



**HAL**  
open science

## **Lack of protective effect of CCR3 blockade during experimental colitis may be related to CCR3 expression by colonic Tregs**

Maroua Ferhat, Julie Hablot, Mahdia Taieb, Fatouma Salem, Patrick Netter, Laurent Peyrin-Biroulet, Jean-Yves Jouzeau, David Moulin

### ► To cite this version:

Maroua Ferhat, Julie Hablot, Mahdia Taieb, Fatouma Salem, Patrick Netter, et al.. Lack of protective effect of CCR3 blockade during experimental colitis may be related to CCR3 expression by colonic Tregs. *Clinical and Translational Medicine*, 2021, 11 (6), pp.e455. 10.1002/ctm2.455 . hal-03356573

**HAL Id: hal-03356573**

**<https://hal.science/hal-03356573>**

Submitted on 28 Sep 2021

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

**Lack of protective effect of CCR3 blockade during experimental colitis may be related to CCR3 expression by colonic Tregs**

Maroua FERHAT<sup>1\*</sup>, Julie HABLOT<sup>1\*</sup>, Mahdia TAIEB<sup>1</sup>, Fatouma SALEM<sup>1</sup>, Patrick NETTER<sup>1</sup>, Laurent PEYRIN-BIROULET<sup>2,3</sup>, Jean-Yves JOUZEAU<sup>1,4#</sup> and David MOULIN<sup>1,5#</sup>

1 IMoPA, UMR7365 CNRS-Université de Lorraine, Vandœuvre Les Nancy, France

Université de Lorraine, CNRS, IMoPA, F-54000 Nancy, France

2 NGERE, UMR-U1256 INSERM-Université de Lorraine, Vandœuvre Les Nancy, France

Université de Lorraine, Inserm, NGERE, F-54000 Nancy, France

3 Service d'hépatogastroentérologie, CHRU de Nancy, Vandœuvre Les Nancy, France

4 Service de Pharmacologie Clinique et de Toxicologie, CHRU-Nancy, Nancy, France

5 CHRU de Nancy, Contrat d'interface, Vandœuvre Les Nancy, France

\* These authors contributed equally to this work

# These authors contributed equally to this work

Running title: CCR3 blockade aggravates DSS colitis

Address for correspondence:

David MOULIN

IMoPA, UMR7365 CNRS-Université de Lorraine, Vandœuvre Les Nancy, France

david.moulin@univ-lorraine.fr to whom requests **for reprints should be addressed**

**+33 3 72 74 65 61**

**Dear Editor,**

while previous findings indicate that blockade of the chemokine receptor CCR3 can be viewed as an attractive therapeutic alternative to actual treatments in inflammatory bowel diseases (IBD), we demonstrate herein that CCR3 deficiency or pharmacological blockade does not protect and even aggravates clinically, histologically and biologically DSS-induced colitis. Interestingly, we show that a subpopulation of colonic regulatory T (Treg) cells express CCR3 providing an explanation for this unexpected deleterious effect of CCR3 inhibition or absence during experimental colitis.

Over the past two decades, several drugs (e.g., biologics and tofacitinib) have changed IBD management. However, a significant proportion of patients remain or become refractory to these drugs, indicating an urgent need for new therapeutic targets.

CCR3 is a chemokine receptor, highly expressed by eosinophils and basophils, that have been implicated in autoimmune disorders and allergic diseases. CCR3 binds a wide range of ligands necessary for chemotaxis and tissue infiltration, including RANTES (CCL5), MCP-3 (CCL7), MCP-4 (CCL13), MIP-5 (CCL15), CCL18 (MIP-4), Eotaxin 1 (CCL11), Eotaxin-2 (CCL24), Eotaxin-3 (CCL26), and MEC (CCL28)<sup>1,3</sup>. Among these, eotaxin-1 is a key regulator of intestinal inflammation as demonstrated experimentally<sup>1,2</sup> and clinically<sup>3</sup>. Similarly, eotaxin-2, is also increased in Ulcerative Colitis (UC) patients<sup>4</sup>.

All these lines of evidence provide a basis for targeting the eosinophil/eotaxin axis in patients with UC. In this prospect, Bertilimumab, an IgG4 monoclonal antibody that targets eotaxin-1, with a very high affinity and specificity, was developed and is currently in phase 2 clinical trial for the treatment of moderate to severe UC (NCT01671956)<sup>5</sup>. In addition, anti-eotaxin-2 antibodies are currently investigated for their potential use in the treatment of IBD (US Patent # 9,067,989).

Experimental and clinical data indicate that RANTES is also a player of IBD<sup>6,7</sup>. Moreover, clinical investigations showed an over-expression of eotaxin-3<sup>4,8</sup> and MIP-2<sup>9</sup> in the tissue and serum of UC patients.

Therefore, to explore the relevance of CCR3 inhibition in colitis, we firstly decided to block CCR3 during colitis using GW766994, a CCR3 selective antagonist, during DSS-induced colitis in mice (Figure 1A). Surprisingly, GW766994 treatment was associated with a higher incidence rate (Figure 1B) and a more severe disease as assessed by DAI (disease activity index), body-weight loss and survival rate (Figure 1C-E) compared to control mice receiving DSS alone.

Moreover, colon length reduction or fecal lipocalin levels, two markers of inflammation remained unaffected by CCR3 antagonist treatment (Figure 1F and G). Colons of GW766994-treated mice showed similar histological damages (crypts, architecture loss, and leukocytes infiltration) to non-treated DSS mice (Figure 1H).

To challenge the results obtained with a pharmacological approach, we used a genetic CCR3 deficiency model. DSS was therefore induced in WT and CCR3 KO BALB/c mice (Figure 2A). Disease severity was significantly higher in CCR3-deficient mice than in WT mice following DSS administration (Figure 2C). Such disease worsening was also reflected by a higher body-weight loss and a trend to a lower survival rate (Figure 2D-E) in CCR3 KO mice compared to WT mice. Moreover, shortening of the colon and fecal lipocalin-2 level confirmed these findings (Figure 2F-G). Finally, histological examination of CCR3 KO mice showed slightly higher tissue damages in response to DSS, as observed above for GW766994-treated mice (Figure 2H-I). Taken together, these results show that CCR3 KO mice are prone to develop a more severe DSS colitis, suggesting a protective role for CCR3 during colitis.

T-helper lymphocytes expressing IL-17 (Th17) and regulatory T cells (Treg) are critical regulators of intestinal barrier function and key players in colitis. To investigate the contribution of CCR3 to DSS colitis susceptibility, we performed flow cytometric analysis of Th17/Treg population in colon and mesenteric lymph nodes (mLNs). No difference in frequency or absolute number of Th17 relative to CCR3 deficiency or blockade was found (Supplementary Figure 1), suggesting that DSS polarizes Th17 response independently of CCR3.

Interestingly, while in basal conditions, both WT and CCR3-deficient mice (left panels) had the same frequency (Figure 3B) and absolute cell number (Figure 3C) of Tregs, abundance of Treg cells was higher in colonic *lamina propria* from WT mice than in CCR3 KO mice (Figure 3C, left panel) after DSS-challenge. Similar results, were obtained for GW766994-treated animals in comparison to DSS-untreated control mice (Figure 3B and 3C, right panels). In mLNs, frequency and absolute number of CD3<sup>+</sup> CD4<sup>+</sup> FoxP3<sup>+</sup> Treg cells were significantly decreased in CCR3 KO mice and GW766994-treated mice (Figure 3D and 3E, right panels) compared to WT control mice. Consequently, a decrease of Treg over Th17 cells ratios was observed in CCR3 KO mice and GW766994-treated mice in comparison to control mice, particularly in mLNs (Supplemental Figure 2B). Our data show that CCR3 deficiency or blockade suppresses Treg response in the large intestine and mLNs of mice, suggesting that CCR3 limits gastrointestinal inflammation *via* the control of Treg cell abundance in these tissues.

As indicated in Figure 4A, ImmGen database indicated that CCR3 is expressed by Treg FoxP3<sup>+</sup> specifically amongst T cells. We thus analyzed CCR3 expression on Treg cells by flow cytometry. As shown in Figure 4, a fraction of colonic resident and mLNs Treg cells (Figure 4B and 4C) express CCR3 at their cell surface under physiological conditions. Such CCR3 expression is consistent with the diminished level of Treg cells observed in colonic *lamina propria* and mLNs of both CCR3-deficient and GW-766994-treated DSS mice. Since some lamina propria Treg cells express RORγt and differ in their function from “classical” Treg, we have investigated the consequence of CCR3 blockade/deletion on these colonic RORγt<sup>+</sup> Treg. Our results indicate that most of the colonic CCR3 positive Tregs were classical RORγt<sup>-</sup> Treg, and that CCR3 blockade or deletion did not affect this RORγt<sup>+</sup> Treg population (Figure 4D and 4E).

Previous reports indicate that CCR3 receptor is also expressed by splenic and adipose tissue Treg cells<sup>10</sup>. Taking altogether, our results support an immunomodulatory role of CCR3 in shaping the intestinal immune response through Treg cell populations in intestinal tissues.

Although all strains were housed in the same animal facility and all precautions were taken to normalize the microbiota of the different strains, we cannot exclude that variation in microbiota composition between the strains could have contributed to the phenotypes observed.

In conclusion, our findings do not support the development of CCR3 blockers as potential therapeutic targets for human IBD and shed a new light on Treg cells contribution to the pathogenesis of IBD.

**Author contribution**

P.N., L.P-B, J-YJ. and D.M. designed the study and supervised the project; J.H. and M.F. designed parts of the study and performed most of the experiments with the help of M.T. and F;S.; M.F wrote the manuscript; D.M., J-Y. J, L.P-B., edited the manuscript.

**Acknowledgments**

We thank Glaxo Smith Kline for kindly providing GW766994. This work was supported by the Region Grand Est and by the Fondation Arthritis. M.F. was supported by a fellowship from french PIA project « Lorraine Université d'Excellence », ANR-15-IDEX-04-LUE. We thank Dr Huguette LOUIS from the Cytometry Core Facility of UMS2008/US40 IBSLor (Université de Lorraine, CNRS, INSERM), F-54000 Nancy, France. The authors declare no competing financial interests.

**Data Availability Statements**

The data underlying this article are available in the article and in its online supplementary material.

## References

1. Ahrens R., Waddell A., Seidu L., Blanchard C., Carey R., Forbes E., et al. Intestinal macrophage/epithelial cell-derived CCL11/eotaxin-1 mediates eosinophil recruitment and function in pediatric ulcerative colitis. *J Immunol* 2008;**181**(10):7390–9. Doi: 10.4049/jimmunol.181.10.7390.
2. Vieira AT., Fagundes CT., Alessandri AL., Castor MGM., Guabiraba R., Borges VO., et al. Treatment with a Novel Chemokine-Binding Protein or Eosinophil Lineage-Ablation Protects Mice from Experimental Colitis. *Am J Pathol* 2009;**175**(6):2382–91. Doi: 10.2353/ajpath.2009.090093.
3. Mir A., Minguez M., Tatay J., Pascual I., Peña A., Sanchiz V., et al. Elevated serum eotaxin levels in patients with inflammatory bowel disease. *The American Journal of Gastroenterology* 2002;**97**(6):1452–7. Doi: 10.1016/S0002-9270(02)04043-1.
4. Manousou P., Kolios G., Valatas V., Drygiannakis I., Bourikas L., Pyrovolaki K., et al. Increased expression of chemokine receptor CCR3 and its ligands in ulcerative colitis: the role of colonic epithelial cells in in vitro studies. *Clin Exp Immunol* 2010;**162**(2):337–47. Doi: 10.1111/j.1365-2249.2010.04248.x.
5. Immune Pharmaceuticals A Randomized, Double-Blind, Placebo-Controlled, Parallel Group, Multi-Center Study Designed to Evaluate the Safety, Efficacy, Pharmacokinetic and Pharmacodynamic Profile of Bertilimumab in Patients With Active Moderate to Severe Ulcerative Colitis. [clinicaltrials.gov](https://clinicaltrials.gov); 2018.
6. Ansari N., Abdulla J., Zayyani N., Brahmi U., Taha S., Satir AA. Comparison of RANTES expression in Crohn's disease and ulcerative colitis: an aid in the differential diagnosis? *Journal of Clinical Pathology* 2006;**59**(10):1066–72. Doi: 10.1136/jcp.2005.034983.
7. Ajuebor MN., Hogaboam CM., Kunkel SL., Proudfoot AEI., Wallace JL. The Chemokine RANTES Is a Crucial Mediator of the Progression from Acute to Chronic Colitis in the Rat. *The Journal of Immunology* 2001;**166**(1):552–8. Doi: 10.4049/jimmunol.166.1.552.
8. Takahashi K., Imaeda H., Fujimoto T., Ban H., Bamba S., Tsujikawa T., et al. Regulation of eotaxin-3/CC chemokine ligand 26 expression by T helper type 2 cytokines in human colonic myofibroblasts. *Clinical & Experimental Immunology* 2013;**173**(2):323–31. Doi: <https://doi.org/10.1111/cei.12117>.
9. Kotarsky K., Sitnik KM., Stenstad H., Kotarsky H., Schmidtchen A., Koslowski M., et al. A novel role for constitutively expressed epithelial-derived chemokines as antibacterial peptides in the intestinal mucosa. *Mucosal Immunology* 2010;**3**(1):40–8. Doi: 10.1038/mi.2009.115.
10. Dioszeghy V., Mondoulet L., Puteaux E., Dhelft V., Ligouis M., Plaquet C., et al. Differences in phenotype, homing properties and suppressive activities of regulatory T cells induced by epicutaneous, oral or sublingual immunotherapy in mice sensitized to peanut. *Cellular & Molecular Immunology* 2017;**14**(9):770–82. Doi: 10.1038/cmi.2016.14.

## Figure legend

**Figure 1. Lack of protective effect of CCR3 pharmacological inhibition in dextran sodium sulfate (DSS)-induced colitis.** (A) Schematic representation of study design: 10-12-week-old BALB/c WT and CCR3-deficient male mice (CCR3 KO) were challenged with 7 days of 3% DSS in drinking water, followed by a 4 day of washout phase. During all the procedure mice were receiving (i.p) a daily dose (10mg/kg) of CCR3 antagonist GW766994. (B) Body weight variation relative to baseline (% of day 0) of mice challenged with DSS. (C-E) Disease activity index (DIA) (C), Incidence of colitis (D) and survival rates (E) have been recorded in control and colitic mice (n= 8 per group). Mice were sacrificed on day 11 and colonic tissues length (F) were measured and feces were collected to assess Lipocalin-2 amount (G) after DSS challenge. (H) Histologic score of one representative experiment based on hemotoxylin and eosin (HE) staining. (I) HE staining of distal colon harvested on day 11 of DSS treatment. Each panel is representative of tissue from at least 3 mice (x40 magnification and 100x magnification). Data were pooled from 2-3 independent experiments. (n = 8-9/group). Data are expressed as mean  $\pm$  SEM, Two-way ANOVA or ANOVA with Tukey posthoc test for multiple comparisons were used for statistical differences between groups. n.s: non-significant; \* p<0.05; \*\* p<0.01 and \*\*\* p<0.001.

**Figure 2. CCR3 genetic deficiency aggravates clinically dextran sodium sulfate (DSS)-induced colitis.** (A) Schematic representation of study design: 10-12-week-old BALB/c WT and CCR3 -deficient male mice (CCR3 KO) were challenged with 7 days of 3% DSS in drinking water, followed by a 4 day of washout phase. (B) Body weight variation relative to baseline (% of day 0) of mice challenged with DSS. (C-E) Disease activity index (DIA) (C), Incidence of colitis (D) and survival rates (E) have been recorded in control and colitic mice (n= 8 per group). Mice were sacrificed on day 11 and colonic tissues length (F) were measured, and feces were collected to assess Lipocalin-2 amount (G) after DSS challenge. (H) Histologic score of one representative experiment based on hemotoxylin and eosin (HE) staining. (I) HE staining of distal colon harvested on day 11 of DSS treatment. Each panel is representative of tissue from at least 3 mice (x40 magnification and 100x magnification). Data were pooled from 2-3 independent experiments. (n = 8-9/group). Data are expressed as mean  $\pm$  SEM, Two-way ANOVA or ANOVA with Tukey posthoc test for multiple comparisons used for statistical differences between groups. n.s: non-significant; \* p<0.05; \*\* p<0.01.



**Figure 3. Regulatory T cell (Treg) population is impaired by CCR3 inhibition in DSS-induced colitis.** (A) Flow cytometry dot plots showing Treg gating strategy. (B) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> FoxP3<sup>+</sup> Treg cells isolated from colonic *lamina* propria of mice challenged or not with DSS. (C) Treg cell count in colonic *lamina* propria of mice challenged or not with DSS. (D) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> FoxP3<sup>+</sup> Treg cells isolated from mesenteric lymph nodes (mLN) of mice challenged or not with DSS. (E) Treg cell count in mLN of mice challenged or not with DSS. (n = 3/group). Data are expressed as mean ± SEM, ANOVA with Tukey posthoc test for multiple comparisons was used for statistical differences between groups. n.s: non-significant; \* p<0.05 and \*\* p<0.01.

**Figure 4. Colonic regulatory T cells (Treg) express CCR3 receptor.** (A) Immune cell expression of CCR3 according to Immgen data base ([www.immgen.org](http://www.immgen.org)). (B) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> FoxP3<sup>+</sup> Treg cells isolated from colonic *lamina* propria of healthy WT and CCR3 KO mice. Representative dot plots of CCR3 expression on colonic *lamina* propria Tregs isolated from healthy WT and CCR3 KO mice. CCR3 expression on colonic *lamina* propria Tregs isolated from healthy WT and CCR3 KO mice. (C) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> FoxP3<sup>+</sup> Treg cells isolated from mesenteric lymph nodes (mLN) of healthy WT and CCR3 KO mice. Representative dot plots of CCR3 expression on mLN Tregs isolated from healthy WT and CCR3 KO mice. (D) Dot plot of RORγt and CCR3 staining in Foxp3 colonic Tregs. (E) Frequency of RORγt<sup>+</sup> Treg in lamina propria of mice challenged or not with DSS and treated with GW766994. Data are expressed as mean ± SEM, ANOVA with Tukey posthoc test for multiple comparisons was used for statistical differences between groups. n.s: non-significant; \*\*\* p<0.001; and \*\*\*\* p<0.001

## Supplemental Figures

**Supplemental Figure 1. CCR3 inhibition does not exacerbate Th17 cell population after DSS challenge.** (A) Flow cytometry dot plots showing Th17 gating strategy. (B) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> ROR $\gamma$ t<sup>+</sup> Th17 cells isolated from colonic *lamina propria* of mice challenged or not with DSS. (C) Th17 cell count in colonic *lamina propria* of mice challenged or not with DSS. (D) Frequency of CD3<sup>+</sup> CD4<sup>+</sup> ROR $\gamma$ t<sup>+</sup> Th17 cells isolated from mesenteric lymph nodes (mLN) of mice challenged or not with DSS. (E) Th17 cell count in mLN of mice challenged or not with DSS. (n = 3/group). Data are expressed as mean  $\pm$  SEM, ANOVA with Tukey posthoc test for multiple comparisons was used for statistical differences between groups. n.s: non-significant; \* p<0.05; \*\* p<0.01 and \*\*\* p<0.001.

**Supplemental Figure 2. CCR3 inhibition impairs Treg/Th17 balance in DSS-induced colitis.** (A) Treg/Th17 ratio in colon *lamina propria* of mice challenged or not with DSS. (B) Treg/Th17 ratio in mesenteric lymph nodes (mLN) of mice challenged or not with DSS. (n = 3/group). Data are expressed as mean  $\pm$  SEM, ANOVA with Tukey posthoc test for multiple comparisons was used for statistical differences between groups. n.s: non-significant; \* p<0.05; \*\* p<0.01 and \*\*\* p<0.001.