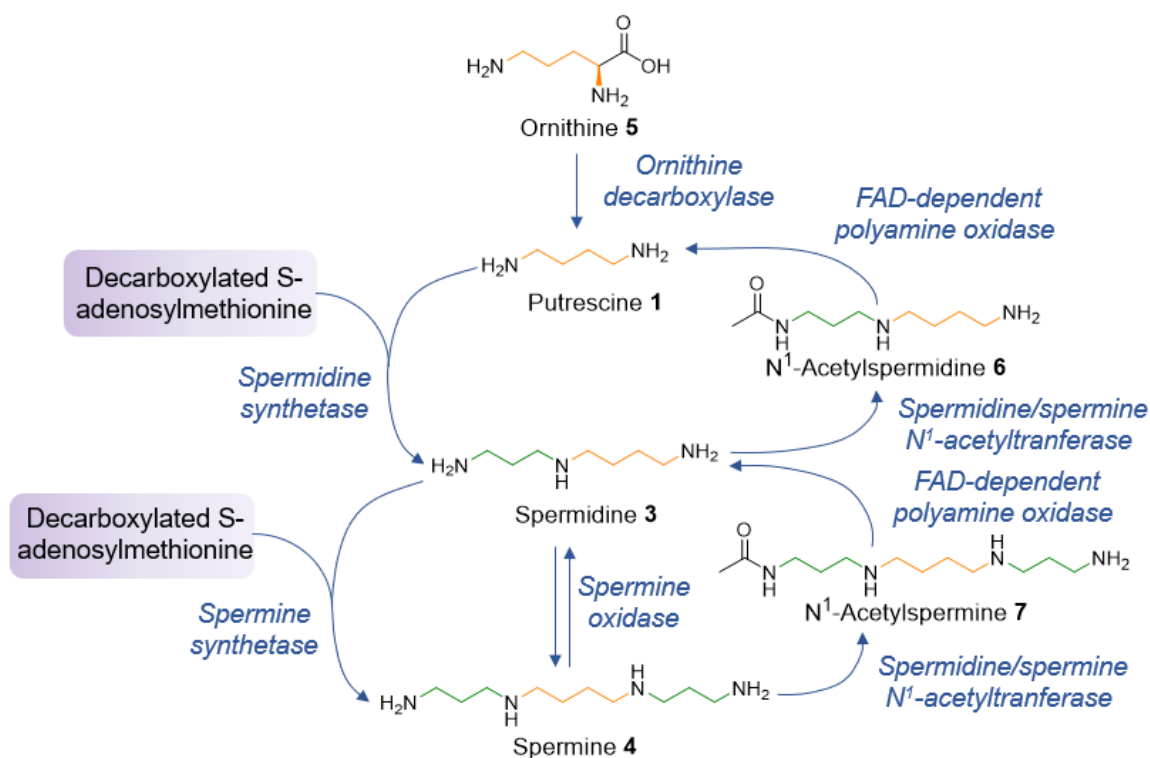


Figure 3. Biosynthesis pathway of putrescine **1**, spermidine **3** and spermine **4**.

All these polyamines **1-4** were also demonstrated possessing antioxidant and anti-apoptotic activities, involved in gut inflammatory processes, allergy as well in tumor growth. [3, 8, 9] In this latter case, the specific biosynthesis pathway involved has been targeted for the development of antineoplastic therapy as well as cancer treatments. [3, 4]

Furthermore, less common polyamines such as norspermidine **8** and norspermine **10** were identified in plants *Canavalia gladiata*, *Viciaradiata*, *V. sativa*, in water-deficit stressed *Medicago sativa* L. (alfalfa) and photosynthetic *Cyanidium caldarium* acidothermophilic alga whereas thermospermine **13**, homocaldopentamine **12** and homocaldohexaamine **15** were found in *Brasenia schreberi* osmotolerant alfalfa plant (Figure 4). [7, 8] All these polyamines have been widely studied and are involved in plant adaptation and development of stress tolerance. [8]

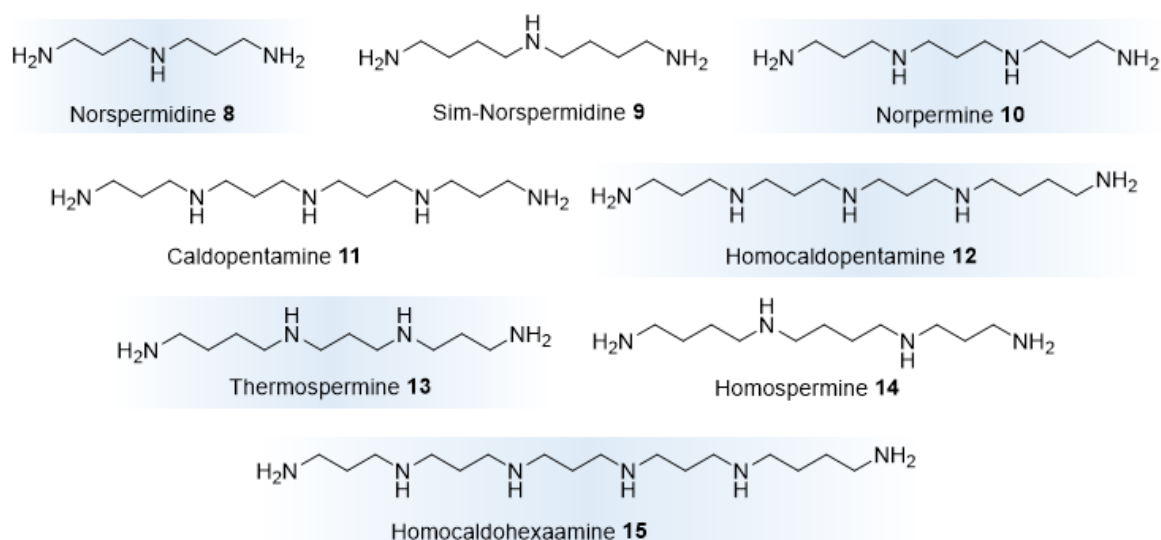


Figure 4. Chemical structures of unusual polyamines.

On the other hand, unicellular microalgae which refer to eukaryotic unicellular organisms containing chlorophyll are widely distributed in marine ecosystems. Their genomes are relatively diverse due to the complexity of their evolutionary origin which is also reflected in microalgal polyamine metabolic genes. [10] It is now well admitted that nitrogen is often the most limiting nutrient for microalgal growth in the oceans. [11] Therefore, the availability of dissolved organic nitrogen (DON) has a great impact on the primary productivity of marine ecosystems. [12] Thus, polyamines such as putrescine, spermidine, and spermine constitute one of the most important sources of DON in the oceans. [13] It is also known that local changes in polyamine concentration are usually related to algal blooms or the appearance of specific marine bacteria. Moreover, putrescine, spermidine, and spermine are typically found in most microalgae as common polyamines (Figure 4). Nevertheless, uncommon polyamines such as long-chain polyamines LCPAs have also been identified (Figure 5). [14-18]

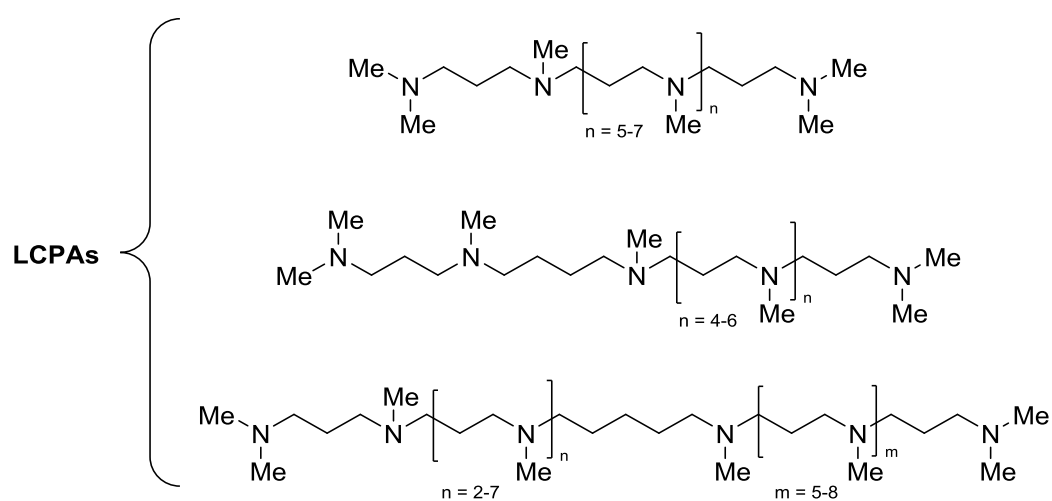


Figure 5. Molecular structure of uncommon long-chain polyamines encountered in diatoms.

In 2014, Burczyk *et al.* analyzed cell wall-conjugated polyamines in *Scenedesmus* and *Chlorella Chlorophyceae* having cell walls like higher plants, as well as similar conjugated polyamine composition including putrescine, spermidine, and spermine. [19] In contrast, the cell walls of diatoms are not made of cellulose. Their unique cell wall also contains very different polyamines compared to higher plants. Few polyamine synthetic enzymes have been well characterized especially from microalgae that produce uncommon polyamines. As encountered in most organisms, putrescine is the common precursor for the synthesis of other polyamines in microalgae. Thus, this diversity of polyamines could be derived from metabolic pathways comprising numerous polyamine synthase enzymes and hypothetically staged as indicated in Figure 6.

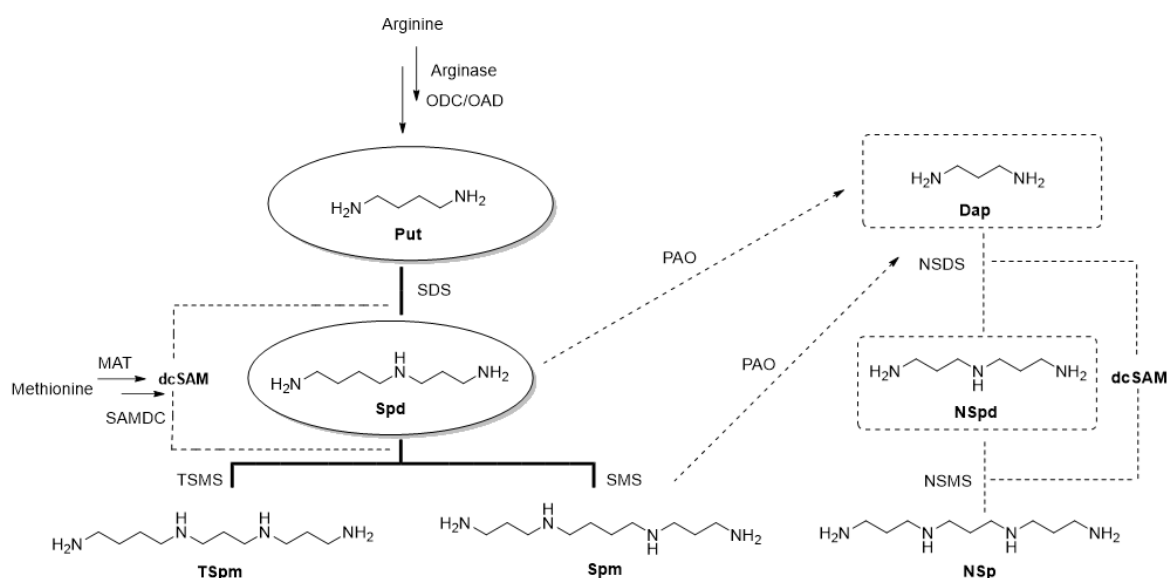


Figure 6. Postulated pathway for polyamine synthesis in microalgal cells. SAM, decarboxylated S-adenosylmethionine; Dap, diaminopropane; Put, putrescine; NSpd, nor-spermidine; Spd, spermidine; NSpm, norspermine; Spm, spermine; TSpm, thermospermine; ADC, arginine decarboxylase; MAT, methionine adenosyl transferase; NACPH, N-carbamoylputrescine amidohydrolase; NSDS, nor-spermidine synthase; NSMS, nor-spermine synthase; ODC, ornithine decarboxylase; OAD, ornithine/arginine decarboxylase; PAO, polyamine oxidase; SAMDC, S-adenosylmethionine decarboxylase; SDS, spermidine synthase; SMS, spermine synthase.

Although norspermidine is widely present in most microalgae, the corresponding synthetic pathway remains unclear and could be synthesized from Dap and aspartate-semialdehyde (ASA) as illustrated in Figure 6. Finally, numerous studies have suggested that these uncommon polyamines may have the ability to kill cancer cells and found applications in the design of silica nanomaterials for potent medical applications. [16-18, 20, 21] Thus, it is

envisioned that the microalgal enzymes involved in the synthesis of distinctive polyamines may be transferred to a target organism for mass production via a synthetic biology approach.

Nevertheless, since the discovery of natural aliphatic polyamines, plenty of other more complex natural polyamines such as toxins, iron scavengers or chemical factors were isolated from varied types of organisms as arthropods, mammals, fishes, fungi, bacteria, sponges and plant and exhibited various biological activities that will be depicted below.

III-Polyamine Toxins

III-1 Argiotoxins from spider venoms

Spider venoms are complex mixtures of peptides, proteins, and low molecular weight organic molecules such as polyamines. Thus, from venoms of orb weaver spiders *Argiope lobata* and *Argiope aurantia* were isolated two polyamine toxins namely Argiotoxin 636 (ArgTX-636, **16**) and Argiotoxin 659 (ArgTX-659, **17**) (Figure 77). [22-24]

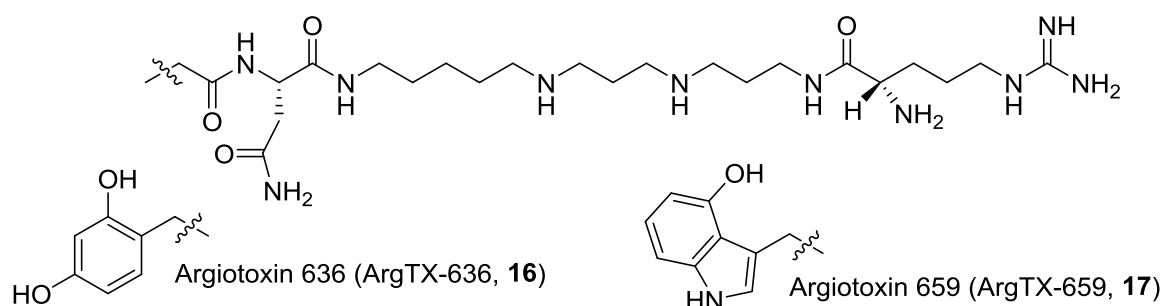


Figure 7. Structure of Argiotoxin 636 (ArgTX-636, **16**) and Argiotoxin 659 (ArgTX-659, **17**)

The synthesis of ArgTX-636 **16** was firstly described by Shih *et al.*, with 2 major building blocks and 10 steps. [25] Later, a chemical pathway achieving both ArgTx-636 **16** and ArgTX-659 **17** synthesis was depicted. [26] In 1995, Blagbrough *et al.* purposed a synthesis of ArgTX-636 **16** in 14 steps including 4 different building blocks. [27] The more recent synthetic pathway using a solid-phase synthesis afforded ArgTX-636 **16** in 2% overall yield as described in Figure 8. [23] Thus, adsorption of the mono protected diamine **18** on a resin followed by two amino acid coupling reaction and a 2-trimethylsilylethoxycarbonyl amine protection provided **20** which lead to the synthesis of **22** through a Fukuyama-Mitsunobu alkylation strategy involving a series of protection and deprotection reactions. The Ns-protected amines were subsequently deprotected and the sample cleaved from the resin with removal of the Pbf, Boc, benzyl and Trt protecting groups to afford the expected ArgTX-636 **16**. [23]

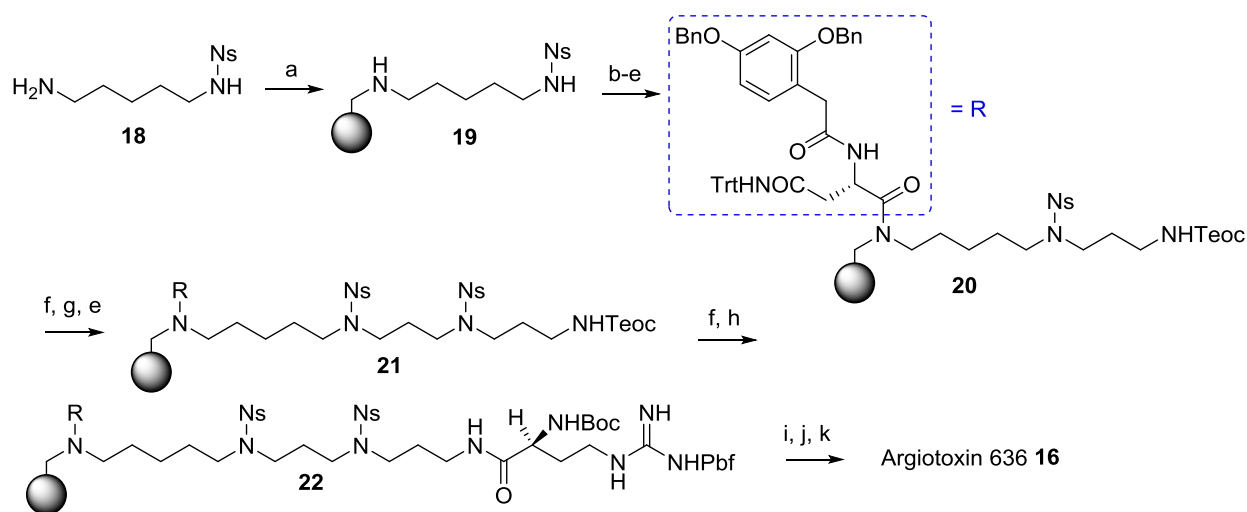


Figure 8. Solid-phase synthesis of ArgTX-636 **16**. Reagents and conditions: a) BAL PS resin (loading: 0.92 mmol/g), NaCNBH₃, DMF/AcOH (99:1), 1h ; b) Fmoc-L-Asn(Trt)-OH, HATU, collidine, DMF, 16h followed by purification/washing and a further 16h ; c) 20% piperidine in DMF, 2min followed by purification/washing and a further 20min ; d) 2-(2,4-bis(benzyloxy)phenyl)acetic acid, HATU, collidine, DMF, 2h ; e) OH(CH₂)₃NHTEoc, Bu₃P, ADDP, CH₂Cl₂/THF (1:1), 3x3h ; f) TBAF, THF, 50°C, 30min ; g) Ns-Cl, THF/CH₂Cl₂ (2:1), 3h ; h) Boc-L-Arg(Pbf)-OH, HATU, collidine, DMF, 2h ; i) DBU, β -mercaptoethanol, DMF, 3x20min ; j) TFA/CH₂Cl₂/TIS/H₂O (47.5:47.5:2.5:2.5), 2h ; k) H₂, Pd/C, MeOH, 16h.

On a biological point of view, it was demonstrated that ArgTX-636 antagonizes a specific class of glutamate receptors and exerts actions on various types of antagonism. [28] Indeed, argitoxin 636 is a potent but nonselective open-channel blocker of iGlu receptor with a selectivity for the NMDA receptor with respect to AMPA or KA receptor with an IC₅₀ of 10, 77 and 300 nM, respectively. [29-31] Thus, ArgTX-636 produces a reversible blockade of excitatory transmission with a full recovery between 2 and 4 h and it acts on muscle flaccidity in 10/20 min, with an EC₅₀ of 2 and 7 μ g for motor and sensory inhibition, respectively. [32, 33] It was enlighten that activity of ArgTX-636 is due to its longer carbon chain which may bind deeper to the ion channel region of iGlu receptors, while the secondary amino groups are important for selectivity with respect to AMPA and NMDA receptors. [28, 34] Thus, in paralysis assays ArgTX-659 **17** was demonstrated to be 4-5-fold more active than ArgTX-636 **16**, with ED₅₀ values ranging from 0.8 pmol/mg to 3.4 pmol/mg, respectively. [24] Additionally, it is noteworthy that ArgTX-636 was found as a potent melanogenesis inhibitor 500 times higher than the reference inhibitor, kojic acid. [35]

In 2017, two other polyamine spider toxins of similar structure bearing an aromatic head group were discovered. Thus, PA₃₆₆ **23** was identified in *Acanthoscuria geniculata*, *Chilobrachyus penang*, *Phlogius sp.* and *Psalmopoeus irminia* and PA₃₈₉ **24** was found in *Ceratogyrus darlingi* and *Atrax robustus* (Figure 9). [4]

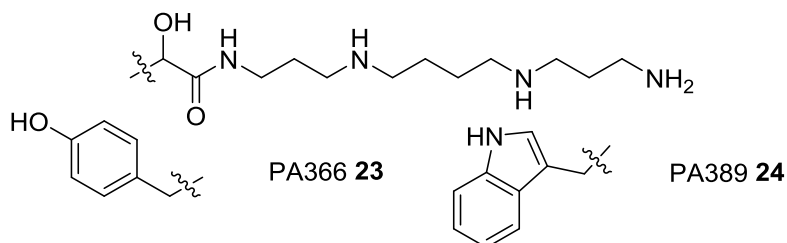


Figure 9. Structures of polyamine spider toxins PA₃₆₆ **23** and PA₃₈₉ **24**.

Both polyamines are composed with a spermine tail functionalized with an aromatic head group: 2-hydroxy-3-(4-hydroxyphenyl) propanal for PA₃₆₆ and 2-hydroxy-3-(1*H*-indol-3-yl)propanal for PA₃₈₉. On a biological point of view, PA₃₆₆ **23** and PA₃₈₉ **24** possess both an activity against MCF-7 breast cancer, SK-MEL-28 melanoma and neonatal foreskin fibroblast (NFF) primary cells, but PA₃₆₆ is generally more active than PA₃₈₉ and is selective against MCF-7 breast cancer cells. Furthermore, it was demonstrated that PA₃₈₉ only displays cytotoxicity at high concentrations contrary to PA₃₆₆ suggesting that the aromatic head group plays an important role for the encountered cytotoxicity. [4]

Some other polyamine spider toxins were collected and identified such as PA3343 **25** and PA33433 **26** in *Agelenopsis aperta*, PA3343 **25** and PA33343 **27** in *Hololena curta* as well as IndAc3334Gu **28** isolated from *Paracoelotes birulai*, with the particularity to possess a guanidino moiety (Figure 10). [36, 37]

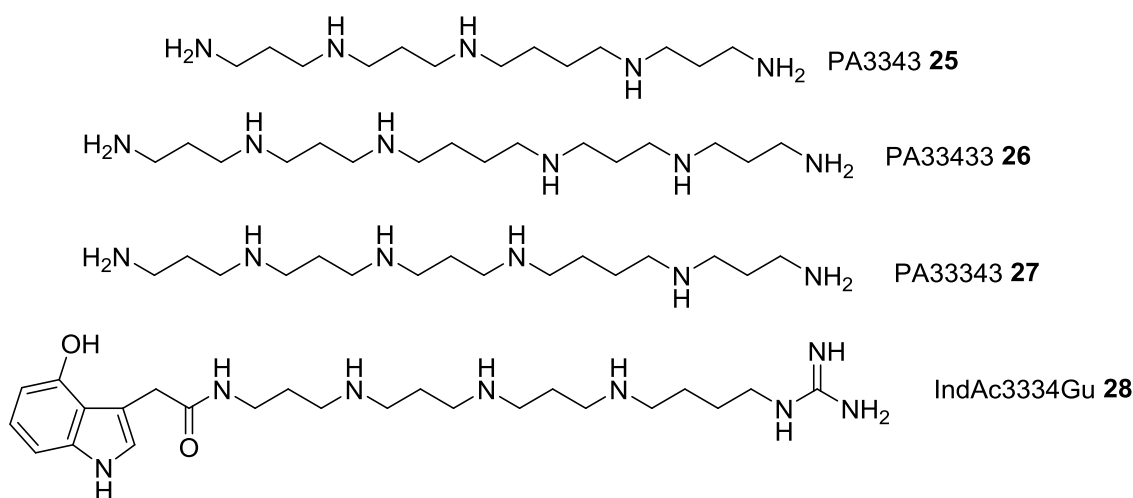


Figure 10. Structure of linear polyamine spider toxins **25-28**.

On the other hand, another class of Nephilinae toxins were identified such as NPTX-466B **29**, NPTX-643F **32** from *N. clavata*, *N. madagascariensis*, *N. cruentata* and *N. borbonica*, NPTX-489B **31** collected in all Nephilinae spiders except *N. clavata* and NPTX-473B **30** encountered in *N. cruentata* and *N. borbonica* (Figure). [38]

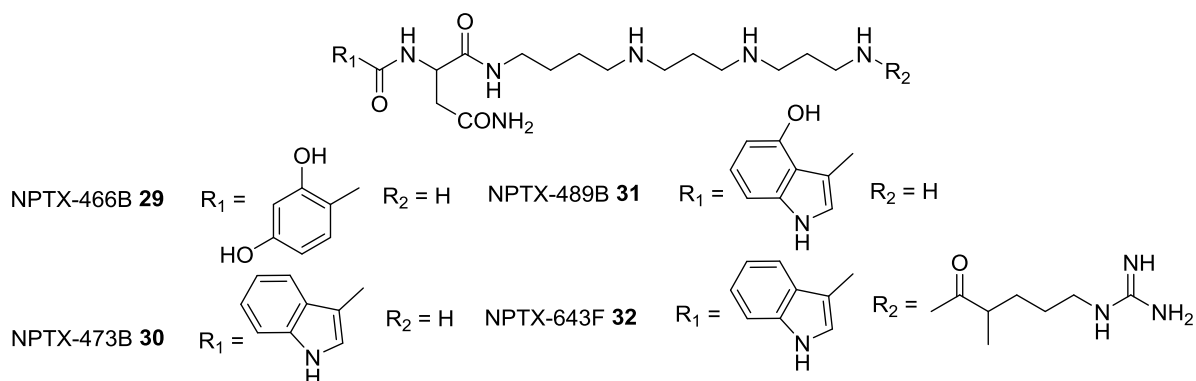


Figure 11. Chemical structures of polyamine toxins from Nephilinae spiders.

Thus, a synthetic pathway was proposed to achieve the preparation of NPTX-473B **30** in 6 steps with an overall yield of 45% and involving a 2-nitrobenzenesulfonamide (Ns) group as both protecting and activating group (Figure 5). [39] First, the intermediate **34** was obtained from the coupling of the mono-Ns-putrescine **33** with N^t-Boc-L-asparagine *p*-nitrophenyl ester. Then, the intermediate **36** was achieved from the mono-Ns-1,3-diaminopropane **35** by the Cbz protection. Finally, intermediates **34** and **36** were coupled to afford the desired polyamine backbone **37** which was subsequently deprotected and condensed with indoleacetic acid N-hydroxysuccinimide ester to provide the fully protected toxin **38** easily deprotected to lead to the formation of NPTX-473B **30**. [40]

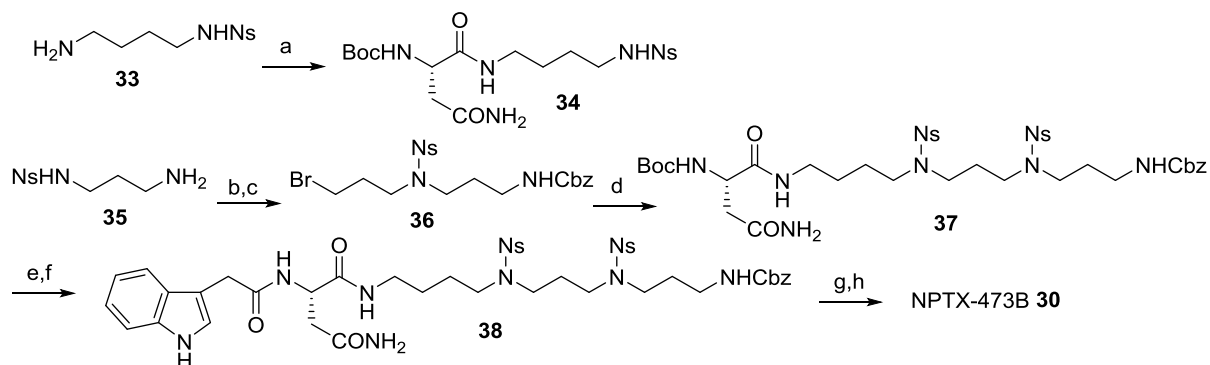


Figure 5. Synthesis of NPTX-473B **30**. Reagents and conditions: (a) N^t-Boc-L-Asn-ONp, DMF, 95%; (b) CbzCl, Et₃N/CH₂Cl₂, 99%; (c) 1,3-dibromopropane, Cs₂CO₃/DMF, 91%; (d)

34, Cs₂CO₃-TBAI/DMF, 94%; (e) TFA/CHCl₂; (f) indoleacetic acid-Osu, Et₃N/DMF, 84% (two steps); (g) 2-mercaptoethanol, DBU/DMF; (h) H₂-Pd-(OH)₂/AcOH, 60% (two steps).

As other polyamines previously isolated from venoms, Nephilinae toxins act as antagonists of iGlu receptors, but the particularity to containing an amino acid influence their biological activity and it was demonstrated that they can distinguish different subtypes of glutamate receptors. [38]

III-2 Alpha agatoxins from spider venoms

The study of *Agelenopsis aperta*, the desert grass spider belonging to the family Agelenidae in dry regions across the Southern United States and into Mexico, allows to isolate α -agatoxins: AG_{416a} **39**, AG₄₇₉ **40**, AG_{395b} **41**, AG₄₁₆ **42**, AG_{416b} **43**, AG_{379a} **44**, AG_{395c} **45**, then AG₄₈₈ **46** and AG₅₀₄ **47** which have the particularity to bear a guanidinoxy moiety (Figure). [41-43]

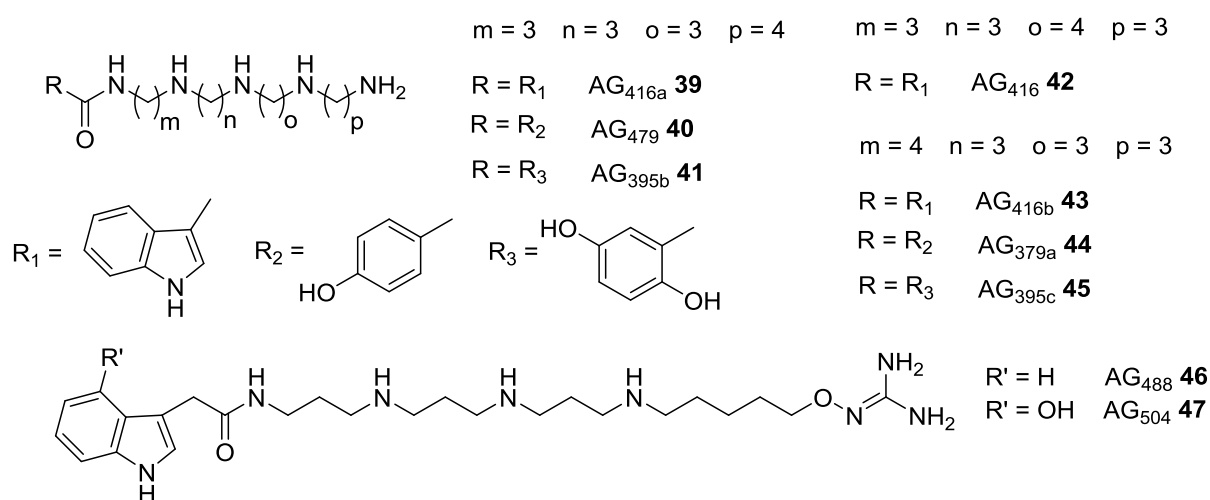


Figure 13. Structures of polyamines **39-47** isolated from the venom the *A. aperta*.

Chemical syntheses of α -agatoxins **39** to **45** were described by Manov *et al.* by using a solid-phase protocol, as described in Figure 6 for the synthesis of AG₄₁₆ **42**. [43] Firstly, Merrifield resin was converted to intermediate **48**, followed by an elongation of the polyamine backbone to afford intermediate **50**. This latter was deprotected and the acylation with TBSIndAcOH provided the intermediate **51**. Finally, the desired product was obtained by removal of **51** from the resin with ACE-Cl followed by MeOH/N₂H₄·H₂O (Figure 14). [43] It is noteworthy that Kan *et al.* described a synthesis of AG_{416b} **43** according to the Ns-

strategy.

[44]

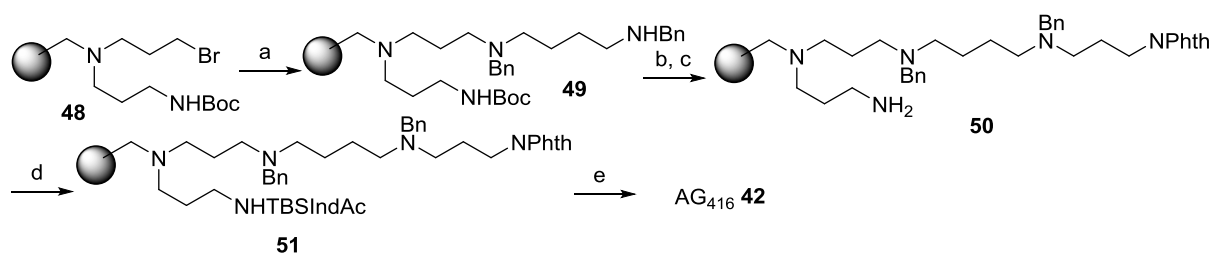


Figure 6. Synthesis of AG₄₁₆ **42**. Reagents and conditions: (a) NMP, N,N'-Dibenzylbutane-1,4-diamine, DIEA; (b) NMP, N-(3-Bromopropyl)phthalimide, DIEA; (c) TFA, CH₂Cl₂; (d) AcylOH, DIC, NMP / CH₂Cl₂, 2-{1-(*tert*-butyl)dimethylsilyl}-1H-indole-3-acetic acid; (e) ACE-Cl, (CH₂Cl₂)₂ then MeOH, N₂H₄·H₂O

On a biological point of view, α -agatoxins block glutamate-sensitive receptor channels in insects, as other polyamines isolated from spider venoms, but they possess the particularity to antagonize NMDA receptor specifically and potently. [42, 45] Thus, it was demonstrated that AG₄₁₆ **42** possesses an IC₅₀ of 36 μ M to antagonize NMDA responses in the hippocampal slice preparation and 0.52 μ M to antagonize in NMDA the cytosolic free calcium concentrations in cerebellar granule neurons. [41] Thus, paralytic action is provided by a polyamine at least the size of spermine which is acylated with an aromatic acid. [42] Furthermore, it was demonstrated that α -agatoxins can exert anticonvulsant effects. [41]

III-3 Jorotoxins from spider venoms

In 2011, the study of the venom of *Nephila clavata* allows to isolate a molecule belonging to the joro spider toxin family, namely JSTX-4 **52** (Figure). [34]

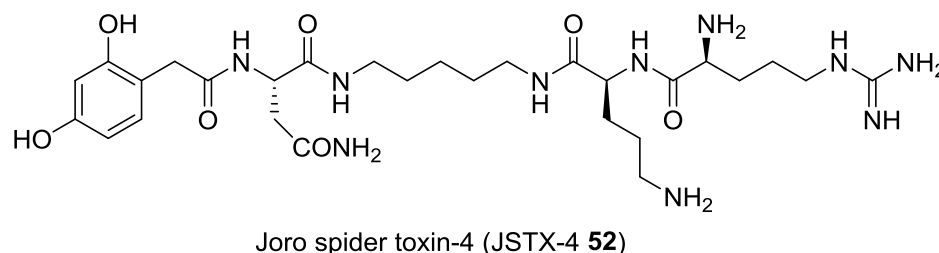


Figure 15. Chemical structure of Joro spider toxin-4 **52**.

A chemical synthesis of JSTX-4 **52** was then proposed using a solid-phase protocol closely related to that used for the synthesis of AgTX-636 **16**, in 10 steps with an overall yield of 39% (Figure 7). [34] First, 2-(trimethylsilyl)ethyl 5-aminopentylcarbamate **53** was attached by a backbone amide linker on the resin through a reductive amination, then the aromatic

headgroup was generated by subsequent couplings of protected asparagine and 2-(2,4-bis(benzyloxy)phenyl)acetic acid to provide the intermediate **54**. The N-Teoc protection group was then selectively cleaved with TBAF and the appropriately protected ornithine was coupled to afford **55** which was subsequently deprotected and coupled with the protected arginine to achieve the fully protected, resin-bound JSTX-4 **56**. Subsequent cleavage by treating resin with TFA, triisopropylsilane, water and subsequent deprotection of the aromatic O-benzyl groups with Ph(OH)₂/C and H₂ in solution afford JSTX-4 **52**. [34]

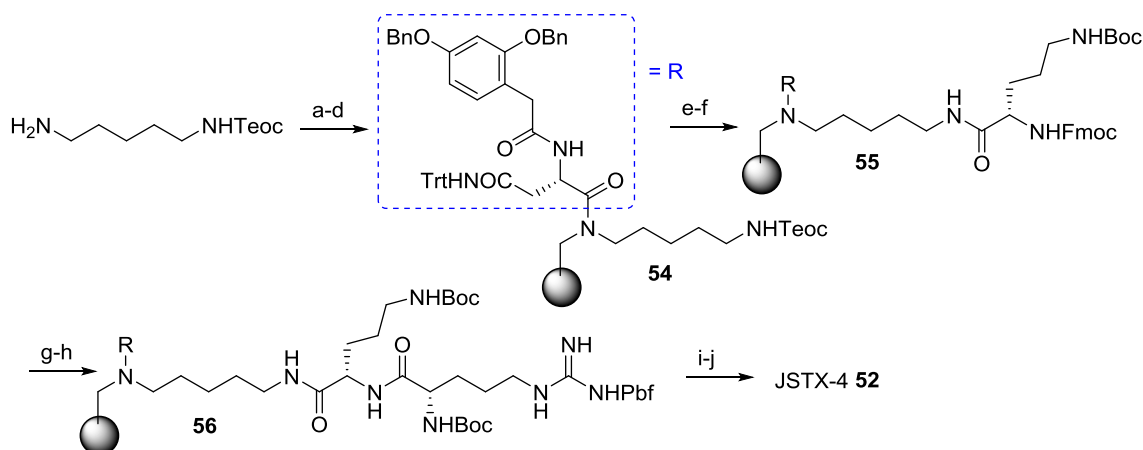


Figure 7. Synthesis of JSTX-4 **52**. Reagents and conditions: (a) (i) DFPE resin, DMF / acetic acid (9:1); (ii) NaBH(OAc)₃; (b) Fmoc-L-Asn(Trt)-OH, HATU, DIPEA, CH₂Cl₂ / DMF (9:1); (c) 20% piperidine in DMF; (d) 2-(2,4-bis(benzyloxy)phenyl)acetic acid, HATU, DIPEA, CH₂Cl₂ / DMF (9:1); (e) TBAF, THF, 55°C; (f) Fmoc-L-Orn(Boc)-OH, HATU, DIPEA, CH₂Cl₂ / DMF (9:1); (g) 20% piperidine in DMF; (h) Boc-L-Arg(Pbf)-OH, HATU, DIPEA, CH₂Cl₂ / DMF (9:1); (i) TFA / CH₂Cl₂ / TIPS / H₂O (75:20:2.5:2.5); (j) Pd(OH)₂/C, H₂, HOAC.

As other toxins isolated from spider venoms, JSTX-4 **52** is an open-channel blocker of iGlu receptors, but less effective than other toxins since a 100 μM concentration is required to inhibit 25% of both receptor subtypes. [34] On the other hand, contrary to other spider venoms toxins, the action of **52** block irreversibly the neuromuscular junction of the walking leg of lobster. [46]

III-4 Philanthotoxins from European beewolf wasp

Except for spiders, polyamine toxins were encountered in the venom of other arthropods such as *Philanthus triangulum*, commonly known as the european beewolf and which is a solitary wasp that lives in the Western Palearctic and Afrotropics. In 1990, a toxin

namely Philanthotoxin-433 (PhTX-433 **57**) and composed by a central tyrosine, a residue amide-linker, a thermospermine moiety and an n-butanoyl chain was isolated from the venom of the female (**Figure 8**. Structure of philanthotoxin-433 (PhTX-433 **57**)Figure 8). [47-50]

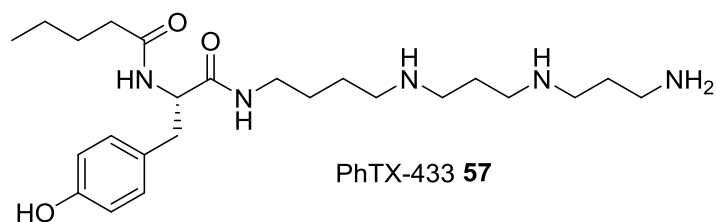


Figure 8. Structure of philanthotoxin-433 (PhTX-433 **57**).

Few chemical syntheses of PhTX-433 **57** were described and always using a multistep solution phase strategy. [51-53] In 2000, a versatile solid-phase synthesis was purposed with 77% of yield and 80% of purity (Figure 9). [54] Starting from the resin **58**, two successive amides coupling and deprotection steps afforded the β Ala- γ Abu-diamide **59**. The reduction of **59**, followed by an oxidative workup and protection with Boc provided the intermediate **61** which was subsequently coupled with Fmoc-L-Tyr(^t-Bu), treated with piperidine, and coupled with butyric acid to afford **62**. Crude toxin **57** was then finally obtained as a tris(trifluoroacetate) ammonium salt after cleavage from the resin. [54] Since this first synthesis, other solid-phase syntheses were described to successfully obtain PhTX-433 **57**. [55-57]

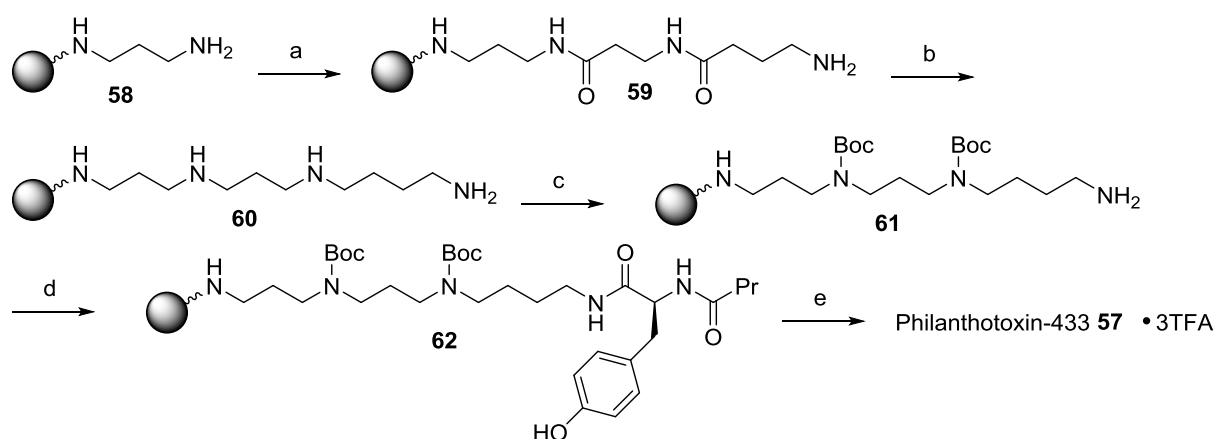


Figure 9. Synthesis of PhTX-433 **57**. Reagents and conditions: (a) (1) (i) $\text{HO}_2\text{C}(\text{CH}_2)_2\text{NHFmoc}$, HBTU, HOBT, *i*-Pr₂EtN, DMF; (ii) 20% Piperidine / DMF; (2) Repeat step (1) with $\text{HO}_2(\text{CH}_2)_3\text{NHFmoc}$; (b) (1) 1M BH_3 / THF; (2) Iodine, THF / *i*-Pr₂EtN / AcOH (7 : 1 : 2); (c) (1) 2-Ac-dimedone, DMF; (2) Boc₂O, *i*-Pr₂EtN, CH_2Cl_2 ; (3) 2% NH_2NH_2 / DMF; (d) (1) (i) HO-Tyr(^t-Bu)-Fmoc, HBTU, HOBT, *i*-Pr₂EtN, DMF; (ii) 20% Piperidine /

DMF; (2) CH₃CH₂CH₂CO₂H, HBTU, HOBt, i-Pr₂EtN, DMF; (e) TFA / H₂O / i-Pr₃SiH (95 : 2.5 : 2.5).

PhTX-433 possesses a neuromuscular activity to immediately paralyze a prey but with reversible effect. [51, 58] It was demonstrated that PhTX-433 **57** is a non-selective and uncompetitive antagonist of ionotropic receptors, such as ionotropic glutamate receptors. [47, 52, 59] Thus, the major difference with spider venoms is that PhTX-433 also antagonizes nicotinic acetylcholine receptors (nACh) (Figure 10A). [56, 60] Furthermore, PhTX-433 has been reported to lead to a selective antagonism of quisqualate, AMPA and kainate at the spinal level (Figure 10B). [61] Otherwise, wasp venom toxin can discriminate Ca²⁺ permeable from impermeable AMPA and kainate receptors subtypes. [34, 62] Moreover, it was demonstrated that PhTX-433 possesses also modulatory effects on nucleotides and proteins. [56]

A	Antagonist	nACh receptor	qGlu receptors	Neurally-evoked twitch contraction of the locust	Muscles
	IC ₅₀ (μM)	1.1	18	18	30

B	Receptors	NMDA	Quisqualate	AMPA	Kainate
	Percentage reduction (%)	3 ± 2	85 ± 4	87 ± 4	96 ± 4

Figure 10. (A) IC₅₀ values (μM) for each antagonist effects of PhTX-433. (B) Table of reduction of excitatory amino acids responses of spinal neurons.

In addition, PhTX-433 possesses an activity against bacteria, by inhibiting the cation-selective porins OmpF channels of *E. coli* by multiples binding sites with polyamines with a half-maximum inhibition for a 200 nM concentration. [63]

Among all the philanthotoxins PhTX-433 is not the most active and some derivatives are now under current investigation to improve their activities for the development of a potent therapeutic use. [47-50, 57-59, 61, 64-69]

IV-Polyamines from the sea

IV-1 Polyamines from sharks

Polyamines are also present in other animals than arthropods, with different chemical structures and biological activities. Thus, sharks which are predatory scavengers present remarkable resistance to bacterial and viral infections. The research of host defense agents from the dogfish shark *Squalus acanthias* and later from the bacterially challenged sea lamprey *Petromyzon marinus*, resulted in the discovery, isolation and characterization of a family of novel aminosterol derivatives (Figure 11). [70, 71] These compounds possess an invariant steroid skeleton with a *trans* AB ring junction, a polyamine attached equatorially at C-3 and a cholestane-related side chain functionalized. Squalamine **70** is the most abundant aminosterol found in the dogfish shark liver (10-20 $\mu\text{g/g}$) whereas seven other compounds (**63-69**) are present in smaller amounts. [70]

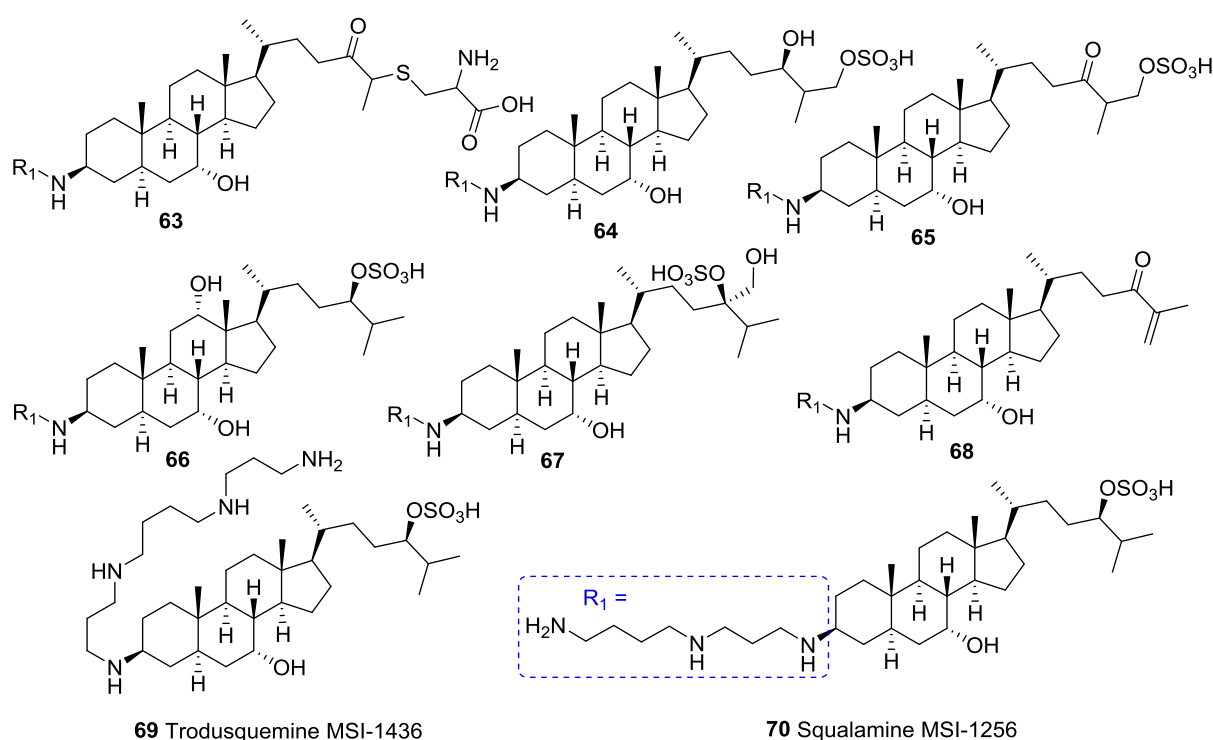


Figure 11. Structure of the different aminosterols **63-70** isolated from the dogfish shark liver.

IV-1-1 Squalamine

The most studied aminosterol is amphiphilic cationic steroid squalamine (Sq) **70**, also called MSI-1256, provided by the white blood cells and possessing a hydrophilic head (polyamine) and a hydrophobic tail (Figure 12). [71-74]

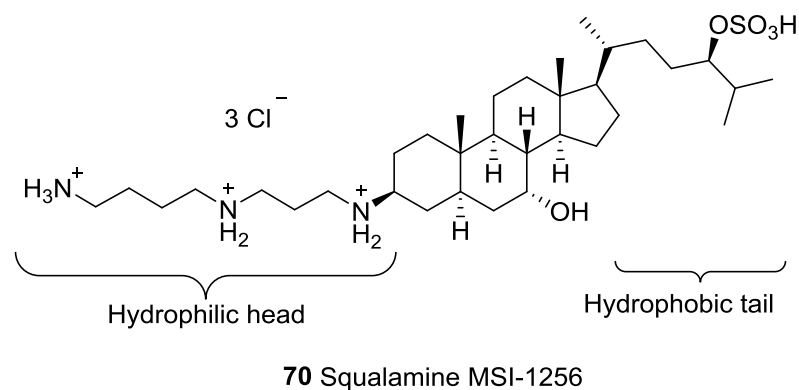


Figure 12. Chemical structure of squalamine **70**.

During the last decades, numerous chemical syntheses were developed to achieve the preparation of squalamine **70** in suitable amounts (Figure 13). Both first syntheses achieved a mixture of diastereomers in C-24 position, in 17 steps. [75, 76]. In 1995, Moriarty *et al.* described the first stereoselective synthesis in 20 steps from stigmasterol **73**. [77] The same starting material was also used by Zhang *et al.* who improved the synthesis in 15 steps [78], with a key intermediate 7α -(benzoyloxy)-3-dioxolane cholestan-24(*R*)-ol prepared in 10 steps. [79] Another strategy based on a biotransformation, allows the researchers to reduce the synthesis to 11 steps. [80] In 2000, a synthetic pathway in 14 steps (19% of overall yield) was developed from methyl-3-keto-5 α -chenodeoxycholanate **75** [81] and this synthesis was subsequently improved in 9 steps with an overall yield of 31% and 99% of diastereoselectivity. [82] It is noteworthy that at the same time, a 13 steps synthesis from desmosterol **76** was reported with a 7% overall yield. [83] Zhang and Zhou improved consecutively their previous work with a 9 steps synthesis and an overall yield of 14%, starting from the easily available methyl chenodeoxycholanate **77**. [84, 85] Finally, in 2019 Zhou *et al.* reported a synthesis of a key intermediate of squalamine 7α , 24*R*-dihydroxy-5 α -cholestan-3-one starting from Δ^5 -3 β -hydroxycholanate **78**, involving a one-pot Grignard reaction key step. [86]

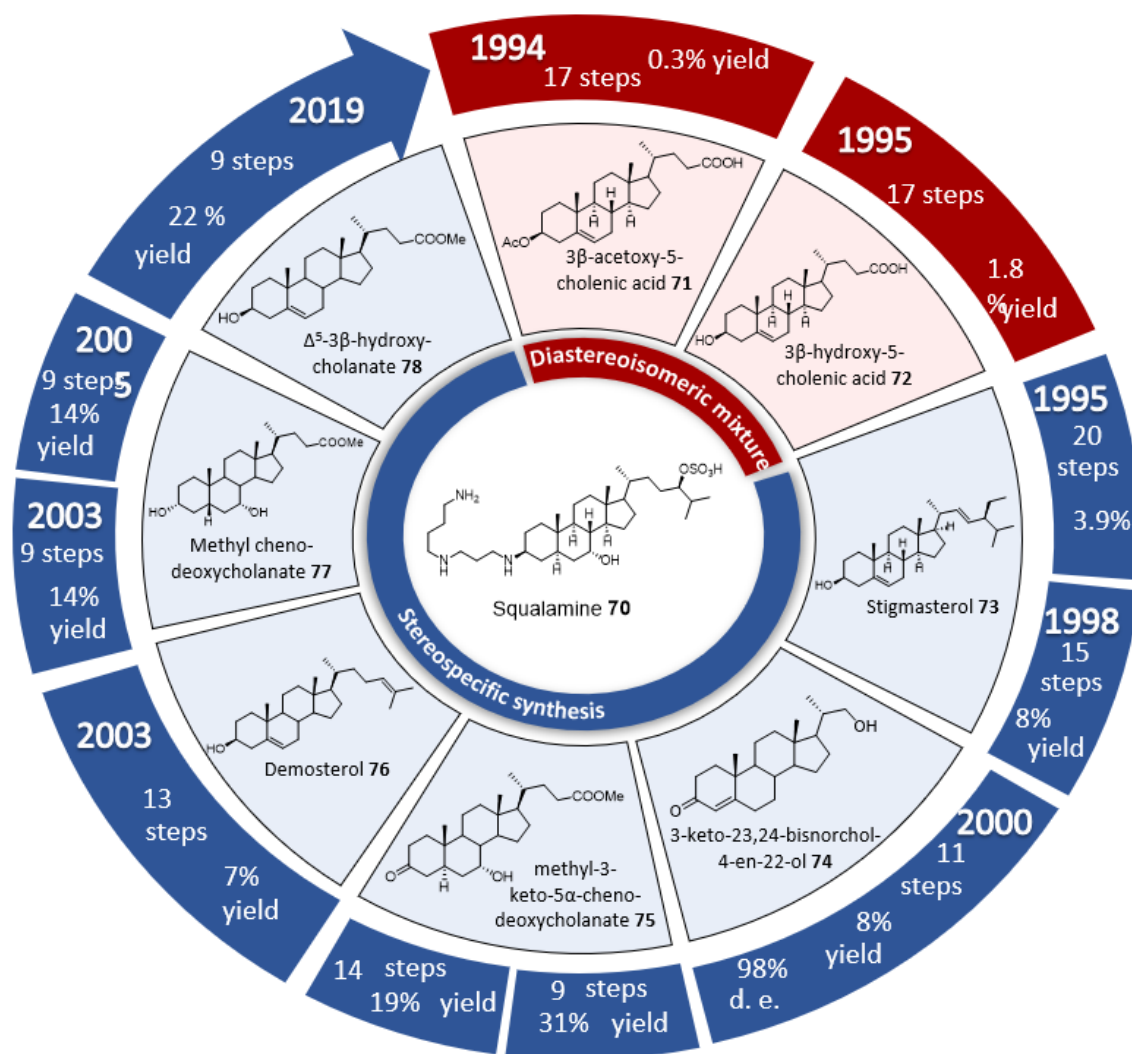


Figure 13. Different synthetic strategies reported for the preparation of squalamine **70**.

In this latter case, the hydroxyl function at the position C-3 was protected with THP, then a reduction of the methyl ester in position C-23 afforded the expected alcohol **79** which was protected with a benzyl moiety. The oxidation of the hydroxyl group in position C-7 lead to the intermediate **80** which was subsequently reduced with dihydrogen and palladium. The deprotection of the hydroxyl function in position C-3 as well as the reduction of the ketone in position C-7 afforded **82**. Selective oxidation at C-3 and deprotection of the hydroxyl group in position C-24 lead to the key intermediate **83** which can be converted into squalamine according well-known procedures (Figure 14). [86]

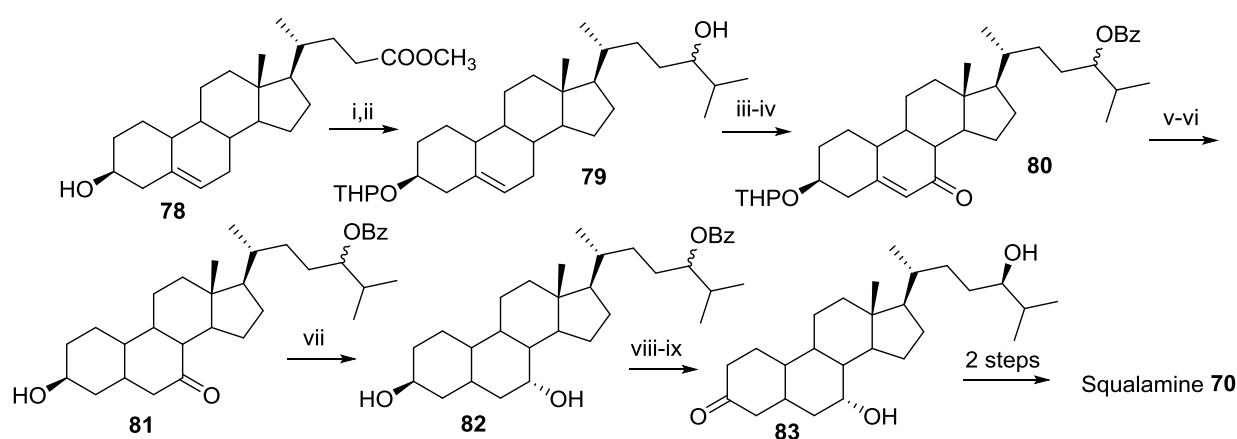


Figure 14. Synthesis of squalamine performed by Zhou *et al.* in 2019. Reagents and conditions: (i) DHP, PPTS, DCM; (ii) *i*-PrMgBr, LiBH₄; (iii) BzCl; (iv) PDC, TBHP; (v) H₂, Pd/C; (vi) HCl; (vii) L-selectride; (viii) AgCO₃/Silica; (ix) KOH, MeOH

IV-1-1-1 Antimicrobial activities of squalamine

It has been demonstrated that squalamine **70** possesses excellent antimicrobial activities against a large panel of both Gram-positive and Gram-negative bacteria, with Minimum Inhibitory Concentrations (MICs) varying from 1 to 10 mg/L and 1 to 64 mg/L, respectively (Table 1).

Gram-negative bacteria	MICs (mg/L)			Gram-positive bacteria	MICs (mg/L)
	Reference strains	Clinical isolates			
		Non mucoid	Mucoid		
<i>Escherichia coli</i>	1-4 [72, 87, 88]	8 [89]	N.D.	<i>Staphylococcus aureus</i>	1-2 [72, 87, 88]
<i>Pseudomonas aeruginosa</i>	4-16 [72, 87, 88]	2-8 [89]	16-32 [89]	<i>Enterococcus hirae</i>	10 [90]
<i>Enterobacter aerogenes</i>	32 [87]	N.D.	32 [89]	<i>Enterococcus faecalis</i>	1-2 [74]
<i>Acinetobacter baumannii</i>	8 [87]	8 [89]	N.D.	Fungi species	MICs (mg/L)
<i>Klebsiella pneumoniae</i>	8 [87]	16 [89]	N.D.	<i>Trichophyton</i>	8-16 [91]
<i>Burkholderia cepacia</i>	16-64 [87]	N.D.	16-64 [89]	<i>Candida</i>	4-16 [72, 87, 88, 90, 92]
<i>Proteus vulgaris</i>	4-8 [72]	N.D.	N.D.	<i>Aspergillus</i>	8-16 [88, 92]

Table 1. MICs values of Sq in mg/L against Gram-positive and Gram-negative bacteria (from reference and clinical strains as well as from mucoid and non-mucoid clinical isolates) and against fungi. N.D.: not determined.

A time-kill study performed at the MIC demonstrated that squalamine **70** can kill, with a direct bactericidal effect, *S. aureus* and *P. aeruginosa* in 1 and 2h, respectively (Figure 15). [93]

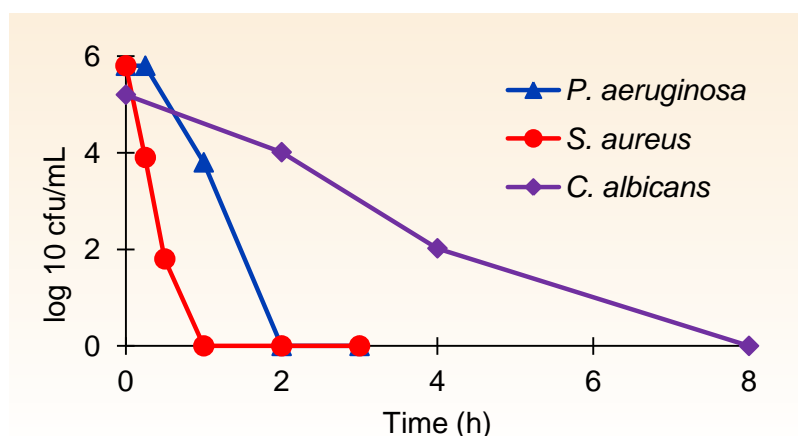


Figure 15. Time-kill curves of squalamine at the MIC against *P. aeruginosa* ATCC 27853, *S. aureus* ATCC 25923 and *Candida albicans* ATCC 90028.

On a mechanistic point of view, it has been well established that squalamine induces the leakage of polar fluorescent dyes from large unilamellar phospholipid vesicles, but do not present any ionophoric activity. [94, 95] The addition of divalent cations (Mg^{2+} or Ca^{2+}) increased the MIC of squalamine against *P. aeruginosa* by 8-fold (

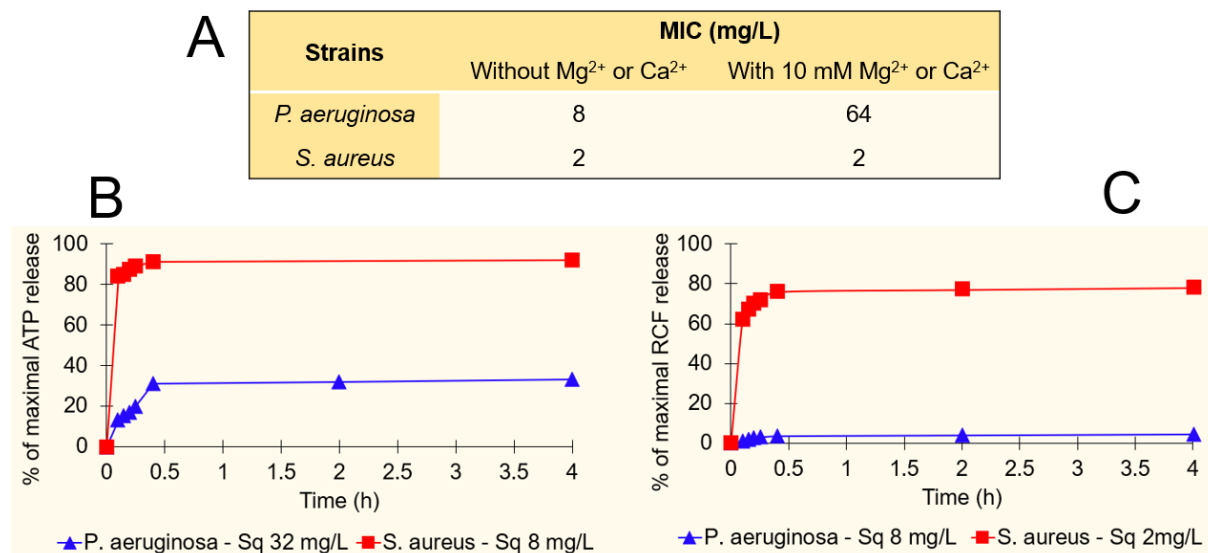


Figure 16A), suggesting that positively charged amine groups interact with negatively charged phosphate groups of lipopolysaccharides and displace divalent cations. [93] Nevertheless, this observation is only valid for Gram-negative bacteria. Furthermore, a time-dependent ATP release was observed for *P. aeruginosa* and a rapid ATP release was induced on *S. aureus*, indicating that membrane lesions appear rapidly (

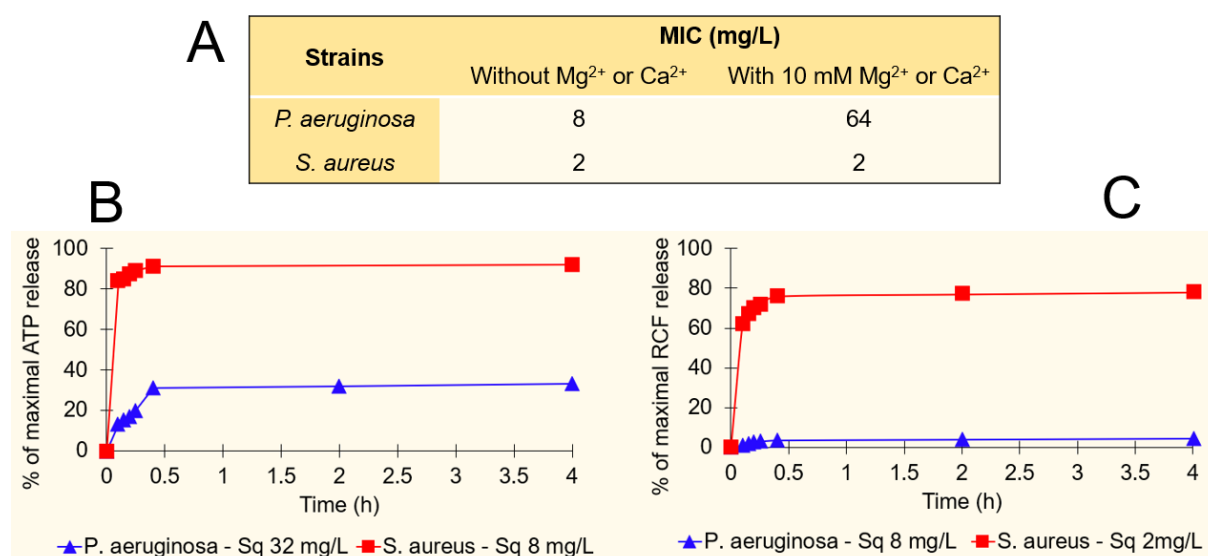


Figure 16B). [93] Finally, a strong depolarization of the Gram-positive bacterial membrane was observed with a rapid and important increased of the RCF values, whereas no depolarizing effect was observed for Gram-negative bacteria (

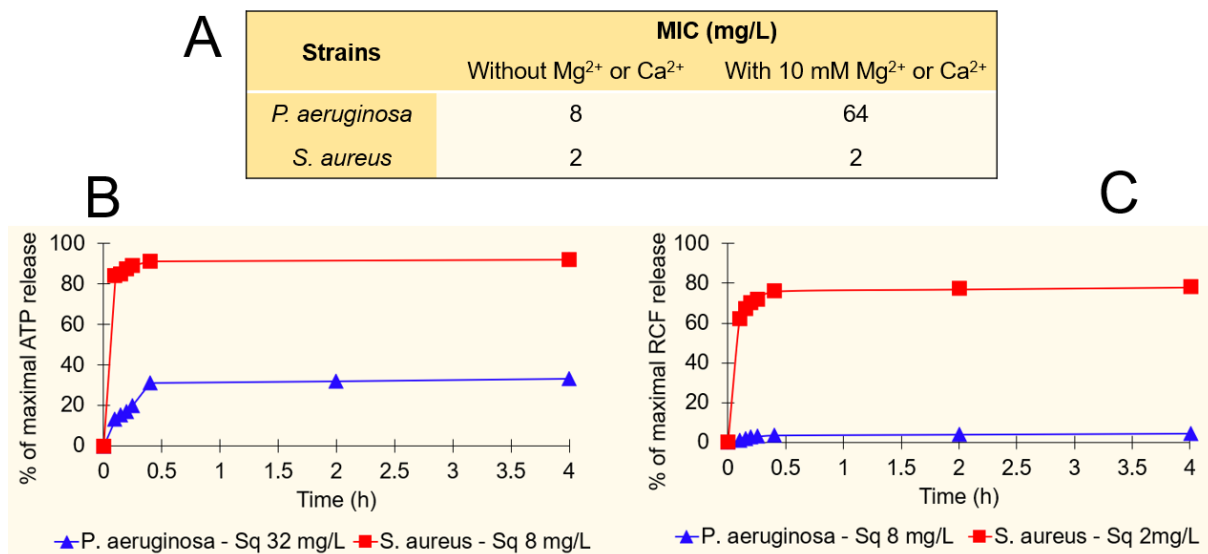


Figure 16C). [93]

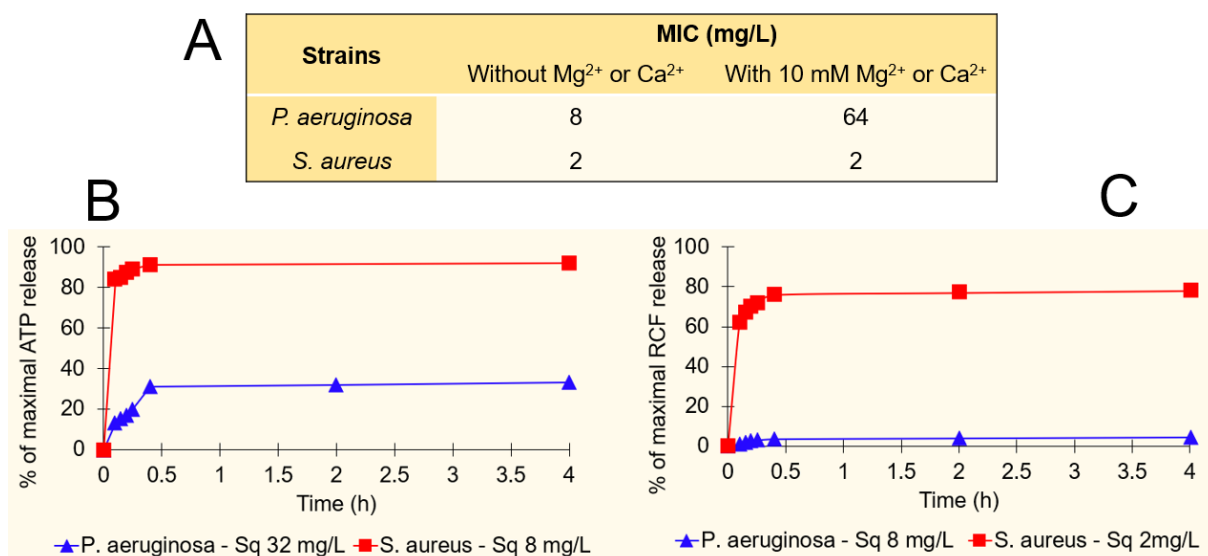


Figure 16. (A) Antimicrobial activity of Sq in the presence or absence of divalent cation salt solution, (B) ATP release kinetics and (C) depolarization of the bacterial membranes of Gram-positive and Gram-negative bacteria.

Based on all these considerations, a mechanism of action of squalamine has been proposed both on Gram-positive and Gram-negative bacteria (Figure 17). [87, 96] On the one hand, treatment of *S. aureus* with squalamine leads to a dramatic disruption of the bacterial membrane, causing a drainage of the cytoplasmic material. [93] On the other hand, squalamine acts as a detergent against Gram-negative bacteria, resulting in bleb-like projections that originated from the outer membrane. [93]

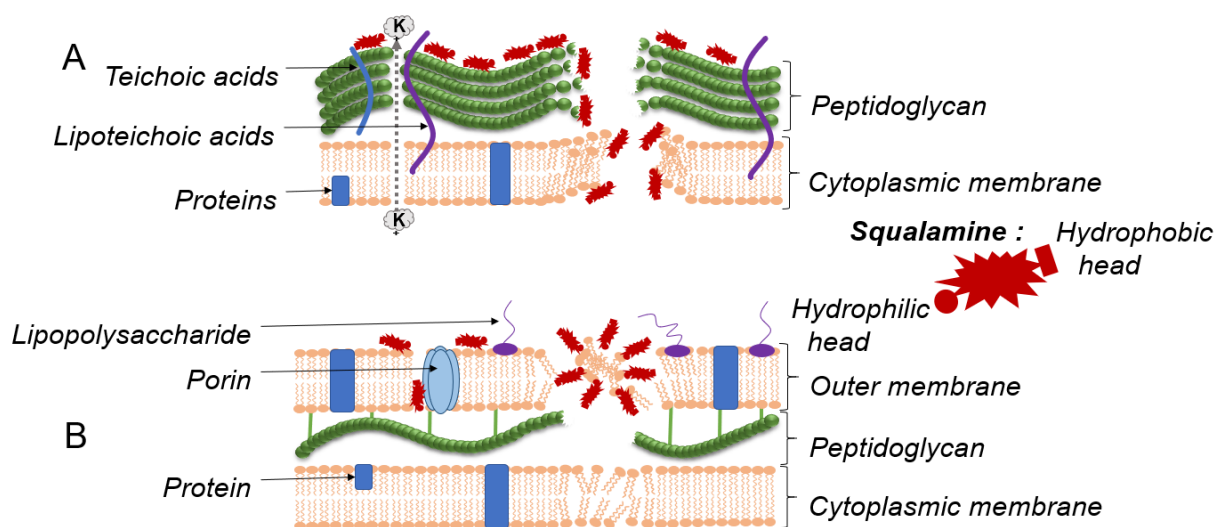


Figure 17. Mechanism of action of squalamine against (A) Gram-positive bacteria and (B) Gram-negative bacteria.

It is noteworthy that squalamine is an efficient host defense against bacteria, but some modifications can occur to protect bacteria. [97] Thus, by studying *E. coli* it was demonstrated that O4 specific antigen confers a protection against squalamine *in vitro* and potentiates urinary tract infection in a mouse model *in vivo*. In contrast the presence of the K54 antigen enhanced the antibacterial activity of squalamine. [97] It was also observed that, resistance mechanism of the bacteria becomes ineffective in presence of squalamine, because of its detergent-like mechanism of action which reduces the possibility of resistance emerging. [72, 90, 93] This was confirmed by attempts to isolate squalamine-resistant *S. aureus* strains, which failed. [98] Otherwise, squalamine was investigated against bacteria isolated from cystic fibrosis patients area (Table 1), leading of moderate to excellent MICs values varying from 2-8 mg/L to 16-32 mg/L against non-mucoid or mucoid isolates, respectively. [89]

Additionally, squalamine was found to be able to kill dormant cells of *A. baumannii* without induced persisters nor viable. [99] Nevertheless, squalamine was determined to be ineffective against *Mycobacterium tuberculosis* certainly due to peculiar lipid composition of the membrane of this class of bacteria. [100, 101] On the other hand, squalamine possesses antibacterial activity against human methanogenic archaea, with MICs ≤ 1 mg/L and electron microscopy observations suggested that squalamine breaks the *Methanobrevibacter smithii* cell wall, inducing cytoplasm leakage and cell death, by a mechanism of action similar to that observed against Gram-negative bacteria. [102-104]

Otherwise, squalamine was found to be used as a chemo sensitizing agent to enhance the activity of antibiotics against Gram-negative susceptible or resistant bacteria (Table 2). [105, 106] Thus, squalamine increases activity of chloramphenicol and ciprofloxacin against *E. coli*, *P. aeruginosa* and *K. pneumoniae* as well as *E. aerogenes*, unlike MICs of erythromycin which are not significantly improved. [105]

Bacteria strains		MICs (mg/L)						
		Sq 70	Chloramphenicol		Ciprofloxacin		Erythromycin	
			without Sq	with Sq	without Sq	with Sq	without Sq	with Sq
<i>E. coli</i>	AG100	2	8	0.5 ^a	0.25	0.125 ^a	512	256 ^a
	AG100A	1	2	1 ^b	0.06	0.06 ^b	128	64 ^b
	AG100At et	2	16	2 ^a	1	0.03 ^a	256	256 ^a
<i>P. aeruginosa</i>	PA01	16	512	16 ^c	16	<1 ^c	512	256 ^c
	PA124	16	256	64 ^c	32	16 ^c	1024	256 ^c
<i>K. pneumoniae</i>	KP63	N.D.	256	64 ^d	16	<1 ^d	256	64 ^d
	KP55	N.D.	32	<1 ^c	32	<1 ^c	256	64 ^c
<i>E. aerogenes</i>	ATCC	N.D.	4	0.25 ^d	0.5	0.015 ^d	512	128 ^d
	13048	N.D.	4	0.25 ^d	0.5	0.015 ^d	512	128 ^d
	CM-64	N.D.	256	16 ^d	0.5	0.03 ^d	512	256 ^d

Table 2. MICs values of antibiotics in presence or absence of Sq and MICs of Sq alone against various Gram-negative bacteria. Sq concentrations: ^a 0.4 mg/L. ^b 0.2 mg/L. ^c 3.2 mg/L. ^d 1.6 mg/L. N.D.: not determined.

As previously mentioned in Table 1, squalamine possesses a remarkable activity against a large panel of fungi with MICs ranging from 4 to 16 mg/L. [91, 107] A time-kill study performed at the MIC against *C. albicans*, demonstrated a fungicidal effect observed after 8 h (Figure 15). [107]

Considering its antimicrobial activities against a broad spectrum of bacteria and fungi, squalamine was studied for numerous applications either in disinfection or in topic local use. First, squalamine was used as disinfecting agent for home nebulizer of cystic fibrosis patients, in form of a soluble tablet and disinfection can be achieved against both Gram-positive and Gram-negative bacteria and also fungi *C. albicans* in 20 min, whereas 6 h are needed for the fungi *A. niger*. [108] In a second application, squalamine delivered by aerosolization was

demonstrated to reduce, in a *P. aeruginosa* animal infected model, the lung bacterial load and bronchopneumonia-induced histological lesions with an efficiency comparable to that encountered for colistin. [109] Finally, a squalamine 1% ointment applied in a *S. aureus* skin decolonization model lead to a reduction of viable bacteria by 4 log in 1 h with a single application whereas a reduction 1.3 log for the standard mupirocin drug was observed. [98, 110] This strategy was also envisioned, with encouraging results, in the evaluation of the ability of squalamine to act as a potent antidermatophyte drug against *Miscrosporium audouinii*. [111]

It is noteworthy that, these promising antimicrobial results lead to the development of efficient synthetic analogs which are not depicted here. [87, 88, 107, 112, 113]

IV-1-1-2 Antiangiogenic activities of squalamine

Cancer is the second most killing disease in the world. In this context, squalamine was demonstrated to be able to inhibit growth of few cancers thanks to its antiangiogenic activity. [114, 115] This effect is mediated by blocking mitogen-induced proliferation and migration of endothelial cells, thus preventing neovascularization of the tumor. [116] Nevertheless, it is noteworthy that squalamine is not directly cytotoxic for tumor cells. [116] This inhibitory effect is time dependent, with no effect after immediate addition and maximum effect with 1 h exposure and fully reversible. [117, 118] Two anti-angiogenic mechanisms were postulated: First, squalamine could selectively inhibit the sodium-hydrogen (Na^+/H^+) antiporter exchange isoform NHE3, which hinders hydrogen efflux out of the endothelial cell, and consequently blocked intracellular alkalization and cellular proliferation (Figure 18A). [117] Moreover, squalamine may alter cellular responses to growth stimuli that increase intracellular calcium by binding to NHE3 and redistributing calmodulin. [119] Secondly, squalamine could inhibit the release of the vascular endothelial growth factor (VEGF) by the tumor cell, which hinders binding and activation of VEGF receptor of the endothelial cell (Figure 18B). [120]

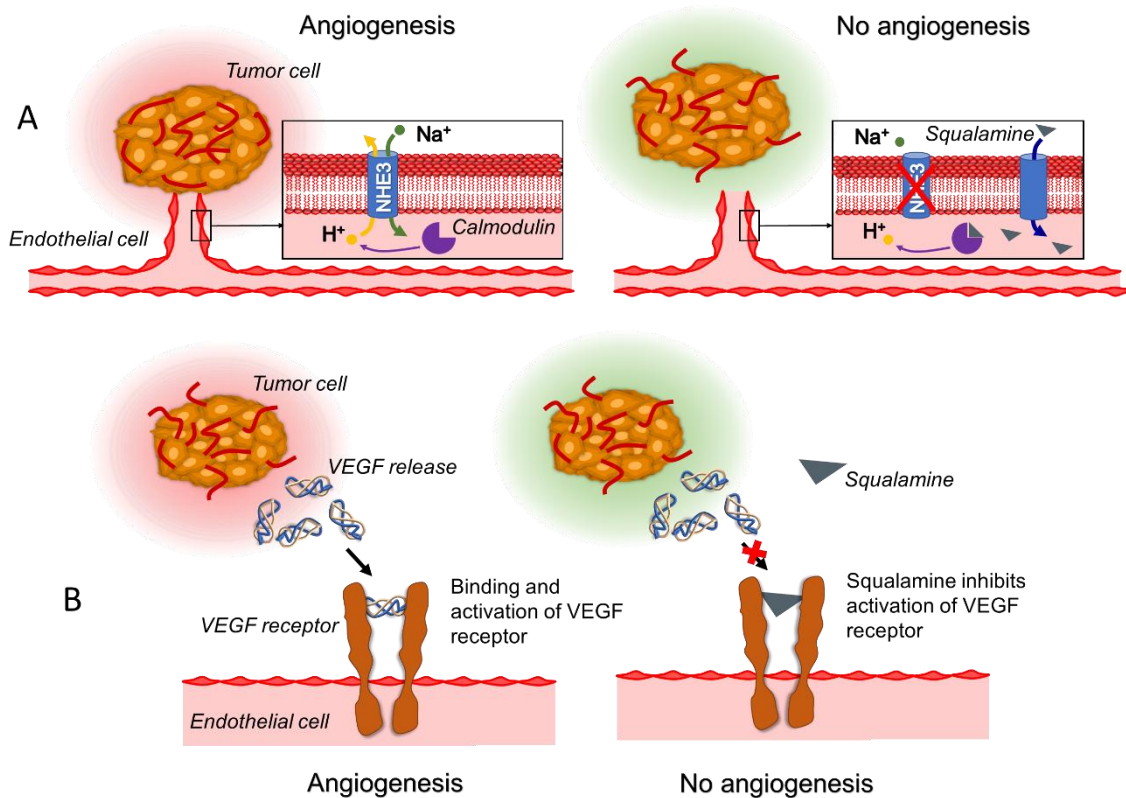


Figure 18. Potent mechanisms of anti-angiogenic activity of squalamine by inhibiting (A) NHE3 protein and (B) VEGF receptor.

On a clinical point of view, the use of squalamine in combination with chemotherapy for the treatment of lung cancer patients was performed. [121] It appears that squalamine alone does not inhibit the tumor growth, whereas its combination with cisplatin or carboplatin is efficient against tumor cells growth and better than the platinum analogues taken alone (Table 3). [121] In this case, squalamine stops new blood vessels growth by disrupting endothelial cell architecture and the integrity of endothelial cell-cell attachment in tumors and consequently prevents further tumor growth. [115]

Tumor	Sq	Tumor growth delays in days			
		Cisplatin		Carboplatin	
		without Sq	with Sq	without Sq	with Sq
H460	< 0.2	2.5	> 5	2.9	> 6.7
Calu-6	< 0.8	5	> 10.7	2.5	> 6.3
NL20T-A	< 0.1	12	> 29	N.D.	N.D.
Lewis lung	< 0.02	2.1	3.6	3.1	5.7

Table 3. Treatment of different tumors with cisplatin and carboplatin in the presence or absence of squalamine. N.D.: not determined

It was then demonstrated that squalamine possesses an activity against prostate and breast cancer as well as neuroblastoma and lung chemoresistant tumors, alone or in combination with chemotherapy. [115, 122, 123] Furthermore, a study *in ovo* enlightens that squalamine has no obvious local or systemic deleterious consequences for vitality and development of the embryo. [124] Therefore, we must mention that several clinical trials were performed by using squalamine alone or in combination with chemotherapy for the treatment of various of cancers. [115, 116, 119, 120, 125-128] It is noteworthy that squalamine is well-tolerated by humans, thereby the US Food and Drug Administration has designated squalamine as an ‘orphan drug’ for development as a new antitumor agent to treat ovarian cancer, even if studies remains necessary concerning the mechanism of action induced. [127]

Age-related macular degeneration (AMD) is the leading cause of vision loss in the industrialized world. [129] In this area, it was demonstrated that squalamine blocks angiogenesis in a noncancerous model, by reducing retinal neovascularization. [130] Indeed, squalamine can inhibit growth factor-mediated endothelial cell proliferation and migration, including that stimulated by VEGF (Figure 18B), at concentrations that are not toxic for endothelial cells *in vitro* and without affecting normal vascular development. [131] Moreover, an assay on monkey demonstrated that systemic administration could prevent and reduce iris neovascularization. [132] The formulation of squalamine was realized as lactate salt, then tested by systematically administration on rat and partially reduced choroidal neovascular membrane development. [133] Squalamine was then tested in phase I/II clinical trials for the treatment of age-related macular degeneration, with a gain of ≥ 3 lines in 26% of case whereas the visual acuity remained stable for 74% of the patients. [129, 130, 134, 135] With the advantage to be administered intravenously, squalamine eliminated the risk of eye infection or injury. [72] The phase II clinical study was made with more than 90% of patients have stabilized or improved vision at 4 months follow up. [136, 137] The squalamine lactate was investigated by Ohr Pharmaceutical under the name of OHR-102, with a phase III trials approved by the FDA. [138] Afterwards, a phase III squalamine/ranibizumab eyes drops assay was performed but was unsuccessful. [139-141]

IV-1-1-3 Antiviral activities of squalamine

Viral infections are one of the major causes of disease worldwide. Thus, according to the WHO in one year 390 millions of person were infected by dengue virus and hepatitis B. [142, 143] In this context, squalamine was demonstrated to be active *in vitro* and *in vivo* against a broad spectrum of human viral pathogens, including both RNA- and DNA-

enveloped viruses. [144] Thus, squalamine possesses an activity against dengue virus at a concentration of 100 mg/L allowing a complete inhibition of the virus, and an activity against hepatitis B virus, leading to an 84% inhibition at a concentration of 20 mg/L (Figure 19). [144]

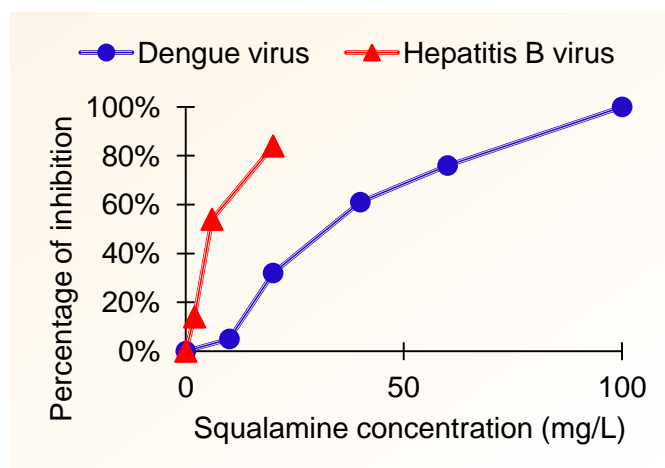


Figure 19. Effect of squalamine against dengue virus infection of human endothelial cells and hepatitis B virus infection of primary human hepatocytes.

The suggested mechanism involves the capacity of squalamine to neutralize the negative electrostatic surface charges of intracellular membranes and to bring the cell less effective in supporting viral replication, which will be detrimental to the emergence of viral resistance. [144]

IV-1-1-4 Antineurodegenerative activities of squalamine

Most common neurodegenerative disorders are Parkinson and Alzheimer, one characterized by motor dysfunction and the other is the first cause of dementia with one new patients affected every 3 seconds. [145, 146] Thus, in a preliminary study it has been demonstrated that squalamine **70** possesses a specific effect against α -synuclein aggregation by acting on a competitive binding model, where squalamine and toxic α -synuclein oligomers compete for binding sites at the surface of vesicles and neurons (Figure 20). [147-149]

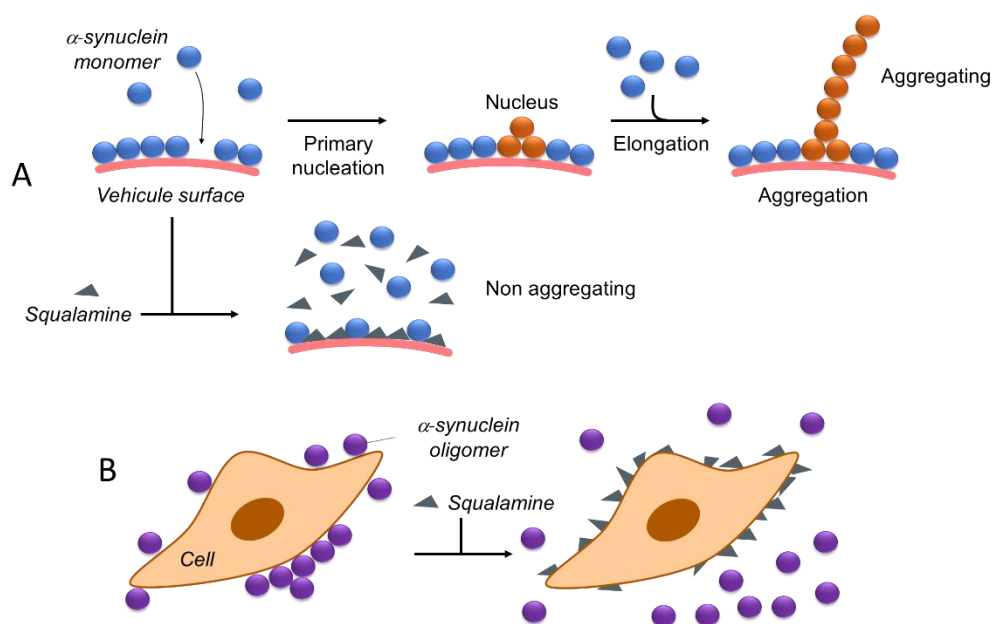


Figure 20. Effects of squalamine on a-synuclein aggregation. (A) Inhibition of the initial events of α -synuclein aggregation. (B) Protection from toxic a-synuclein oligomers in cells via competitive binding.

IV-1-1-5 Stability and cytotoxicity of squalamine

It was demonstrated that squalamine induces blood hemolysis at 320 μ M, which is a higher concentration than those requested to observe biological activities. [107] Squalamine was found to be non-genotoxic until at a concentration of 160 μ M (solubility limit), a proliferative index upper than 80% was observed. [150] Furthermore, during clinical trial a hepatotoxicity was observed, but in reversible way after discontinuation of the treatment. [130] To determine all the toxicological and pharmacokinetic parameters, a bioanalytical method for quantitative determination of squalamine in patient, as well as in animal plasma was developed. [151] Then, a study of stability of squalamine lactate in different conditions was applied, to determined eventual degradation products, and demonstrated that squalamine lactate is stable in refrigerated conditions as well as in basic solution. [152] Furthermore, the half-life of squalamine is approximately 1 h and terminal elimination occurs after 7.2 h in humans. [130] Thanks to all these studies, it is now well admitted that squalamine administration to humans is safe. [112]

IV-1-2 Trodusquemin

Trodusquemin **74** also called MSI-1436 (Figure 11), possesses the same chemical structure than squalamine **75**, and only differs by the replacement of the spermidine moiety by

a spermine one. This small structural difference has huge consequences on its potent biological activities.

IV-1-2-1 Appetite suppressor activity

Trodsuquemine was well studied to afford reversible suppression of food and fluid intake in mammals, resulting in profound weight loss with the maintain of high-energy expenditure and on the other hand to promote insulin sensitivity and glycemic control without any symptoms of illness. [153-155] Furthermore, the intracerebroventricular administration was 1,000 times more potent than an intraperitoneal treatment, certainly since trodsuquemine can cross the blood brain barrier and has a half-life of 7 days. [154]

A preliminary study suggested that trodsuquemine acts on central targets by knockout the MC4 receptor. [153] It was subsequently demonstrated that trodsuquemine targets the brain by inducing a Fos immunoreactivity in the paraventricular hypothalamic nucleus, but without affecting dopamine transporter function. [154, 156] Thus, screening assays revealed that trodsuquemine inhibits protein-tyrosine phosphatase 1B (PTP1B) a key enzyme regulating insulin and leptin signaling, with an IC_{50} of 1.3 μM in a reversible, non-competitive and dose dependent manner (Figure 21). [155, 157-159]

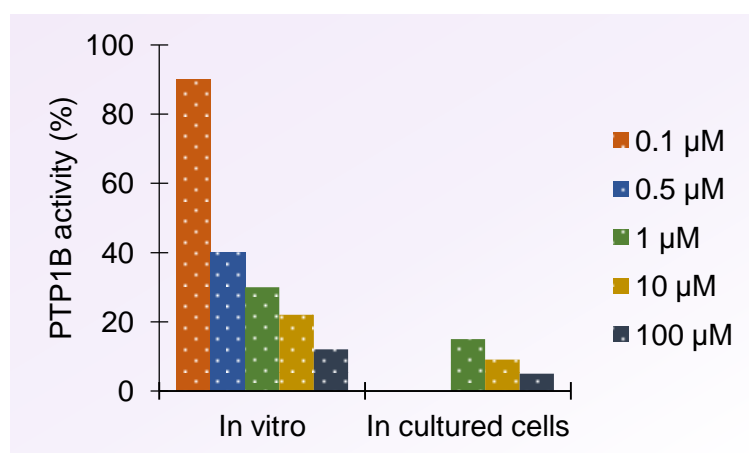


Figure 21. Inhibition activity of trodsuquemine on PTP1B *in vitro* and in cultured cells.

A study identified two binding sites for trodsuquemine in the long form of PTP1B. Hence, a first molecule of trodsuquemine sandwiched between $\alpha 7$ and the C-terminal $\alpha 9'$ may bring the $\alpha 7$ helix close to $\alpha 3$ and $\alpha 6$, forming the second site where another molecule of **69** would bind, which induced a conformational change in PTP1B enzyme and cause its inhibition by a cytostatic effect (Figure 22). [160]

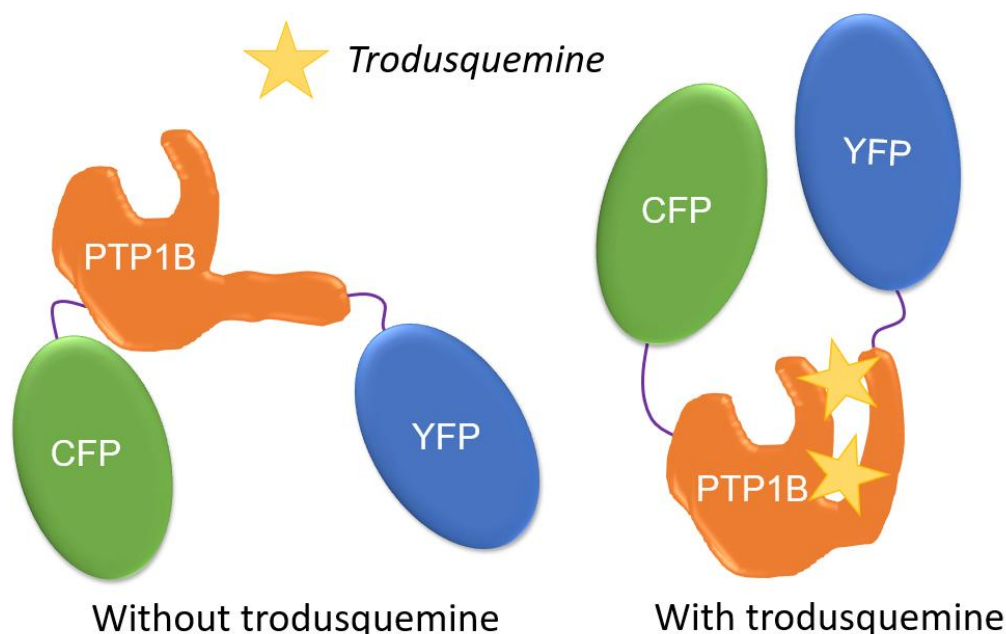


Figure 22. PTP1B fusion protein tagged with CFP and YFP on the N- and C- terminus respectively with and without action of trodusquemine.

It has been demonstrated that the administration of trodusquemine to the hypothalamus, restored central insulin signaling and improved the response of peripheral tissues to insulin on mice. [161] Furthermore, the inhibition of PTP1B by **69** prevents and reverses atherosclerotic plaque formation on mice and allows to demonstrate that PTP1B plays a central role in the regulation of endoplasmic reticulum stress in the endothelium. [162, 163]

IV-1-2-2 Trodusquemine activity against non-alcoholic fatty disease

On account of global obesity epidemic, non-alcoholic fatty disease (NASH) became the first cause of chronic disease of the liver, characterized by an excessive accumulation of fat in the liver. [164] In this context, trodusquemine possesses an activity against NASH which is associated with insulin resistance and diabetes by decreasing hepatic lipid synthesis, stimulating lipolysis and demonstrating its role as an insulin sensitizer (Figure 23). [165]

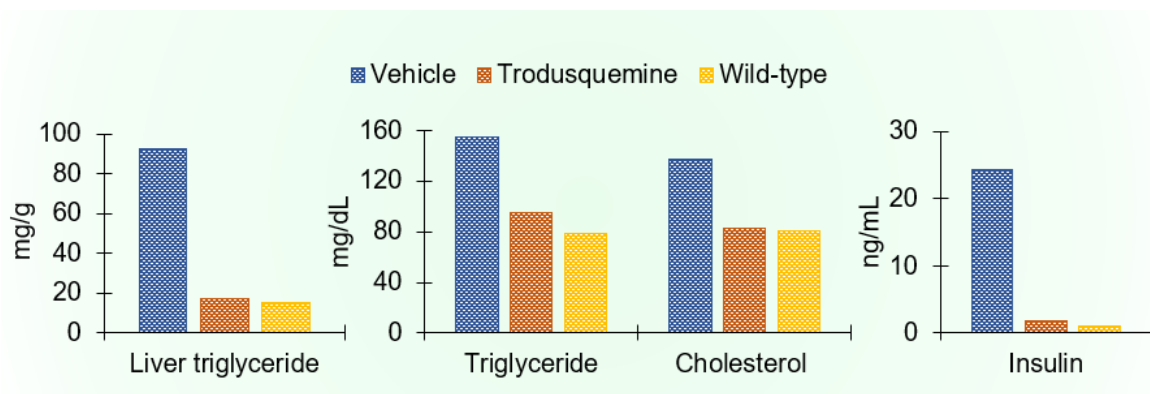


Figure 23. Comparison of the biochemical parameters in wild-type and *Lep^{ob/ob}* mice with or without administration of trodusquemine.

IV-1-2-3 Trodusquemine activity in regenerative medicine

Regenerative medicine, the most recent and emerging branch of medical science, deals with functional restoration of tissues or organs for the patients suffering from severe injuries or chronic disease. [166] In this context, it was demonstrated that the treatment of adult zebrafish with trodusquemine increased the rate of regeneration of the amputated caudal fin. [167] Furthermore, the administration of trodusquemine to adult mice after induction of myocardial infraction, increased survival and cardiomyocyte proliferation, improved heart function, reduced infarct size and ventricular wall thinning. [167] Studies are now under current investigation for a therapeutic human use.

IV-1-2-3 Trodusquemine activity against neurodegenerative diseases

As its closely related compound squalamine, trodusquemine possesses an activity against aggregation of the α -synuclein and inhibits also a secondary nucleation pathway of α -synuclein (fibril-dependent) (

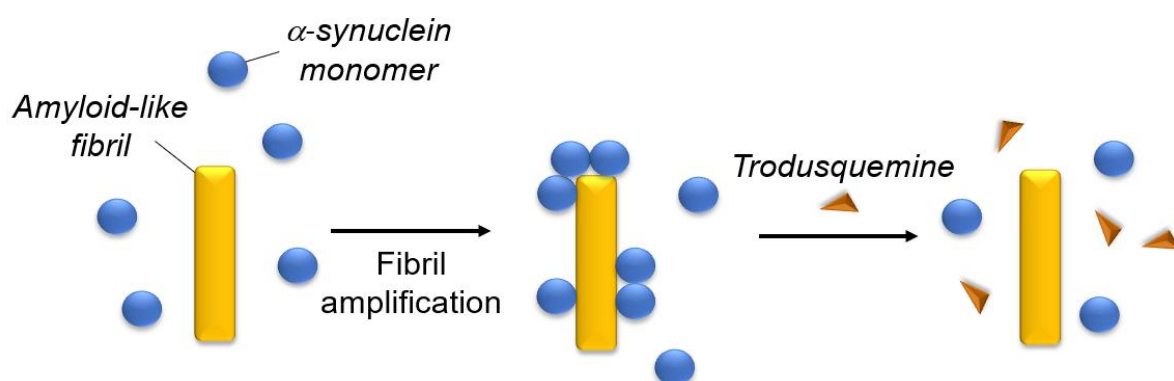


Figure 24), leading to a decrease of the toxicity of α -synuclein oligomers in neuronal cells. [168] Moreover, its administration after the initial growth phase, lead to a dramatical reduction in the number of α -synuclein inclusions, therefore in Parkinson disease it eliminated the related muscle paralysis and increased lifespan. [168] Thus, it was demonstrated that trodusquemine enhances the rate of aggregation of residue 42 from the amyloid- β -peptide, a key pathogenic agent in Alzheimer disease, but significantly reduces the toxicity of the resulting oligomers. [169]

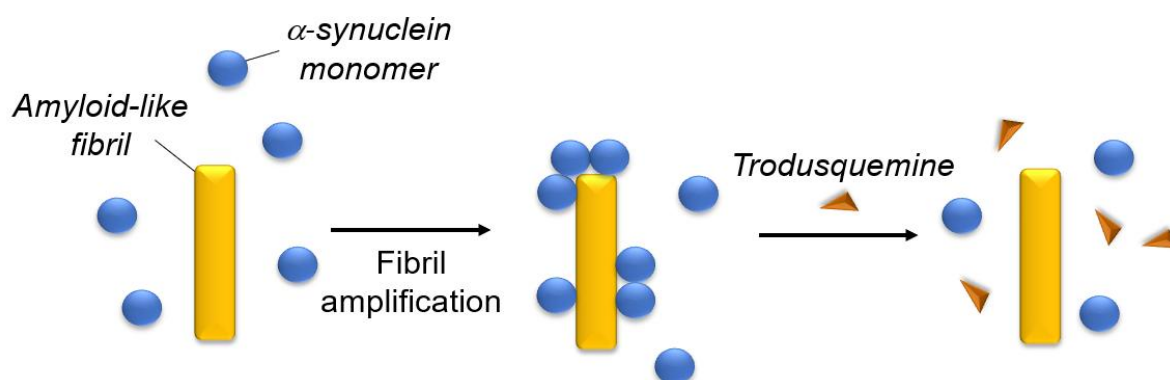


Figure 24. Effect of trodusquemine on fibril amplification.

IV-2 Polyamines from sponges

Marine environment constitutes an amazing store house of novel bioactive natural compounds, with structural and chemical features generally not encountered in terrestrial natural ones. [170] Thus, recent investigations devoted to the search of polyamine derivatives in marine sponges were successful as illustrated above.

IV-2-1 Ianthelliformisamines

The study of marine sponges *Suberea ianthelliformis* and *Pseudoceratina purpurea* allowed to isolate bromotyrosine-derived chemicals namely ianthelliformisamine A, B and C and tokaradine C, respectively **84**, **85**, **86** and **87** (Figure 25). [171, 172] These compounds are all characterized by a 3,5-dibromo-4-methylanisole group linked to a spermine for ianthelliformisamine A **84**, a spermidine for ianthelliformisamine B **85** and tokaradine C **87** whereas a spermidine coupled with two 3,5-dibromo-4-methylanisole group for ianthelliformisamine C **86**. [171]

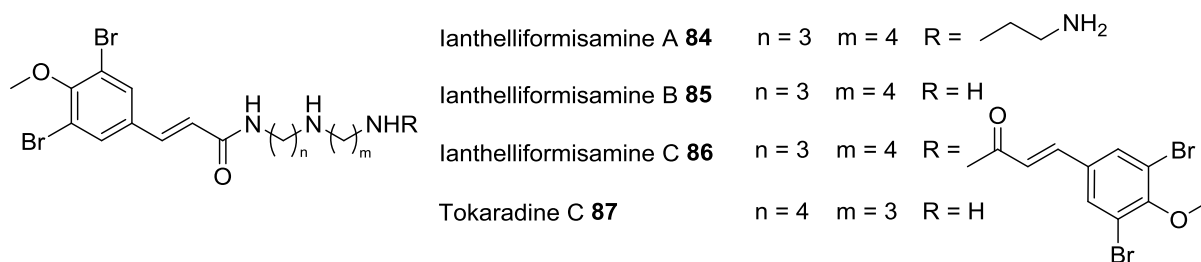


Figure 25. Chemical structures for ianthelliformisamines A-C **84-86** and tokaradine C **87**.

In 2014, two different synthetic pathways were described to achieve ianthelliformisamines A-C and tokaradine C synthesis. [173, 174] The one described here used the same intermediate **90** obtained from 3,5-dibromo-4-hydroxybenzaldehyde **88** through methylation of the hydroxyl group then followed by a Wittig reaction with the phosphonium **89**. [173] **90** was then coupled with spermine **4** to afford ianthelliformisamine A and C with overall yields of 25% and 58%, respectively. [173] On the other hand, the intermediate **90** was coupled with either spermine intermediates bis protected **91** or **92** to afford ianthelliformisamine B and tokaradine C in 67 and 76% of yield, respectively (Figure 26). [173]

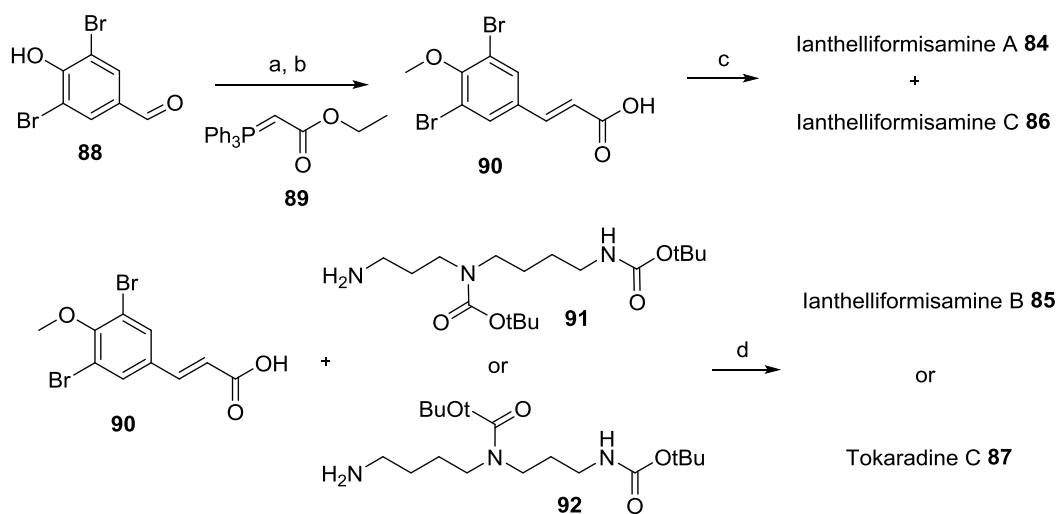


Figure 26. Synthetic pathway for the preparation of ianthelliformisamines A-C and tokaradine C. Reagents and conditions: (a) MeI (1.2 equiv), K_2CO_3 , DMF, 96%; (b) 1) **88**, toluene, 2) LiOH, THF/ H_2O , 87%; (c) Spermine **4**, HOBt/DCC (1.1 equiv), CH_2Cl_2 , yields: 30% **84**, 70% **86**; (d) 1) HOBt/DCC (1.1 equiv), CH_2Cl_2 , 2) HCl, 1M in Et_2O , yields: 80% **85**, 91% **87**.

On a biological point of view, ianthelliformisamines **84** and **86** were reported to possess antibacterial activities against *P. aeruginosa* and *S. aureus* with MIC values of 35 μM , and 17.5 μM , respectively. [171, 173, 174] Furthermore, **84** and **85** possess an activity

against *E. coli* with MICs of 11.04 μM and 14.5 μM , respectively. [174] It is noteworthy that Khan *et al.* demonstrated no activity of **86** against *S. aureus* ATCC 25923, whereas Pieri *et al.* reported a MIC of 12.5 μM against *S. aureus* DSM 799. [173, 174] On the other hand, **86** possesses an antibacterial activity against Gram-negative bacteria *P. aeruginosa* and *K. pneumoniae* with MICs of 25 μM and 12.5 μM , respectively. [173]

Afterwards, ianthelliformisamines and tokaradine C tested in combination with classic antibiotics, were studied for their ability to bypass the different mechanisms of resistance of Gram-negative bacteria and thus improve the activity of inactive antibiotics. [173] Thus, compounds **85-87** used at a 3.12-12.5 μM concentration can restore the activity of doxycycline used at 2 $\mu\text{g/mL}$ against *E. aerogenes*, *P. aeruginosa* and *K. pneumoniae*. [173] In the same way, compounds **86** and **87** at 1.6 and 3.12 μM concentration, respectively, can restore the activity of cefepime used at a 2 $\mu\text{g/mL}$ concentration against *P. aeruginosa*. [173]

Additionally, the parasitic-specific activity of ianthelliformisamines against *Plasmodium falciparum* (3D7) was investigated demonstrating that ianthelliformisamines C **86** is the most active with an IC_{50} of 4.4 μM , while ianthelliformisamines A and B are less active with IC_{50} ranging from 14.5 to 12.1 μM , respectively. [175]

IV-2-2 Protoaculeines

The study of another marine sponge *Axinyssa aculeata* allowed to discover protoaculeine B **93** and a long-chain polyamine LCPA-Aa **94** (Figure 27). [176, 177] Protoaculeine B **93** is structurally composed of a tryptophan-derived hetero tricycle and a long-chain polyamine that is a linearly extended 14 monomers of 1,3-propanediamine and belongs to the toxin peptide aculeines family whereas LCPA-Aa **94** is a linear compound consisting of 5 to 15 monomers of 1,3-propanediamine. [176-178]

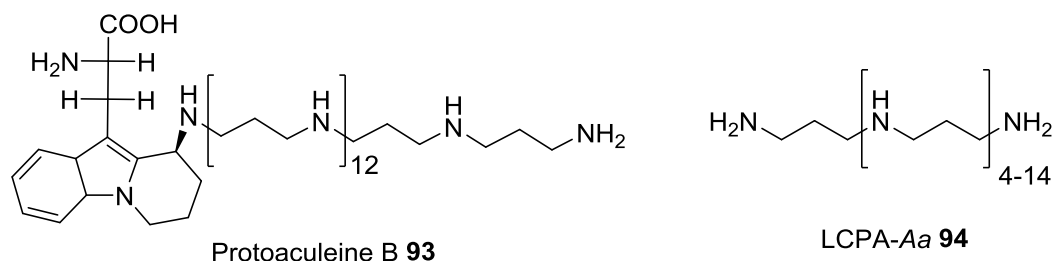


Figure 27. Chemical structures of protoaculeine B **93** and LCPA-Aa **94**.

To achieve the chemical synthesis of protoaculeine B **93**, a synthetic strategy was developed involving the preparation of the fully protected product **102**, in 21 steps with an overall yield of 0.70% (Figure 28). [178] Protoaculeine B **93** was thus prepared from two key

fragments: heterotricyclic N-Ns amine and hydroxylated LCPAs, which were assembled through a Mitsunobu reaction. [178] The heterotricyclic N-Ns amine fragment **98** was achieved in 14 steps, starting from DL-tryptophan **95**, while the hydroxylated LCPAs was afforded in 7 steps from *N*-NPEC-N-Ns amine **99** by an iterative sequence including a split, a NPEC photodegradation, a Tr deprotection and finally a Mitsunobu coupling. [178]

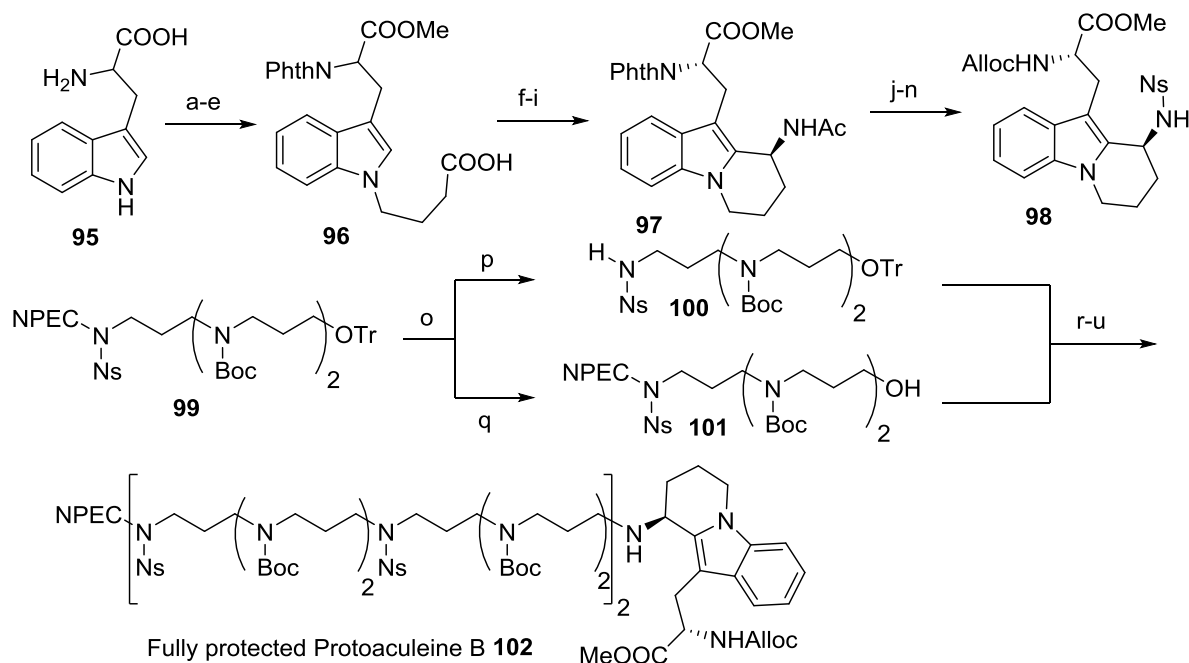


Figure 28. Synthetic pathway to afford the fully protected protoaculeine B **102**. Reagents and conditions: (a) 1) phthalic anhydride pyridine, reflux, 2) AcCl, MeOH, 91%; (b) allyl bromide Cs₂CO₃, DMF, 93%; (c) *tert*-butyl acrylate Zhan catalyst-1B, benzene, 65%; (d) H₂, 5% Pd/C, EtOAc, 93%; (e) TFA, CH₂Cl₂, 100%; (f) P₂O₅, *p*-xylene, reflux, 72%; (g) NH₂OH·HCl, pyridine, reflux, 95%; (h) Fe(OAc)₂, Ac₂O, AcOH, THF, 65%; (i) H₂, 5% Pd/C, EtOH (0.03M), 89%; (j) Boc₂O, DMAP, THF, reflux, 79%; (k) H₂NNH₂·H₂O, THF, reflux, 85%; (l) AllocCl, Et₃N, CH₂Cl₂, 87%; (m) 1) TBSOTf, 2,6-lutidine, CH₂Cl₂, 2) Bu₄NF, THF, 100%; (n) NsCl, Et₃N, CH₂Cl₂, 99%, (o) 1) split; (p) 2) hn (high-pressure Hg lamp, 435 W), MeOH (0.0023 M), 0°C, 94%: NPEC photodegradation; (q) 2') TFE, reflux, 88%: Tr deprotection; (r) 3) DEAD, PPh₃, PhH, 82%: Mitsunobu coupling; (s) iterative sequence: 1) split, 2) NPEC photodegradation, 81%, 2') Tr deprotection, 55% (87% brsm), 3) Mitsunobu coupling, 78%; (t) Tr deprotection: TFE, 40% (90% brsm); (u) heterotricycle fragment **98**, DEAD, PPh₃, PhH, 63%

It was demonstrated that LCPA-Aa **94** is a chemical factor involved in silica depositions in sponges. [177] LCPA was also reported to interact with bilayers to form three-dimensional

stacks on the surface of artificial lipid membranes, but the stack formation is highly dependent on the number of polyamine units present in the LCPA. [178, 179]

IV-2-3 Motuporamines

Marine sponges are a rich source of structurally novel metabolites of interest as potential lead compounds for the development of new drugs. [180] In this context, extracts of the tropical sponge *Xestospongia exigua* (Kirkpatrick) lead to the isolation of macrocyclic polyamines, namely motuporamines A to I **103-111** (Figure 29). [180, 181]

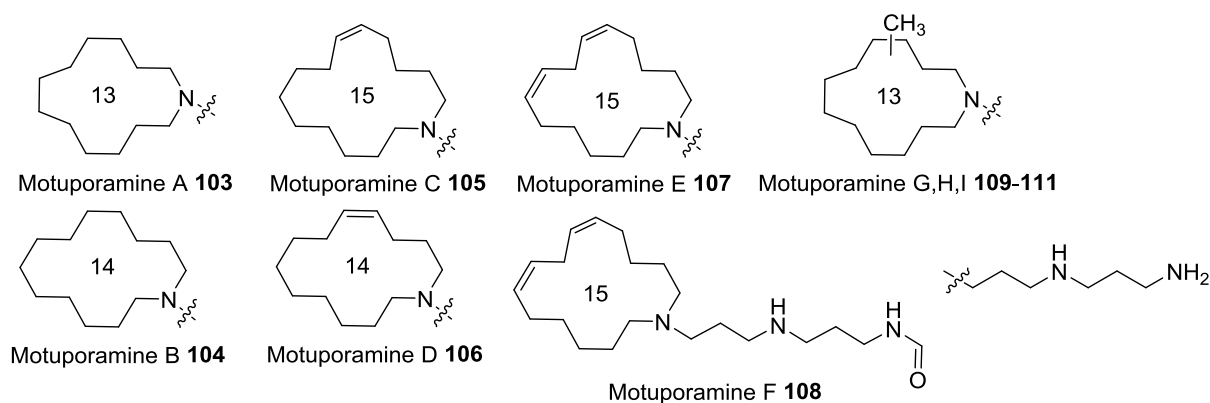


Figure 29. Chemical structures of motuporamines A to I **103-111**.

Since their isolations, different synthetic pathways were described to achieve motuporamines A **103**, B **104**, C **105** and G **109** preparation (Figure 30). Thus, the synthesis of motuporamines A **103** and B **104** were performed from either 2-azacyclotridecanone **112** or a mixture of 2-azacyclotetradecanones **116** and **117** in 4 steps, with overall yields of 85% and 75%, respectively. [182] 2-azacyclotridecanone **112** was also used to afford **103** in 8 steps (25% overall yield) whereas cyclododecanone **115** was involved to achieve the synthesis of **104** in 11 steps (8% overall yield), both involving a key step of reductive amination using sodium triacetoxyborohydride. [183] Afterwards, an iterative ring-expansion method was developed to achieve **103** from 1-benzyl-2-vinylpyrrolidine **113** and **104** from 1-benzyl-2-vinylpiperidine **114**, by a tandem conjugate addition to (*p*-toluenesulfonyl) ethyne and a formal 3-aza-Cope rearrangement. [184, 185] The motuporamine C **105** was obtained in 6 steps synthesis with 45% overall yield, starting from N-(dec-9-en-1-yl)hex-5-enamide **118** and involving a ring-closing metathesis reaction, a Michael addition and an amide formation reaction to introduce the spermine-like unit. [182] Thus, the same starting material was used to afford **105** using a Z-selective catalyzed ruthenium macrocyclization. [186] Otherwise, the synthesis of **105** was achieved from 9-undecyn-1-ol **119** in 8 steps with an overall yield of 12%. [187] In 2012, the synthesis of motuporamine G **109** was achieved through a three steps

synthesis from 2-(prop-1-en-2-yl)cyclononan-1-one **120** and by using a tandem nucleophilic addition as well as an oxy-2-aza-cope rearrangement. [188]

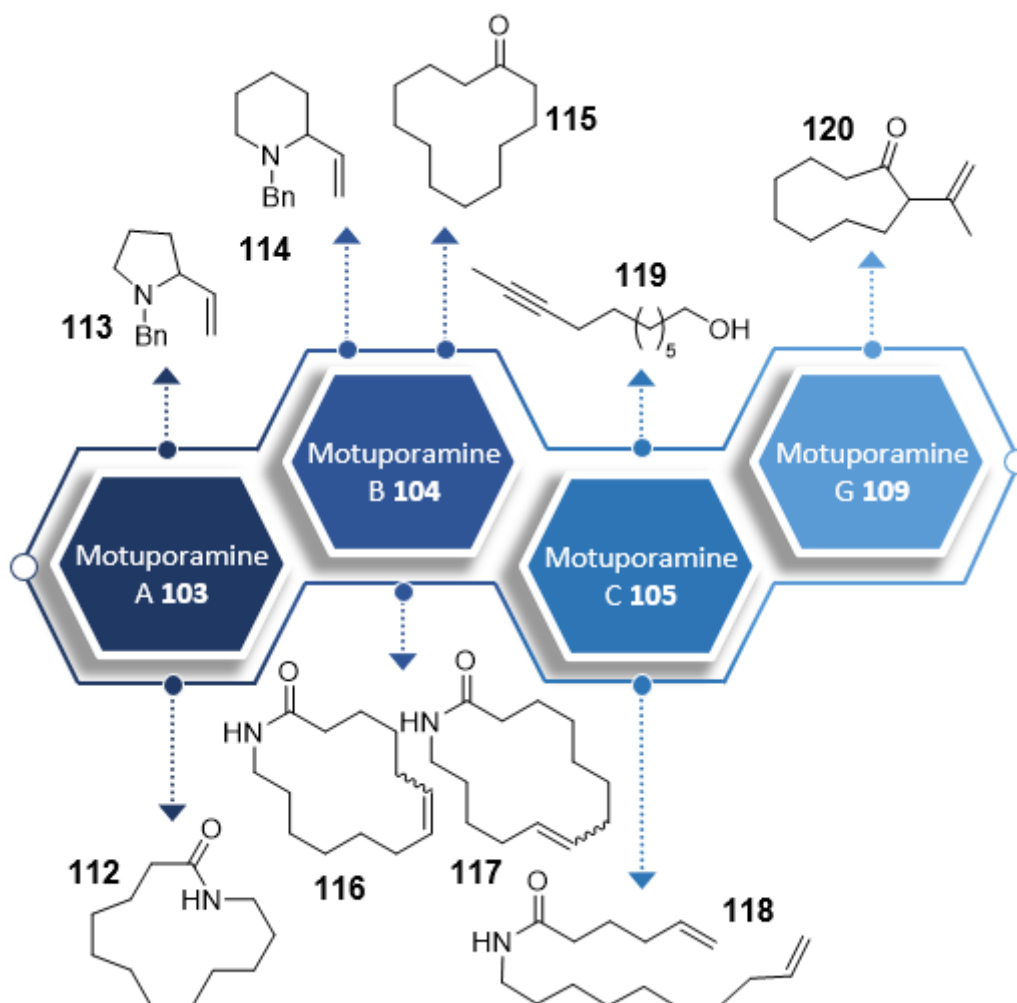


Figure 30. Synthetic pathways involved to prepare motuporamines A **103**, B **104**, C **105** and G **109**.

On a biological point of view, it was demonstrated that motuporamines possess anti-invasive and anti-angiogenic activities against cancer cells with a mean IC_{50} of 0.6 $\mu\text{g/mL}$ (Figure 31). [180] On the other hand, they were identified as inhibitors of invasion of breast carcinoma cells and the more potent anti-invasive compound is motuporamine C which interferes with the migration and leading edge ruffling of human breast cancer cells, prostate carcinoma cells and glioma cells. [189] Furthermore, motuporamine C inhibits angiogenesis in both an *in vitro* sprouting assay and an *in vivo* chick chorioallantoic membrane assay, with a low cytotoxicity. [189] Afterwards, it was demonstrated that motuporamine C inhibits the motility of MDA-231 human breast cancer cells *in vitro* and it is believed that this is the basis of its anti-invasive and anti-angiogenic properties. [181]

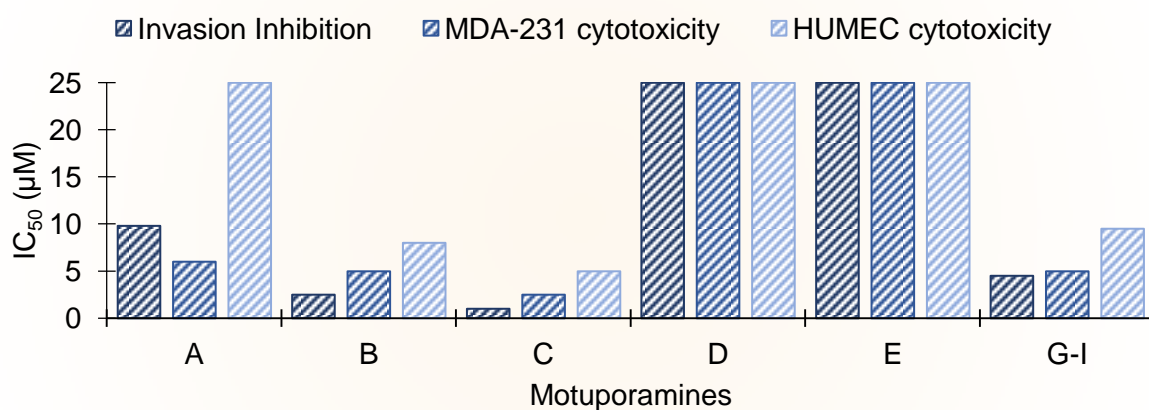


Figure 31. Anti-invasion and cytotoxicity data for motuporamines A-E and G-I.

It is also noteworthy that non-natural motuporamine derivatives were recently prepared and are now widely studied for their interesting anti-angiogenic and antimicrobial activities. [190-195]

V-Polyamines from microorganisms

Microorganisms play an essential role in almost all ecosystem and some of them use polyamines to adapt and survive in all these environments. In this section we will focus our attention on the recent discovery of new polyamine derivatives from bacteria and fungi presenting interesting biological activities.

V-1-Petrobactins

In 2002, spermidine containing compounds namely petrobactin **121-123** were isolated from an oil-degrading marine bacterium *Marinobacter hydrocarbonoclasticus* as well as from *Marinobacter aquaolei* (Figure 32). It is also noteworthy that petrobactin **89** was found in *Bacillus anthracis* the causative agent of anthrax and some other bacteria from *Bacillus* group. [196-201]

The first isolation and characterization of petrobactin supposed a structure possessing 2,3-dihydroxybenzoyl functions. [196] Nevertheless, its synthesis was not conclusive and the preparation of the compound with 3,4-dihydroxybenzoyl groups permitted to confirm the correct structure of naturally isolated petrobactin **121**. [202]

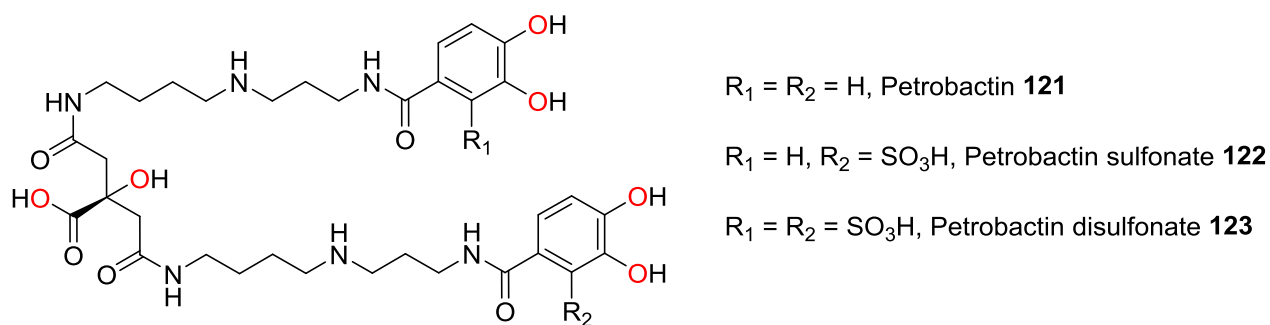


Figure 32. Chemical structures of petrobactin **121**, petrobactin sulfonate **122** and petrobactin disulfonate **123**. The iron-coordinating atoms are indicated in red.

Petrobactin is a siderophore i.e. a highly specific Fe(III) transport ligand secreted by bacteria under conditions of iron stress to scavenge and transport iron. [196, 203] For a siderophore, possess a 2,3-dihydroxybenzoyl function is classical, whereas the presence of a 3,4-dihydroxybenzoyl group is quite unusual. [202] The siderophore part is composed by a bis-catechol α -hydroxy acid which is bound to iron to form the Fe(III)-petrobactin complex **124**. [196, 204] On account of the presence of a citryl group, the ferrated form might be photolabile. Thus, when FePB **124** is exposed to the light it readily performed a decarboxylation reaction, followed by the reduction of Fe(III) into Fe(II) (Figure 33). [196, 203] Furthermore, petrobactin has also an unexpected binding affinity for boron through the catechol groups of petrobactin to form a borate-petrobactin complex permitting the development of methods to study petrobactin either in the bacteria or in seawater. [205-210]

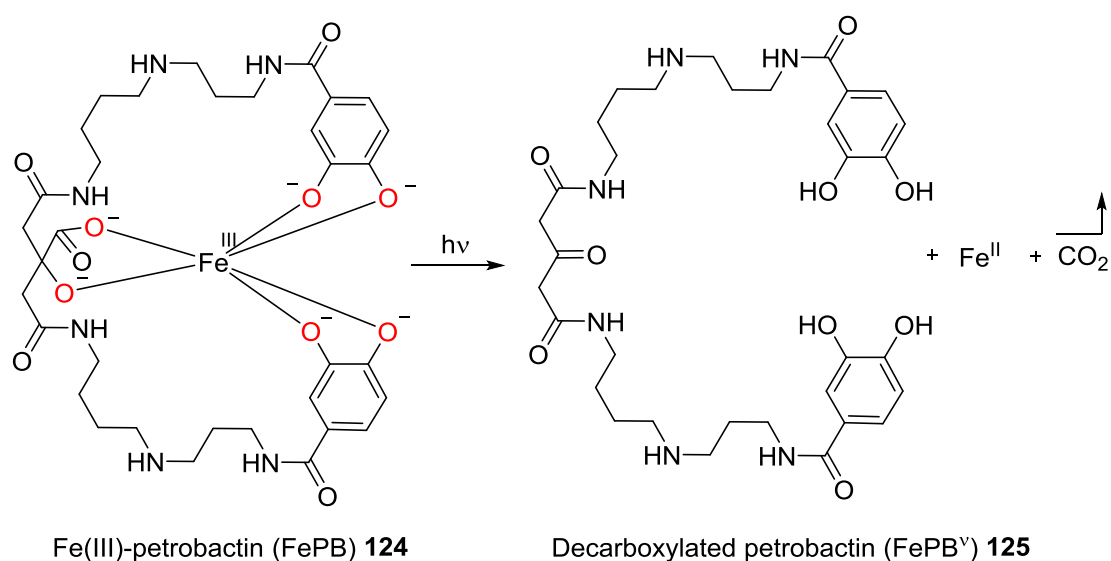


Figure 33. Photochemical transformation of FePB **124** into FePB^V **125**.

Since its discovery, few chemical syntheses of petrobactin were performed. [202, 211, 212] Thus, the synthetic pathway described below was purposed by Bugdanh *et al.* in 2014 in 9 steps with 19% overall yield (Figure 34). [207, 213]

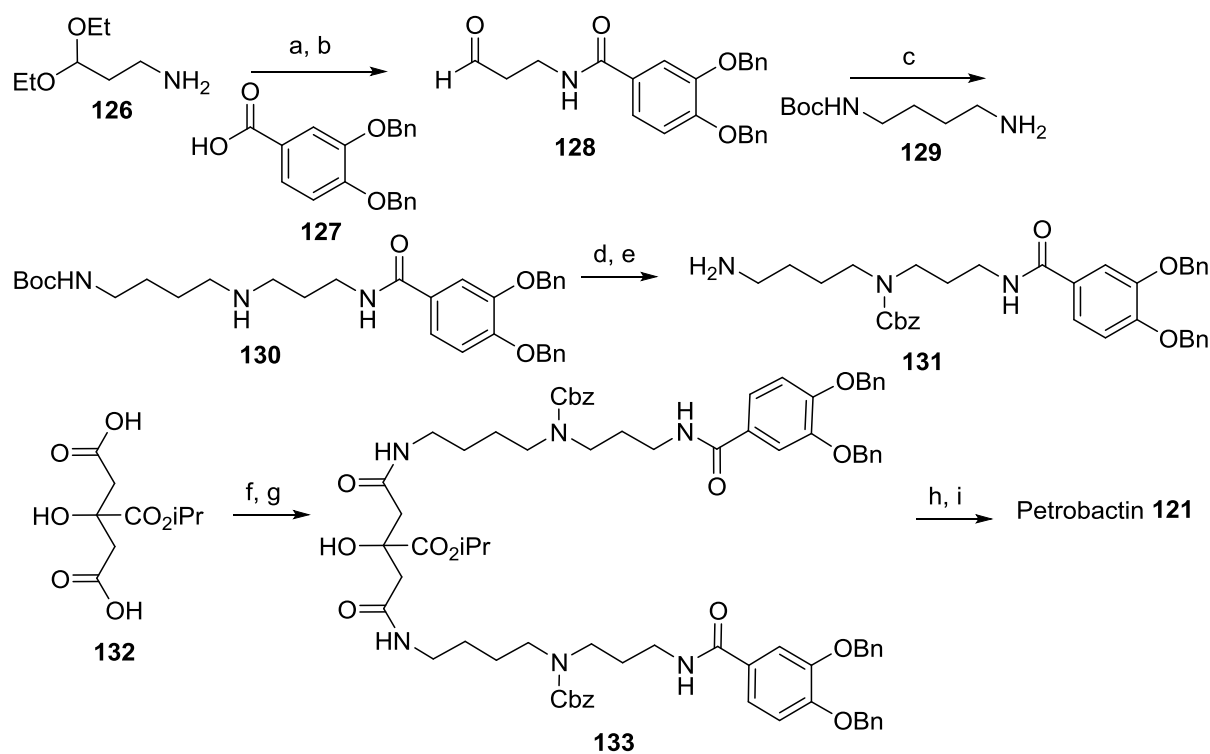


Figure 34. Synthesis pathway for the preparation of petrobactin **121**. Reagents and conditions: (a) **127**, EDC, HOBT, Et₃N, DMF, 92%; (b) PPTS, acetone/H₂O, 40°C, 96%; (c) **129**, NaBH(OAc)₃, Et₃N, MeOH, 71%; (d) CbzCl, Et₃N, MeOH, 94%; (e) TFA, CH₂Cl₂, 0°C, 95%; (f) PNP, DCC, 74%; (g) **131**, Et₃N, 69%; (h) LiOH, 0°C; (i) H₂, Pd/C, 62% (2 steps)

The synthesis of the petrobactin spermidine side chain **131** was realized from commercially available diethyl acetal of 3-aminopropanal **126** and the bis protected 3,4-bis(benzyloxy)benzoic acid **127** easily obtained from 3,4-dihydroxybenzoic acid. [207, 211] The isopropyl-protected citric acid building block **132** was activated by treatment with *para*-nitrophenol and DCC for amide coupling reaction with the petrobactin spermidine side chain **131** to provide the fully protected petrobactin **133**. The isopropyl ester was then removed by treatment with aqueous lithium hydroxide and followed by the removal of the Cbz and Bn groups by a subsequent hydrogenolysis to afford petrobactin **121** (Figure 34). [213]

On a biological point of view, it was demonstrated that *B. anthracis* uses petrobactin to scavenge iron from its environment during the infection and increases its full pathogenicity. [214, 215] Thus, petrobactin is produced by pathogenic strains but also innocuous isolates

alike suggesting that it is not itself indicative of virulence but certainly required for pathogenesis (Figure 35). [216] Furthermore, petrobactin evades innate immune response of mammalian, because neither petrobactin nor FePB are substrates for sequestration by siderocalin, a mammalian protein binding with siderophore to prevent iron acquisition by pathogenic bacteria. [214] Nevertheless, a reshaping of siderocalin was developed to specifically bind to FePB with a high affinity, in the aim to reactivate immune response against the pathogenic bacteria. [217]

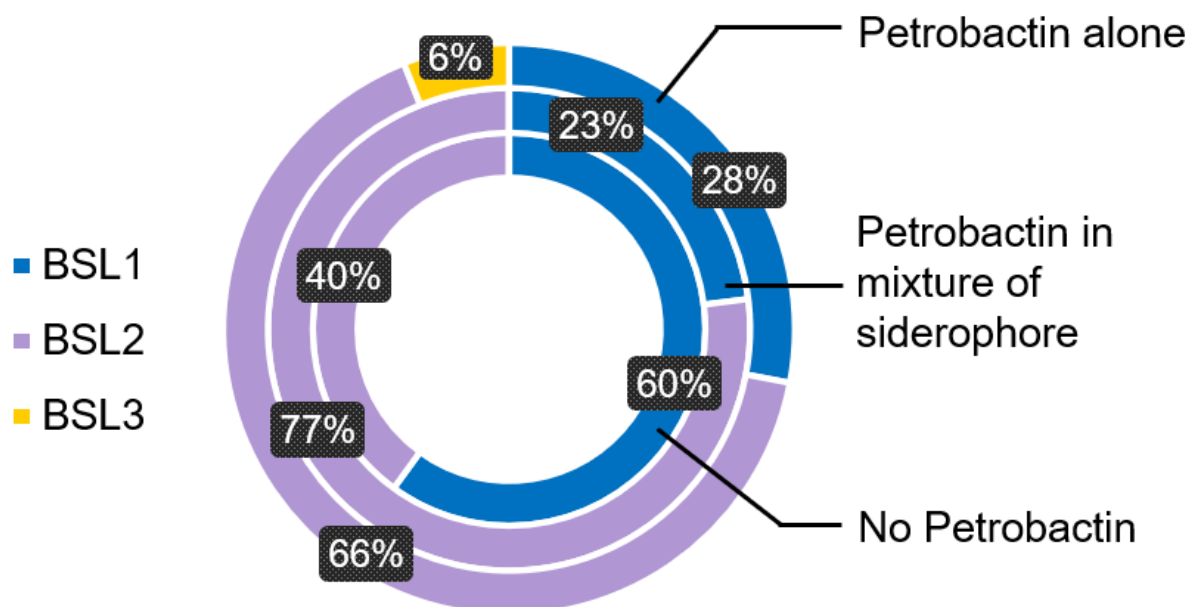


Figure 35. Percentage of strains in each biological safety level (BSL) designation and their siderophore production phenotypes.

The biosynthesis of petrobactin was performed by the *asb*ABCDEF (anthrochelin biosynthesis) gene cluster from *B. anthracis*, starting from 3,4-dihydroxybenzoic acid **134**, spermidine **3** and citric acid **135** (Figure 36). [215, 218-226] In addition, it was demonstrated that the whole or a part of the gene cluster that encoded petrobactin was presented in other bacteria. [227, 228]

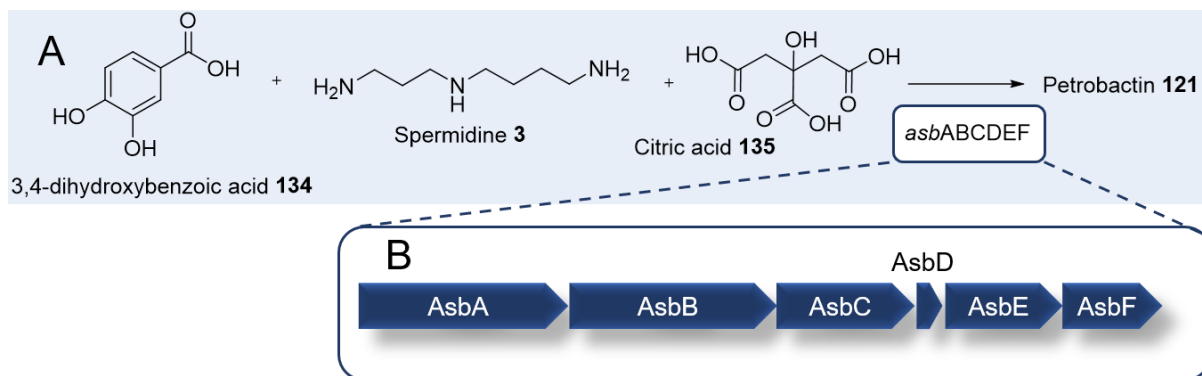


Figure 36. Biosynthesis of petrobactin **121**. (A) Different components enter the biosynthesis of petrobactin **121**. (B) Enzymes involved in the assembly of petrobactin **121** are encoded by the genes from the *asbABCDEF* cluster.

The study of the mechanism of transport of petrobactin demonstrated the implication of the *yclNOPQ* operon for *B. subtilis* and the protein FpuA for the import of petrobactin for *B. anthracis*. [229-231] Indeed, from $\Delta fpuA$ bacterial strains, spores were significantly attenuated, and lethal dose was 3,000 times higher than for wild type. [231] It was then demonstrated that for *B. subtilis* the responses of the ferric uptake regulator were performed by FsrA and PerR regulons. [232]

Numerous investigations allow to enlighten a part of the mechanism of transport of petrobactin in *B. anthracis* (Figure 37). Firstly, petrobactin was exported into peptidoglycan through the apo-petrobactin exporter (ApeX) a member of the resistance-nodulation-cell division family of transporters. [208] In a second step, petrobactin will bind to ferric iron ions to form FePB complex, which is recognized by petrobactin receptor FpuA linked with two permeases and three ATPases to import petrobactin. [233] The last step of the mechanism dealing with the siderophore degradation or the iron release into the bacteria remains still unclear and two hypotheses either metal reduction or siderophore hydrolysis are still under current investigations. [234]

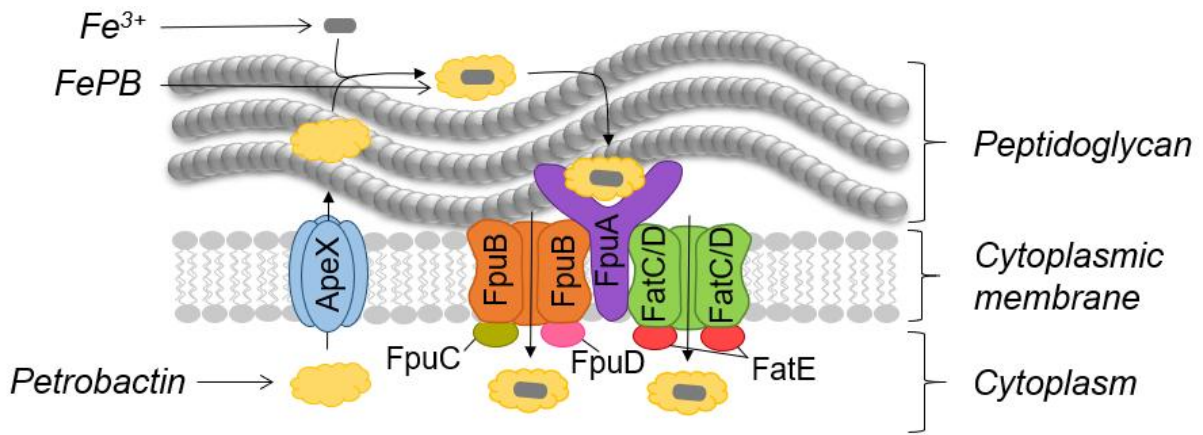


Figure 37. Model for *B. anthracis* petrobactin export and import.

It was demonstrated that petrobactin is involved in germination of *B. anthracis*, growth in macrophage, required for sporulation and may be for transmission. [235] Furthermore, petrobactin is accumulated inside the bacteria when medium contains more than 20 μM of iron and accumulation is higher in presence of oxygen. [236] Moreover, it was demonstrated that petrobactin is synthesized even in iron-rich conditions and during sporulation, but instead of being exported, petrobactin remains intracellular to protect against oxidative stress and improve sporulation (Figure 38). [235]

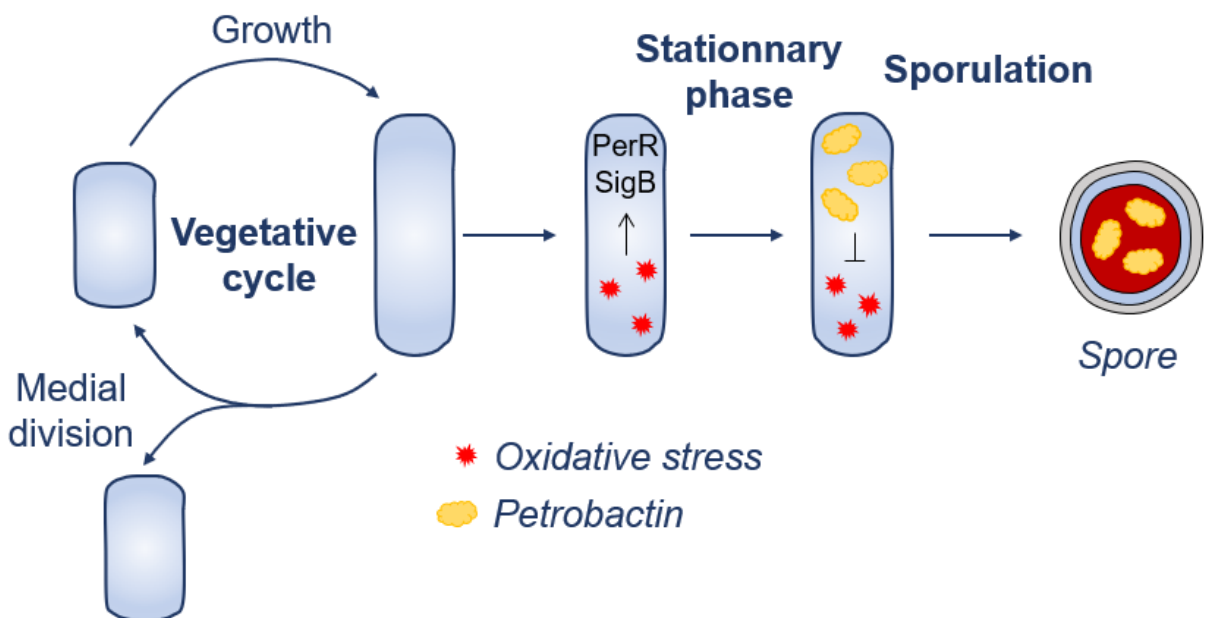


Figure 38. Petrobactin role in *B. anthracis* during late-stage growth and early sporulation.

V-2-Pistillarins

Pistillarins constitute another class of polyamine derivatives isolated from fungi and widely studied. Thus, pistillarin **136** presenting two 3,4-dihydroxycatechol groups linked to a spermidine moiety, was encountered in fungi such as *Clavariadelphus pistillaris*, several *Ramaria spp.* [237], *Penicillium bilaii* [238], *Rubinoboletus ballouii* [239], *Gomphus floccosus* [240] and *Gomphaceae* family [241] as well as *Clavaria pistillaris* and *Clavaria fennica* (Figure 39). [242]

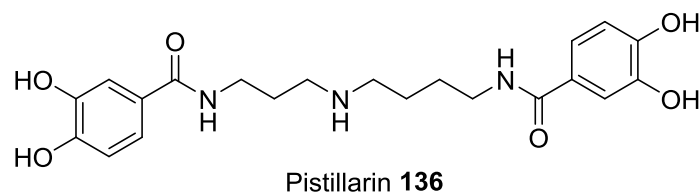


Figure 39. Chemical structure of pistillarin **136**.

The chemical synthesis of pistillarin **136** was achieved in two steps from 3,4-dimethoxybenzoic acid **137** and spermidine **3** involving a peptide coupling reaction using CDI, while methylated groups were subsequently deprotected with boron tribromide (Figure 40). [243, 244]

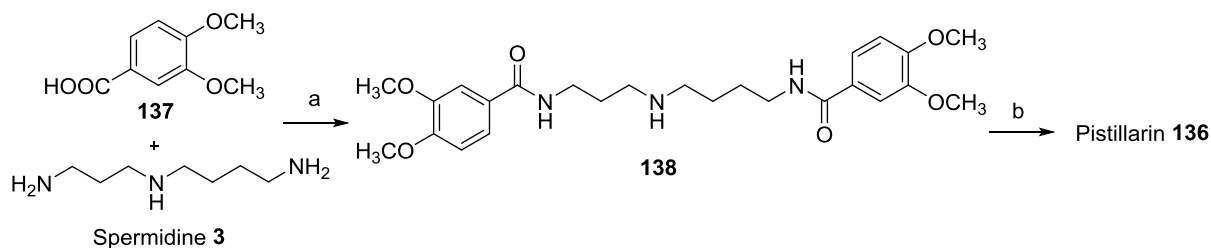


Figure 40. Chemical synthesis of pistillarin **102**. Conditions and methods: (a) CDI, CH₂Cl₂, overnight, room temperature; (b) BBr₃ / CH₂Cl₂, 97%

It is noteworthy that pistillarin contains the same unusual 3,4-dihydroxycatechol moieties than petrobactin and was demonstrated to act as a siderophore for the Gram-negative bacteria *Yersinia enterocolitica*. [244, 245] Thus, the formation of the complex occurs rapidly in acidic media with an affinity constant for Fe(III) of 29, indicating that pistillarin is a very good ligand for iron(III). [246] Otherwise, pistillarin was demonstrated exhibit significant anti-inflammatory effects on phytohemagglutinin-stimulated human peripheral blood mononuclear cells, by inhibiting [methyl-³H]-thymidine uptake and inflammatory cytokines productions such as tumor necrosis factor (TNF)- α , interleukin (IL)-10, interferon (IFN)- γ and IL-1 β (Figure 41). [239]

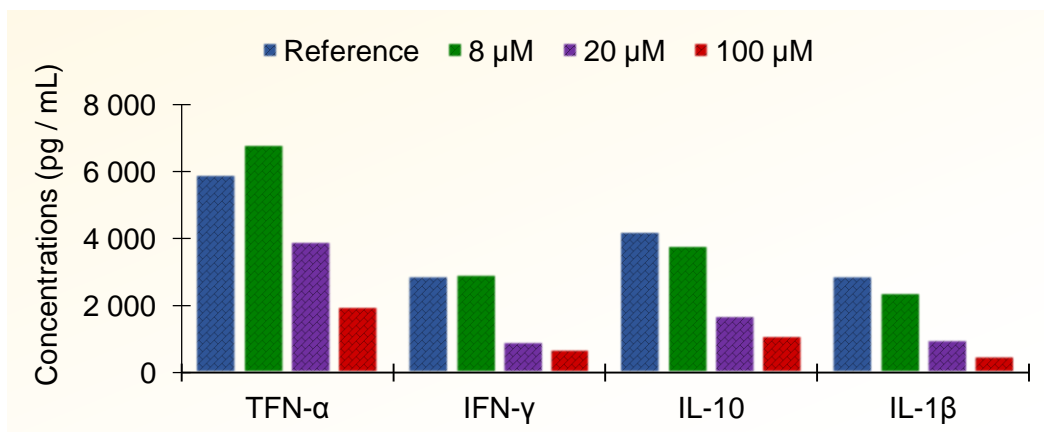


Figure 41. Immunomodulating effect of pistillarin **136** on (TNF)- α , (IL)-10(IFN)- γ and IL-1 β .

On another biological point of view, pistillarin was then tested against a panel of Gram-positive and Gram-negative bacteria as well as fungi and no antimicrobial activity was detected even at high concentrations (MICs > 100 μ M). [247] Additionally, pistillarin does not present any cytotoxic effect against the mammalian cell HEK293 and HepG2, with CC₅₀ > 50 μ M. [247] Nevertheless, pistillarin **136** possesses an antimalarial activity *in vitro* and *in vivo* against *Plasmodium falciparum* (3D7) with an IC₅₀ of 1.9 μ M. [175] Since it is well-known that iron is essential for parasitic growth and multiplication, hence chelation within infected erythrocytes could constitute the mechanism of action of pistillarin to control *P. falciparum* infection. [175] Finally, pistillarin exhibits also significant protective effects against DNA damage, by iron chelation towards hydroxyl radicals generated by the Fenton reaction involving iron and hydrogen peroxide. [240]

VI-Polyamines from plants

Secondary metabolites of plant origin have long been of interest to man, one of the major reasons in recent years being their use, either directly as precursors or as lead compounds in the pharmaceutical industry. [248]

VI-1-Kukoamine A

Lycii Cortex, the root bark of *Lycium chinensis* is a famous traditional Chinese herb and a functional food widely consumed in health-preserving cuisines. [249] Thus, from this Chinese medicinal plant was isolated the active compound, a bis(dihydrocaffeoyl) spermine derivative namely kukoamine A **139** and later it was also encountered from the potato *Solanum tuberosum* (Figure 42). [248, 250]

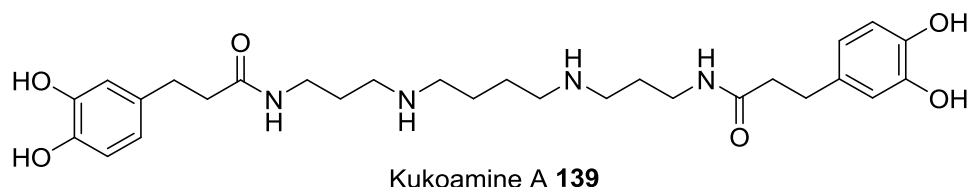


Figure 42. Chemical structure of kukoamine A **139**.

Few synthetic pathways were reported in literature to achieve the synthesis of kukoamine A **139**, using either a selective alkylation of spermine or a solid-phase chemistry. [251, 252] Then, Garnelis *et al.* described a synthesis in 5 steps starting from the 3,4-dihydroxybenzaldehyde **140** and involving a Wittig reaction followed by a saponification step and affording the expected intermediate **141**. The corresponding crystalline active esters **142** was prepared through a reaction with *N*-hydroxysuccinamide in the presence of *N,N'*-dicyclohexylcarbodiimide and the bishydroxycuccimidate salt was easily obtained by reacting with spermine (Figure 43). [253]

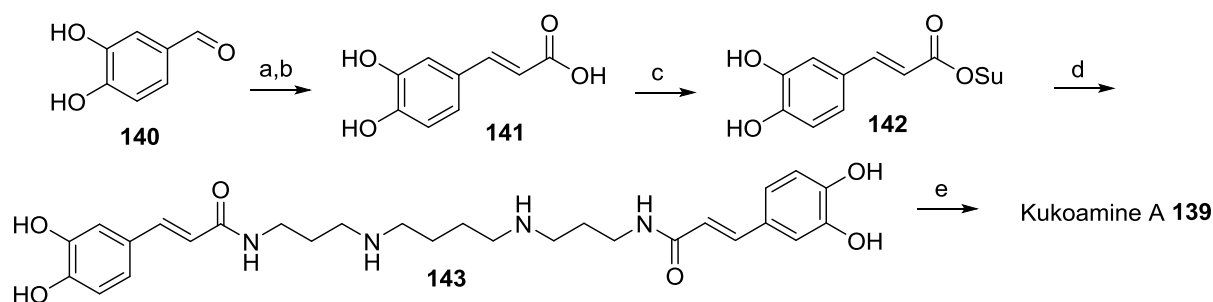


Figure 43. Synthesis of kukoamine A **139**. Reagents and conditions: (a) $\text{Ph}_3\text{C}=\text{CHCO}_2$, MeCN, reflux, 10 h; (b) 4N NaOH, DMSO / MeOH (2:1), (c) HOSu / DCC, THF / FMD (3:1), 0°C, (d) Spermine / Pr_2Net , (e) H_2 (1atm) / 20% Pd-C, MeOH / AcOH / H_2O (4:1:0.1).

An extractive method of Kukoamine A from the potato peels was also reported permitting to obtain the product with a purity of 90%. [254]

Kukoamine A possesses a widely range of different biological activity such as antihypertensive activity [250], inhibitor of *Crithidia fasciculata* trypanothione reductase [255], anti-oxidative stress and protection against Fenton-induced damage (Figure 44A). [256-258] Kukoamine A was demonstrated to inhibit human glioblastoma cell growth and migration *in vitro* and *in vivo* through apoptosis induction and epithelial-mesenchymal transition attenuation (Figure 44B). [259] Thanks to these both activities, **139** was studied to prevent the detrimental effect of ionizing radiation used during radiotherapy on hippocampal neurons and demonstrated to be neuroprotective compound against radiation-induced brain

injury though inhibiting neuronal oxidative stress and apoptosis (Figure 44C). [260, 261] Furthermore, it was enlightened that the protection of the brain by kukoamine A occurs through mitochondria apoptosis signaling pathway. [262] Moreover, **139** was used with success as neuroprotective against Parkinson and Alzheimer disease [263, 264] as well as against NASH by attenuating insulin resistance and fatty liver (Figure 44D). [265]

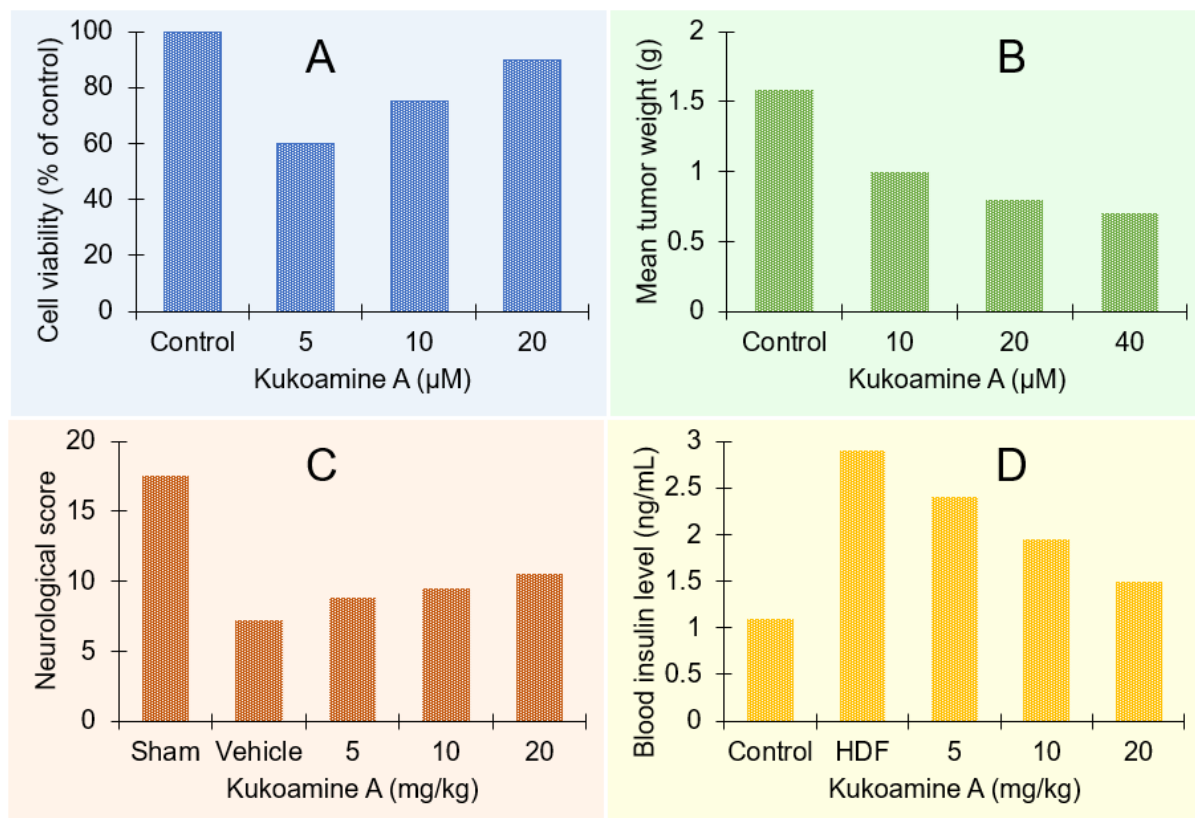
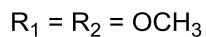
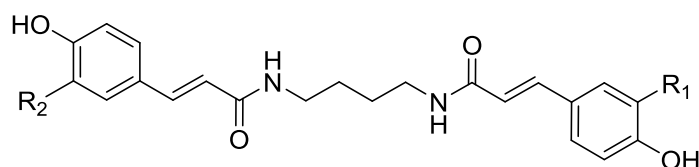


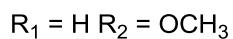
Figure 44. (A) Neuroprotection of kukoamine A in SH-SY5Y cells, dose dependent protective effect of pretreatment with **139** against H₂O₂ (600 μM) induced cytotoxicity. (B) Inhibition of human glioblastoma growth by Kukoamine A. (C) Dose response study of the neuroprotection afforded by **139**, column chart showed corresponding the neurological scores of rats developed by Garcia *et al.* (D) Effect of **139** on insulin resistance in HFD-fed mice.

VI-2-Putrescine derivatives

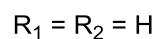
From the corn bran were isolated three compounds *N,N'*-Diferuloylputrescine **144**, *N,p*-Coumaroyl-*N'*-feruloylputrescine **145** and *N,N'*-Dicoumaroylputrescine **146**, all possessing a putrescine core (Figure 45). [266, 267]



N,N'-Diferuloylputrescine **144**



N,p-Coumaroyl-*N'*-feruloylputrescine **145**



N,N'-Dicoumaroylputrescine **146**

Figure 45. Chemical structure of polyamines **144-146** found in corn bran.

It was demonstrated that these compounds possess antioxidant activity against 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical as well as superoxide and hydroxyl radicals generated by enzymatic and nonenzymatic reactions. [267] Thus, **144** showed potent DPPH ($\text{IC}_{50} = 38.46 \mu\text{M}$) and superoxide ($\text{IC}_{50} = 291.62 \mu\text{M}$) radical scavenging activities, while **146** exhibited the strongest hydroxyl radical scavenging activity ($\text{IC}_{50} = 120.55 \mu\text{M}$). [267]

Conclusion

Natural polyamines appear as essential for life and are widely involved for growth cells, cellular reparation, gene transcription, proteins, and nucleic acids synthesis. On the other hand, due to their structure and physical properties, polyamine derivatives open the way to the development of new class of anticancer, antifungal, or antimicrobial agents against MDR pathogens and could become a last-line therapeutic drug in the coming fifty years. Indeed, development in these fields could lead to the isolation or design of various polyamine derivatives possessing original mechanism of action towards cancer cells or microbes. In this latter case, during the last decades, the search and development of new antibiotics did not follow the rhythm of emergence of Multidrug resistant bacteria. Thus, the search for antimicrobial polyamine derivatives acting by membrane depolarization and/or integrity membrane disruption could constitute an efficient alternative to the well-known classes of drugs available. Furthermore, the finding of active molecules preferably with a weak antibiotic activity and that in combination with antibiotics could enhance or synergize the antimicrobial activity of inefficient antibiotics. In this case, the use of antibiotic adjuvants will

lead to two beneficial outcomes: enhancement of the antimicrobial effect and reduction of the occurrence of mutations resulting in resistance. Finally, it is reasonable to envision that the continuous advances in the development of new and potent high-throughput technologies will definitively allow the discovery of new polyamine compounds possessing specific biological properties against cancer, degenerative diseases...

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