



# Activity-Regulated Cytoskeleton-Associated Protein Accumulates in the Nucleus in Response to Cocaine and Acts as a Brake on Chromatin Remodeling and Long-Term Behavioral Alterations

Marine Salery, Marc dos Santos, Estefani Saint-Jour, Lara Moumné, Christiane Pagès, Vincent Kappès, Sébastien Parnaudeau, Jocelyne Caboche, Peter Vanhoutte

## ► To cite this version:

Marine Salery, Marc dos Santos, Estefani Saint-Jour, Lara Moumné, Christiane Pagès, et al.. Activity-Regulated Cytoskeleton-Associated Protein Accumulates in the Nucleus in Response to Cocaine and Acts as a Brake on Chromatin Remodeling and Long-Term Behavioral Alterations. *Biological Psychiatry*, 2017, 81 (7), pp.573-584. 10.1016/j.biopsych.2016.05.025 . hal-03365869v2

**HAL Id: hal-03365869**

**<https://hal.science/hal-03365869v2>**

Submitted on 5 Oct 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Activity-Regulated Cytoskeleton-Associated Protein Accumulates in the Nucleus in Response to Cocaine and Acts as a Brake on Chromatin Remodeling and Long-Term Behavioral Alterations

Marine Salery, Marc Dos Santos, Estefani Saint-Jour, Lara Moumné, Christiane Pagès, Vincent Kappès, Sébastien Parnaudeau, Jocelyne Caboche, and Pèter Vanhoutte

**BACKGROUND:** Addiction relies on persistent alterations of neuronal properties, which depends on gene regulation. Activity-regulated cytoskeleton-associated protein (Arc) is an immediate early gene that modulates neuronal plasticity underlying learning and memory. Its role in cocaine-induced neuronal and behavioral adaptations remains elusive.

**METHODS:** Acute cocaine-treated mice were used for quantitative reverse-transcriptase polymerase chain reaction, immunocytochemistry, and confocal imaging from striatum. Live imaging and transfection assays for Arc overexpression were performed from primary cultures. Molecular and behavioral adaptations to cocaine were studied from Arc-deficient mice and their wild-type littermates.

**RESULTS:** Arc messenger RNA and proteins are rapidly induced in the striatum after acute cocaine administration, via an extracellular-signal regulated kinase-dependent de novo protein synthesis. Although detected in dendrites, Arc accumulates in the nucleus in active zones of transcription, where it colocalizes with phosphorylated histone-H3, an important component of nucleosomal response. In vitro, Arc overexpression downregulates phosphorylated histone-H3 without modifying extracellular-signal regulated kinase phosphorylation in the nucleus. In vivo, Arc-deficient mice display decreased heterochromatin domains, a high RNA-polymerase II activity and enhanced c-Fos expression. These mice presented an exacerbated psychomotor sensitization and conditioned place preference induced by low doses of cocaine.

**CONCLUSIONS:** Cocaine induces the rapid induction of Arc and its nuclear accumulation in striatal neurons. Locally, it alters the nucleosomal response, and acts as a brake on chromatin remodeling and gene regulation. These original observations posit Arc as a major homeostatic modulator of molecular and behavioral responses to cocaine. Thus, modulating Arc levels may provide promising therapeutic approaches in drug addiction.

**Keywords:** Addiction, Arc, Cocaine, Extracellular-signal regulated kinase, ERK, Signaling, Striatum

<http://dx.doi.org/10.1016/j.biopsych.2016.05.025>

Long-term behavioral alterations induced by drugs of abuse rely on molecular adaptations within specific brain areas that belong to the reward circuitry (1). Within these structures, early changes in gene expression occur soon after cocaine exposure and set the stage for long-lasting modifications of neuronal activity and behavior (2). This early transcription is characterized by the induction of immediate early genes (IEGs), which encode either transcription factors, including c-Fos, and Zif268, regulating a second wave of genes, or effector proteins acting directly on cellular functions and homeostasis. Activity-regulated cytoskeleton-associated protein (Arc), which belongs to this second category of IEGs, is rapidly induced by cocaine in cortical and striatal regions and cocaine-associated stimuli (3–9). However, the role of striatal

Arc in the development of neuronal and behavioral adaptations to cocaine is not known.

In the hippocampus, Arc is highly regulated by changes in neuronal activity, including high-frequency stimulation of the perforant path or electroconvulsive shock (10–12), thus positioning Arc as a reliable index of activity-dependent synaptic modifications. Because of its characteristic dendritic localization, where it controls glutamate alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid receptor (AMPA) trafficking, synaptic strength, and long-term neuronal plasticity (13–17), much attention has been paid on Arc's functions at synapses. However recent studies indicate that it accumulates within the nucleus (18–20), where its functions remain controversial with some evidence in favor of its positive

impact on transcription (20), although others support a negative role of Arc on transcription (19).

Herein, we identified unexpected functions of Arc in response to cocaine. By studying the dendritic versus nuclear localization of Arc, we made the original observation that cocaine dramatically increases the expression of the protein in the nucleus of medium-sized spiny neurons (MSN), where it is localized within active zones of transcription. Using either overexpression of Arc *in vitro*, or its knockout *in vivo*, we show that Arc acts as a brake on gene regulation and long-term behavioral adaptations induced by cocaine. We conclude that the rapid and transient induction of Arc within the nucleus contributes to the dynamic of chromatin remodeling, and modulates behavioral responses to cocaine. This new facet of Arc's functions in the nucleus is likely to contribute to its homeostatic role within neurons.

## METHODS AND MATERIALS

### Animals and Behavior

Experiments were carried out on 8-week-old C57BL/6 mice, green fluorescent protein (GFP)-Arc knockin (KI) mice, and their wild-type (WT) littermates in accordance with the European Community guidelines on the Care and Use of Laboratory Animals (86/609/EEC). Experiments were approved by the local ethic committee C2EA-05. Locomotor activity was measured as the number of interruptions by the mice of two adjacent beams in a circular corridor (Imetronic, Pessac, France) containing four infrared beams placed at every 90°. After 3 days of habituation, mice were subjected to the psychomotor sensitization protocol consisting of two cocaine injections separated by a one-week interval. Spontaneous activity was recorded for 60 min before the first cocaine injection and locomotor activity was measured for 1 hour. One week later, a second session was performed as described for the first cocaine injection. The conditioned place preference (CPP) was evaluated in a Plexiglas Y-Shaped apparatus (Imetronic) consisting of two compartments distinguished by different patterns on floors and walls, separated by a small neutral area. After a preconditioning during which mice were placed in the neutral area and allowed to explore both chambers, mice were treated for 6 days consecutively with alternate injections of cocaine (2.5 mg/kg) or saline during the so-called conditioning phase. The postconditioning phase was conducted exactly as the preconditioning phase with free access to both chambers. The CPP score was calculated as the difference between the time spent in cocaine-paired chamber during postconditioning minus preconditioning. See the [Supplement](#) for details on behavioral tests.

For measurements of messenger RNA (mRNA) levels, mice were sacrificed at the indicated time after cocaine or saline injections, and dissected striata were snap-frozen before being processed for quantitative reverse-transcriptase polymerase chain reaction. For immunohistochemistry, mice were anesthetized and perfused transcardially with 4% paraformaldehyde. See the [Supplement](#) for details.

### Image Acquisition and Analysis

Immunoreactive cells were quantified in the dorsomedial striatum (DM) and nucleus accumbens shell (ShNAcc). Confocal images

(SP5; Leica, Wetzlar, Germany) were acquired bilaterally with a 40× oil immersion objective. Quantifications were performed using ImageJ software (National Institutes of Health, Bethesda, MD), taking into account the cells with immunofluorescence above a fixed threshold.

### Nucleus Versus Neuropil Fluorescence Intensity Assessment.

Images were acquired as described above. Image analyses were performed in ImageJ using custom-built procedures. Nuclei are segmented by a binarization of the Hoechst signal. The fluorescence was measured inside and outside the nuclei masks to discriminate protein levels in neuropile or in the nucleus. The percent of positive nuclei represents the percent of nuclei above a threshold defined by user relative to the total number of nuclei.

### Arc and Phospho-H3 Colocalization Within the Nucleus.

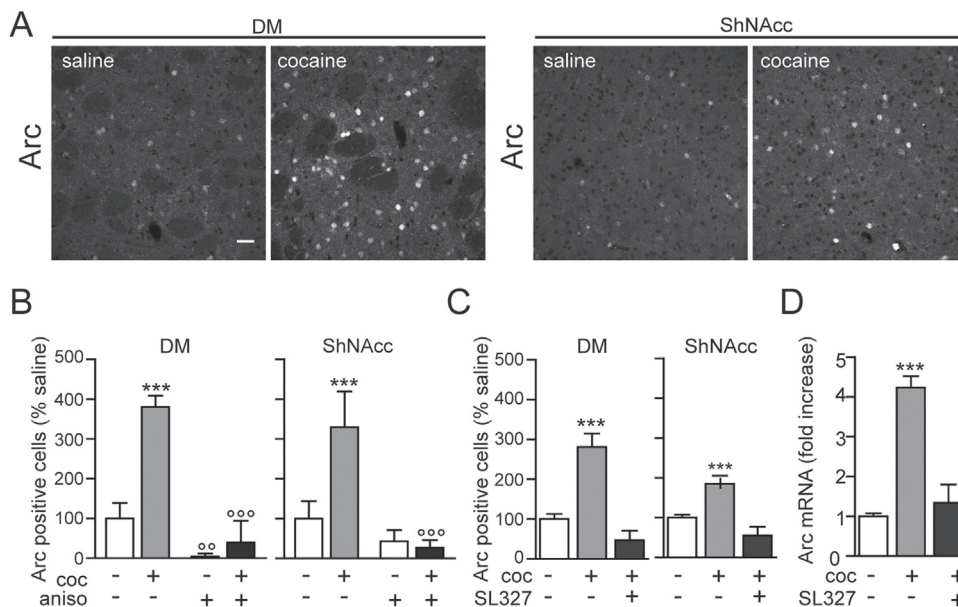
Confocal images were acquired in the DM bilaterally and images were taken with a 63× oil immersion objective, Zoom 4, pixel size:  $x = 0.06 \mu\text{m}$ ,  $y = 0.06 \mu\text{m}$ ,  $z = 0.21 \mu\text{m}$ . A deconvolution step was performed using the maximum likelihood estimation algorithm with Huygens 3.6 Software (Scientific Volume Imaging, Wetzlar, Germany) as described in Heck *et al.* (21). This treatment aims at limiting light diffraction, which increases the accuracy of colocalization analyzes. Three-dimensional segmentation of intranuclear Arc and phosphorylated histone-H3 (pH3) spots at high resolution has been computed thanks to the 3D ImageJ Suite plugins (22) as already described in Heck *et al.* (23). The local maxima inside the nucleus are detected and defined as seeds of the spots. Then an adaptive threshold is automatically calculated for each object based on their intensity by measuring the signal in concentric spheres created around the seeds. This allows detecting the contour in three dimensions automatically without a user-defined threshold (23). Colocalization of Arc and pH3 spots is automatically analyzed using the same plugins, and two spots with overlapping voxels are considered as colocalizing.

## RESULTS

### Cocaine-Induced Arc Expression in the Striatum Relies on Extracellular-Signal Regulated Kinase-Dependent Transcription and De Novo Protein Synthesis

To study the role of Arc in cocaine-induced cellular and molecular adaptations, we first established a precise time-window of its expression by acute cocaine in mice. Both Arc mRNA and protein were transiently induced by cocaine in the DM and the ShNAcc (Figure 1A) and core (not shown) with a peak from 30 to 60 minutes and a return to basal levels at 2 hours (Supplemental Figure S1A–D). This induction occurred downstream from both D<sub>1</sub> (D1R) and N-methyl-D-aspartate glutamate receptors (NMDAR) in MSN expressing c-Fos (Supplemental Figure S1E–G).

The signaling pathways that regulate Arc expression in the striatum in response to cocaine remain to be established. A protein synthesis inhibitor (anisomycin) was administered 30



\*\*\* $p < .01$ , \*\*\* $p < .001$ , vehicle vs. anisomycin pretreated animals. (C) Impact of a pretreatment with SL327 (50 mg/kg) on cocaine-induced Arc protein:  $n = 8$  mice per group; one-way ANOVA in the DM:  $F_{2,21} = 23.54$ ,  $p < .001$ ; and in the ShNAcc,  $F_{2,16} = 13.67$ ,  $p < .001$ ; followed by post hoc comparisons (Bonferroni). \*\*\* $p < .001$ , saline or SL327 pretreated cocaine groups vs. saline pretreated saline group. (D) Arc messenger RNA:  $n = 5$  mice per group; one-way ANOVA:  $F_{2,12} = 32.47$ ,  $p < .001$ , followed by post hoc comparisons (Bonferroni). \*\*\* $p < .001$ , saline or SL327 pretreated cocaine groups vs. saline pretreated saline group and at 45 or 60 min cocaine postinjection, respectively.

minutes before cocaine and mice sacrificed 1 hour later. Cocaine-induced Arc expression was abolished by anisomycin in both the DM and ShNAcc (Figure 1B). Of note, Arc immunoreactivity was also strongly reduced by anisomycin in saline-treated mice, which is consistent with the short half-life of this IEG. A major pathway involved in cocaine-mediated long-term cellular adaptations is the extracellular-signal regulated kinase (ERK) cascade, which behaves as an integrator of D1R and NMDAR signaling (24–27). The administration of SL327, a selective inhibitor of the ERK pathway, before cocaine abolished the induction of Arc mRNA and proteins (Figure 1C, D). Altogether, these data show that Arc mRNA and proteins are transiently induced by acute cocaine in the striatum via an ERK-dependent de novo protein synthesis downstream from both D1R and NMDAR.

### Cocaine Preferentially Increases Arc Expression in the Nucleus of Striatal Neurons

There are accumulating evidences that Arc induction in the dendritic compartment plays a key role in neuronal plasticity (13). However, whether it is induced locally in dendrites in response to cocaine remains unknown. Confocal imaging was performed to determine the impact of cocaine on the cellular localization of Arc. One hour after cocaine, Dil labeling of the dendritic shaft and spines (21) coupled to Arc staining showed that Arc-positive puncta were present in the dendrites and spines (Figure 2A). Nevertheless, a strong immunolabeling was also visible in the nucleus (visualized by Hoechst staining) of striatal neurons (Figure 2B). Arc cellular distribution within the nucleus and outside of it, in the soma and dendritic tree (referred to as neuropile; see Figure 2C), was analyzed using a

computer-based routine that was set up from confocal images of Arc and Hoechst costaining. The Arc signal that colocalized with Hoechst was subtracted from total Arc staining to obtain the neuropile staining (neuropile in Figure 2C). By contrast, the signal colocalized with Hoechst allowed the determination of Arc-positive nuclei (nuclei in Figure 2C). In the DM, total Arc (Figure 2D) and neuropile (Figure 2E) fluorescence intensity significantly increased 60 minutes postcocaine, and then returned to basal levels. Cocaine appeared to have a stronger effect on the nuclear expression of Arc (Figure 2F), which was increased as soon as 30 minutes postcocaine and further augmented at 60 minutes to return to basal levels within 2 hours. Of interest, in the ShNAcc, cocaine only increased the nuclear pool of Arc albeit fewer Arc positive cells were detected than in the DM (Supplemental Figure S2A–D). This indicates that Arc proteins accumulate earlier and preferentially in the nucleus when compared with soma and neuropile in response to cocaine.

### Arc Is Localized at the Vicinity of Active Chromatin Markers in the Nucleus

We studied the precise localization of Arc within the nucleus at high magnification and observed a perfect exclusion of Arc from heterochromatin, which is characterized by puncta with a bright Hoechst staining (Figure 3A). Heterochromatin corresponds to inactive zone of transcription where DNA is highly packed into chromatin, a DNA/protein complex comprising histones and components of the transcriptional machinery. Activity-dependent remodeling of chromatin towards a permissive state for transcription involves posttranslational modifications of histones, also called nucleosomal response.

**Figure 1.** Cocaine-induced activity-regulated cytoskeleton-associated protein (Arc) expression in the striatum relies on extracellular-signal regulated kinase-dependent transcription and de novo protein synthesis. (A) Illustrative confocal images of the Arc staining in the dorsomedial striatum (DM) (left) and nucleus accumbens shell (ShNAcc) (right) from the areas delineated in Supplemental Figure S1B. Scale bar = 30  $\mu$ m. (B) Quantifications of Arc-positive cells in the DM and ShNAcc from mice treated with saline or cocaine (coc) for 1 hour with or without a 30-min pretreatment with 100 mg/kg of anisomycin (aniso);  $n = 4$ –5 mice per group; two-way analysis of variance (ANOVA): interaction between treatment and pretreatment,  $F_{1,12} = 45.47$ ,  $p < .001$ ; effect of treatment,  $F_{1,12} = 74.46$ ,  $p < .001$ ; effect of pretreatment,  $F_{1,12} = 142.64$ ,  $p < .001$ ; followed by post hoc comparisons (Bonferroni's test). \*\*\* $p < .001$ , cocaine group vs. saline group;

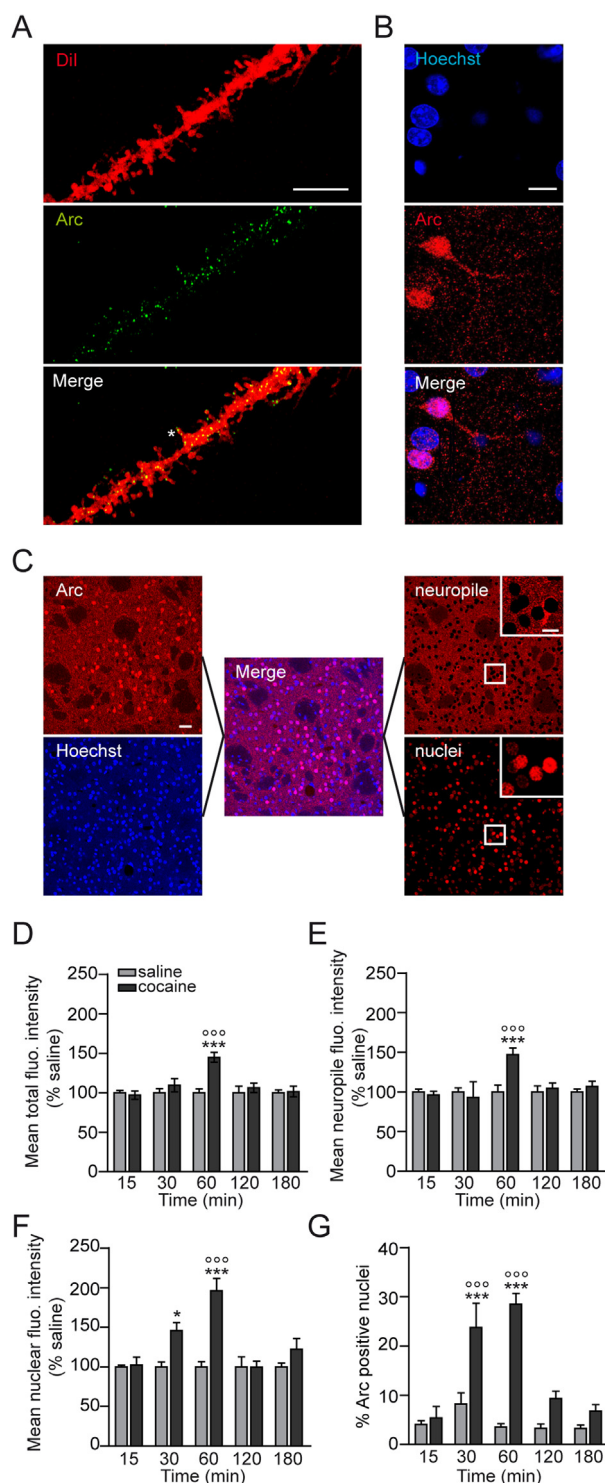


Ser10-Histone pH3 labeling, a marker of nucleosomal response is rapidly (30 minutes) activated downstream ERK and mitogen and stress-activated kinase 1 (MSK-1) in response to cocaine (28). The distribution of pH3 and Arc puncta was analyzed in 3D within the nuclei. At 30 minutes

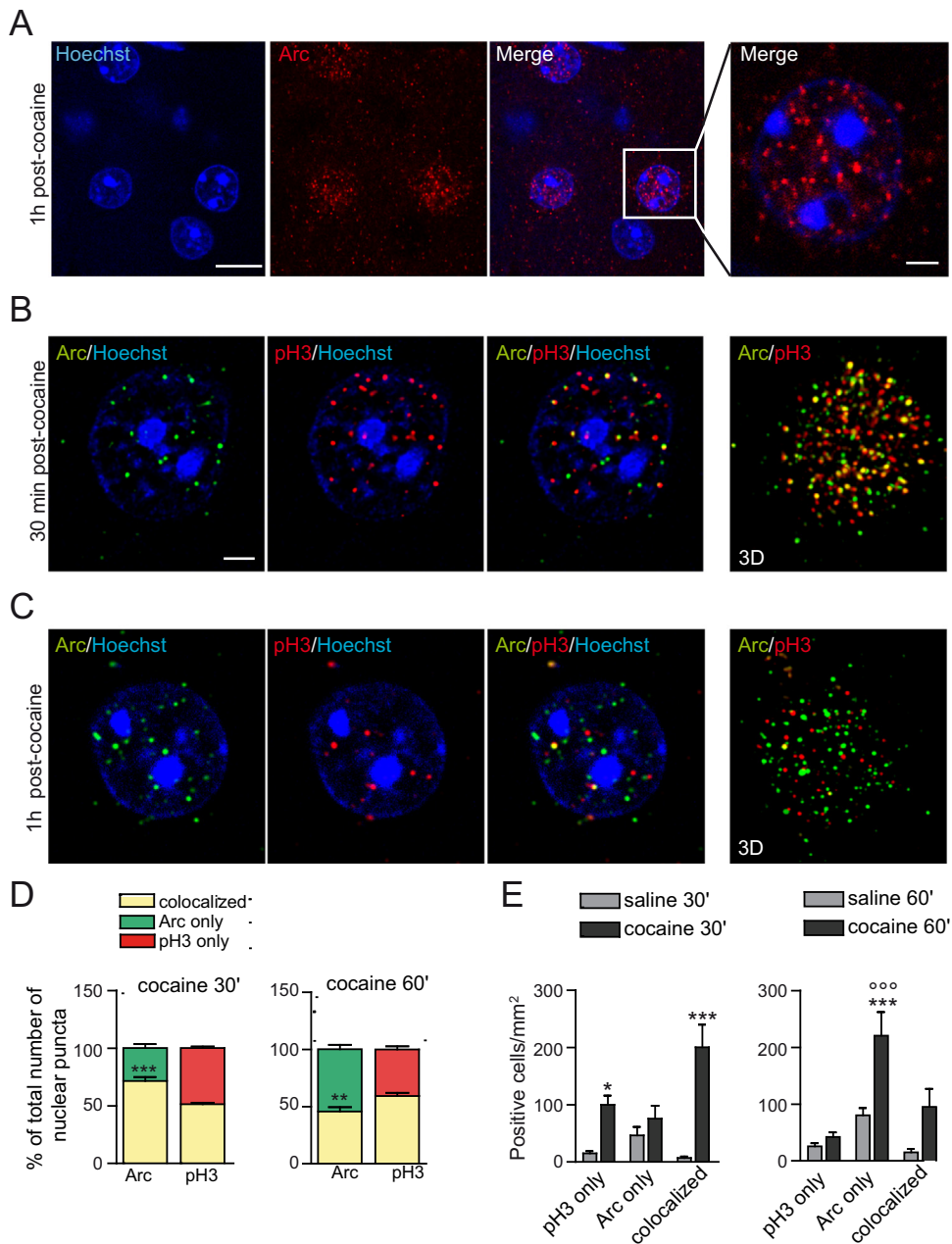
postcocaine, Arc and pH3 strongly colocalized, with 70% of Arc-positive puncta colocalized with pH3 (Figure 3B, C). One hour postcocaine, these Arc/pH3 positive puncta were significantly reduced, with only 50% of colocalization (Figure 3D, E). Consistently, the comparative kinetics of these markers showed that both signals were detectable at 30 minutes postcocaine in the same cells (Figure 3F, left panel), whereas pH3 was decreased at 1 hour meanwhile Arc was maintained (Figure 3F, right panel), in accordance with the decrease of Arc/pH3- colocalization.

### Unraveling the Role of Nuclear Arc in Striatal Neurons In Vitro

Because our data indicated that cocaine-induced Arc accumulation in the nucleus preferentially occurred within active zones of transcription in striatal neurons, we hypothesized that Arc could be a regulator of chromatin remodeling. To test this, we used striatal cultured neurons treated with glutamate (10  $\mu$ M), which induces increases in pH3 levels downstream from ERK-MSK-1 activation (29,30). The impact of Arc expression on these cellular events was studied after transfection of a complementary DNA encoding Arc fused to GFP, followed by live imaging. Under basal conditions, the GFP-Arc signal was detectable in all cellular compartments, with higher



**Figure 2.** Cocaine induces a preferential increase of activity-related cytoskeleton-associated protein (Arc) in the nucleus of striatal neurons. (A) Maximum z projection of a stack of confocal images of a dendritic fragment visualized from a medium-sized spiny neurons stained with Dil (red) and the corresponding Arc labeling (green) detected 1 hour after cocaine administration. The merge of the two signals illustrates that Arc is expressed in the dendritic shaft and in dendritic spines (\*) as expected. Scale bar = 10  $\mu$ m. (B) Single confocal plan from a striatal slice prepared 1 hour after a cocaine administration. Nuclei are counterstained with Hoechst (blue) and Arc is detected in red. The merge shows that Arc is expressed in somatic, dendritic, and nuclear compartments. Scale bar = 10  $\mu$ m. (C) Diagram summarizing the principle of the custom routine used to separate the Arc signal outside and within nuclei. A single confocal plan of Arc (red) and Hoechst (blue) signals is acquired after double staining from the same striatal slice (left panel). Scale bar = 30  $\mu$ m. After automatic detection of the nuclei, the corresponding signal is subtracted from the image acquired in the red channel. The resulting image corresponds to the Arc signal in all subcellular compartments except the nucleus (neuropile) as illustrated in the insert. Scale bar = 10  $\mu$ m. To isolate the Arc signal in the nucleus, the signal corresponding to the nucleus (detected in the blue channel) is kept in the red image. Based on this automated approach the mean intensity of the (D) total, (E) neuropile, and (F) nuclear Arc signals were measured in the dorsomedial striatum and expressed as percentage of increase relative to the corresponding saline condition at each time point.  $n = 6-8$  mice per group; two-way analysis of variance (ANOVA): for total Arc signal: interaction between treatment and time,  $F_{5,54} = 3.56$ ,  $p < .01$ ; effect of treatment,  $F_{1,54} = 6.88$ ,  $p < .05$ ; effect of time,  $F_{5,54} = 4.65$ ,  $p < .01$ , for non-nuclear Arc signal: interaction between treatment and time,  $F_{5,55} = 3.18$ ,  $p < .05$ ; effect of treatment,  $F_{1,55} = 10.25$ ,  $p < .01$ ; effect of time,  $F_{5,55} = 3.18$ ,  $p < .05$ ; for nuclear Arc signal: effect of interaction  $F_{5,55} = 6.072$ ,  $p < .001$ ; effect of treatment,  $F_{1,55} = 20.25$ ,  $p < .001$ ; effect of time,  $F_{5,55} = 6.072$ ,  $p < .001$  followed by post hoc Bonferroni comparisons. \*\*\* $p < .001$ , \* $p < .05$  cocaine group vs. saline group; °°° $p < .001$ , each time point vs. 15-min group. (G) The percentage of Arc-positive nuclei (i.e., above a fixed intensity threshold) has been calculated at each time point after saline or cocaine administration.  $n = 6-8$  mice per group; two-way ANOVA: interaction between treatment and time  $F_{4,48} = 5.95$ ,  $p < .001$ ; effect of treatment,  $F_{1,48} = 34.01$ ,  $p < .001$ ; effect of time,  $F_{4,48} = 12.66$ ,  $p < .001$ , followed by post hoc comparisons (Bonferroni). \*\*\* $p < .001$ , cocaine group vs. saline group; °°° $p < .001$ , each time point vs. 15-min group.



**Figure 3.** Within the nucleus activity-regulated cytoskeleton-associated protein (Arc) is excluded from heterochromatin and localized at the vicinity of an active chromatin marker. **(A)** Representative confocal images obtained from a striatal slice prepared from a mouse that has been sacrificed 1 hour postcocaine injection. Nuclei were counterstained with Hoechst (blue) and Arc is labeled in red. Scale bar = 10  $\mu$ m. Right panel is a close-up of the nucleus with double labeling. Scale bar = 2  $\mu$ m. The fusion of the two channels illustrates the exclusion of Arc puncta from nuclear domains with dense Hoechst labeling that corresponds to inactive zones of transcription (referred to as heterochromatin). Mice were sacrificed 30 min **(B)** or 1 hour **(C)** after saline or cocaine administration and confocal sections were acquired from slices counterstained with Hoechst (blue), Arc (green) and phospho-Ser<sup>10</sup> histone H3 (pH3) (red). Scale bar = 2  $\mu$ m. Right panels are three-dimensional (3D) views of a pile of confocal Arc and pH3 images at these two time points postcocaine. **(D)** The proximity of Arc and pH3 was analyzed in 3D at 30 min postcocaine 5 (left graph);  $n = 17$  nuclei per group; unpaired  $t$  test.  $***p < .001$ , Arc colocalized with pH3 vs. pH3 colocalized with Arc or 1 hour (right graph);  $n = 21$  nuclei per group; unpaired  $t$  test.  $**p < .01$ , Arc colocalized with pH3 vs. pH3 colocalized with Arc. **(E)** The number of cells that are immunoreactive for pH3 or Arc only and the ones that are positive for both Arc and pH3 was assessed by immunohistochemistry from mice that have been sacrificed 30 min (left) or 1 hour (right) after cocaine administration.  $n = 4$ –5 mice per group; unpaired  $t$  test for each marker.  $**p < .01$ ,  $*p < .05$ , cocaine group vs. saline group.

levels in the soma (Figure 4A, left panel). Incubation with glutamate triggered a progressive accumulation of GFP-Arc in the nucleus (Figure 4A, B), thus rendering this in vitro model

ideally suited to evaluate the potential impact of Arc on glutamate-mediated increase in pH3. In response to glutamate, striatal neurons transfected with GFP alone displayed

increased pH3 labeling (Figure 4C [upper panel], D). This response was decreased in neurons expressing GFP-Arc (Figure 4C [lower panel], D), thus indicating that Arc overexpression alters the nucleosomal response and inhibits pH3. This effect was not due to an attenuation of glutamate-induced neuronal activity, as neither ERK1/2 phosphorylation nor its nuclear translocation was altered in the presence of GFP-Arc (Figure 4E, F).

### Deletion of Arc Alters Heterochromatin Domains In Vivo

Our results support that Arc is a potential modulator of chromatin remodeling. To test this in vivo, we took advantage of the GFP-Arc KI mice in which the coding sequence of Arc has been replaced by the GFP gene (31). In homozygous KI mice, Arc is absent and GFP serves as a reporter of Arc promoter's activity. One hour postinjection, WT mice presented an induction of Arc in the DM (Supplemental Figure S3A, B) with no detectable levels of GFP (Supplemental Figure S3C, D). The mirror image was observed in homozygous GFP-Arc KI mice in which cocaine increased GFP but not Arc expression. To study the role of Arc on cocaine-induced chromatin remodeling, we focused on heterochromatin domains that can be detected through Hoechst staining (32). In WT mice, cocaine reduced the number of Hoechst-positive puncta in MSN (Figure 5A–C), along with the area occupied by these puncta in Arc-positive nuclei (Figure 5D, E), thus illustrating that cocaine induces DNA decompaction. In Arc-GFP KI mice a significant reduction of the number (Figure 5A–C) and size (Figure 5D, E) of Hoechst positive puncta were found in saline mice. The number of punctate was even further decreased after cocaine treatment (Figure 5B). This indicates that the removal of Arc alters the basal organization of chromatin, reduces DNA compaction and hence potentially sensitizes neuronal cells to favor transcription.

RNA polymerase II (RNA-Pol II) is a central player of the transcriptional machinery, which activity is influenced by chromatin organization (33). To evaluate the role of cocaine on RNA-Pol II activity, we used an antibody that recognizes the phosphorylated form of RNA-Pol II on its c-terminal domain (34). In WT mice, cocaine triggered a strong increase of RNA-Pol II phosphorylation (Figure 5F, G), which is consistent with the wide spectrum of activity-dependent gene induced by cocaine (2). By contrast, KI mice showed a constitutively high level of RNA-Pol II activity with no further increase detectable upon cocaine exposure (Figure 5F, G). This indicates that Arc has an unexpected broad inhibitory effect on chromatin remodeling and RNA-Pol II-mediated transcription.

### Arc Deficiency Favors the Development of Cocaine-Induced Long-Term Behavioral Alterations

Chromatin remodeling is critical for behavior alterations induced by drugs of abuse (2). Hence, we studied the role of Arc in the development of long-lasting adaptations at the behavioral level in GFP-Arc mice. We first measured the basal locomotor activity of these mice and found that KI mice

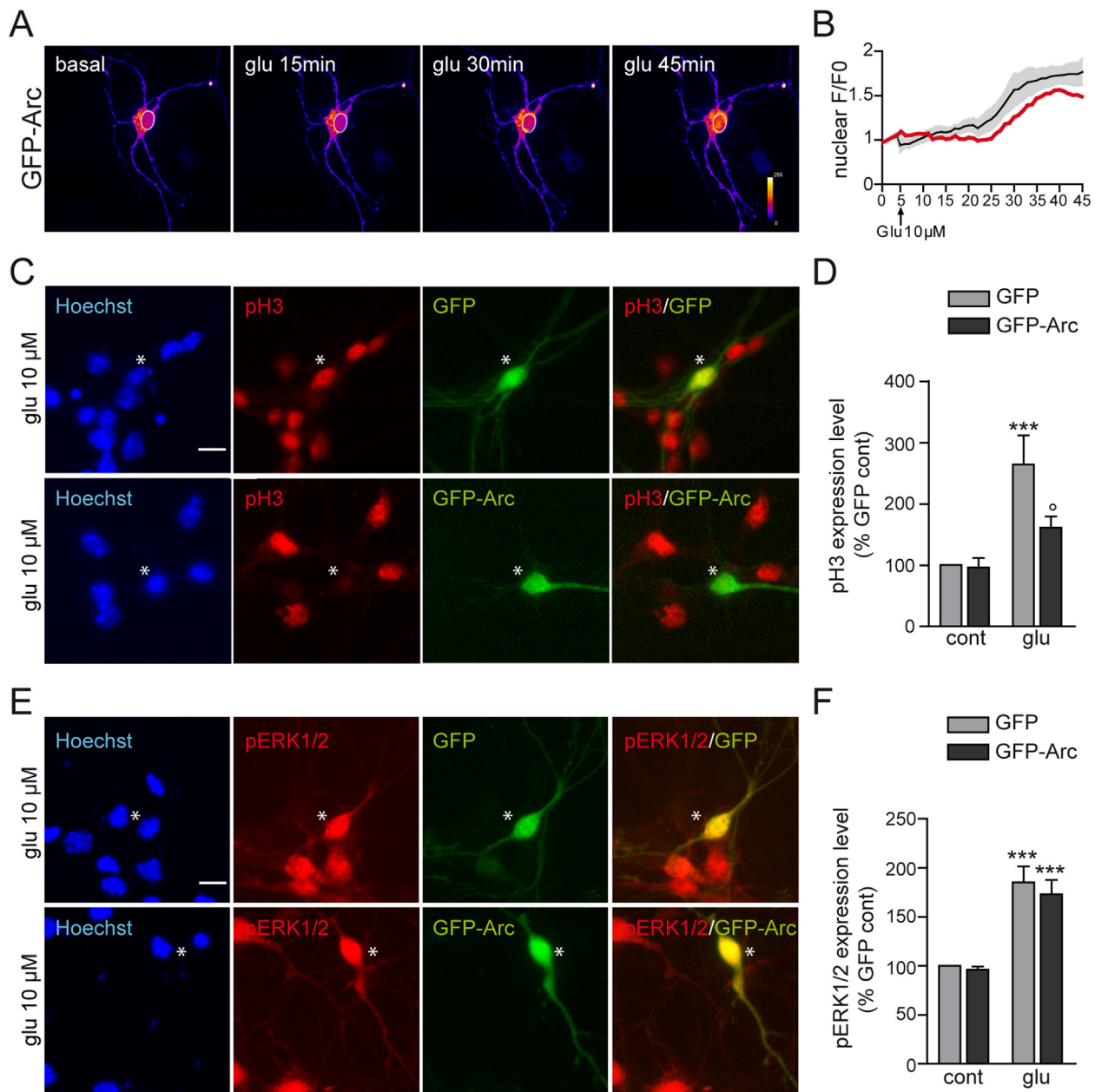
displayed a significantly lower spontaneous locomotion than WT littermates (Figure 6A, B). Mice were then subjected to a protocol of psychomotor sensitization induced by two injections of cocaine at 15 mg/kg with a 1-week interval, which triggers a robust psychomotor sensitization in WT mice (25) (Figure 6C). After the first administration of cocaine (15 mg/kg), the acute locomotor response increased in both genotypes with a higher response in KI mice (Figure 6D). After the second injection, both groups of mice showed a behavioral sensitization with a significantly higher locomotion measured in KI mice (Figure 6D). We then used a lower dose of cocaine (7.5 mg/kg), which triggers a moderate behavioral sensitization in WT mice and may unmask a facilitating effect of Arc deficiency on the development of psychomotor sensitization. In these conditions, KI mice displayed an acute response that was undistinguishable from WT mice, but their sensitized response to the second injection was significantly greater than in WT mice (Figure 6E). These data indicate that the absence of Arc favors the development of psychomotor sensitization. At this low dose of cocaine, pH3 immunoreactivity was not significantly increased, whatever the genotypes (Supplemental Figure S3F). However, the expression of c-Fos, which is known to facilitate psychomotor sensitization (34), was sensitized at this dose, with a stronger increase in KI when compared with WT mice (Figure 6F). This may be due to the observed decrease of DNA compaction in these KI mice, which is further decrease upon exposure to 7.5 mg/kg of cocaine (Figure 6G). The impact of Arc deficiency on behavioral alteration induced by cocaine, was further assessed in the CPP paradigm, which measures the ability of mice to associate the rewarding properties of the drug with environmental cues. With a low dose of 2.5 mg/kg of cocaine, which does not trigger CPP in WT mice, KI mice displayed a clear preference for cocaine-paired chamber in KI mice (Figure 6H, I). Altogether, these data demonstrate that Arc deficiency exacerbates molecular and behavioral alterations induced by cocaine.

## DISCUSSION

The present work demonstrates a nuclear accumulation of Arc in MSN in response to cocaine. Arc was restricted to active zones of transcription where it colocalized with pH3. In vitro, Arc overexpression in cultured striatal neurons altered glutamate-induced pH3, while sparing activation of ERK. In vivo, mice deficient for Arc showed a decrease of heterochromatin domains associated to a high RNA-Pol II activity. These mice showed an exacerbated psychomotor sensitization and CPP induced by low doses of cocaine.

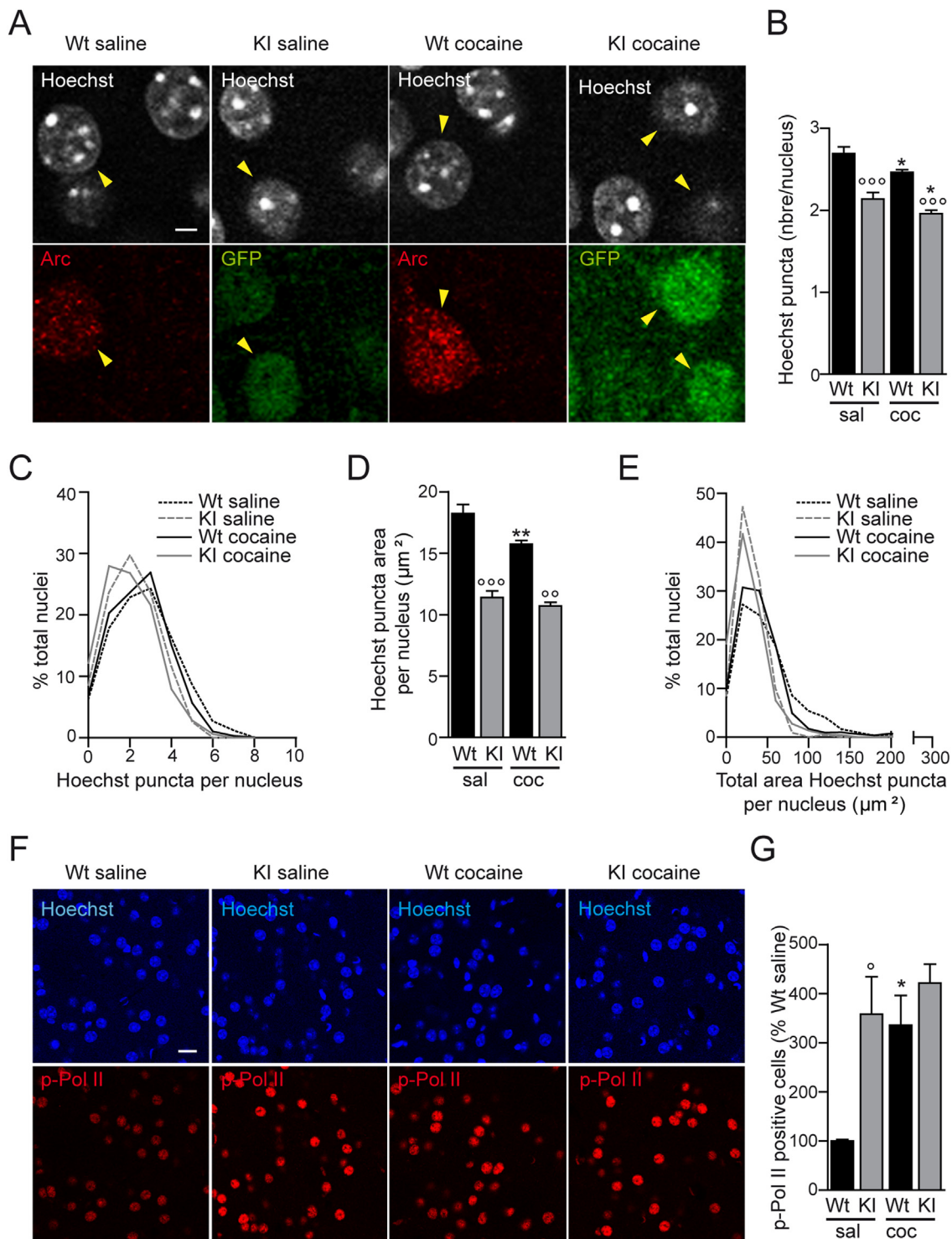
With regard to signaling cascades driving Arc expression, a D1R antagonist was previously shown to abolish Arc induction by acute cocaine (3). Herein, we confirm this observation and further demonstrate that Arc induction also strictly depends on NMDAR stimulation. Cocaine-mediated expression of Arc is thus at the crossroad of dopamine and glutamate inputs converging onto MSN. Consistently, Arc transcription and translation occurred downstream from ERK activity, which behaves as a key integrator of D1R and NMDAR signaling in response to cocaine (24–27,36). The promoter of Arc comprises





**Figure 4.** Overexpression of activity-regulated cytoskeleton-associated protein (Arc) alters histone H3, but not extracellular-signal regulated kinase (ERK) 1/2, phosphorylation in vitro. **(A)** Cultured striatal neurons were transfected with complementary DNA encoding Arc fused to green fluorescent protein (GFP). Live imaging was performed and images were acquired before (basal) and at the indicated time point after application of glutamate 10  $\mu$ M. **(B)** Quantifications of the GFP-Arc fluorescence profile overtime (F/F<sub>0</sub>; error bars are SEM) within the nucleus (white dotted line). The black curve is the average intensity of the nuclear GFP-Arc from 4 transfected neurons including the one shown in **(A)**, represented by the red curve. Note the progressive nuclear accumulation of the GFP-Arc signal in response to glutamate. **(C)** Shown are representative pictures of neurons transfected with GFP (top panels) or GFP-Arc (bottom panels), treated with glutamate (glu) (10  $\mu$ M) for 30 minutes to induce histone Ser<sup>10</sup>-H3 phosphorylation (pH3). Nuclei are counterstained with Hoechst (blue), pH3 is red, GFP in green and the merge of pH3 and GFP is show on the right panels. Scale bar = 10  $\mu$ m. **(D)** Quantifications of the intensity of the pH3 labeling in transfected cells stimulated or not with glutamate. Results are expressed as percentage increase when compared with control neurons transfected with GFP.  $n = 3$ –4 independent experiments per group; two-way analysis of variance (ANOVA): interaction between treatment and transfection,  $F_{1,16} = 3.48$ , not significant (ns); effect of treatment,  $F_{1,16} = 18.54$ ,  $p < .001$ ; effect of transfection,  $F_{1,16} = 4.01$ , ns; followed by post hoc comparisons (Bonferroni).  $***p < .001$ , control group (cont) vs. glu group;  $^{\circ}p < .05$ , GFP vs. GFP-Arc plasmid. **(E)** Illustrations of neurons transfected as described in **(B)** and treated with glu for 10 min to induce the phosphorylation of ERK1/2 (pERK1/2) (red). Scale bar = 10  $\mu$ m. **(F)** Analyses of the intensity of the pERK1/2 labeling in transfected cells stimulated or not with glutamate.  $n = 3$ –4 independent experiments per group; two-way ANOVA: interaction between treatment and transfection,  $F_{1,12} = 0.15$ , ns; effect of treatment,  $F_{1,12} = 54.94$ ,  $p < .001$ ; effect of transfection,  $F_{1,12} = 0.52$ , ns; followed by post hoc comparisons (Bonferroni).  $***p < .001$ , cont group vs. glu group.





two serum response element (SRE) DNA regulatory elements that binds a ternary complex formed by the transcription factor Elk-1 and two molecules of serum response factor. Mutation of one of these SRE sites blunted the late phase of cerebellar long-term depression (37), indicating that SRE-dependent transcription of *Arc* is critical for neuronal plasticity. We previously showed that the phosphorylation of Elk-1 by ERK, which is necessary and sufficient to trigger SRE-dependent transcription (38), was involved in cocaine-mediated *Arc* induction (39). Our data thus support that cocaine-mediated *Arc* transcription critically relies on the ERK/Elk-1 signaling module.

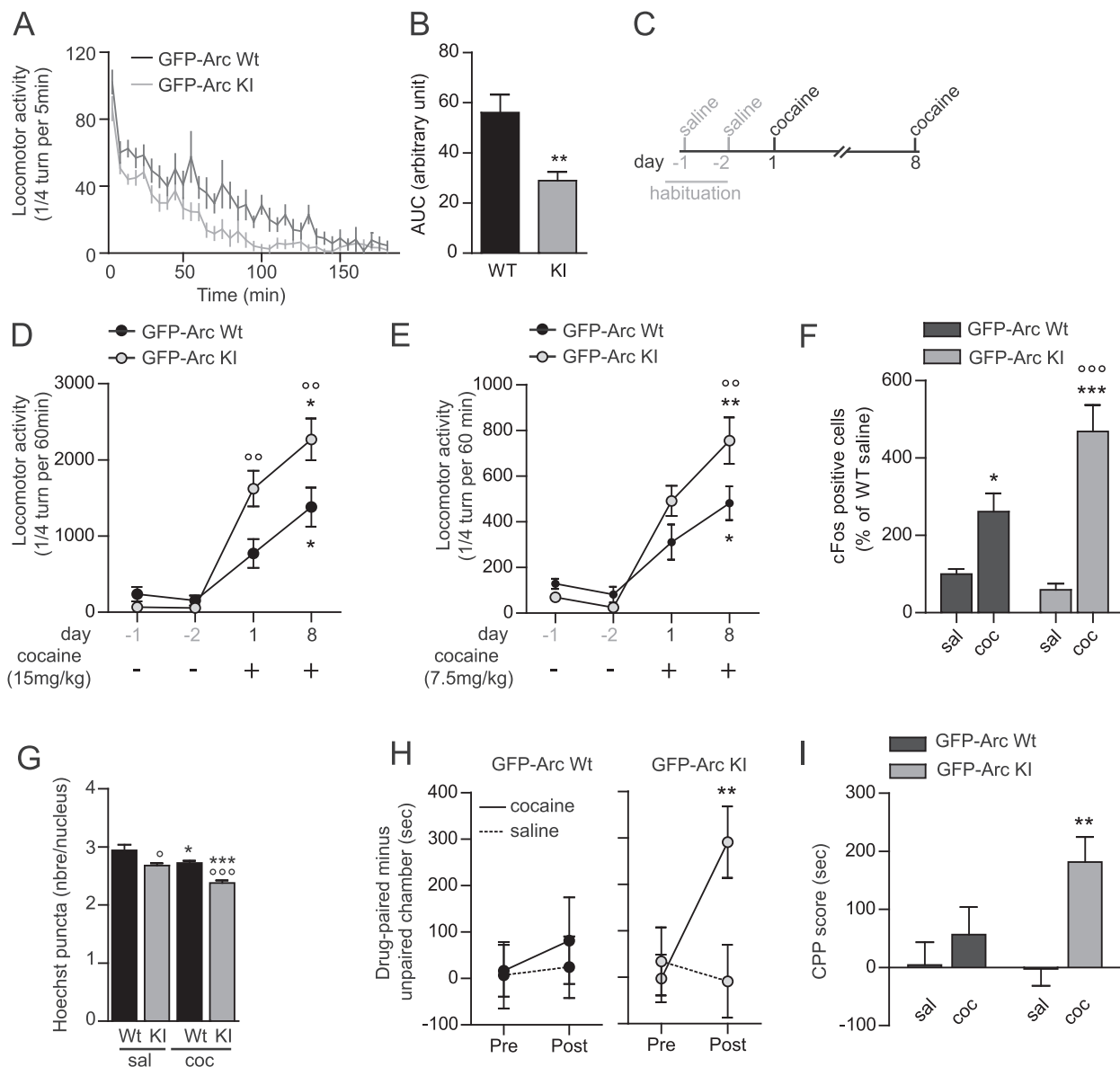
Herein, we demonstrated that *Arc* accumulates in the nucleus of MSN *in vivo* in response to cocaine. *Arc* has been detected in the nucleus of cultured hippocampal neurons (18) where it translocates in an activity-dependent manner (19). Accordingly, we also observed an activity-dependent nuclear accumulation of *Arc* by live imaging of cultured MSN. The role of this nuclear pool of *Arc* remained controversial and emerged only recently, although its functions at synaptic sites have been extensively explored. Initial observations indicated that changes in neuronal activity increased *Arc* in both nuclei and soma, but *Arc* was enriched in dendrites where it coprecipitates with F-actin (40,41). Since then, most of its functions in neuronal plasticity have been described at the level of synapses and involve regulation of cytoskeleton dynamics, AMPAR trafficking and synaptic strength (14–17). Although cocaine increases dendritic *Arc* expression in the DM, we surprisingly found that the major impact of cocaine on *Arc* expression occurred in the nucleus. This was even clearer in the ShNAcc where nuclear *Arc* expression was increased in the absence of change in dendrites. With respect to its biochemical properties and a lack of DNA interaction consensus sequence, *Arc* is not likely to bind DNA or act as a transcription factor in the nucleus (40). What is known so far is that *Arc* binds to nuclear spectrin and promyelocytic leukemia (PML) bodies (19). PML bodies regulate transcription, notably by controlling the availability and histone acetyl transferase activity of cyclic adenosine monophosphate response element binding protein (CREB)-binding protein (CBP) and p300 (42). In cultured hippocampal neurons, increased neuronal activity triggers a nuclear localization of *Arc* within PML bodies, which decreases CBP-mediated transcription of GluA1 AMPAR subunit (19). By contrast, by interacting with the histone acetyl transferase Tip60 at PML bodies *Arc* increases the acetylation of H4K12 (20), a histone mark associated with increased transcription and learning and memory. We report here that *Arc* colocalizes with pH3, within active zones of transcription in the nucleus of MSN after exposure to cocaine. In MSN, this phosphorylation event involves the nuclear kinase MSK-1

downstream from ERK and is critical for gene regulation at specific loci (28,29). Cocaine-induced PH3 *in vivo* also results from DARPP32-induced inhibition of phosphatases (43). *Arc* overexpression in cultured MSN significantly decreased glutamate-induced pH3, thus supporting that *Arc* plays a negative retrocontrol on transcription. This negative feedback is unlikely, due to an effect of *Arc* overexpression on AMPAR trafficking because ERK activation and nuclear translocation, which involve glutamate-induced endocytosis of AMPAR (44), were preserved. By contrast, *Arc* slightly inhibited glutamate-induced MSK-1 activation in the nucleus (Supplemental Figure S2E). However, this inhibitory effect was weaker than the decrease of pH3 itself, and independent on DARPP32 phosphorylation and nuclear translocation (data not shown) supporting that additional mechanisms are engaged by *Arc* to alter pH3. Given the tight relation between *Arc* and actin dynamics in the cytoplasm (41), a tentative explanation could be that *Arc* also interacts with nuclear actin and as such controls the cytoarchitecture of the nucleus and hence chromatin remodeling. Such mechanisms could also account for the altered accessibility of MSK-1 to ERK-mediated phosphorylation in the nucleus.

*In vivo*, acute cocaine triggered a decrease in heterochromatin domains in the nucleus of WT MSN. Persistent decreases in heterochromatin also occur after repeated cocaine, which suggests a potential role of heterochromatin organization in the long-term effects of cocaine (32). The decompaction of DNA induced by cocaine in WT mice was associated with an increased RNA-Pol II activity, which is in agreement with the boost of activity-dependent transcription induced by cocaine (1,2). In both saline- and cocaine-treated KI mice, we found a significant decrease of heterochromatin domains, thereby indicating a relaxing of chromatin in the absence of *Arc*. Upon cocaine administration, the constitutive decrease of heterochromatin was combined with high levels of RNA-Pol II activity, an intracellular event that favors transcription. Altogether, these data support that *Arc* exerts a negative retro-control on transcription, by limiting chromatin decompaction and RNA-Pol II activity. Such impact of *Arc* on these nuclear events may have important implications for its well-known homeostatic functions.

*Arc* expression is correlated to neuronal processing underlying learning and memory storage (45,46) and *Arc*-deficient mice have severe deficit in the consolidation of long-term synaptic plasticity and memory (47). In the striatum, *Arc* induction is associated with early phase of learning as well as inversion phase during reversal learning, suggesting a role in striatal plasticity underlying learning acquisition (48,49). In the context of addiction, *Arc* is increased by cocaine (3–5) or upon re-exposure to a context previously associated with the

**Figure 5.** Deletion of activity-regulated cytoskeleton-associated protein (*Arc*) alters heterochromatin domains and promotes the phosphorylation of polymerase II *in vivo*. **(A)** Homozygous knockin green fluorescent protein (GFP)-*Arc* (KI) and corresponding wild-type (WT) mice were treated with saline (sal) or cocaine (coc) (20 mg/kg) and sacrificed 1 hour postinjection. Shown are representative confocal images acquired in the dorsomedial striatum of nuclei counterstained with Hoechst and immunolabeling of *Arc* or GFP performed from WT and KI mice, respectively. Arrowheads indicate the nuclei that were used for subsequent analyses in which low (saline-treated mice) or high (cocaine-treated mice) levels of *Arc* or GFP were detectable. Scale bar = 2  $\mu$ m. The number of Hoechst-positive puncta per nuclei was analyzed **(B)** as well as the distribution **(C)** of the different categories of nuclei according to the number of Hoechst puncta they display.  $n = 800$ – $1000$  neurons per group; Mann-Whitney *U* test between each group.  $^{*}p < .05$ , saline group vs. cocaine group;  $^{***}p < .001$ , WT vs. KI. The total area corresponding to Hoechst-positive puncta within nuclei (in  $\mu$ m<sup>2</sup>) was measured **(D)** as well as the distribution of nuclei **(E)** depending on the area occupied by heterochromatin domains. 800–1000 neurons per group; Mann-Whitney *U* test between each group.  $^{**}p < .01$ , saline group vs. cocaine group;  $^{***}p < .001$ ,  $^{**}p < .01$ , WT vs. KI. **(F)** WT and KI mice were treated with saline or cocaine and sacrificed 1 hour later to study the phosphorylation of the polymerase II on the Ser<sup>2</sup> of its C-terminal domain (p-Pol II) by immunohistochemistry. Scale bar = 15  $\mu$ m. **(G)** Quantifications p-Pol II positive cells;  $n = 3$ – $4$  mice per group; two-way analysis of variance: interaction between treatment and genotype,  $F_{1,11} = 2.27$ , not significant; effect of treatment,  $F_{1,11} = 6.88$ ,  $p < .05$ ; effect of genotype,  $F_{1,11} = 9.12$ ,  $p < .05$ ; followed by post hoc comparisons (Bonferroni).  $^{*}p < .05$ , saline group vs. cocaine group;  $^{*}p < .05$ , WT vs. KI.



**Figure 6.** Activity-regulated cytoskeleton-associated protein (Arc) deficiency augments behavioral sensitization to low doses of cocaine. **(A)** Time course of basal locomotor activity of green fluorescent protein (GFP)-Arc wild-type (WT) (black), and homozygous knockin (KI) (gray) mice and **(B)** corresponding area under the curve (AUC) in arbitrary unit.  $n = 11$  mice per group; unpaired  $t$  test.  $^{**}p < .01$ , WT vs. KI mice. **(C)** Diagram depicting the psychomotor sensitization protocol induced by two injections in which mice received a first injection of cocaine after a period of habituation and a second injection a week later of the same dose of cocaine. Mice of the two genotypes have been subjected to a protocol of psychomotor sensitization induced by two injections of cocaine at 15 mg/kg **(D)** or 7.5 mg/kg **(E)**.  $n = 11$  mice per group; two-way analysis of variance (ANOVA) (repeated measure over time for matching data): for 15 mg/kg: interaction between genotype and time,  $F_{3,54} = 6.76$ ,  $p < .001$ ; effect of genotype,  $F_{1,54} = 5.62$ ,  $p < .05$ ; effect of time,  $F_{3,54} = 56.25$ ,  $p < .001$ ; for 7.5 mg/kg: interaction between genotype and time,  $F_{3,54} = 5.53$ ,  $p < .01$ ; effect of genotype,  $F_{1,54} = 2.29$ , not significant (ns); effect of time,  $F_{3,54} = 54.94$ ,  $p < .01$  followed by post hoc Bonferroni comparisons.  $^{*}p < .05$ ;  $^{**}p < .01$ , WT vs. KI. **(F)** Homozygous KI mice and corresponding WT littermates treated with saline (sal) or cocaine (coc) (7.5 mg/kg) were sacrificed 30 minutes postinjection and the number of c-Fos positive cells were analyzed in the striatum. Data expressed as percent of the mean of the WT saline group.  $n = 4$ –5 mice per group; two-way ANOVA: interaction between treatment and genotype,  $F_{1,14} = 8.97$ ,  $p < .01$ ; effect of treatment,  $F_{1,14} = 47.59$ ,  $p < .001$ ; effect of genotype,  $F_{1,14} = 4.03$ , ns; followed by post hoc comparisons (Bonferroni).  $^{*}p < .05$ ;  $^{***}p < .001$ , saline vs. cocaine-treated group;  $^{ooo}p < .001$ , WT vs. KI. **(G)** DNA compaction state was assessed in these animals by analyzing the number of Hoechst-positive puncta per nuclei according to the number of Hoechst puncta they display.  $n = 800$ –1000 neurons per group; Mann-Whitney  $U$  test between each group.  $^{*}p < .05$ ,  $^{***}p < .001$ , saline group vs. cocaine-treated group;  $^{*}p < .05$ ;  $^{ooo}p < .001$ , WT vs. KI. **(H, I)** Conditioned place preference (CPP) induced by cocaine ( $n = 11$  mice per group) was performed from WT or homozygous KI mice in a three-pairing CPP paradigm. **(H)** Initial preference during preconditioning session (pre) and place preference on the test session (post) in WT and homozygous KI mice were analyzed using two-way ANOVA (repeated measure over time for matching data): for WT mice: interaction between genotype and time,  $F_{1,44} = 0.11$ , ns; effect of time,  $F_{1,44} = 0.21$ , ns; effect of treatment,  $F_{1,44} = 0.32$ , ns; for KI mice: interaction between genotype and time,  $F_{1,42} = 5.72$ ,  $p < .05$ ; effect of time,  $F_{1,42} = 3.49$ , ns; effect of treatment,  $F_{1,42} = 3.18$ , ns; followed by post hoc comparison (Bonferroni).  $^{*}p < .01$ , cocaine vs. saline group. **(I)** The CPP score (time spent in cocaine-paired chamber during postconditioning minus preconditioning) was analyzed using two-way ANOVA (repeated measure over time for matching data): interaction between genotype and time,  $F_{1,43} = 2.59$ , ns; effect of time,  $F_{1,43} = 2.09$ , ns; effect of treatment,  $F_{1,43} = 8.33$ ,  $p < .01$ ; followed by post hoc comparison (Bonferroni).  $^{**}p < .01$ , cocaine vs. saline group.

drug (6,7). In rats trained to self-administer cocaine, the local inhibition of Arc in the dorsolateral part of the striatum did not affect cocaine seeking but blocked extinction of this behavior (9), a result in agreement with the increased Arc expression during reversal learning and changes of rules (49).

To the best of our knowledge, no causal link has been established between Arc expression and the development of long-term behavioral alterations to cocaine. Arc induction has been correlated to the level of psychomotor sensitization and the rate of cocaine delivery (50). With regard to self-administration, individuals considered as vulnerable showed lower Arc mRNA expression in the DM, whereas resilient rats displayed higher levels (51). Accordingly, we showed here that psychomotor sensitization by low doses of cocaine was higher in KI mice when compared with their WT littermate, revealing a presensitization to the drug in the absence of Arc. This effect was unlikely due to global changes in excitability since KI mice did not show any increase in striatal GluA1 mRNA (Supplemental Figure S3E). By contrast, the expression of c-Fos that facilitates for psychomotor sensitization (35), was significantly higher in KI mice when compared with WT littermate at these low doses of cocaine. Furthermore, at doses of cocaine that did not induce CPP in WT animals, KI mice displayed a clear preference for the cocaine-paired chamber. Altogether these data are in agreement with an inhibitory role of Arc on chromatin decompaction and RNA-Pol II activity at specific loci. These observations strongly support that, upon cocaine administration, the rapid induction of Arc acts as a brake on chromatin remodeling and could contribute to the behavioral homeostatic response to cocaine by limiting activity-dependent transcription of genes. These new facets of Arc's functions suggest that modulating levels of Arc may provide interesting therapeutic approaches in drug addiction.

## ACKNOWLEDGMENTS AND DISCLOSURES

This work was supported by Centre National de la Recherche Scientifique (CNRS), Institut National de la Santé et de la Recherche Médicale (INSERM), University Pierre et Marie Curie Paris VI (UPMC), Agence Nationale pour la Recherche (ANR), Fondation Jérôme Lejeune, Fondation pour la Recherche Médicale (FRM), the Bio-Psy labex cluster of excellence and the Ecole des Neurosciences de Paris (ENP). MS and MDS were the recipients of a fourth-year fellowship from the labex Bio-Psy. ESJ was the recipient of fellowship from the ENP network. LM had a postdoctoral fellowship from the Association Française contre la Myopathie (AFM).

We wish to thank the imaging and animal house facilities of the Institute of Biology Paris Seine as well as Dr. Nicolas Heck for his helpful advice on confocal imaging and Dr. Antoine Besnard for instrumental comments on immunohistochemistry experiments. Authors are also grateful to present and past members of the laboratory for their technical help with the primary cultures, for helpful discussions and critical reading of the manuscript. We also warmly thank CP for all these years spent handling brain slices for all the members of the laboratory with a lot of dexterity and achievement.

The authors report no biomedical financial interests or conflicts of interest.

## ARTICLE INFORMATION

From the INSERM, UMR-S 1130, Neuroscience Paris Seine, Institute of Biology Paris Seine; and Sorbonne Universités, UPMC Université Paris 06, UM CR18, Neuroscience Paris Seine, Paris, France.

JC and PV contributed equally to this work.

Address correspondence to Peter Vanhoutte, Ph.D., Université Pierre et Marie Curie Paris VI, CNRS/UMR 8246, INSERM/UMR-S 1130, Paris, 75005 France; E-mail: peter.vanhoutte@upmc.fr.

Received Dec 21, 2015; revised May 17, 2016; accepted May 31, 2016.

Supplementary material cited in this article is available online at <http://dx.doi.org/10.1016/j.biopsych.2016.05.025>.

## REFERENCES

- Lüscher C, Malenka RC (2011): Drug-evoked synaptic plasticity in addiction : from molecular changes to circuit remodeling. *Neuron* 69: 650–663.
- Robison AJ, Nestler EJ (2011): Transcriptional and epigenetic mechanisms of addiction. *Nat Rev Neuroscience* 12:623–637.
- Fosnaugh JS, Bhat RV, Yamagata K, Worley PF, Baraban JM (1995): Activation of arc, a putative "effector" immediate early gene, by cocaine in rat brain. *J Neurochem* 64:2377–2380.
- Fumagalli F, Bedogni F, Frasca A, Di Pasquale L, Racagni G, et al. (2006): Cortico-striatal up-regulation of activity-regulated cytoskeletal-associated protein expression after repeated exposure to cocaine. *Mol Pharmacol* 70:1726–1734.
- Fumagalli F, Franchi C, Caffino L, Racagni G, Riva MA, Cervo L (2009): Single session of cocaine intravenous self-administration shapes goal-oriented behaviours and up-regulates Arc mRNA levels in rat medial prefrontal cortex. *Int J Neuropsychopharmacol* 12:423–429.
- Hearing MC, See RE, McGinty JF (2008): Relapse to cocaine-seeking increases activity-regulated gene expression differentially in the striatum and cerebral cortex of rats following short or long periods of abstinence. *Brain Struct Funct* 213:215–227.
- Hearing MC, Miller SW, See RE, McGinty JF (2008): Relapse to cocaine seeking increases activity-regulated gene expression differentially in the prefrontal cortex of abstinent rats. *Psychopharmacology (Berl)* 198:77–91.
- Zavala AR, Osredkar T, Joyce JN, Neiswander JL (2008): Upregulation of Arc mRNA expression in the prefrontal cortex following cue-induced reinstatement of extinguished cocaine-seeking behavior. *Synapse* 62:421–431.
- Hearing MC, Schwendt M, McGinty JF (2011): Suppression of activity-regulated cytoskeleton-associated gene expression in the dorsal striatum attenuates extinction of cocaine-seeking. *Int J Neuropsychopharmacol* 14:784–795.
- Link W, Konietzko U, Kauselmann G, Krug M, Schwanke B, Frey U, et al. (1995): Somatodendritic expression of an immediate early gene is regulated by synaptic activity. *Proc Natl Acad Sci U S A* 92:573–578.
- Steward O, Wallace CS, Lyford GL, Worley PF (1998): Synaptic activation causes the mRNA for the IEG Arc to localize selectively near activated postsynaptic sites on dendrites. *Neuron* 21: 741–751.
- Larsen MH, Olesen M, Woldbye DP, Hay-Schmidt A, Hansen HH, Ronn LC, et al. (2005): Regulation of activity-regulated cytoskeleton protein (Arc) mRNA after acute and chronic electroconvulsive stimulation in the rat. *Brain Res* 1064:161–165.
- Bramham CR, Alme MN, Bittins M, Kuipers SD, Nair RR, Pai B, et al. (2010): The Arc of synaptic memory. *Exp Brain Res* 200:125–140.
- Chowdhury S, Shepherd JD, Okuno H, Lyford G, Petralia RS, Plath N, et al. (2006): Arc/Arg3.1 interacts with the endocytic machinery to regulate AMPA receptor trafficking. *Neuron* 52:445–459.
- Shepherd JD, Rumbaugh G, Wu J, Chowdhury S, Plath N, Kuhl D, et al. (2006): Arc/Arg3.1 mediates homeostatic synaptic scaling of AMPA receptors. *Neuron* 52:475–484.
- Guzowski F, Lyford GL, Stevenson GD, Houston FP, McGaugh JL, Worley PF, et al. (2000): Inhibition of activity-dependent arc protein expression in the rat hippocampus impairs the maintenance of long-term potentiation and the consolidation of long-term memory. *J Neurosci* 20:3993–4001.
- Wang MW, Pfeiffer BE, Nosyreva ED, Ronesi JA, Huber KM (2008): Rapid translation of Arc/Arg3.1 selectively mediates mGluR-dependent LTD through persistent increases in AMPAR endocytosis rate. *Neuron* 59:84–97.



18. Bloomer WA, VanDongen HM, VanDongen AM (2007): Activity-regulated cytoskeleton-associated protein Arc/Arg3.1 binds to spectrin and associates with nuclear promyelocytic leukemia (PML) bodies. *Brain Res* 1153:20–33.
19. Korb E, Wilkinson CL, Delgado RN, Lovero KL, Finkbeiner S (2013): Arc in the nucleus regulates PML-dependent GluA1 transcription and homeostatic plasticity. *Nat Neurosci* 16:874–883.
20. Wee CI, Teo S, Oey NE, Wright GD, VanDongen HMA, *et al.* (2014): Nuclear Arc interacts with histone acetyltransferase tip60 to modify H4K12 acetylation. *eNeuro* <http://dx.doi.org/10.1523/ENEURO.0019-14.2014>.
21. Heck N, Betuing S, Vanhoutte P, Caboche J (2012): A deconvolution method to improve automated 3D-analysis of dendritic spines, application to a mouse model of Huntington's disease. *Brain Struct Funct* 217:421–434.
22. Ollion J, Cochenne J, Loll F, Escudé C, Boudier T (2013): TANGO: A generic tool for high-throughput 3D Image analysis for studying nuclear organization. *Bioinformatics* 29:1840–1841.
23. Heck N, Dos Santos M, Amairi B, Sallery M, Besnard A, Herzog E, *et al.* (2014): A new automated 3D detection of synaptic contacts reveals the formation of cortico-striatal synapses upon cocaine treatment in vivo. *Brain Struct Funct* 220:2953–2956.
24. Pascoli V, Besnard A, Hervé D, Pagès C, Heck N, Girault JA, *et al.* (2011): Cyclic adenosine monophosphate-independent tyrosine phosphorylation of NR2B mediates cocaine-induced extracellular signal-regulated kinase activation. *Biol Psychiatry* 69:325–336.
25. Cahill E, Pascoli V, Trifilieff P, Savoldi D, Kappès V, Lüscher C, *et al.* (2014): D1R/GluN1 complexes in the striatum integrate dopamine and glutamate signaling to control synaptic plasticity and cocaine-induced responses. *Mol Psychiatry* 19:1295–1304.
26. Pascoli V, Cahill E, Bellivier F, Caboche J, Vanhoutte P (2014): Extracellular signal-regulated protein kinases 1 and 2 activation by addictive drugs: a signal towards pathological adaptation. *Biol Psychiatry* 76:917–926.
27. Cahill E, Sallery M, Vanhoutte P, Caboche J (2014): Convergence of dopamine and glutamate signaling onto striatal ERK activation in response to drugs of abuse.
28. Bami-Cherrier K, Valjent E, Hervé D, Darragh J, Corvol JC, Pagès C, *et al.* (2005): Parsing molecular and behavioral effects of cocaine in mitogen- and stress-activated protein kinase-1 deficient mice. *J Neurosci* 25:11444–11454.
29. Bami-Cherrier K, Lavour J, Pagès C, Arthur JS, Caboche J (2007): Glutamate induces histone H3 phosphorylation but not acetylation in striatal neurons: role of mitogen- and stress-activated kinase-1. *J Neurochem* 101:697–708.
30. Lavour J, Bernard F, Trifilieff P, Pascoli V, Kappès V, *et al.* (2007): A TAT-DEF-Elk-1 peptide regulates the cytonuclear trafficking of Elk-1 and controls cytoskeleton dynamics. *J Neurosci* 27:14448–14458.
31. Wang KH, Majewska A, Schummers J, Farley B, Hu C, Sur M, *et al.* (2006): In vivo two-photon reveals a role of arc in enhancing orientation specificity in visual cortex. *Cell* 126:389–402.
32. Maze I, Feng J, Wilkinson MB, Sun H, Shen L, Nestler EJ (2011): Cocaine dynamically regulates heterochromatin and repetitive element unsilencing in the nucleus accumbens. *Proc Natl Acad Sci U S A* 108:3035–3040.
33. Sims RJ 3rd, Mandal SS, Reinberg D (2004): Recent highlights of RNA-polymerase-II-mediated transcription. *Curr Opin Cell Biol* 16:263–271.
34. Meinhart A, Kaminski T, Hoepfner S, Baumli S, Cramer P (2005): A structural perspective of CTD function. *Genes Dev* 19:1401–14015.
35. Zhang J, Zhang L, Jiao H, Zhang Q, Zhang D, Lou D, *et al.* (2006): c-Fos facilitates the acquisition and extinction of cocaine-induced persistent changes. *J. Neurosci.* 26:13287–13296.
36. Girault JA, Valjent E, Caboche J, Hervé D (2007): ERK2: a logical and gate critical for drug-induced plasticity? *Curr Opin Pharmacol* 7:77–85.
37. Smith-Hicks C, Xiao B, Deng R, Ji Y, Zhao X, Shepherd JD, *et al.* (2010): SRF binding to SRE 6.9 in the Arc promoter is essential for LTD in cultured Purkinje cells. *Nat Neurosci* 13:1082–1089.
38. Besnard A, Galan-Rodriguez B, Vanhoutte P, Caboche J (2011): Elk-1 a transcription factor with multiple facets in the brain. *Front Neurosci* 5:35.
39. Besnard A, Bouveyron N, Kappès V, Pascoli V, Pagès C, Heck N, *et al.* (2011): Alterations of molecular and behavioral responses to cocaine by selective inhibition of Elk-1 phosphorylation. *J Neurosci* 31:14296–14307.
40. Lyford GL, Yamagata K, Kaufmann WE, Barnes CA, Sanders LK, Copeland NG, *et al.* (1995): Arc, a growth factor and activity-regulated gene, encodes a novel cytoskeleton-associated protein that is enriched in neuronal dendrites. *Neuron* 14:433–445.
41. Messaoudi E, Kanhema T, Soulé J, Tiron A, Dagyte G, da Silva B, *et al.* (2007): Sustained Arc/Arg3.1 synthesis controls long-term potentiation consolidation through regulation of local actin polymerization in the dentate gyrus in vivo. *J Neurosci* 27:10445–10455.
42. Boisvert FM, Kruhlak MJ, Box AK, Hendel MJ, Bazett-Jones DP (2001): The transcription coactivator CBP is a dynamic component of the promyelocytic leukemia nuclear body. *J Cell Biol* 152:1099–1106.
43. Stipanovich A, Valjent E, Matamalas M, Nishi A, Ahn JH, Maroteux M, *et al.* (2008): A phosphatase cascade by which rewarding stimuli control nucleosome response. *Nature* 453:879–884.
44. Trifilieff P, Lavour J, Pascoli V, Kappès V, Bami-Cherrier K, Pagès C, *et al.* (2009): Endocytosis controls glutamate-induced nuclear accumulation of ERK. *Mol Cell Neurosci* 41:325–336.
45. Guzowski JF, McNaughton BL, Barnes CA, Worley PF (1999): Environment-specific expression of the immediate-early gene Arc in hippocampal neuronal ensembles. *Nat Neurosci* 2:1120–1124.
46. Guzowski JF, Setlow B, Wagner EK, McGaugh JL (2001): Experience-dependent gene expression in the rat hippocampus after spatial learning: a comparison of the immediate-early genes Arc, c-fos, and Zif 268. *J Neurosci* 21:5089–5098.
47. Plath N, Ohana O, Dammermann B, Errington ML, Schmitz D, Gross C, *et al.* (2006): Arc/Arg3.1 is essential for the consolidation of synaptic plasticity and memories. *Neuron* 52:437–444.
48. Kelly MP, Deadwyler SA (2003): Experience-dependent regulation of the immediate-early gene Arc differs across brain regions. *J Neurosci* 23:6443–6451.
49. Daberkow DP, Reidy MD, Kesner RP, Keefe KA (2007): Arc mRNA induction in striatal efferent neurons associated with response learning. *Eur J Neurosci* 26:228–241.
50. Samaha AN, Mallet N, Ferguson SM, Gonon F, Robinson TE (2004): The rate of cocaine administration alters gene regulation and behavioral plasticity: implications for addiction. *J Neurosci* 24:6362–6370.
51. Quinn RK, Brown AL, Goldie BJ, Levi EM, Dickson PW, Smith DW, *et al.* (2015): Distinct miRNA expression in dorsal striatal subregions is associated with risk for addiction in rats. *Transl Psychiatry* 5:e503.