



**HAL**  
open science

## **A possible alternative method for collecting mosquito larvae in rice fields**

Vincent Robert, Gilbert Le Goff, Frédéric Arieu, Jean-Bernard Duchemin

### ► **To cite this version:**

Vincent Robert, Gilbert Le Goff, Frédéric Arieu, Jean-Bernard Duchemin. A possible alternative method for collecting mosquito larvae in rice fields. *Malaria Journal*, 2002, 1 (1), pp.4. <10.1186/1475-2875-1-4>. <hal-03362645>

**HAL Id: hal-03362645**

**<https://hal.science/hal-03362645v1>**

Submitted on 1 Oct 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



HAL Authorization

Methodology

## A possible alternative method for collecting mosquito larvae in rice fields

Vincent Robert\*<sup>1,2</sup>, Gilbert Le Goff<sup>1,3</sup>, Frédéric Arie<sup>1</sup> and Jean-Bernard Duchemin<sup>1</sup>

Address: <sup>1</sup>Groupe de Recherche sur le Paludisme, Institut Pasteur de Madagascar, <sup>2</sup>UR Paludologie afro-tropicale, Institut de Recherche pour le Développement and <sup>3</sup>UR Caractérisation et contrôle des populations de vecteurs, Institut de Recherche pour le Développement

E-mail: Vincent Robert\* - robert@pasteur.mg; Gilbert Goff - legoff@pasteur.mg; Frédéric Arie - arie@pasteur.mg; Jean-Bernard Duchemin - duchemin@pasteur.mg

\*Corresponding author

Published: 8 April 2002

*Malaria Journal* 2002, 1:4

Received: 1 March 2002

Accepted: 8 April 2002

This article is available from: <http://www.malariajournal.com/content/1/1/4>

© 2002 Robert et al; licensee BioMed Central Ltd. Verbatim copying and redistribution of this article are permitted in any medium for any purpose, provided this notice is preserved along with the article's original URL.

### Abstract

**Background:** Rice fields are efficient breeding places for malaria vectors in Madagascar. In order to establish as easily as possible if a rice field is an effective larval site for anophelines, we compared classical dipping versus a net as methods of collecting larvae.

**Results:** Using similar collecting procedures, we found that the total number of anopheline larvae collected with the net was exactly double (174/87) that collected by dipping. The number of anopheline species collected was also greater with a net.

**Conclusions:** The net is an effective means of collecting anopheline larvae and can be used for qualitative ecological studies and to rapidly determine which rice fields are containing malaria vectors.

### Introduction

The role of rice fields as breeding places for anophelines has already been established in the highlands of Madagascar [1–3] (Figure 1 and 2). However, the method of collecting anopheline larvae in rice fields has not been addressed. We compared two simple methods to determine which was the most efficient way to collect anopheline larvae from the rice fields.

### Materials and Method

The first method was based on the classical dipping method with a white tray (20 x 15 x 3 cm) [4] (Figure 3). The second design consisted of using a small nylon gauze net mounted on a circular frame (15 cm diameter) and attached to a wooden handle (1 m long) (Figure 4). The fine

gauze prevents the loss of small larvae. We had initially intended to draw the net through the water for measured distances to estimate larval densities, but the time taken for the bag to become clogged was highly variable, depending on the substrate and the depth of the water. In practice then, the net was drawn through the water until it became clogged with particles and sludge. It was then removed from the water and emptied into a large white tray (30 x 22 x 4 cm). The next-step of both methods was to use a pipette to remove all the mosquito larvae that were at least at the second instar stage.

The study was carried out by two experienced technicians who simultaneously collected the same rice field for 20 minutes. The larvae were then immediately fixed in 70%



**Figure 1**  
General view of rice fields at the end of rainy season in Madagascar.



**Figure 2**  
General view of rice fields at the end of dry season in Madagascar.

ethanol before being studied under the microscope (note that we were unable to classify six of the 2<sup>nd</sup> instar larvae in the genus *Culex* to species level). This study was carried out in Madagascar, in the Fianarantsoa province and both methods were repeated three times in different rice fields: Rice field A, 3 km W Amborompotsy, S 20° 36.6', E 46° 16.1', alt 1068 m, Rice field B, 15 km E Amborompotsy, S 20° 35.9', E 46° 19.4', alt 1266 m, and Rice field C, 10 km W Ambositra, 1 km N Andina, S 20° 30.8', E 47° 09.0', alt 1326 m. No vector control programme exists in locations A and B, but the third rice field is located in an area in which DDT has been sprayed for several years. The approximate depth of the water was 5–10 cm.



**Figure 3**  
Illustration of the dipping method.



**Figure 4**  
Illustration of the net method.

## Results

A total of 363 mosquito larvae belonging to three genera were collected. The results for both collection methods are summarised in Table 1 and exhaustively presented in Table 2 (see additional file: tabl 2 vr.xls).

Given similar conditions a net is a much more efficient way to collect anophelines than dipping. This was the case for the total number of mosquitoes collected, the total number of anophelines collected, the number of each anopheline instar, the efficiency of collecting anophelines and the total number of Culicinae collected but not for the total number of *Culex* collected. This was also the case for the total number of pupae (results not shown). The total number of anopheline larvae collected with the net was exactly double (174/87) that collected by dipping.

**Table 1: Number of mosquito larvae collected by dipping or with a net in 3 rice fields (locations of the A, B and C rice fields are indicated in the Materials and Method).**

Rice field	Aspect of the rice	Method of collect	Total <i>Anopheles</i>	Total <i>Culex</i>	Total <i>Uranotaenia</i>	Total mosquitoes	Anophelines / total mosquitoes
A	green ears	Dipping	6	20	8	34	18%
		Net	18	6	15	39	46%
B	yellow ears	Dipping	13	3	0	16	81%
		Net	48	20	0	68	71%
C	no ears	Dipping	68	18	1	87	78%
		Net	108	10	1	119	91%
Total		Dipping	87	41	9	137	64%
		Net	174	36	16	226	77%

Nine *Anopheles* species and eight *Culex* species were collected.

The anopheline species collected were *An. squamosus*, *An. coustani* (or *coustani* group), *An. pretoriensis*, *An. cydippis*, *An. gambiae* s.l., *An. maculipalpis*, *An. funestus*, *An. mascarensis* and *An. rufipes*. The first five species were obtained by both collecting methods. *An. maculipalpis* was only obtained with the net (5 larvae). The three remaining anopheline species were collected at low number: 2 larvae of *An. funestus* with the net, 1 larva of *An. mascarensis* with the net, and 2 larvae of *An. rufipes* among which 1 with the net and 1 with the dipping. It is also noteworthy that the two *An. funestus* larvae were both collected from rice field B, which contained dense and high rice plants. Conversely the four *An. gambiae* s.l. larvae were all found in rice field C, which did not contain emergent vegetation and was exposed to the sun. This is fully consistent with the classical characteristics of larval habitats for these malaria vector species.

Different results were observed with *Culex* larvae. Only two of the eight *Culex* species collected were collected by both methods (*Cx. antennatus* and *Cx. striatipes*, with 36 and 2 larvae, respectively), four species were only collected by dipping (*Cx. univittatus*, *Cx. quinquefasciatus*, *Cx. tigripes* and *Cx. bitaeniorhynchus*, with 5, 3, 2 and 2 larvae, respectively) and two species were only collected with the net (*Cx. argenteopunctatus* and *Cx. poecilipes*, with 19 and 3 larvae, respectively). In other words the two collecting methods are complementary for the collection of *Culex* larvae, and we recommend that both methods be used when focusing on this genus.

Data on age structure of larvae were detailed in Table 2 (see additional file: tabl 2 vr.xls). Age structure was significantly different between *Anopheles* and *Culex* with the dipping ( $\chi^2 = 6.317$ ,  $df=2$ ,  $P = 0.042$ ) and with the net ( $\chi^2$

$= 16.701$ ,  $df=2$ ,  $P = 0.0002$ ) strongly suggesting that both methods collected older *Culex* and/or missed younger *Anopheles*. Age structure was also analysed within each genus by collecting methods. In *Anopheles*, the comparison of the proportions of each instar relatively to the total collected were always non significant ( $P = 0.087$  by exact Fisher's test for the 4<sup>th</sup> instar,  $P = 0.096$  for the 3<sup>rd</sup> instar and  $P = 0.77$  for the 2<sup>nd</sup> instar). The comparison of the relative proportion of 4<sup>th</sup> and 3<sup>rd</sup> instars was near significance ( $P = 0.053$  by exact Fisher's test) suggesting that the net would collect smaller anopheline larvae that were being missed with the dipping; however, the comparison of the relative proportion of 3<sup>rd</sup> and 2<sup>nd</sup> instars did not support this interpretation ( $P = 0.45$ ). In *Culex*, these analyses were always non significant, indicating a similar efficiency of both methods per age group of larvae.

Four species of *Uranotaenia* sp. were obtained, but the low numbers collected prevented any valuable analysis.

## Discussion

These results, which clearly favour the using of the net for collecting anopheline larvae, can be explained by two non-exclusive hypotheses. The volume of water examined was higher with the net. Consequently, the number of materials, including larvae, per tray was much higher (once remembers the net content was emptied into a large tray) and the time spent per tray by the observer was much longer which gave the larvae more time to reach the surface of the water, facilitating their detection with the net method.

The question of differences in collecting *Anopheles* versus *Culex* remains unclear, but clearly it cannot be related to instars. Because *Culex* are relatively underrepresented with the net method, an explanation may reside in the specific escape behaviour of these mosquitoes.

In conclusion, the net is an efficient and simple way of collecting anopheline larvae. Further studies are necessary to validate this method for quantitative evaluation of larval populations. However, the net can be used for qualitative ecological studies and to rapidly determine which rice fields are containing malaria vectors such as *An. gambiae* s.l., *An. funestus*, and at least in Madagascar, *An. mascarensis*.

## Additional material

### Table 2

Table 2. Exhaustive data presented by method of collection, species and instars.

tabl 2 vr.xls

[<http://www.biomedcentral.com/content/supplementary/1475-2875-1-4-S1.xls>]

## Acknowledgements

We thank Donadieu Randrianambinina and Edmond Rajaonarivelo for their helpful contributions in the field and in the laboratory. Dr Isabelle Jeanne kindly provided Figure 1.

## References

1. Fontenille D, Rakotoarivony I: **Reappearance of *Anopheles funestus* as a malaria vector in the Antananarivo region, Madagascar.** *Trans R Soc Trop Med Hyg* 1988, **82**:644-645
2. Laventure S, Mouchet J, Blanchy S, Marrama L, Rabarison P, Andrianaivolambo L, Rajaonarivelo E, Rakotoarivony I, Roux J: **Le riz source de vie et de mort sur les plateaux de Madagascar.** *Cahiers Santé* 1996, **6**:79-86
3. Marama L, Rajaonarivelo E, Laventure S, Rabarison P: ***Anopheles funestus* et la riziculture sur les plateaux de Madagascar.** *Cahiers Santé* 1995, **5**:415-419
4. Awono-Ambene HP, Robert V: **Survival and emergence of immature *Anopheles arabiensis* in market-gardener in Dakar, Senegal.** *Parasite* 1999, **6**:179-184

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMedcentral will be the most significant development for disseminating the results of biomedical research in our lifetime."

Paul Nurse, Director-General, Imperial Cancer Research Fund

Publish with **BMC** and your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours - you keep the copyright

Submit your manuscript here:

<http://www.biomedcentral.com/manuscript/>



**BioMedcentral.com**

[editorial@biomedcentral.com](mailto:editorial@biomedcentral.com)