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Cav1.4 calcium channels control cytokine production by human peripheral Th17-cells and psoriatic skin-infiltrating T cells

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32 Abstract

33 Background: Type-17 inflammation characterizes psoriasis, a chronic skin disease. As several inflammatory cytokines contribute to psoriasis pathogenesis, inhibiting the simultaneous 34 production of these cytokines in Th17-cells may be beneficial in psoriasis. We found that 35 Ca_v1.4, encoded by CACNA1F, was the only Ca_v1 calcium channel expressed in Th17-cells. 36 **Objective**: We investigated the role of Ca_v1.4 expression in early Th17-activation events and 37 effector functions, as well as its association with Th17 signature genes in lesional psoriatic (LP) 38 skins. Methods: Transcriptional gene signatures associated with CACNAIF expression were 39 examined in LP skins by RT-PCR and in situ hybridization. Cav1 inhibitor and/or shRNA 40 41 lentivectors were used to assess the contribution of Ca_v1.4 on Th17 activation and effector functions in a 3D skin reconstruction model. Results: CACNA1F expression correlated with 42 inflammatory cytokine expression that characterizes LP skins and was preferentially associated 43 44 with *RORC* expression in CD4⁺ and CD4⁻ cells from LP biopsies. Nicardipine, a Ca_v1 channel antagonist, markedly reduced inflammatory cytokine production by Th17-cells from blood or 45 LP skin. This was associated with decreased TCR-induced early calcium events at cell 46 membrane and proximal signaling events. The knockdown of Cav1.4 in Th17-cells impaired 47 cytokine production. Finally, Cav1 inhibition reduced the expression of the keratinocyte genes 48 characteristic of Th17-mediated psoriasis inflammation in human skin equivalents. 49 **Conclusion**: Ca_v1.4 channels promote Th17-cell functions both at the periphery and in 50 inflammatory psoriatic skin. Clinical implication: These data open a new field of therapy based 51 on the development of drugs that inhibit Ca_v1.4 calcium channels in the treatment of psoriasis. 52

53 Capsule Summary

- 54 Ca_v1.4 calcium channels selective for Th17 among effector T-cells are detected in skin psoriasis
- 55 lesions and their inhibition reduces inflammatory cytokine production, opening a new field in
- 56 psoriasis treatment.
- 57
- 58 Key words
- 59 Psoriasis, calcium channels, Ca_v1.4, inflammatory cytokine, Th17 cells, TCR activation

60 Abbreviations

- 61 AD: atopic dermatitis
- 62 Ca^{2+} : calcium
- 63 CsA: ciclosporin A
- 64 DC: dendritic cells
- 65 ECE: Elementary calcium events
- 66 GMFI: geometric mean fluorescence intensity
- 67 IHF: immunohistofluorescence
- 68 ISH: in situ hybridization
- 69 HD: healthy donor
- 70 LP: lesional psoriasis skin
- 71 NLP: non-lesional psoriasis skin
- 72 PBMC: peripheral blood mononuclear cell
- 73 PP: psoriasis patient
- 74 SE: skin equivalent

75 Introduction

Psoriasis is a chronic inflammatory skin disease frequently associated with comorbidities, 76 including arthritis, cardiometabolic disease, and a high emotional burden. It affects 2-3% of the 77 world's population. The skin lesions present as erythematous scaly patches characterized by 78 keratinocyte hyperproliferation and infiltration of the epidermis and dermis by inflammatory 79 cells^{1, 2}. Psoriasis results from the abnormal dialog between innate and adaptive immune cells, 80 with an important role for T-helper 17 (Th17) cells³. Skin injury causes cell death and the 81 release of self DNA-anti-microbial products such as LL37, which can activate plasmacytoid 82 dendritic cells (DC) in the skin, resulting in interferon α (IFN α) production and DC activation. 83 Activated DCs migrate into the draining lymph node cells, where they polarize a population of 84 self-reactive CD4⁺ T lymphocytes producing interleukin (IL) 17, IL22, IFNy and tumor necrosis 85 factor α (TNF α) through the secretion of IL23 mainly and IL12⁴. These effector T-cells are 86 recruited into the skin and produce inflammatory cytokines that induce keratinocyte 87 proliferation and promote a positive feedback loop between the epithelial and immune cells, 88 resulting in sustained inflammation^{3, 5}. 89

In accordance with this pathophysiological scheme, targeting the TNFα/IL23/IL17 cytokine axis is effective in the treatment of psoriasis⁶⁻⁸. Although neutralizing anti-cytokine antibodies are protective in psoriasis, they present some disadvantages in terms of cost, administration route, and potential adverse events^{9, 10}. As Th17-derived cytokines are key effectors of psoriasis pathogenesis¹¹, developing new therapies to reduce their productions could be beneficial in psoriasis.

In this context, we focused on the singularities of signaling pathways that control cytokine production in T-helper cells. We previously reported the discriminative expression of Ca_v1 calcium (Ca²⁺) channel family members in different T-cell populations¹²⁻¹⁴. Ca_v1 channels are a multi-protein complex composed of a pore-forming α 1 subunit encoded by four genes 100 (*CACNA1S*, *CACNA1C*, *CACNA1D* and *CACNA1F* encoding for the Ca_v1.1 to Ca_v1.4 channels, 101 respectively), and auxiliary subunits β and $\alpha 2-\delta$ (each encoded by four genes)¹⁵. In T 102 lymphocytes, Ca_v1 expression is plastic depending on their activation and differentiation 103 status^{13, 14, 16}. The Ca_v1.2 and Ca_v1.3 channels are selectively expressed in mouse and human 104 Th2-cells¹²⁻¹⁴ and Ca_v1.4 is the only Ca_v1 calcium channel to be expressed in Th17 cells from 105 the peripheral blood of healthy donors (HD)¹⁴. Nicardipine, a Ca_v1 channel antagonist, 106 decreased TCR-induced IL17A production by human Th17 (CCR6⁺) cells¹⁴.

In the present study, we highlighted the presence of $Ca_v 1.4$ calcium channels in ROR γt^+ CD4⁺ T cells from lesional psoriatic (LP) skin biopsies. $Ca_v 1$ inhibition dampened inflammatory cytokine production by Th17, regardless of their origin (blood or LP). $Ca_v 1$ channels acted early during Th17 activation contributing to the elementary Ca^{2+} events (ECE) preceding the global Ca^{2+} response. Finally, the $Ca_v 1$ blocker, nicardipine was beneficial in a 3D model of psoriasis-like reconstructed skin induced by the infiltration of activated Th17cells.

114 Methods

115

116 Healthy donors and patients

This study complied with the ethical principles of the Declaration of Helsinki and with 117 applicable French regulations. All human participants provided written informed consent. 118 PBMCs from blood of anonymous, healthy donors (HD, 18 to 65 year-old male) were from the 119 Toulouse blood transfusion center (Etablissement Français du Sang). Study (n°2016-A00863-120 48) from psoriasis patients (PP) and atopic dermatitis (AD) patients were accepted by the 121 Committee for the Protection of Persons in the South-West and Overseas France III (Bordeaux) 122 and The National Agency for the Safety of Medicines and Health Products (ANSM). For more 123 details see the Online Repository. 124

125

126 Th17 cell sorting and expansion

From PBMC, CD4⁺CXCR3⁻ T cells were enriched using EasySepTM Human Th17 Cell Enrichment Kit (STEMCELL Technologies). Then, CD4⁺CD45RA⁻CCR6⁺CXCR3⁻ (Th17) cells were cell sorted on a FACS Aria cell sorter (BD Biosciences). Cells ($0.2x10^6$) were expanded during 14 days with ImmunoCultTM Human CD3/CD28/CD2 T Cell Activator (STEMCELL) in Immunocult-XF (STEMCELL) medium containing IL2 (5 ng/mL), anti-IFN γ antibody (5 µg/ml, B27, BD Biosciences, to limit Th1 bias) and etanercept (10μ g/ml, to favor cell proliferation).

134

135 Total internal reflection fluorescence (TIRF) microscopy

136 Cells were loaded with 5 μ M Fluo-5-AM in culture medium containing 5% heat-inactivated

137 FBS and 100 μ M EGTA-AM (buffering Ca²⁺ at the channel mouth) for 30 minutes at 37°C.

138 After washing and plating cells in medium, we recorded elementary calcium events (ECE) at

the cell membrane every 2 ms before and after stimulation with ImmunoCultTM Human CD3/CD28 T Cell Activator in presence or not of 10 μ g/ml of nicardipine. We analyzed spatiotemporal fluorescence changes expressed as F/F0, the number of opened channels and the ECE opening frequencies. Complete procedures are given in the Online Repository.

143

ELISA, qPCR, Cytometry, calcium measurements, western-blotting, 144 immunohistofluorescence (IHF) and in situ hybridization (ISH) on skin sections, Th17-145 cell transduction with shRNA, T-cell recovery and amplification from LP biopsies, the 146 generation of human inflammatory psoriasis skin equivalents populated with Th17 cells, 147 murine imiquimod-induced psoriasis like model and statistical analyses were detailed in 148 the Online Repository. 149

150 **Results**

151

CACNA1F expression, encoding the Cav1.4 calcium channel, expression is higher in lesional psoriatic compared with non-lesional psoriatic or atopic dermatitis biopsies

We first characterized gene expression profile in lesional (LP) versus non-lesional (NLP) 154 psoriatic biopsies and for comparison we included lesional atopic dermatitis (AD) biopsies. As 155 expected, the LP biopsies were characterized by the upregulation of genes known to have a 156 pathological role in psoriasis, including genes encoding the cytokines related to the IL17/IL22 157 axis, (IL17A, IL26, IL23A, IFNG, CSF2, IL22, IL6), chemokines (CCL20 and CXCL8) involved 158 in both Th1/ Th17 lymphocytes and neutrophil recruitment, and skin barrier function (elafin 159 (PI3), ß-defensin2 (DEFB4B), late cornified envelop (LCE3A), psoriasin (S100A7), MPR6 160 (S100A8), mKi67, keratin16 (KRT16) and involucrin (IVL)). These genes were significantly 161 162 more expressed in LP than in NLP skins (Figure 1A R1 and see Table E1 in the Online repository) and in AD biopsies for most of them (Figure 1A R1 and see Table E2 in the 163 164 **Online repository**). The AD biopsies were characterized by the upregulation of genes involved in type 2-cell function/recruitment (IL13, CCL26, carbonic anhydrase 2 (CA2) and neural EGFL 165 like 2 (NELL2)), (Figure 1A R1 and see Table E2 in the Online repository). Hierarchical 166 clustering based on gene expression levels showed two clusters with respect to inflammation 167 status: one containing the LP and AD biopsies and the other the NLP biopsies and two AD 168 samples with low levels of anti-microbial peptide expression. 169

We observed significant *CACNA1F* overexpression in the LP versus NLP or AD skin (four and five-fold, respectively), (**Figure 1B R1**). The cytokines (*IL17A, IFNG*, and *IL26*) expression were significantly upregulated in LP compared with NLP and AD samples (**Figure E1A R1 in the Online repository**), and the expression of *CACNA1F* positively correlated with these cytokines (**Figure 1C R1**), but not in the different disease-states taken separately. 175

In LP skin, Ca_v1.4 calcium channel is preferentially detected in *RORC*⁺ compared with *RORC*⁻ cells

178 Using *in situ* hybridization (ISH), we next analyzed whether $Ca_v 1.4$ expressing ROR $\gamma t^+ CD4^+$ (Th17) lymphocytes can be detected in LP skins. CD4⁺ T cells co-expressing CACNA1F and 179 RORC (encoding for the transcription factor RORyt) mRNAs were detected in all LPs (Figure 180 **1D R1**). Among $CD4^+$ cells, which represented 21% (20.7 ± 9.1) of nucleated cells, 29% (28.5) 181 \pm 10.1) were *RORC*⁺ (Figure 1D and 1E R1). *CACNA1F* was found in 47% (47.2 \pm 18.9) of 182 $RORC^+ CD4^+$ cells versus 28% (27.8 ± 17.3) of $RORC^- CD4^+$ cells (Figure 1E R1). Thus, there 183 was a strong enrichment in CACNA1F⁺ cells among RORC⁺ CD4⁺ cells compared with RORC⁻ 184 $CD4^+$ (Figure E1B R1 in the Online repository). Among the $CD4^-$ cells, 19% (19.2 ± 10) 185 were *RORC*⁺, with 40% (39.8 ± 15.6) being *CACNA1F*⁺ versus 21% (21.2 ± 10) of *CACNA1F*⁺ 186 187 in RORC⁻ (Figure 1E R1). As in the CD4⁺ cells, there was an enrichment in CACNA1F⁺ cells among RORC⁺ CD4⁻ cells compared with RORC⁻ CD4⁻ (Figure E1C R1 in the Online 188 **repository**). There was a positive correlation between the percentage of $CD4^+$ RORC⁺ 189 CACNA1F⁺ and CD4⁻ RORC⁺ CACNA1F⁺ (Figure E1D R1 in the Online repository 190), suggesting a robust association between CACNA1F and RORC gene expression within the 191

same cells.

At the protein level, we confirmed the detection of cells co-labeled with anti-Ca_v1.4, anti-CD4 and anti-ROR γ t antibodies in the dermis (**Figure 1F R1**). Notably, higher numbers of Ca_v1.4⁺ ROR γ t⁺ CD4⁺ cells were associated with T cell clustering in LP skins (**Figure 1F R1 and New Figure 1G R1**). Ca_v1.4 staining was not detected in the epidermis.

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TCR-induced cytokine production by T cells infiltrating psoriatic lesions depends on Ca_v1
 channels

To investigate the role of Ca_v1.4 in LP skins, infiltrating T-cells were recovered and amplified 200 by stimulation with anti-CD3/CD28/CD2 in the presence of IL2 and IL23, as previously 201 described¹⁷, (**Figure 2A**). Fourteen days later, all cells were CD3⁺, dividing nearly equally into 202 CD4⁺ and CD8⁺ T cells (Figure 2B). We defined Th1, Th1/Th17, and Th17 cells as 203 CD4⁺CD45RA⁻ cells that were CXCR3⁺CCR6⁻, CXCR3⁺CCR6⁺ and CXCR3⁻CCR6⁺ 204 respectively according to previous works¹⁸. Expanded skin T-cells were composed of Th17 205 (30%), Th1/Th17 (30%) and Th1 (20%) cells (Figure 2C). Upon anti-CD3/CD28 stimulation, 206 T cells produced IL17A, IL22, IFN γ and TNF α (Figure 2D). Nicardipine, a Ca_v1 channel 207 antagonist (used at 10 μ g/ml, a concentration that inhibits Ca_v1 function in human Th2 cells¹⁴) 208 reduced the production of IL17A (36%), IL22 (35%), IFN_Y (27%) and TNF_a (53%) upon TCR 209 stimulation (Figure 2D and Figure E2A in the Online repository). Cytokine gene 210 transcription depended on Cav1 channels, as nicardipine inhibited the TCR-induced 211 transcription of IL17A (52%), IL22 (57%) and at a lower level IFNG (37%) and TNFA (27%) 212 (Figure 2E and see Figure E2B in the Online repository). In addition, IL17F and CSF2 213 transcript expressions induced by TCR-stimulation were reduced by nicardipine (by 54% and 214 42% respectively, Figure 2E and Figure E2B in the Online repository). Altogether, these 215 data suggest that TCR-induced cytokine expression and production were partially controlled by 216 Ca_v1 calcium channels in LP-infiltrating T-cells. 217

218

219 **TCR-induced cytokine transcription and production by Th17 cells expanded from healthy**

and psoriatic patient PBMCs are also partially controlled by Cav1 channel activation

Based on chemokine receptor expression, we showed that the only population to be significantly increased in PP compared with HD was the Th17 cells (see Figure E3 in the **Online repository**). To investigate the role of $Ca_v 1.4$ in Th17 cell functions of PP and HD, we set up a protocol of amplification to obtain sufficient Th17 cells from the peripheral blood. We sorted the Th17 cells (CD4⁺CD45RA⁻CCR6⁺CXCR3⁻) and expanded them by anti-CD3/CD28/CD2 stimulation for 14-21 days in the presence of IL2, anti-IFN γ and anti-TNF α antibodies. The expanded Th17 cells still displayed their ability to produce IL17A, IL22, and IFN γ (although at a lower level for the two latter cytokines) when compared with *ex vivo* production (**see Figure E4A in the Online repository**). The expression of *RORC* and *CACNA1F* remained detectable in expanded Th17-cells (**see Figure E4B in the Online repository**).

The Th17 cells from PP and HD produced IL17A, IL22, IFNy, and TNFa after anti-232 CD3/CD28 stimulation (Figure 3). Nicardipine decreased not only the percentage of cytokine-233 producing cells (Figure 3) but also the amount of cytokines per cells (GMFI) and in culture 234 supernatants, and their transcription (see Figure E5 and E6 in the Online repository). The 235 inhibitory effect of nicardipine on cytokines was similar in HD and PP, with respect to cytokine 236 secretion, production, or transcription (see Figure E5 and E6 in the Online repository). 237 Together, these results show that $Ca_v 1 Ca^{2+}$ channels contribute to optimal cytokine production 238 by TCR-activated Th17 lymphocytes from HD and PP. 239

240

Cav1 channels are involved in TCR-driven initial calcium events and transduction signaling in Th17 cells

To demonstrate that $Ca_v 1$ channels play a role in the global Ca^{2+} response of Th17 cells, we analyzed changes in the intracellular Ca^{2+} concentration ($[Ca^{2+}]_i$) induced by TCR stimulation of expanded Th17 cells from the peripheral blood of HD. As shown in **Figure 4A-B**, nicardipine strongly reduced the TCR-induced increase in $[Ca^{2+}]_i$. We also analyzed the potential role of $Ca_v 1$ channels in the regulation of endoplasmic reticulum (ER) Ca^{2+} stores and store-operated Ca^{2+} entry (SOCE) by examining the effect of nicardipine on the Ca^{2+} signal elicited by thapsigargin in the presence or absence of calcium in the extracellular medium. Nicardipine

neither affected intracellular Ca²⁺ store release into the cytoplasm, nor SOCE resulting from 250 the ER calcium depletion induced by thapsigargin (see Figure E7 in the Online repository). 251 In order to deepen the role of $Ca_v 1$ channels in the influx of Ca^{2+} following TCR engagement, 252 we developed total internal reflection fluorescence microscopy (TIRFM) imaging to record 253 rapid Ca²⁺ changes close to the plasma membrane. This allows the imaging of peaks 254 corresponding to spatiotemporal single elementary calcium events (ECE) likely due to the 255 activity of one channel or a cluster of Ca²⁺ channels¹⁹. Under resting conditions, the expanded 256 Th17-cells from the peripheral blood of HD displayed few ECEs and their number significantly 257 increased by 1.82 fold after 2-10 seconds of stimulation with anti-CD3-biotin/streptavidin 258 complexes (Figure 4C-E). In addition, the ECE opening frequency significantly increased by 259 TCR stimulation (Figure 4F), suggesting that both an increase in the number of open channels 260 and higher opening frequencies are responsible for initial TCR-driven localized Ca²⁺ fluxes. 261 262 Nicardipine decreased the number of spontaneous ECEs in resting cells (Figure 4C-E) and precluded the increase in ECE numbers and opening frequencies upon TCR stimulation (Figure 263 **4C-F**). Thus, $Ca_v 1$ channels support early Ca^{2+} influx at the plasma membrane in both resting 264 and TCR-activated Th17 cells, even before the global $[Ca^{2+}]_i$ rise. 265

We then studied the consequences of this early decrease in calcium entry after TCR engagement on proximal transduction signaling. The TCR-induced phosphorylated-tyrosine protein pattern (**Figure 4G**) and phosphorylation of the MAPKinase, ERK2, were decreased by nicardipine (**Figure 4H**). Nicardipine also inhibited the nuclear translocation of the Ca^{2+} dependent transcription factor, NFATc2 (**Figure 4I**). Altogether, these data suggest that Ca_v1 channels control an early calcium entry after TCR engagement, which is essential for the full TCR-driven activation of Th17 cells.

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274 Knockdown of Ca_v1.4 decreases IL17A and IL22 production by Th17 cells

As nicardipine antagonizes all Cav1 channels, we tested the specific role of Cav1.4 in Th17 275 functions by knocking down its expression using GFP reporter shRNAs. Th17-cells from the 276 peripheral blood of HD transduced with gene-specific shRNAs resulted in a strong reduction 277 of CACNA1F expression (50%, Figure 4J) associated with a reduction of $Ca_v 1.4$ channel at the 278 protein level (87%, Figure 4K) in GFP⁺ sorted cells compared with cells transduced with 279 control shRNA. We did not detect compensatory CACNA1C and CACNA1D expression in the 280 Th17 cells transduced with shRNAs against CACNA1F (Figure 4J). The decreased Cav1.4 281 expression was associated with the reduced production of IL17A and IL22 after TCR 282 stimulation by 55 and 59%, respectively (Figure 4L). These data provide evidence for the 283 selective role of Ca_v1.4 in IL17A and IL22 production by Th17 cells. 284

285

Ca_v1 channels are involved in the development of psoriatic inflammation in a human 3D skin reconstruction model and in murine imiquimod-induced psoriasis like model

To study the role of Cavl channels in the pathogenesis of Th17 cells in psoriasis, we established 288 a three-dimensional (3D) skin reconstruction model, in which the expression by keratinocytes 289 of psoriasis inflammatory markers is induced by populating the skin with activated Th17-290 cells²⁰. To evaluate the role of $Ca_v 1$ channels in the development of psoriasis inflammation, 291 nicardipine was added to Th17 cells before stimulation and on each day of the culture (Figure 292 5A). Ciclosporin A (CsA), used as a control, was only added the last two days of the culture, 293 as previously described²⁰. Activation of Th17 cells was confirmed by CD69 expression and was 294 not modified by nicardipine (data not shown). Nicardipine did not prevent Th17 cells from 295 migrating into skin equivalents (SE), as shown by the number of infiltrating CD3⁺ T-cells after 296 four days of culture (Figure 5B-C). According to²⁰, addition of Th17-cells did not modify the 297 epidermis morphology, with no effect of nicardipine or CsA (see Figure E8 in the Online 298 repository). Nicardipine tended to reduce IL17A production and significantly decreased IL22 299

production (**Figure 5D**). The infiltration of Th17 cells in SE induced a significant increase in the expression of psoriasis-associated genes, such as *KRT16*, *P13*, and *DEFB4* (encoding for cytokeratin 16, elafin, and β -defensin 2, respectively), after four days of culture (**Figure 5E**). Nicardipine decreased the expression of these genes to the same level as CsA (**Figure 5E**). These data suggest that inhibition of Ca_v1 channels can releave the psoriatic inflammation in a 3D skin reconstruction model, probably by acting on Th17-cells functions, in accordance with locally decreased IL17A and IL22 production.

Finally, we investigated the role of Ca_v1 calcium channels in local lesions in murine 307 imiquimod-induced psoriasis like model. We applied Imiquimod to the shaved back and on one 308 ear of BALB/c mice that received or not intraperitoneal injections of nicardipine. As shown in 309 the New Figure E9A R1 below, imiquimod-treated mice presented dermal inflammation and 310 epidermal acanthosis compared with untreated mice, in treated-ear and -back skin (see New 311 312 Figure E9A and 1B R1 in the Online repository) and a significant increase of IMQ-treatedear and -back skin thickness (see New Figure E9C R1 in the Online repository). The 313 314 inhibition of Cav1 channels by nicardipine greatly reduced both dermal inflammation, epidermal acanthosis and decreased the ear and back-skin thickness in this experimental model. 315

316 **Discussion**

This study aimed to investigate the singularities of the Ca²⁺ toolkit of human Th17 CD4⁺ T cell 317 activation and its possible involvement in psoriasis, a Th17 skewed chronic skin disease. We 318 focused on $Ca_v 1.4$, since this channel, which is present in naive CD4⁺ T cells, disappears upon 319 activation and differentiation of effector human T cells, except in Th17 cells¹⁴. Our results 320 showed a preferential association of CACNA1F with RORC expression in CD4⁺ and CD4⁻ cells, 321 and the detection of CD4⁺ ROR γ t⁺ Ca_v1.4⁺ triple-positive cells in the LP skin dermis. The 322 inhibition of Cav1 channels decreased Th17 cytokine transcription and production. The Cav1 323 channels acted early during Th17-cell activation, allowing optimal Th17 cell activation, as 324 indicated by the effect of nicardipine on global early tyrosine phosphorylation, MAP Kinase 325 activation and NFAT nuclear translocation. The Cav1.4 knockdown decreased IL17A and IL22 326 production, supporting the role of this channel in the Th17-cells. Moreover, the Ca_v1 channel 327 328 blocker decreased psoriatic inflammation in a model of skin equivalent model populated with Th17 cells. 329

The expression of CACNA1F was higher in LP than in NLP or AD skin and correlated 330 with the expression of the inflammatory cytokines IL17A, IFNG, and IL26 only when the 331 different groups (lesional and non-lesional psoriasis and atopic dermatitis) were analyzed 332 together. Moreover, CACNA1F expression was enriched among RORC⁺ over RORC⁻ cells in 333 both CD4⁺ and CD4⁻ dermis-infiltrating cells from PP. We did not check the expression of 334 CACNA1F in the regulatory T cells, but these cells have been shown to turn into RORC 335 expressing cells which produce IL17 in psoriatic skin²¹. The Ca_v1.4 channel was also present 336 in RORC⁺ CD4⁻ cells (which might include CD8 T cells, neutrophils, and ILC3), supporting the 337 role of $Ca_v 1.4$ in the IL17-associated inflammatory $axis^{22}$. $Ca_v 1.4$ expression was not observed 338 in the epidermis, suggesting that the keratinocytes do not express this Ca^{2+} channel. In the 339 dermis, Cav1.4 was essentially found in inflammatory zones containing clusters of CD4⁺T cells. 340

We expanded T cells from the biopsies and showed that they transcribed the pro-inflammatory 341 cytokines IL17A, IL17F, IL22, IFNG, CSF2 and TNFA after TCR stimulation. Nicardipine, a 342 Ca_v1 channel antagonist, reduced both TCR-induced cytokine transcription and production by 343 the CD4⁺ T cells infiltrating the PP skin. Although the CD4⁺ T cells were the main producers 344 of IL17A and IL22 (the CD8⁺ T cells produced very few IL17A and IL22; data not shown), 345 IFNy and TNF α were also produced by infiltrating CD8⁺ T cells, and nicardipine inhibited their 346 production (not shown), in accordance with $Ca_v 1.4$ expression by the CD8⁺ T cells¹⁴. It is 347 interesting to note the potential inhibitory action of nicardipine on all cytokines and in the 348 349 majority of PPs, since these cytokines act synergistically and amplify the inflammatory loop in psoriatic skin inflammation^{23, 24}. 350

The CCR6⁺CXCR3⁻ CD4⁺ T cells that expressed *RORC*, the master switch transcription 351 factor of Th17 cells, were the only subset to increase in the blood of PP compared to HD, in 352 accordance with Blauvelt et al.²². The Th17 cells expressed CACNA1F and maintained this 353 expression as well as their pattern of TCR-induced cytokine expression after expansion, 354 indicating that they represent a valuable tool for studying Th17 functions. TCR stimulation of 355 Th17 cells induced similar amounts of cytokines (transcription and production) from both PP 356 and HD, which were inhibited to a similar extent by nicardipine. These data confirm the 357 quantitative bias toward more Th17 cells in psoriasis and show that nicardipine reduced 358 cytokine production by the Th17 cells obtained from the blood or even from inflamed tissues. 359 Among the four Ca_v1 channels, $Ca_v1.4$ was the only Ca_v1 channel expressed in the Th17 cells. 360 Furthermore, the transduction of these cells with specific shRNA reduced inflammatory 361 cytokine production. Altogether, these results support the role of Ca_v1.4 in Th17-cell functions. 362 Nicardipine decreased the transcription and production of all inflammatory cytokines 363 tested, indicating that Cavl channels regulate common pathways upstream of cytokine gene 364 transcription. The crucial role of Ca^{2+} in Th17 functions and their pathological potential²⁵ 365

explains why ciclosporin A, which inhibits Ca^{2+} -dependent phosphatase calcineurin, is 366 effective in the treatment of PP²⁶. Our investigations described an early role of Ca_v1 channels 367 during TCR activation as nicardipine fully abrogated early ECE induced by TCR stimulation 368 and even decreased the few Ca²⁺ fluxes observed under resting conditions, leading to the 369 conclusion that $Ca_v 1.4$ is involved in both Ca^{2+} homeostasis and the TCR response. Contrary 370 to what is shown in murine naive T lymphocytes¹⁶, the Ca_v1.4 channels do not regulate the 371 intracellular Ca²⁺ stores nor the functionality of STIM/ORAI (SOCE) since the response to 372 thapsigargin was not affected by nicardipine in human Th17 cells. However, nicardipine 373 reduced the TCR-driven global Ca²⁺ response suggesting a link between the ECE and the 374 cellular Ca²⁺ response and that the initial defect cannot be overcome to induce a full response 375 in the absence of functional Cav1 channels. It is conceivable that the ECEs locally modify the 376 cell environment in the vicinity of the immune synapse allowing an optimal assembly of the 377 378 TCR transduction machinery. This is in accordance with the role of local ECE preceding full TCR activation, required to initiate cell transduction, possibly through an effect on cytoskeleton 379 rearrangement^{27, 28}. In murine CD8⁺ T cells, Ca_v1.4 can associate with p56^{lck} and vav1, which 380 both play a key role in TCR-dependent signal transduction²⁹. Nicardipine reduced the early 381 profile of tyrosine phosphorylation, ERK2 activation and NFATc2 nuclear translocation, 382 supporting the role of Ca_v1 channels in the initiation of global signal transduction. Our data 383 suggest that Ca_v1.4, which is known to activate at a relatively negative cell membrane potential, 384 closer to the resting cell membrane potential than the other Cav1 channels, could deliver Ca²⁺ 385 in a very fast and localized manner facilitating the transduction machinery assembly. Several 386 groups have shown that ORAI channels play a major role in the calcium response induced by 387 TCR engagement in human pathogenic Th17 cells^{25, 30}, raising the question of the respective 388 places of Ca_v1 and Orai Ca²⁺ channels in Th17 cell Ca²⁺ signaling. TIRF experiments performed 389 here favor the earliest involvement of Cav1 channels in the initiation of T-cell activation (very 390

few seconds) followed by STIM/ORAI channel activation. Accordingly, the Ca_v1 channels were described to act earlier than the ORAI channels following cell activation and to convey ten more calcium ions than the ORAI channels³¹.

In order to establish a potential beneficial use of Cav1.4 inhibitors in the treatment of 394 PP, we tested the effects of nicardipine on a human psoriatic inflammation 3D skin model 395 populated with Th17 cells. Nicardipine did not prevent the migration of Th17 cells into the 396 dermis, but it tended to diminish the expression of specific psoriasis keratinocyte markers with 397 the same effects as CsA. As the cytokines produced by T cells appeared to be the main factor 398 responsible for the psoriatic inflammatory phenotype²⁰ and that nicardipine diminished IL17A 399 and IL22 cytokines in the skin-equivalent infiltrated with Th17 cells, we propose that Ca_v1 400 channel blockade reduced psoriatic gene expression by inhibiting the cytokine production by T 401 cells. Moreover, we showed Cavl calcium channel blocker was beneficial in the treatment of 402 403 psoriasis-like inflammation in mice since nicardipine inhibits the dermal inflammation, epidermal acanthosis and the ear and back-skin thickening in murine imiquimod-induced 404 psoriasis like model. 405

The effect of nicardipine on the cytokine production by Th17 cells was partial, but affected all tested cytokines that participate in a positive feedback inflammatory $loop^{26}$. *CACNA1F* was also associated with *RORC* expression in the CD4⁻ cells that also participate in psoriasis pathogenesis³². Therefore, targeting this channel might be beneficial to treat psoriasis. Finally, psoriasis is associated with cardiovascular comorbidities such as blood hypertension³³; therefore, Ca_v1 channel blockers could be also beneficial in this regard.

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502 Author contributions

MMa performed all the experiments. IN, CL, SB and CR performed ISH and HIF of psoriatic 503 skin biopsies and confocal analysis. MMo and LP performed TIRF experiments. MMo 504 developed software for TIRFM analysis and analyzed the number of open channels and their 505 opening frequency. SL helped to analyze the nuclear translocation of NFAT. CP and MT 506 designed and implemented the clinical study, recruited patients with psoriasis and atopic 507 dermatitis. EG and CD developed analysis of qPCR data and performed heatmap. JCG, HD and 508 DR contributed to the design of experiments and criticized the results. MB and MDT managed 509 and supervised the clinical project. FL did the experimental design, performed and analysed the 510 results of qPCR and criticized the results. LP and MS did the experimental design, analysed the 511 results, and wrote the paper. MS supervised the work. All authors critically reviewed the 512 manuscript. 513

514 Figure legends

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New Figure 1 R1. Cav1.4 calcium channel expression in Th17-cells infiltrating psoriasis skins

(A) Hierarchical gene expression heat map in lesional (LP), non-lesional (NLP) psoriasis and 518 atopic dermatitis (AD) skins. (B) Violin plots showing CACNA1F expression. One symbol by 519 patient; the median and the interquartile ranges are indicated. (C) Correlation between 520 CACNA1F and cytokine expression in LP, NLP and AD skins. (D) Representative images with 521 CD4 (red), RORC (green) and CACNA1F (white) encoding mRNAs by ISH of LP skins. (E) 522 Quantifications of cell numbers per field as described in the Online Repository. (+/-) groups 523 together cells that are positive or negative for the marker. Means \pm sem; one symbol by patient. 524 (F) Representative images with CD4 (red), RORyt (green) and Cav1.4 (white) proteins by IHF 525 526 of LP skins. Bar = 50 μ m for general view of skin section and 10 μ m for inserts. (New G) IHF scores of LP skins. $0 = any CD4^+ ROR\gamma t^+ Ca_v 1.4^+$ cells in any zone; $1 = one CD4^+ ROR\gamma t^+$ 527 $Ca_v 1.4^+$ cells in one zone; 2 = several zones containing one CD4⁺ RORyt⁺ Ca_v 1.4⁺ cells ; 3 = 528 several zones containing at least three CD4⁺ ROR γ t⁺ Ca_v1.4⁺ cells. Means ± sem; one symbol 529 by patient. 530

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Figure 2. Ca_v1-dependent inflammatory cytokine production by T-cell-infiltrating lesional psoriasis skin

535 (A) Experimental procedure. (B) Lymphocyte frequencies after T-cell expansion. (C) Th17,

536 Th1/Th17 and Th1-cell frequencies in expanded T-cells. (D-E) Cytokine production (ELISA,

537 D) and expression (qPCR, E) in expanded T-cells after anti-CD3/CD28 stimulation with or

538 without nicardipine. One symbol by patient. P values were determined by one-way ANOVA

followed by Turkey's multiple comparison test. * p<0.05; ** p<0.01; ***p<0.005;
****p<0.0001.

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Figure 3. Cav1-dependent inflammatory cytokine production by peripheral blood Th17 cells

Th17 cells expanded from healthy donor (HD) and psoriasis patient (PP) blood were stimulated with anti-CD3/CD28/CD2 antibodies plus or minus (nicar) for 4h before intracellular cytokine staining. Comparisons with one-way ANOVA followed by Turkey's multiple comparison tests. p<0.05; ***p<0.005; ***p<0.005; ***p<0.0001.

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New Figure 4. Cav1 channels contribute to TCR-induced [Ca²⁺]_i rise and transduction 549 signaling in Th17 cells and knocking-down CACNA1F reduces Th17 cytokine production 550 551 (A-B) HD Th17 cells were loaded with Fura2-AM and fluorescence was recorded before (F0) and after (F) TCR stimulation \pm nicardipine (nicar). (A) Mean \pm SEM from 50-100 cells from 552 3 HD. (B) the area under the curve. (C-F) 3-D images of ECE from Th17 cells loaded with 553 Fluo-5AM (C), number of ECE (one symbol by cell; pool from 2 HD) (D), relative ECE number 554 (E) and opening frequency (one symbol by ECE; 5-7 cells from 2 donors) (F). (New G-H) 555 Th17 cells were stimulated with ImmunoCult™ Human CD3/CD28/CD2 T Cell Activator for 556 2 min (Stim) or not (NS) in presence or absence of nicardipine (nicar). Equal amounts of lysate 557 proteins were blotted and probed with antibody against phosphorylated tyrosine (P-Tyr) (New 558 G), P-ERK2 (New H) or β-actin (Actin, loading control). Graphs show the quantification of 559 staining intensities relative to actin intensity and to unstimulated samples of 3-4 donors. (New 560 I) Th17 cells were stimulated with ImmunoCult[™] Human CD3/CD28 T Cell Activator for 4 561 hours (Stim) or not (NS) in presence or absence of nicardipine (nicar). Nuclear translocation of 562 NFATc2 is analyzed by confocal. Bar = $5 \mu m (J-L) HD Th 17$ cells were transduced with control 563

(shCtr) or *CACNA1F* GFP reporter shRNAs. *CACNA1* expression (**J**), Ca_v1.4 protein level (New K, Bar = 5 μ m) and TCR-induced cytokine production (**L**) in GFP⁺-sorted cells; 2-3 biological replicates from 3 experiments. P values were determined by Student's t-test (**B**, **D**-**F**, New I and New K) and by paired Wilcoxon test (**L**). * p<0.05; ** p<0.01; ***p<0.005; ****p<0.0001.

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Figure 5. Antagonizing Cav1 channels diminishes cytokine and psoriasis-like gene expression in skin equivalent infiltrated by Th17 cells

(A) Experimental procedure. (B-C) Enumeration of CD3⁺ T cells 4 days after introducing Th17 cells under the skin equivalent (SE). The mean is indicated and the dotted line shows the background without T-cells. (D) IL17A and IL22 production 24h after Th17 cells introduction in SE Each point corresponds to one SE. (E) *KRT16*, *PI3* and *DEFB4B* expression after 4 days of culture. P values were determined by paired Wilcoxon test (D) and Student's t-test (E). * p<0.05; **p<0.01 Bar = 50µm.