

Environmental and life-history factors influence inter-colony multidimensional niche metrics of a breeding Arctic marine bird

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2 metrics of a breeding Arctic marine bird Reyd A. Smith^{1*}, David J. Yurkowski², Kyle J.L. Parkinson¹, Jérôme Fort³, Holly L. Hennin⁴, H. 3 Grant Gilchrist⁴, Keith A. Hobson⁵, Mark L. Mallory⁶, Jóhannis Danielsen⁷, Svend E. Garbus⁸, 4 5 Sveinn A. Hanssen⁹, Jón Einar Jónsson¹⁰, Christopher J. Latty¹¹, Ellen Magnúsdóttir¹⁰, Børge Moe⁹, Glen J. Parsons¹², Christian Sonne⁸, Grigori Tertitski¹³, and Oliver P. Love¹ 6 7 8 ¹ University of Windsor, Windsor, Ontario, Canada, N9B 3P4 9 ² Fisheries and Oceans Canada, Winnipeg, Manitoba, Canada, R3T 2N6 ³ French National Centre for Scientific Research, Paris, France, 75016 10 ⁴ Environment and Climate Change Canada, Ottawa, Ontario, Canada, K0A 1H0. 11 ⁵ Western University, London, Ontario, Canada, N6A 3K7 12 13 ⁶Acadia University, Wolfville, Nova Scotia, Canada, B4P 2R6 14 ⁷ Faroe Marine Research Institute, Tórshavn, Faroe Islands, FO-110 ⁸ Aarhus University, Roskilde, Denmark, DK-4000 15 ⁹ Norwegian Institute for Nature Research, Tromsø, Norway, N-9296 16 ¹⁰ University of Iceland's Research Centre at Snæfellsnes, Hafnargata 3, 340, Stykkishólmur, 17 18 19 ¹¹ Arctic National Wildlife Refuge, U.S. Fish and Wildlife Service, Fairbanks, Alaska, United 20 States, 99701 21 ¹² Nova Scotia Department of Lands and Forestry, Kentville, Nova Scotia, Canada, B4N 4E5 ¹³ Institute of Geography of the Russian Academy of Sciences, Moscow, Russia, 119017 22 23 24 *Corresponding Author 25 26 27 Keywords

Environmental and life-history factors influence inter-colony and multidimensional niche

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Stable isotopes; carbon-13; nitrogen-15; Hg; isotopic niche; climate change; common eider

Abstract

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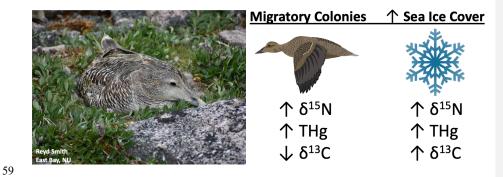
30 Human industrialization has resulted in rapid climate change, leading to wide-scale environmental shifts. These shifts modify food web dynamics by altering the abundance and 31 32 distribution of primary producers (ice algae and phytoplankton), as well as animals at higher 33 trophic levels. Methylmercury (MeHg) is a neuro-endocrine disrupting compound which 34 biomagnifies in animals as a function of prey choice, and, as such, bioavailability is affected by 35 altered food web dynamics and adds an important risk-based dimension in studies of foraging 36 ecology. We determined the multidimensional (δ^{13} C, δ^{15} N, THg) niche dynamics (MDND) among breeding common eider (Somateria mollissima) ducks sampled from 10 breeding 37 38 colonies distributed across the circumpolar Arctic and subarctic. Results show high variation in MDND among colonies as indicated by niche size and ranges in δ^{13} C, δ^{15} N and THg values in 39 40 relation to spatial differences in primary production inferred from sea-ice presence and colony 41 migratory status. Colonies with higher sea-ice cover during the pre-incubation period had higher median colony THg, δ^{15} N and δ^{13} C. Individuals at migratory colonies had relatively higher THg 42 43 and δ^{15} N and lower δ^{13} C, suggesting a higher trophic position and a greater reliance on 44 phytoplankton-based prey. We conclude that variation in MDND exists amongst eider colonies 45 which influenced individual blood THg concentrations. Further exploration of spatial 46 ecotoxicology and niche characteristics at each individual site is important to examine the relationships between anthropogenic activities, foraging behaviour, and the related risks of 47 48 contaminant exposure. Multidimensional niche analysis that incorporates multiple isotopic and contaminant metrics could help identify those populations at risk to rapidly altered food 49 web dynamics, notably important to the Indigenous populations that rely on eiders as a 50 stable resource. 51

52 Highlights

- 1. Common eider colonies vary in their multidimensional ($\delta^{15}N$, $\delta^{13}C$, THg) niche size.
- 54 2. Colonies with higher sea-ice cover had higher $\delta^{15}N$, $\delta^{13}C$ and THg.
- 55 3. Colonies considered migratory had higher average $\delta^{15}N$ and THg, but lower $\delta^{13}C$.
- 56 4. Individuals with lower δ^{13} C and higher trophic positions had higher THg.

58 Graphical Abstract

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1. Introduction

Global anthropogenic activity has resulted in environmental shifts within Arctic systems including rising air and ocean temperatures (Zhang 2005; Screen and Simmonds 2010; Najafi et al. 2015), changes in wind and ocean circulation (Timmermans et al. 2011), a dramatic modification of sea-ice cover in polar marine systems (Johannessen et al. 2004; Hoegh-Guldberg and Bruno 2010; IPCC 2019), and an elevated transport and deposition of biologically harmful contaminants (Dietz et al. 2009; Liu et al. 2012; Foster et al. 2019). These multiple stressors generate cumulative effects which have direct and indirect influences on biological systems, ultimately having the potential to affect food web characteristics including prey availability and selection (Vasseur and McCann 2005; Frederiksen et al. 2006; Parmesan 2006). Arctic marine food-webs have been particularly impacted, leading to dramatic abiotic shifts resulting in alterations of prey abundance and availability which have modified the foraging niche of higher trophic-level organisms (Moline et al. 2008; Pecuchet et al. 2020).

The foraging niche of an organism includes both dietaryc and environmental components, and therefore has been used to discern both trophic roles by consumers and changes in their

and therefore has been used to discern both trophic roles by consumers and changes in their resource use (Newsome et al. 2007). Studies examining foraging niche size (the spatial and trophic-level range at which a group forages) have been used to investigate the effect of phenological changes in primary production on predators (Rabosky 2009). In polar regions, changes in the abundance, distribution, and phenology of primary producers such as phytoplankton and sea ice-algae can have effects on higher trophic-level wildlife (Frederiksen et al. 2006; Kohlbach et al. 2016; Renaut et al. 2018; Lewis et al. 2020). Sympagic-pelagic-benthic coupling drives energy flow between the surface and benthic habitats and provides a foundation for Arctic ecosystem functioning involving benthic consumers but is being decoupled due to

changing sea-ice dynamics (Søreide et al. 2012; Kohlbach et al. 2016; Yurkowski et al. 2020a). 83 84 These shifts can generate bottom-up effects on the foraging ecology of higher trophic species, especially those that rely on consuming resources that inhabit the sea floor (Leu et al. 2011; Post 85 86 et al. 2013; Post 2017; Cusset et al. 2019). Stable isotopes of carbon (δ^{13} C) and nitrogen (δ^{15} N) are tracers that provide time-87 integrated information on habitat use and diet (Bearhop et al. 2006; Cherel and Hobson 2007; 88 Inger and Bearhop 2008). In costal polar environments, δ^{13} C provides dietary information on 89 90 sources of primary productivity and foraging habitat, for example, between ¹³C -depleted phytoplankton-derived carbon and ¹³C -enriched sea-ice derived carbon (Hobson et al. 2002; 91 92 Kohlbach et al. 2016; Yurkowski et al. 2020a; Lewis et al. 2020). In addition, benthic or nearshore environments are typically enriched in ¹³C compared to pelagic or offshore sources 93 (Hobson and Welch 1992; Hobson et al. 1995; France 1995). Values of δ^{15} N indicate relative 94 trophic level, with higher trophic levels associated with increased δ¹⁵N values (Hobson and 95 96 Welch 1992). 97 One further consequence of changing Arctic marine ecosystems is shifts in marine 98 wildlife exposure to contaminants (Muir et al. 1999; Macdonald et al. 2005; Stern et al. 2012). It 99 is well established that Arctic marine food webs are influenced by long-range transport of 100 contaminants (Macdonald et al. 2000; Braune et al. 2005; Kirk et al. 2012). On top of this, the 101 melting of permafrost, ice caps and glaciers is releasing stored contaminants from years of higher 102 contaminant output into the current system (Rydberg et al. 2010; Schuster et al. 2018). 103 Methylmercury (MeHg) is a biologically converted form of mercury (Hg) and is a contaminant 104 of concern due to its high bioavailability in organic tissues and toxic effects at even low, sub-105 lethal concentrations (Wiener et al. 2003; Whitney and Cristol 2017; Evers 2018). Uptake of

MeHg into wildlife can disrupt endocrine functioning, behaviour, and reproductive success (Cardona-Marek et al. 2009; Chen and Hale 2010; Whitney and Cristol 2017). Specifically, reproductive effects in birds include reduced clutch size, altered parental breeding behaviour and reduced hatching and fledgling success (Braune et al. 2012; Tartu et al. 2013; Goutte et al. 2014; Hartman et al. 2019). Together, these impacts may result in slowed or negative population growth rates and reproductive success in individuals exposed as juveniles (Schoch et al. 2014; Paris et al. 2018).

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Climate change and human activity in Arctic ecosystems affect the distribution and accumulation of Hg (Stern et al. 2012; McKinney et al. 2015). Further, changes in Arctic food web dynamics and trophic relationships may shift the flow of Hg between organisms (Braune et al. 2014). To study this, δ^{13} C and δ^{15} N measurements are tools that help determine environmental and dietary sources of Hg due to varying Hg uptake with carbon-source use and trophic level (Atwell et al. 1998; Cardona-Marek et al. 2009; Pomerleau et al. 2016). While δ^{15} N indicates trophic level, with greater Hg concentrations through food web biomagnification, high variability in Hg still occurs among individuals despite similar δ^{15} N values (Atwell et al. 1998; Bearhop et al. 2000; DiMento et al. 2019). Furthermore, isotope biplots consisting of just δ^{13} C and $\delta^{15}N$ obviously capture an incomplete portrayal of ecological niche due to the inclusion of only two isotopic variables. A multidimensional niche, including three or more total variables, can provide more information to assess foraging ecology (Swanson et al. 2015, Hobson et al. 2015, O'Donovan et al. 2018). Therefore, including Hg in niche analyses along with δ^{13} C and δ^{15} N provides a broader method to quantify an individual and population's foraging ecology and multidimensional niche dynamics (Yurkowski et al. 2020b). This analytical approach, combining stable isotopes and contaminants has been applied to multiple taxa including mammals, fish, and

reptiles (Guzzo et al. 2016; Jackson et al. 2016; Purwandana et al. 2016; Yurkowski et al. 2020b). However, a multidimensional approach to determine niche size has not been investigated in seabirds despite them being considered sentinels of ecosystem change (Le Bohec et al. 2013). Arctic seabirds demonstrate varying foraging specializations on a large diversity of prey items reflecting climate-induced trophic shifts and inform spatial variation and temporal changes in sea-ice and ocean dynamics that propagate up the food web, making them an ideal model to research multidimensional niche dynamics (Pratte et al. 2019; Albert et al. 2020; Renedo et al. 2020).

We used an inter-colony and inter-individual approach to examine spatial variation in the 3-dimensional niche of common eiders (*Somateria mollissima*, hereafter eiders). Eiders are a long-lived, colonial-nesting marine bird with high site fidelity and widely dispersed breeding populations across the circumpolar Arctic. Across their range, eiders are likely exposed to diverse environmental conditions that influence colony demographics at varying intensities that may influence foraging decisions and, although typically at low concentrations in eiders, Hg exposure (Mallory et al. 2004; 2017; Jónsson et al. 2013; Goudie et al. 2020; Noel et al. 2021). Eider prey primarily includes a diversity of benthic invertebrates (e.g., urchins, mussels, and gastropods) as well as pelagic macroinvertebrates to a lesser extent (e.g., amphipods; Sénéchal et al. 2011; Kristjánsson et al. 2013; Waltho and Coulson 2015). The abundance and distribution of these prey groups vary spatio-temporally and correlate with both predator Hg concentrations and phenological shifts in primary production (Mouritsen et al. 2005; Barber et al. 2015; Fort et al. 2016; Savoy et al. 2017). Thus, examining δ^{15} N, δ^{13} C and Hg niche dynamics of a costal, primarily benthic foraging species provides an avenue to investigate the future impact of prey shifts and the resulting influence on Hg exposure (Sénéchal et al. 2011), as well as relative

differences in ice algae and phytoplankton-based food webs in relation to sea-ice cover (Søreide et al. 2012; Kohlbach et al. 2016).

We used a 3-dimensional approach combining δ^{15} N, δ^{13} C and total Hg (THg) measurements collected from individuals at 10 eider breeding colonies located across their circumpolar range to evaluate variation in eider foraging ecology and Hg exposure. We anticipated that colonies would show differences in 3-dimensional niche size and ranges of each axis and be influenced by spatial differences in primary productivity (i.e., sea-ice algae and phytoplankton) and migratory behaviour. To examine these possible drivers of colony-level variation in niche metrics, we first examined colony sea-ice cover during a period reflecting periods of isotopic integration in blood. Specifically, we predicted that colonies with higher sea-ice cover would have smaller niche sizes than those with no sea ice present due to more restricted foraging habitat and likely less diverse prey availability (Yurkowski et al. 2016; Pratte et al. 2019). We also examined whether a colony is considered primarily migratory or resident and predicted that eiders from resident, non-migratory colonies would have smaller niche sizes compared to migratory colonies, given that residents remain at the same geographical location year-round and should therefore be continually exposed to a narrower range of environmental conditions than migrants (Herrera 1978; Gómez et al. 2016).

In our second objective, as colony-corrected individual values and colony ranges of δ^{13} C, δ^{15} N and THg provide important and unique information about foraging ecology, we first related these values to colony sea-ice cover in the spring during the period when birds would have been foraging at high rates to obtain body condition needed to breed. We predicted that colonies with higher sea-ice cover during the pre-breeding period would have lower δ^{13} C, δ^{15} N and THg ranges due to a restricted number of foraging locations available due to higher sea-ice concentrations, as

well as higher individual δ^{13} C, δ^{15} N and THg. Second, we related these metrics to the migratory status of a colony. We predicted that migratory colonies would have wider δ^{13} C, δ^{15} N and THg ranges due to a presumed broader distribution and use of habitat, in addition to higher median THg, δ^{15} N and lower δ^{13} C. Median values were used to avoid data skewed by high/low individual values within a mean and better represent general colony values.

Finally, we aimed to assess individual Hg exposure based on possible differences in foraging behaviour and prey selection. We examined the relationship between trophic position (the vertical placement of an individual in the food-web based on prey and individual δ^{15} N) and inter-individual δ^{13} C values on THg concentrations. We predicted that THg would increase with higher trophic position due to the biomagnification of THg, and lower δ^{13} C values because of a greater consumption of resources derived from phytoplankton (Atwell et al. 1998; McMahon et al. 2006; Stern et al. 2012). By examining variation in 3-dimensional niche size among colonies of this Arctic seabird, we aimed to identify the environmental and behavioural factors influencing niche dynamics. Furthermore, multidimensional niche can be used to infer population-level variability in resource use and overall niche diversity which allows for broad predictions about predicted flexibility and resiliency to environmental change (Vander Zanden et al. 2010; Paredes et al. 2012; Smith and Reeves 2012; Sydeman et al. 2012). As such, we also considered how our measurement of niche size may help predict the resiliency of eider colonies to environmental change (Paredes et al. 2012; Smith and Reeves 2012; Sydeman et al. 2012).

2. Methods

2.1. Study sites and sample collection

We collected whole blood samples from pre-incubating or incubating eiders in 2018 at 10 breeding colonies (total number of individuals = 240) across the Arctic and subarctic (spanning a

longitude from -147.776 to 35.774 and latitude from 78.918 to 43.645; Figure 1, Table 1). We captured eiders using colony- and breeding stage-specific techniques. Pre-incubating eiders were sampled at East Bay Island (within in the Qaqsauqtuuq (East Bay) Migratory Bird Sanctuary, Nunavut, Canada), whereas incubating eiders were sampled at the other nine locations.

At East Bay Island, eiders were captured using large flight nets. We collected birds from the nets and drew a 1-mL tarsal blood sample from each female eider (Hennin et al. 2015). These samples were used for isotopic analyses and were collected using a 23G thin-wall, 1-inch needle attached to a sodium-heparinized 1 mL syringe (Lemons et al. 2012). After transferring to a heparinized 1.5 mL Eppendorf tube, samples were kept cool (~10°C), and within 8 h, were centrifuged at 10,000 rpm for 10 min to separate red blood cells (RBCs) and plasma. Plasma was then transferred by pipetting into a separate cryovial and stored along with RBC samples at - 20°C until isotopic analysis. During the same capture period as the first sample, a second blood sample of 1-mL was obtained from the jugular vein using 23G thin wall, 1-inch needles attached to heparinized 3mL syringes to be used for Hg analysis. The whole blood sample was placed in acid-rinsed cryovials and kept at ~10°C, and frozen within 6 hours of collection until analysis.

At the other nine colonies, female eiders were captured on their nest during incubation using either a bownet, noose pole or dogs (John's Island location only; Milton et al. 2016). The specific incubation stage could not be determined at all sites or for all individuals and was excluded from analysis. For most sites, we collected 200-1000 μL of blood from the tarsal vein using a 23G thin-wall, 1-inch needle attached to a heparinized 1 mL syringe. At the Alaskan site, up to 5mL of blood was collected from the jugular vein using a non-heparinized syringe, while at the Icelandic site, blood was collected from the brachial vein. After collection, samples were kept cool and transferred to heparinized 1.5mL cryovials. Within 8-12 h, samples were

centrifuged at approximately 10,000 rpm, dependent on centrifuge model, for 10 min to separate red blood cells (RBCs) and plasma. Plasma was then placed into a separate cryovial and stored along with RBC samples at a minimum of -20°C until analysis.

2.2. Stable isotope analysis

Stable isotopes of elements turnover at different rates based on dietary composition, metabolic rates, and specific metabolic activity of the tissue (Hobson and Clark 1992). Isotopic half-life is dependent on body size and can be affected by temperature exposure (Carleton and Del Rio 2005). For avian blood components, plasma has fast turnover rate that reflects recent diet (days), while the turnover rate of RBCs is slower and reflects diet over a longer period (weeks; Hobson and Clark 1992; Barquete et al. 2013). Further, δ^{15} N and δ^{13} C usually have similar turnover times, hence are comparable (Bearhop et al. 2002). Stable isotope ratios were measured in plasma at East Bay Island and in RBCs at the other nine colonies. Therefore, despite sampling colonies at different life history stages, by using different tissues for analysis we were still able to compare them within a similar stage including the pre-incubation period while eiders are heavily foraging, either at resident sites or in the last period of migration to build up stored resources for their incubation fast.

Samples were freeze-dried for approximately 72 h after collection. Samples were then ground into a fine, homogenized powder using a metal spatula cleaned with acetone. Plasma samples were then lipid extracted using a 2:1 chloroform:methanol solution (Søreide et al. 2006). Specifically, 2:1 chloroform:methanol solution (1.9mL) was added to 100µL of each plasma sample, which were then placed in a water bath at 30°C for 24 h. Samples were then centrifuged at 15,000 rpm for 10 min to separate the lipid solution from the plasma pellet. A p1000 pipette was used to remove the lipid solution, leaving the plasma pellet behind. The pellet was washed

244 again with an additional 1.9 mL of the chloroform:methanol solution and centrifuged for a final 245 10 min at 15,000 rpm. The remaining lipid solution was removed, leaving only a plasma pellet. 246 Samples were left open in the fume hood for 24 h to allow for any remaining 247 chloroform:methanol solution to evaporate. Since RBCs have minimal lipids present, they did 248 not undergo lipid extraction (Hobson and Clark 1992). 249 Plasma and RBC samples were weighed into individual tin-capsules using a 4-digit 250 balance to obtain 0.3-0.5 mg of sample. Plasma samples were prepared at the University of 251 Windsor, and plasma stable isotopes were analysed at the Environment and Climate Change 252 Canada Stable Isotope Laboratory in Saskatoon, Saskatchewan, using continuous-flow isotope-253 ratio mass spectrometry (CFIRMS; Harris et al. 1997). Samples were weighed into pre-254 combusted tin capsules. Encapsulated plasma was combusted at 1030°C in a Carlo Erba NA1500 255 or Eurovector 3000 elemental analyser. The resulting N2 and CO2 were separated 256 chromatographically and introduced to an Elementar Isoprime or a Nu Instruments Horizon 257 isotope ratio mass spectrometer. Two reference materials were used to normalize the results to 258 VPDB and AIR: BWBIII keratin (δ^{13} C = -20.18, δ^{15} N = +14.31 ‰, respectively) and PRCgel $(\delta^{13}C = -13.64, \delta^{15}N = +5.07 \%$, respectively). Within run (n = 5) precisions as determined from 259 260 both reference and sample duplicate analyses were \pm 0.1 ‰ for both $\delta^{13}C$ and $\delta^{15}N$. 261 The RBC samples were prepared for isotope analysis at La Rochelle University, France, 262 and were analysed at the LIENSs institute (La Rochelle, France) as detailed in Fort et al. (2014). 263 Plasma and RBC samples were combusted using a Eurovector 3000 (Milan, Italy) elemental 264 analyser which results in production of N2 and CO2 gases. These were separated by gas 265 chromatography and introduced into a NU Horizon (Nu Instruments, Wrexham, UK) triple-266 collector isotope-ratio mass-spectrometer via an open split. Ratios of carbon (13C/12C) and

267 nitrogen ($^{15}N/^{14}N$) were expressed in typical delta notation (δ) as per mil (‰) deviation from the 268 primary standards (Vienna Pee Dee Belemnite (VDPB) and atmospheric nitrogen (AIR), 269 respectively). Replicate measurements (n = 20) per run of laboratory standards (USGS-61 and 270 USGS-62) indicated that the measurement accuracy was <0.2% for both $\delta^{15}N$ and $\delta^{13}C$ values. 271 Blanks were run at the beginning of each sample set. Detection limit was set to $0.005 \mu g/g dw$. 272 Due to biogeochemical factors, baseline stable isotopes of Arctic marine food webs vary 273 spatiotemporally (de la Vega et al. 2019). Variation in food web δ^{13} C of marine environments is 274 driven largely by primary production growth rate and inorganic substrate (e.g., water 275 temperature, amount and isotopic signature of dissolved CO₂) δ^{13} C and these, in turn, can be 276 influenced by sea-ice cover (de la Vega et al. 2019). Stable nitrogen isotope values in marine 277 food webs are influenced by spatial and temporal changes in temperature, increasing abundance 278 of phytoplankton that fix ¹⁵N, trophic level as well as inputs of nitrogen from terrestrial sources 279 (Smith et al. 1999; Tamelander et al. 2009; McMahon et al. 2013). Importantly, since Arctic food webs are expected to vary isotopically, measured δ^{13} C and δ^{15} N values of biota from different 280 281 regions can only be compared if corrected for such baseline variance (Bowen 2010; Hobson et al. 282 2012; de la Vega et al. 2019). Based on previously published δ^{13} C and δ^{15} N values of known 283 eider bivalve prey at the different breeding sites, we corrected our data by subtracting baseline 284 bivalve isotope values from the eider tissue values to obtain "corrected" isotopic values (Table 285 2). However, Hg values were not corrected to baseline values. Additionally, we were unable to account for the potential impact of incubation stage on δ^{15} N, δ^{13} C or Hg values. Both Hg and 286 287 δ^{15} N values can be elevated in blood due to mobilization of internal nutrient stores during the 288 incubation fast (Hobson et al. 1993; Wayland et al. 2005). However, our use of RBCs instead of 289 plasma to determine incubating female δ^{15} N values minimized this effect (Cherel et al. 2005).

Furthermore, our sampling of eiders during incubation, when Hg is known to increase (Wayland et al. 2005), lessens but does not mitigate the potential bias of East Bay Island pre-breeding eiders having elevated Hg relative to the other colonies as the pre-breeding eiders have not yet depurated Hg to their eggs during laying (Akearok et al. 2010). Hence, caution should be taken when comparing Hg values between East Bay Island and the other nine colonies.

Diet-tissue isotopic discrimination factors allow modeling of isotopic trophic positions or nutrient source tracing (e.g., Wolf et al. 2009; Federer et al. 2010; Bond and Diamond 2011). Such factors have not been established for common eiders and so we used those reported by Federer et al. (2010) for spectacled eider (*Somateria fischeri*) (plasma: +4.9‰; RBC: +4.0‰). Trophic position was calculated using baseline $\delta^{15}N$ bivalve values for the individual's colony ($\delta^{15}N_{base}$), discrimination factors from the respective tissue of spectacled eiders ($\delta^{15}N_{TDF}$; Federer et al. 2010), trophic position (TP) of baseline prey values (TP_{base}; value of 2 consistent with eider prey), and the non-colony corrected $\delta^{15}N$ values for that individual eider ($\delta^{15}N_{eider}$; Hobson and Welch 1992; Vander Zanden et al. 1997; Vander Zanden and Rasmussen 1999).

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$$TP = ((\delta^{15}N_{eider} - \delta^{15}N_{base})/\delta^{15}N_{TDF}) + TP_{base}$$

2.3. THg analysis

Whole blood collected at East Bay Island was sent to the Research and Productivity Council (RPC) in New Brunswick, Canada for THg analysis. Each sample was prepared by microwave-assisted digestion in nitric acid (SOP 4.M26). Mercury was then analysed by cold vapour atomic absorption spectroscopy (AAS; SOP 4.M52 & SOP 4.M53) to obtain THg concentrations.

Quality assurance/control procedures included analysis of four reagent blanks, and reference materials from dogfish liver (DOLT-2) and lobster hepatopancreas (TORT-2) from the National

Research Council (NRC), Canada, as well as four randomly selected duplicate samples. All samples followed quality control procedures. Mercury concentrations were converted from wet weight to dry weight for comparison with the other colonies following known equations and moisture values for avian blood at 79% (Eagles-Smith et al. 2008).

RBCs separated from plasma collected from incubating females. These eiders were captured while incubating (Hanssen et al. 2002; Bottitta et al. 2003; Sénéchal et al. 2011), thus by collecting and analysing RBCs alone, the timeframe which the RBCs represent aligns with the same timeframe represented in the East Bay Island eiders (pre-incubation period for all colonies) since whole blood has a turnover rate approximately intermediate of plasma and RBC (Cherel et al. 2005). Therefore, this provides justification for including the colony in our analysis despite potential differences in physiology due to their reproductive stages. Freeze-dried blood was analysed using an Advanced Hg Analyser spectrophotometer (Altec AMA 254). A quality control program included running blanks prior to the analysis, as well as the same certified reference materials used to analyze the samples from East Bay Island (DOLT-2 and TORT-2) analysed every 15 samples. Samples were analyzed for THg since Hg exists as primarily MeHg in avian blood (near 1:1 ratio; Wiener et al. 2003; Rimmer et al. 2005).

2.4. Statistical analysis

We estimated 3-dimensional niche size (using THg, δ^{13} C and δ^{15} N) and ranges along the THg, δ^{13} C and δ^{15} N axes for 10 eider colonies using the R package nicheROVER v1.0 (Swanson et al. 2015) in R version 3.6.2 (R Development Core Team). NicheROVER uses Bayesian statistical methods to calculate a multidimensional niche region in multivariate space that represents the spatial breadth and placement of a group's (i.e., colony's) niche in relation to other groups and

infers the niche size of each group based on the group's individual values (Swanson et al. 2015). Since the variables are in different units (δ^{13} C and δ^{15} N: ‰ vs. THg: $\mu g/g^{-1}$), we scaled and centred all values by subtracting the mean for each variable and dividing it by the standard deviation, thereby standardizing all data. To determine if there were any correlations among variables prior to analysis, we tested for independence by calculating Spearman's correlation coefficient between THg and colony-corrected δ¹⁵N which showed a significant but weak correlation (R=0.429, p<0.001). Thus, THg and δ ¹⁵N, while related, do possess independent variation that allows us to use both in our analysis to account for spatial variation in THg concentrations. We calculated a 95% probability niche region in multivariate space at 10,000 iterations using diffuse priors (Swanson et al. 2015; Yurkowski et al. 2020b). We ran additional script at 10,000 iterations to obtain δ^{13} C, δ^{15} N and THg ranges (difference between the highest and lowest values; Swanson et al. 2015; Yurkowski et al. 2020b). Three-dimensional models of the niche ranges were visualized using the scatter3D function in the car package v.3.0-9 (Fox and Weisberg 2019) and the rgl package v.0.100.54 (Adler and Murdoch 2017; see Supplemental Materials). Primary migratory status of each colony, not including the potential for individual variation, was determined by consulting previous studies (Schamel 1997; Bønløkke et al. 2006; Hanssen et al. 2016; Steenweg et al. 2017; Mallory et al. 2020) and through spatial tracking data collected by the SEATRACK program (https://seapop.no/en/seatrack/). Moreover, we determined the proportion of sea-ice cover surrounding each colony within a timeframe reflecting turnover rates of isotope sampling (plasma within a week of sampling, RBC within a month of sampling; Hobson and Clark 1992; Barquete et al. 2013). Sea-ice cover was determined

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via a satellite image of a 100 km area around each colony on a cloud-less day from NASA

Worldview dataset (EOSDIS, https://worldview.earthdata.nasa.gov). Land pixels were removed from the image manually and the remaining pixels were categorized into two groups (open water or sea ice) using a K-means clustering procedure via RGB values of the pixels (http://mkweb.bcgsc.ca/color-summarizer/). The resulting proportion of pixels was used as a proxy for the proportion of sea ice around the colony during the time of isotopic turnover for each colony (Cusset et al. 2019).

We ran a preliminary 2-tailed t-test to determine independence between colony sea-ice cover and migratory behaviour and found they were not significantly correlated (t_5 =2.11, p=0.09). Therefore, to determine sources of niche variation with sea ice cover, we first ran a general linear model (GLM) to determine how log-transformed niche size varied by colony sea ice cover. Next, we conducted six GLMs to analyze the relationship between sea-ice cover and colony δ^{15} N, log-transformed δ^{13} C and THg ranges, as well as median individual colony-corrected δ^{15} N, and log-transformed δ^{13} C and THg values for each of the 10 colonies.

To examine variation in niche dynamics between migratory and resident colonies we conducted a 2-tailed t-test to analyze how log-transformed colony niche size varied with migratory status (migratory or resident being the two groups). Following this, we ran six, 2-tailed t-tests to analyze whether migratory status of a colony resulted in different colony δ^{15} N, and log-transformed δ^{13} C and THg ranges, as well as individual colony-corrected eider δ^{13} C, δ^{15} N and log-transformed THg values.

Finally, we used a general linear mixed model (GLMM) to examine the relationship between colony-corrected isotopes indicating carbon source (δ^{13} C), trophic position of each individual, and log-transformed THg. The model consisted of THg as the dependent variable, with δ^{13} C and trophic position as independent variables, as well as colony as a random variable.

381 All models conducted throughout the study met statistical assumptions, and all log-382 transformations were conducted using natural log. 383 3. Results 384 3.1. Niche size and underlying sources of variation 385 Three-dimensional niche size was highly variable among colonies, ranging from 1.4 (Grindøya) 386 to 21.7 (Iceland), with an average niche size among all colonies of 9.2 ± 7.8 (Figure 2, see 387 Supplemental Materials). However, niche size was not correlated with colony sea ice cover (t₈=-388 0.92, p=0.54) or migratory status ($t_4=-0.61$, p=0.57). The Christiansø colony in Denmark had the 389 lowest colony-corrected δ^{13} C values, suggesting higher phytoplankton-derived carbon in their 390 diet, while the Alaskan colony had the highest colony-corrected δ^{13} C suggesting eiders were 391 feeding on prey reflecting more ice algae- or inshore-derived carbon (see Supplemental 392 Materials). The Alaskan colony also had the highest colony-corrected $\delta^{15}N$ values, with the 393 Faroe Islands having the lowest values, suggesting that Faroese eiders forage at lower trophic 394 levels, whereas the Alaskan eiders forage at higher trophic levels. 395 3.2. Relationship between colony migratory behaviour and sea-ice cover with isotopes and 396 THg397 Inter-colony variation in sea-ice presence was positively correlated with median eider THg 398 concentrations, as well as δ^{15} N and δ^{13} C values (Table 3). However, δ^{15} N, δ^{13} C as well as THg 399 ranges did not vary with colony sea-ice cover (Table 3). Migratory colonies had higher 400 individual δ^{15} N and THg, as well as lower individual δ^{13} C compared to individuals captured at resident colonies (Table 3). However, δ^{15} N range, δ^{13} C range and THg range did not vary 401

between the migratory and resident colonies (Table 3).

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403 3.3. Relationship between isotopic niche and THg concentrations 404 Both colony-corrected δ^{13} C and trophic position predicted THg concentration within individuals. 405 Specifically, individuals with a lower δ^{13} C value (phytoplankton-based foraging) had higher THg 406 values (Table 3, Figure 3). Additionally, THg significantly increased with trophic position, 407 where individuals with higher trophic positions had greater THg values (Table 3, Figure 3). 408 4. Discussion 409 Using data collected from ten eider colonies located throughout their circumpolar range, 410 including both Arctic and subarctic sites, we quantified 3-dimensional niche size using $\delta^{15}N$, 411 δ^{13} C and THg to determine colony niche size and ranges. Determination of 3-dimensional niche 412 size allowed us to broadly compare a snapshot of diet breadth at multiple eider colonies in 413 relation to environmental and behavioural differences, as well as infer their potential for 414 flexibility in response to environmental change. We demonstrate the benefits of using multiple 415 chemical tracers when assessing the effects of spatial variation and environmental gradients on 416 the foraging ecology of highly mobile consumers (Ramos and González-Solís 2012). This study 417 represents the first inter-colony, multidimensional niche size analysis for any seabird and the 418 resulting colony-level relationships with THg exposure, an area identified as a key knowledge 419 gap (Dietz et al. 2019). 420 4.1. Colony foraging flexibility and resiliency predictions 421 Generally, colonies that utilize broader, more generalist diets are expected to have larger 422 foraging niches, thus are expected to be inherently less vulnerable to changes since they have 423 greater flexibility in their prey source as they forage on a wide variety of organisms and habitats 424 (Jakubas et al. 2017; Both et al. 2010). Colonies with restricted, specialist diets are expected to 425 have smaller foraging niches, influenced by both prey availability and foraging distance, and are

generally viewed as more vulnerable to environmental changes (Bolnick et al. 2003; Araújo et al. 2011; Pratte et al. 2019). We examined the 3-dimensional niche size of eider breeding colonies, representing prey selection and trophic connections through stable isotopes and Hg exposure. Multidimensional niche analysis may be useful in predicting the resiliency of common eider colonies to ongoing climate change across their range, although numerous other factors should also be considered including multiple biogeochemical metrics, colony recruitment and individual fitness (Paredes et al. 2012; Smith and Reeves 2012; Sydeman et al. 2012). However, our analysis provides a snapshot of niche characteristics at a singular time frame that may assist with colony-wide resiliency predictions in combination with future research on changing niche dynamics at these colonies.

When grouped for similarity, niche size generated the following pattern in our study: Iceland, East Bay Island and Faroe Islands > Christiansø and John's Island > Kongsfjorden, Alaska and Russia > Tern Island and Grindøya (Figure 1). Using our 3-dimensional view of niche size, we predict that colonies with wider (i.e., more generalist) 3-dimensional niches such as those in Iceland and Faroe Islands, or colonies comprised of migrants from multiple locations

Iceland, East Bay Island and Faroe Islands > Christiansø and John's Island > Kongstjorden,
Alaska and Russia > Tern Island and Grindøya (Figure 1). Using our 3-dimensional view of
niche size, we predict that colonies with wider (i.e., more generalist) 3-dimensional niches such
as those in Iceland and Faroe Islands, or colonies comprised of migrants from multiple locations
such as East Bay will show greater resiliency to shifts in food web dynamics compared to
colonies with a smaller (i.e., more restricted, specialist) niche similar to Tern Island and
Grindøya. Smaller niches are predicted to be a disadvantage in a changing climate as these
alterations may change or eliminate food sources, leaving a colony or species more vulnerable if
they do not have the flexibility to adapt and shift their diet with this changing prey base (Both et
al. 2010; Le Bohec et al. 2013; Ceia and Ramos 2015). However, climate change may present an
opportunity for Arctic colonies with smaller niches to expand their prey sources with advancing
phenology of spring phytoplankton blooms and more open water sources. For those with broader

niches at southern latitudes, that niche size may be critical as we see further changes in the distribution and availability of key prey (Staudinger et al. 2019). Nevertheless, more research that integrates measures of breeding success and colony demographics is necessary to test these resiliency hypotheses over long temporal scales to identify seasonal and annual variation in interand intra-colony foraging ecology. 4.2. Variation in niche size, niche metrics and THg to colony sea-ice cover Colony niche size was not correlated with sea-ice cover, possibly attributable to several factors. First, despite the presence of sea ice likely restricting foraging locations, eiders may still be able to access a diversity of resources, including the potential for prey sources such as ice-associated amphipods as niche size was similar to southern locations that have more accessible, ice-free areas (Karnovsky et al. 2008). Second, northern colonies may support a variety of individuals that utilize both generalist and specialist strategies, thus adding to the flexibility and diversity of the colony in the face of environmental change (Woo et al. 2008; Ceia and Ramos 2015; Pratte et al. 2019). Notably, the relative proportion of generalist and specialist individuals in populations dramatically affects dietary niche size with generalist-based populations having wider niches (Newsome et al. 2007). We found that eider colonies with greater sea-ice cover had higher δ^{13} C values, which is not consistent with the expected gradient based on ocean temperature (Sackett et al. 1965; Goericke and Fry 1994; McMahon et al. 2013). However, this pattern can be most likely explained by eiders foraging on more resources derved from ice algae with higher ice cover versus more phytoplankton-derived resources in areas with lower sea-ice presence (Hobson et al. 1995; Tamelander et al. 2006; McMahon et al. 2006). As well, macroalgal carbon has a higher

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 δ^{13} C value than phytoplankton and therefore could be a contributing carbon source to these

spatial differences. Nonetheless, we show spatial differences in carbon-source use amongst eider colonies, where some colonies use more phytoplankton-derived carbon than others. Furthermore, our results suggest that in general, eider colonies with greater sea-ice cover also had higher median individual THg and δ^{15} N, suggesting that sea-ice presence results in eiders foraging at higher trophic levels, thus contributing to higher overall THg exposure. Potential reasons for higher trophic-level foraging include eiders feeding on ice-associated sources of prey prior to laying, such as amphipods, due to restricted access to benthic prey with sea ice present (Karnovsky et al. 2008). Amphipods seasonally forage on ice algae and this would decouple them from the pelagic food web and result in higher δ^{13} C values (Werner 1997; Brown et al. 2017). Furthermore, waters with sea ice present are supersaturated with dissolved THg, thus further contributing to the elevation of THg in food webs (DiMento et al. 2019). Individual variation in foraging behaviour is known to have a role in Hg accumulation (Anderson et al. 2009; Braune et al. 2014; Le Croizier et al. 2019) and in our study, individual variation exists within colonies as demonstrated by the colony ranges. 4.3. Role of migration in shaping niche dynamics and THg exposure We expected migratory populations (see Table 1, Figure 1) to have larger niches due to broader exposure to a variety of environmental conditions and prey types experienced at overwintering, migratory stopover, and breeding grounds (Herrera 1978; Gómez et al. 2016). In the present

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(Kristjánsson et al. 2013). Waters around Iceland are a mix of Arctic (East-Greenland Current),

study we found no pattern with migratory behaviour and niche size. However, two of the largest

niche sizes found at two resident colonies (Iceland and Faroe Islands) may stem from colony-

wide expansion in dietary choice due to increased prey availability. This has been reported in

Iceland where eiders within the colony selected a wide array of both benthic and pelagic prey

as well as Atlantic origins (North Atlantic Current), and this, together with greater lack of sea ice, seem to provide a diverse range of prey options to pre-breeding eiders (Vincent 2010; Kristjánsson et al. 2013; Casanova-Masjoan et al. 2020). The East Bay Island colony, consisting of migrants from Greenland and Newfoundland, had the second largest niche size potentially in part due to a mixing of individuals from both overwintering sites (Steenweg et al. 2017). Additionally, individual variation in migration distance and strategy within a colony, although not included in the present study, may lead to broader colony niche ranges (Mallory et al. 2020).

Colonies supporting migratory eiders had higher THg concentrations and foraged at higher trophic levels compared to colonies supporting residents, suggesting that colonies of migratory individuals have access to higher trophic-level prey, or rely on these prey to store resources necessary for migration. Colonies with resident eiders with higher individual δ^{13} C levels partially indicate diets with greater foraging based on ice algae, potentially due to residing at their Arctic breeding sites year-round while migratory colonies move to more suitable areas with presumably lower sea ice. Combined with our results on colony latitude, these dynamics suggest that changes in environmental conditions and variability may have different effects on migratory and resident seabird colonies dependent on future colony flexibility, thus should be factored into future analyses especially regarding climate change affects within Arctic ecosystems.

4.4. Relationship between trophic position, foraging location and THg concentrations

With increasing industrial activity and contaminant deposition due to long-distance transport in northern latitudes, as well as the release of stored Hg from melting ice and increased erosion of terrestrial sources (Sorenson et al. 2016, Cossa et al. 2018), Arctic-inhabiting species are potentially becoming more at risk for contaminants exposure and a suite of possible negative

518 neurological, physiological and reproductive impacts (Dietz et al. 2013; Scheuhammer et al. 519 2015). This is especially a potential population stability concern for harvested species, such as 520 common eider, which are an important, harvested species for many Indigenous communities in 521 the North (Nakashima and Murray 1988; Priest and Usher 2004). Concentrations of THg for the 522 eider colonies in this study are comparable to those observed in previous eider research 523 (Provencher et al. 2016; Albert et al. 2019; Ma et al. 2020; Dietz et al. 2021). The mean 524 concentrations of THg in the Alaska, Christiansø and East Bay colonies were above 0.95 µg/g 525 dw, suggesting that three of the 10 colonies studied are therefore considered to be above general 526 environmental background concentrations (Eagles-Smith et al. 2008; Ackerman et al. 2016). 527 Only 40% of eiders had higher concentrations of THg (above recognized background 528 concentrations of 0.95 ug/g dw), but these concentrations assign them a general "low risk" status 529 regarding concerns for health, physiology, behaviour and reproductive effects (Ackerman et al. 530 2016; Dietz et al. 2019; 2021). The individual eider with the highest Hg level was from the 531 Iceland colony (2.55 μg/g dw compared to the colony mean of 0.95 μg/g dw), which features 532 less sea ice than other regions such as Alaska, which had the highest colony THg average (1.23 533 $\mu g/g dw$). 534 With a rapidly declining sea-ice cover, a rise in phytoplankton production and 535 consumption by eiders could be related to higher, individual THg concentrations (Stern et al. 536 2012). Combined with our results in section 4.2 showing higher median THg concentrations at 537 colonies with greater sea-ice cover, our results demonstrate that within colonies, individuals with 538 greater phytoplankton-based consumption had higher THg concentrations. The relationship 539 between individual δ^{13} C and THg, combined with effects of trophic position (determined using 540 δ^{15} N) on THg may potentially have future effects on seabird health, behaviour and reproduction

in those with at-risk Hg exposure. However, it is likely that any possible effects would not be consistent across the entire range of a species, especially given the regional difference in which climate change is altering the Arctic (Muir et al. 1999; Mallory and Braune 2012; Swart et al. 2015). Nevertheless, we did detect variation among colonies, suggesting regional differences in the relationship between δ^{13} C, trophic position and THg; for instance, the Christiansø and Kongsfjorden colonies showed opposite relationships between THg and δ^{13} C (Figure 3). Thus, while we conducted a broad-scale study into these relationships, more in-depth analyses of the environmental factors at each site would allow for site-specific investigation into the relationships between foraging behaviour and THg uptake. Additionally, further knowledge of baseline variation (THg, δ^{15} N, δ^{13} C) in colony-specific prey at a varying spatio-temporal scale, parallel with eider sampling, would aid future interpretation of eider niche size and sources of individual and colony THg. The contribution of terrestrial carbon sources and agricultural runoff to marine areas around some of these colonies may also contribute to variation in individual δ^{13} C and $\delta^{15}N$ values and affect the rate of THg accumulation (Dunton et al. 2006; Laursen et al. 2018; Renedo et al. 2020). Overall, the relationship between environmental conditions and THg concentrations presents a complex system and paired with the rapid changes in ecosystem dynamics currently being observed (i.e., sea ice abundance), underscores the necessity of future, and consistent annual monitoring, especially in key ecosystem indicators like seabirds.

5. Conclusion

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We examined inter-colony, 3-dimensional niche size, isotopic and THg ranges, and the relationships between δ^{13} C, δ^{15} N and THg within individuals. Our results suggest that common eider colonies across the Arctic and subarctic have a wide degree of diet variation, potentially influenced by environmental changes including spatio-temporal differences in primary

productivity. We found that consideration of colony-level variation in life histories (migratory vs resident) was important for assessing the level of risk to which a colony may be exposed. Further, inter-annual multidimensional niche analyses would provide greater insight into the temporal variation in niche size that exists at both an inter- and intra-colony scale. Taken together, exploring multidimensional niche metrics is an increasingly valuable tool to provide insight into how diet breadth differs across a species' range, especially when combined with temporal and spatial variation in environmental conditions and key components to variation in life history such as migration.

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Tables and Figures

Table 1. Locations of common eider colonies included in this study, sample size (number of individual birds from each colony included in the study), month samples were obtained, coordinates (latitude and longitude), whether a colony is known to be predominately migratory or resident, and the proportion of sea ice cover in a 100km area around the colony during the appropriate timeframe reflecting isotope data.

Location	n	Sampling month	Latitude	Longitude	Migratory behaviour	Sea ice cover
Kaktovik, Alaska	33	July	70.340	-147.776	Migratory	43.45
Breiðafjörður, Iceland	23	June	65.078	-22.736	Resident	3.10
Christiansø, Denmark	25	May	55.330	15.188	Migratory	0
Grindøya, Norway	17	June	69.633	18.844	Resident	0
John's Island, Canada	19	July	43.645	-66.041	Migratory	0
Kirkjubøhólmur, Faroe Islands	16	July	61.950	-6.799	Resident	0
Kongsfjorden, Norway	16	June	78.918	11.910	Migratory	0
East Bay Island, Canada	43	June	64.023	-81.790	Migratory	34.34
Onega Bay, Russia	24	June	65.048	35.774	Resident	0
Tern Island, Canada	24	June	69.547	-80.812	Migratory	36.47

Table 2. Isotopic signatures (δ^{13} C, δ^{15} N in ‰) of typical common eider bivalve prey as determined by the most spatio-temporally appropriate literature values available at the time of analysis. Prey stable isotope data was used to correct common eider isotope data to allow for inter-colony comparison.

Location	Prey Species	δ ¹³ C	$\delta^{15}N$	Reference
Beaufort Sea, Alaska	Cyrtodaria kurriana	-25.20	7.90	Dunton et al. 2012
Breiðafjörður, Iceland	Mytilus edulis	-19.60	7.40	Sarà et al. 2007
Christiansø, Denmark	Limecola balthica	-20.40	7.20	Ek et al. 2018
Grindøya, Norway	Hiatella arctica	-19.32	7.26	Fredriksen 2003
John's Island, Canada	Mytilus edulis	-19.99	7.17	English et al. 2015
Kirkjubøhólmur, Faroe Islands	Mytilus edulis	-19.20	8.41	Bustamante, unpub.
Kongsfjorden, Norway	Hiatella arctica	-20.30	6.90	Vieweg et al. 2012
East Bay Island, Canada	Hiatella arctica	-18.22	8.64	Sénéchal et al. 2011
Onega Bay, Russia	Styela rustica	-21.60	6.49	Yakovis et al. 2012
Tern Island, Canada	Hiatella arctica	-18.22	8.64	Sénéchal et al. 2011

Table 3. Results of three sets of statistical analyses: a) seven 2-tailed t-tests relating migratory status to colony niche size, isotope and THg ranges, as well as individual colony-corrected isotope and THg values; b) seven GLM results relating colony sea ice cover to colony niche size, isotope and THg ranges as well as median colony isotope and THg values; c) GLMM results relating common eider trophic position, calculated using baseline and individual $\delta^{15}N$ and colony-corrected $\delta^{13}C$ to individual THg values. All log-transformed values were determined using the natural log and used to meet model assumptions.

Variable	Estimate (β)	SE	df	t	p				
a) 2-tailed t-test – Migratory Status									
log(Niche size)			4	-0.609	0.573				
$log(\delta^{13}C Range)$			8	-0.339	0.743				
$\delta^{15}N$ Range			7	0.996	0.353				
log(THg Range)			5	-0.600	0.576				
Individual $\delta^{13}C$			238	2.218	0.028				
Individual $\delta^{15}N$			220	-6.858	<0.001				
log(Individual THg)			124	-6.329	<0.001				
	b) <u>G</u> l	LM – Sea l	ce Cover						
log(Niche size)	-0.0125	0.019	8	-0.639	0.541				
$log(\delta^{13}C Range)$	-0.013	0.008	8	-1.635	0.141				
$\delta^{15}N$ Range	-0.002	0.014	8	-0.106	0.919				
log(THg Range)	-0.004	0.009	8	-0.459	0.658				
$log(Median \delta^{13}C)$	0.028	0.010	8	2.936	0.032				
Median δ ¹⁵ N	0.052	0.022	8	2.332	0.048				
log(Median THg)	0.014	0.006	8	2.525	0.036				
c) GLMM – Individual log(THg)									
Intercept	-1.923	0.304	142.233	-6.330	< 0.0001				
Trophic position	0.563	0.096	234.614	5.862	<0.001				
$\delta^{13}C$	-0.063	0.022	232.943	-2.813	0.005				

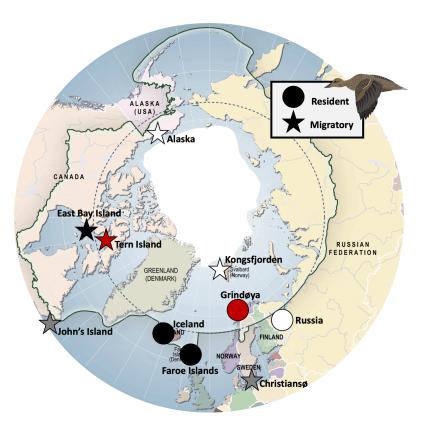


Figure 1. Map of the circumpolar-Arctic, and locations of the 10 common eider colonies used in this study (stars) designated by whether a colony is primarily resident (circles) or migratory (squares). Shape colour represents four groupings of similar median 3-dimensional niche size (95% Bayesian credible intervals). By ascending niche size, colours represent: 1 (smallest; red), 2 (white), 3 (grey), and 4 (largest; black). Dashed line indicates the Arctic Circle, and the solid line indicates the Arctic boundary according to the Arctic Monitoring and Assessment Programme (AMAP; map provided by globalcitymap.com).

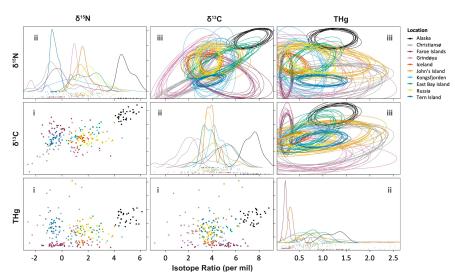


Figure 2. 2-dimensional projections of ten 3-dimensional niche regions produced using the R package nicheROVER v1.0 (Swanson et al. 2015). Corrected stable isotope (δ^{13} C and δ^{15} N in %) and THg data (in μg/g dw) were used from individuals at 10 pan-Arctic or subarctic common eider colonies. Sections show, i) The raw stable isotope and THg data in pairs, ii) density estimates of δ^{15} N, δ^{13} C and THg individually, and iii) pairings of δ^{15} N, δ^{13} C and THg showing 2-dimensional projections of 95% probabilistic niche regions based on 3-dimensional data.

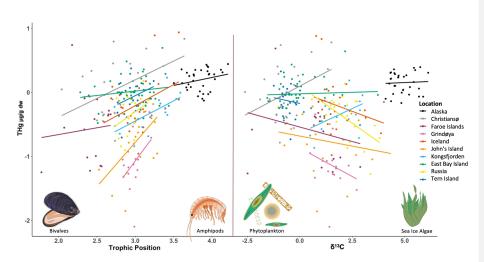


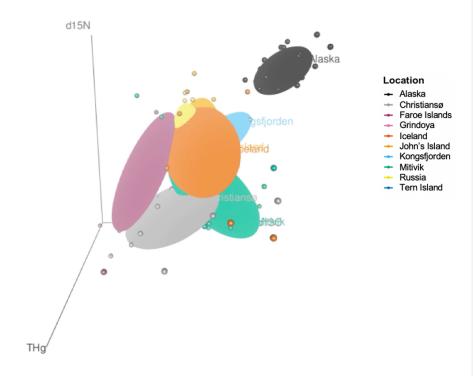
Figure 3. Relationship between THg (natural log-transformed) with trophic position (calculated based on $\delta^{15}N$; see Methods) and colony-corrected $\delta^{13}C$ (in ‰) for individuals at 10 pan-Arctic or subarctic common eider colonies, identified by colour.

Supplemental Materials

Sample sizes (n), means \pm SD, Min-Max (in ‰) for δ^{13} C, δ^{15} N, and total Hg (THg in μ g/g dw) (values in brackets corrected for colony baseline (δ^{13} C, δ^{15} N) and standardized for units (δ^{13} C, δ^{15} N and THg)), medians and range of 3-dimensional niche size (95% Bayesian credible intervals), and δ^{13} C, δ^{15} N and THg ranges (maximum-minimum values using corrected and standardized values) of 10 pan-Arctic and subarctic common eider colonies.

Location		\$13 <i>C</i>	δ^{13} C δ^{15} N THg	THe	Niche size	δ ¹³ C	$\delta^{15}N$	THg
Location	n	0 °C		Niche size	range	range	range	
Alaska	33	$\text{-}20.1 \pm 0.5$	15.3 ± 0.7	1.2 ± 0.2	3.3 (2.3, 5.2)	0.866	1.461	1.689
Alaska	33	(1.8)	(1.8)	(0.9)	3.3 (2.3, 3.2)	0.800		
Christiansø	25	$\textbf{-21.2} \pm 0.9$	10.0 ± 1.3	1.1 ± 0.5	0.7 (((16.7)	1.379	2.923	4.590
Cili istialisø	23	(-1.0)	(-0.9)	(0.6)	9.7 (6.6, 16.7)	1.379	2.923	
Faroe Islands	16	$\textbf{-}18.5 \pm 1.4$	10.1 ± 1.0	0.7 ± 0.4	18.8 (11.5, 36.8)	2.059	2.106	4.508
raioe islands	10	(-0.3)	(-1.5)	(-0.6)	18.8 (11.3, 30.8)	2.039		
Grindøya	17	$\textbf{-}17.8 \pm 0.6$	11.1 ± 0.4	0.4 ± 0.1	1.4 (0.8, 2.6)	0.834	0.907	1.528
Grindøya	1 /	(0.1)	(-0.3)	(-1.4)		0.834	0.907	1.326
Iceland	23	$\textbf{-}17.8 \pm 0.9$	11.6 ± 1.0	0.9 ± 0.5	21.7 (14.2, 37.7)	1.407	2.257	5.410
rceiand	23	(0.3)	(-0.1)	(0.1)		1.40/	2.231	3.410
John's Island	19	$\textbf{-}16.9 \pm 1.5$	12.0 ± 0.8	0.5 ± 0.2	9.1 (5.7, 16.5)	2.250	1.633	1.688
Joini S Island	1)	(-0.1)	(-0.1)	(-0.9)				
Kongsfjorden	16	$\textbf{-}18.4 \pm 0.8$	11.8 ± 1.0	0.8 ± 0.2	4.6 (2.8, 9.1)	1.217	2.128	2.068
Kongsijorden	10	(0.3)	(0.3)	(-0.3)	4.0 (2.0, 7.1)			
East Bay Island	43	$\textbf{-}18.2 \pm 1.1$	13.5 ± 1.3	1.0 ± 0.3	18.5 (13.6, 27.9)	1.904	2.897	3.571
Last Day Island	73	(-0.6)	(0.3)	(0.4)		1.504	2.077	3.371
Russia	24	$\textbf{-}19.4 \pm 0.8$	10.2 ± 0.4	0.7 ± 0.2	3.3 (2.2, 5.8)	1.175	0.962	2.278
Russia	27	(0.4)	(-0.4)	(-0.4)	3.3 (2.2, 3.6)	1.1/3	0.702	2.276
Tern Island	24	$\textbf{-}18.8 \pm 0.3$	12.4 ± 0.5	0.9 ± 0.2	1.4 (0.9, 2.4)	0.475	1.141	2.398
1 CHI ISIGHU	24	(-1.0)	(-0.4)	(0.1)	1.4 (0.5, 2.4)	0.7/3	1.171	2.370

3-dimensional representation of ten 3-dimensional niche regions produced using the R package rgl v.0.100.54 (Adler and Murdoch 2017). Corrected stable isotope values (δ^{13} C and δ^{15} N in ‰) and THg concentrations (in $\mu g/g$ dw) were used from individuals at ten pan-Arctic or subarctic common eider colonies to obtain 3-dimensional niche regions using the R package nicheROVER v1.0 (Swanson et al. 2015).



1180 (Still image of a video showing the 3-dimensional plot in rotating view).