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Review **Hygienic Perspectives of Wood in Healthcare Buildings**

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Abstract: Wood materials are being adopted as nature-based architectural themes inside the healthcare buildings. Concern is raised that the organic and porous character of wood might support microbial survival. Therefore, this review discusses the hygienic properties of wood including the antimicrobial potential and its cleanability in comparison to smooth surface materials. In general, wood has antimicrobial properties owing to its chemical composition and physical structure. However, the hygienic potential of wood is influenced by the type of wood, age of wood, the cleaning method, surface treatment, and its moisture content. This information is intended to guide decision-makers regarding the use of wood in hygienically sensitive places and researchers to help them identify the variables for better utilizing the hygienic potential of this material.

Keywords: surface hygiene; wood; antimicrobial; biofilms; green buildings

1. Introduction

The construction of hospitals and other healthcare facilities has a significant impact on the environment in terms of material usage and consumption of energy and waste generation [1]. Therefore, sustainable construction of healthcare buildings demands the use of eco-friendly materials which can be recycled and have low embodied energy and environmental impact [2,3].

The occupants of healthcare buildings including the patients and the staff, have a higher level of stress as compared to general populations [4]. In the patients, stress is mainly linked to the illness, while for the staff is linked to long working hours and the need to take life-saving decisions [4]. Past research has shown that the interior environment could reduce the level of this stress in the hospital building occupants [5]. One of the main modifications is the use of nature-based architectural themes which provide a healing and relaxing environment to the patients and the staff of healthcare buildings [6].

Wood is an organic and renewable resource of nature that is abundantly used for eco-friendly constructions and is an important component of nature-based themes [7].

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Therefore, using wood material in the construction and designing of healthcare buildings can contribute to the reduction of stress in occupants and the improvement of sustainability markers of healthcare buildings $[1,8]$. This is the reason that the growing number of healthcare buildings are adapting this material.

There is still hesitancy in widely adopting wood for indoor applications in healthcare buildings because this material is perceived as unhygienic owing to its porosity and organic nature [9]. However, some research has shown that untreated wood materials have antimicrobial properties against a wide range of pathogens responsible for healthcare-associated infections (HAI) [10]. Moreover, the most common microbes, including the SARS-CoV-2, survive least on porous materials as compared to smooth solid materials [11–13]. Meanwhile, the cleanability of wood is no worse than other materials such as plastic, glass, and steel [9].

This literature review aims to gather knowledge on the potential uses of wood material in healthcare buildings. It discusses the antimicrobial and hygienic properties of wood along with the influencing variables. This information is intended to identify the knowledge gaps and provide guidelines to the stakeholders regarding the hygienically safe uses of wood in healthcare buildings.

2. Wood as an Indoor Material for Healthcare Buildings

Wood, as indoor material, is generally used for two purposes, i.e., construction and aesthetics, and mostly both go along together, for example, in making furniture, floors, ceiling, and walls [14]. The structure may involve untreated wood, however, a treatment is often applied to protect against wood degrading pests and fungi depending upon the type of wood, geographic location, and the desired look. Indoor surfaces of wood are also frequently coated to protect them from scratches, stains, and weathering [15]. Indeed, various chemicals are used for processing of wood, such as adhesives, pesticides, solvents, resin binders, waterproofing compounds, stains, pigments, paints, and varnishes [9,16]. Some of these chemicals can be toxic [17], as some countries allow the use of, creosote, pentachlorophenol (PCP), or chromated copper arsenate (CCA) [18]. In addition, various volatile organic compounds (VOC) are emitted during such treatments (wood processing) and can cause respiratory or other health-related issues/illnesses. Therefore, nontoxic compounds are favored for wood treatments [18].

Therefore, in this review, we mostly discuss 'untreated wood', which is free from surface treatment and coating, because this condition best represents the natural properties of wood. In addition, the application of surface finishes could make wood materials more difficult to recycle. Although some studies discuss the antimicrobial properties of wood coatings [19,20], that is a different subject. In addition, untreated wood is sometimes used as an indoor material in form of non-contact furniture surfaces, ceilings, floors, and some wall frames. Such untreated materials are more preferred to be recycled as compared to treated wood.

3. Healthcare-Associated Infections (HAI) and Role of Environmental Surfaces

Nosocomial or healthcare-associated infections (HAI) are transmitted to patients when they receive medical care in healthcare facilities. The World Health Organization (WHO) remarks HAI as the most common adverse event among hospitalized patients [21]. In general, the HAI concern up to 7% of the hospitalized patients in developed and 10% in developing countries [22]. They are responsible for increasing the economic burden on the healthcare system by prolonged hospital staying time, disability, and treatment cost [23,24].

Generally, HAI are characterized as ventilator-associated pneumonia (VAP), central line-associated bloodstream infections, catheter-associated urinary tract infections, and surgical site infections [25]. The severity of these infections can range from mild to fatal depending upon the causative microorganism, e.g., bacteria, fungi, or viruses, and the type of infection or system infected.

The prevalence of these microbial agents differs depending upon patient populations, available medical facilities and the condition of the indoor environment. The most notable etiological agents causing HAI can be listed as: *Staphylococcus aureus*, *Enterococcus* spp. (e.g., E. faecalis, E. faecium), Escherichia coli, Candida spp. (e.g., C. albicans, C. glabrata), Klebsiella pneumoniae, Pseudomonas aeruginosa, Acinetobacter baumannii and Aspergillus fumigatus [26]. The viruses are also very important in the context of HAI, especially, the seasonal influenza virus, and the recent Severe Acute Respiratory Syndrome associated Coronavirus (COVID- 19 virus).

surgical site infections $\overline{25}$. The second mild to fatalogue from mild to fatal

Environmental surfaces are the non-clinical items including medical devices (catheters, surgical instruments, stethoscopes, electronic thermometers, blood pressure cuffs, infusion pumps, hemodialysis machines, etc.), and the household surfaces of the patient room, washroom, and surrounding objects (Figure 1). The household surfaces can be further classified based on the frequency of touch as the high contact (bed railings, sink, tap, door handle, curtain, etc.), medium contact (furniture, window frame, floor, etc.), and low door handle, curtain, etc.), medium contact (furniture, window frame, floor, etc.), and low contact surfaces (ceilings, walls, pillars, etc.) [27]. contact surfaces (ceilings, walls, pillars, etc.) [27].

Figure 1. Classification of indoor environmental surfaces in healthcare buildings.

Figure 1. Classification of indoor environmental surfaces in healthcare buildings. agents and can play a role in transmission (Figure 2). Primarily, patients contaminate the environmental surfaces when they directly touch them $[22,28]$. Meanwhile, the settling down of sneezed and coughed droplets can also contaminate surfaces [29]. Some aerosol particles generated during washing and toilet flush can also contaminate the sur-
 rounding surfaces [30]. These environmental surfaces are a continuous risk as long as the contaminating microbes survive on them, and this survival time can be short, from minutes, to long, for months, [31] depending upon the type of microorganism and the surface material (Table 1). The transmission occurs when a susceptible individual comes in
 $\frac{1}{2}$ contact with these contaminated surfaces [27,32]. Healthcare personnel's hands can also Environmental surfaces inside healthcare buildings could be a reservoir of infectious get contaminated from surfaces and pass pathogens to susceptible patients [32].

 $\frac{1}{2}$ Low to non-contact surfaces and pass paints can also get contact surfaces for all pass paints can also get contact surfaces, such as ceilings, walls, floors, window frames, and some furniture are not considered as an important source of microbial transmission. Therefore, they are not cleaned as often as the frequent contact surfaces [33]. However, they may sustain the microbes and can result in transmission by accidental contact, or by indoor air contamination during cleaning, construction, and renovation operations [34,35]. Such air contamination is less likely, and if it happens, can cause respiratory and surgical site wound infections in susceptible patients [27].

Figure 2. Role of environmental surfaces in the transmission of healthcare-associated infections. **Figure 2.** Role of environmental surfaces in the transmission of healthcare-associated infections.

Table 1. Survival of the most common nosocomial pathogens on different substrate conditions.

Escherichia coli Escherichia coli in 15 months 21.5 km

1–28 days (strain dependent), 1–3 **4. Hygienic and Antimicrobial Behavior of Wooden Surfaces** 144

In the outdoor and humid environment, many microbes are present on wood material. This flora is normally found in soil and on plants $[48]$ and they are responsible for the processing industry can have a higher microbial load as compared to outside air [50]. degradation of organic materials [49]. This is the reason that the indoor air of the wood Therefore, the organic, porous, and moisture-absorbing nature of wood is perceived as

days (flood water) [41], 28

microbe heaven. However, in the low moisture and indoor environments, the survival of microbes on wood material is difficult [9]. Moreover, the persistence of hygienically important bacteria and fungi is also low because of the antimicrobial activity of wood material [51]. The theory behind the antimicrobial behavior is linked to the chemical composition and porous hygroscopic nature of wood materials [42].

4.1. Biochemical Composition and Antimicrobial Activity

Wood contains non-structural compounds known as extractives. These chemicals protect wood against microbial degradation and many of them have antimicrobial properties, e.g., tannins, phenolic acids, flavonoids, terpenoids, etc. As these compounds have different chemical natures they may have a different mode of action against various microorganisms as shown in Table 2 [9].

Table 2. Antimicrobial actions of wood chemicals against microbes *.

* Reproduced from Munir et al. [9] with permission.

Types and quantity of wood extractives may vary among different wood species and parts of trees, therefore the antimicrobial properties also vary among different wood species (Table 3) [52] and extractives from different parts of wood, i.e., sapwood and heartwood [53]. However, there is only a minor difference of types of extractives present in the same parts of different trees of a wood species growing in different regions [54,55]. Likewise, no significant difference in antimicrobial activity was observed among the sessile oak trees collected from different regions in France that had different soil fertilities and, therefore, presumably different growing conditions [56].

Table 3. Grading of wood species depending upon the hygienic suitability *.

* Reproduced from Munir et al. [9] with permission.

Various treatment variables can influence the composition of extractives in wood. For example, heat treatments are the part of wood processing, and they are used to increase the durability of this material [68]; meanwhile, heating can accelerate the emissions of volatile organic compounds [64], and as these VOC are also responsible for the antimicrobial activity of wood [69], the antimicrobial behavior may change with heat treatments [56,64].

Logically, some VOC are lost during the usage and storage life of wood, therefore, an influence on the VOC-based antimicrobial activity is expected, however, in our previous study, we observed that the antimicrobial activity of various wood samples stored for two years did not change [52]. Schönwälder et al. [62] also reported that the age and storage time of pinewood did not influence its antimicrobial behavior.

Wood extractives are also used to treat the surfaces of less durable wood to increase their durability against microbial biodegradation [70]. Meanwhile, the removal of extractives has been reported to increase the thermal durability of wood [71]. In such cases, extracting these compounds for surface treatment of wood could give desirable results of durable and antimicrobial wooden surfaces. However, further studies are needed to evaluate the influence of such treatments on indoor VOC emissions.

Wood is an acidic material due to the presence of acidic functional groups in the polymers and extractives of wood. The pH of different wood species ranges from slightly acidic, e.g., pH of poplar and fir is 6, to very acidic, for example, pH of oak wood species can be 3.5 [72]. This acidic environment can pose a challenge to microbial growth when compared to inert solid inanimate surfaces.

4.2. Role of Physical Structure in the Antimicrobial Activity of Wood

The physical structure of wood contributes as a microbial counter mechanism mainly by moisture regulation. The porosity and hygroscopic properties of wood are the main characteristics involved in this behavior; however, controversies surround the microbial survival and hindrance in cleaning due to these physical properties.

4.2.1. Porosity of Wood

Wood is a product of a living tree, therefore it contains the dead elongated cells and they appear in the form of pores at the surface. The porosity varies with the anatomy of wood species, part of tree, and cutting plane of wood. It is sometimes perceived that the open pores of untreated wood surfaces could contain the microbes, ultimately making surface cleaning and microbial removal difficult. Microbes can for sure enter into the porous surfaces of wood, however, they should not necessarily be considered as a hygienic risk. Many studies have shown that the porosity of wood can be a challenge to recover microbes from wood and it can influence the interpretation of microbial survival results. In a previous review, we reported that the recovery of microbes was dependent upon the method of recovery and the moisture content of samples, however, the best results were obtained when the elution-based methods were used [51]. Using the elution-based recovery method to study the microbial survival, we observed that microbial recovery from wood and non-porous materials was similar, however, the survival of most common HAI bacteria was lowest on wood compared to other surfaces such as plastic, steel, or aluminum [12]. In a similar study, Vainio-Kaila et al. [73] observed that lower CFU of *E. coli* and *L. monocytogenes* were recovered from pine heartwood as compared to a glass surface. To further test whether the cultivable bacetria were still hiding in pores or not, they resuspended the wood pieces in the culture medium to let the hidden microbes grow. After one day of incubation, no microbial growth was observed, proving that no culturable microbes were hidden inside the pores [73].

The porosity of wood does not necessarily impact its cleaning, for example, studies of wooden cutting boards have shown that the in-use methods of cleaning were equally efficient for wood as they were for plastic surfaces [59,74–79]. In addition, washing does not decrease the cleanability of wood over time [80] and the action of disinfectants in cleaning equally efficient for cleaning *E. coli, L. monocytogenes, P. aeruginosa, S. aureus, C. jejuni, Salmonella* Typhimurium, *E. coli* O157:H7 on both the plastic and wooden boards [81–84]. Another recent study from indoor hospital surfaces showed the wipe cleaning with disinfectant was sufficient to decrease the microbial numbers on wooden surfaces [8].

Porous surfaces have a greater surface area as compared to non-porous surfaces and it may help in faster drying. Therefore, wood dries faster as compared to non-porous materials [85]. A study by Chiu et al. [86] observed that *Vibrio parahaemolyticus* survived less on porous material (bamboo and wood) as compared to smooth surfaces (plastic, stainless steel, and glazed ceramic), and this difference in survival is more likely because the smooth surfaces retain surface moisture for a longer time [87].

As described above, the pores are the long cells of wood and they can have a length of 3 mm, likewise, the microbes can descend deep inside the pores on wood. However, most porosity is on transversally cut (RT) wood, which is not commonly used for construction operations [12]. Prechter et al. [88] reported that *E. coli* cells and spores of *Bacillus subtilis* descended deeper (around 3 mm) in RT cuttings than in longitudinal cut (LT) wooden boards. In contrast, boards with LT faces are easier to clean because of shallow and wider openings on the surface [89]. The persistence of porosity of wooden surfaces may change over time, for example, Kotradyova et al. [8] reported that the pores of wooden surfaces were filled with organic debris and dust during the usage life of wooden surfaces inside the hospital buildings. However, it still remains to be investigated, how long it takes for the filling of pores in various indoor environments and how it can influence the surface hygiene.

Overall, the transfer of microbes from wooden surfaces to food or hands is also lower as compared to non-porous surfaces [90–93]. For example, a recent study reported that *E. coli* and *Penicillium expansum* not only survived less on wood compared to plastic and cardboards but their transfer from wood surface to apples was also lower [42]. However, if the microbes survive inside the surfaces, they are a continuous risk, in this scenario, innovative microscopic tools can be handy for risk assessment and microbial bio-distribution studies [77,89].

4.2.2. Hygroscopicity and Capillary Action

Hygroscopicity of a material is the property of absorbing and adsorbing moisture from the environment. This ability is influenced by environmental humidity and temperature, and free water, bound water, and fiber saturation point of the wood [94]. Most bacteria and some fungi are sensitive to desiccation and need a water potential of ≤−2.8 MPa for growth, motility, and nutrient uptake [9,58]. However, properly dried wood (12% moisture) has a higher water potential that does not offer enough water for microbial growth and multiplication [9,58]. This is the reason that the hygroscopic nature of wood leads to faster absorption of moisture as compared to other non-porous contact surfaces, consequently, microbes survive least on wood compared to non-absorptive smooth surfaces such as plastics and metals [12,86,95–97]. However, after reaching the fiber saturation point, wood does not adsorb more moisture and the hygroscopic antimicrobial activity may diminish. For example, Gehrig et al. [85] reported very high *E. coli* counts on both polyethylene and wood, while the bacterial number was lower on wood in a drier environment.

Coating of wood materials may decrease or diminish the surface hygroscopicity, thus influencing microbial adhesion and survival [67]. Hedge [16] studied the survival of *E. coli, P. aeruginosa*, *S. aureus* on untreated and varnished beechwood, and plastic. The microbes decreased fastest on natural wood, followed by varnished wood and plastic. Here, the difference of survival can be attributed to the moisture absorbing ability of surfaces, especially the difference between varnished and no-varnished wood.

The lower microbial recovery and moisture retention of wooden surfaces also lead to the assumption that the microbes are strongly attached to wooden surfaces which could help the biofilm formation [98]. However, many studies have shown that microbial attachment to wood is not stronger compared to other smooth surfaces [67]. In fact, in our recent study, we found that the bacteria attached more to non-porous melamine surfaces and formed biofilm while no biofilm was found on oak wood surfaces in similar experimental conditions [99].

5. Conclusions and Future Outlook

Wood material is used in the green healthcare buildings, mostly as a medium to non- contact surfaces. Therefore, it very unlikely that wooden surfaces would increase HAI transmission.

Many wood species have antimicrobial properties due their chemical nature and physical structure. In addition, various microbes responsible for HAI survive less on wooden surfaces compared to other porous or non-porous surfaces. However, future studies should investigate whether the use of wooden surfaces inside the healthcare buildings can influence the prevalence and transmission of HAI.

Porosity of wood material can offer some hindrance in cleaning, however, with proper cleaning methods the cleanability of wooden and non-wooden surfaces is similar. Meanwhile, there is no evidence that the microbes would hide inside the pores of wood and become a risk of transmission after cleaning. In addition, the filling of wooden pores with debris and dust in the indoor healthcare environment also needs exploration to see if it can influence the microbial survival or cleanability of surfaces.

As the antimicrobial properties of wood materials are influenced by various factors, future studies should consider the variables of the species, treatment, cutting, and age of wood while studying the hygienic properties of this material.

There are no current estimates on the amount and economy of wood usage in the healthcare buildings. Therefore, respective stakeholders should investigate these figures to estimate the real value addition of wood usage in the healthcare buildings to the economy.

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References

- 1. Chías, P.; Abad, T. Green Hospitals, Green Healthcare. *Int. J. Energy Prod. Manag.* **2017**, *2*, 196–205. [\[CrossRef\]](http://doi.org/10.2495/EQ-V2-N2-196-205)
- 2. Kumari, S.; Kumar, R. Green Hospital—A Necessity and Not Option. *J. Manag. Res. Anal.* **2020**, *7*, 46–51. [\[CrossRef\]](http://doi.org/10.18231/j.jmra.2020.010)
- 3. Kotradyová, V.; Kaliňáková, B. Wood as Material Suitable for Healthcare and Therapeutic Facilities. *Adv. Mater. Res.* 2014, 1041, 362–366. [\[CrossRef\]](http://doi.org/10.4028/www.scientific.net/AMR.1041.362)
- 4. Chou, L.-P.; Li, C.-Y.; Hu, S.C. Job Stress and Burnout in Hospital Employees: Comparisons of Different Medical Professions in a Regional Hospital in Taiwan. *BMJ Open* **2014**, *4*, e004185. [\[CrossRef\]](http://doi.org/10.1136/bmjopen-2013-004185) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24568961)
- 5. Andrade, C.C.; Devlin, A.S.; Pereira, C.R.; Lima, M.L. Do the Hospital Rooms Make a Difference for Patients' Stress? A Multilevel Analysis of the Role of Perceived Control, Positive Distraction, and Social Support. *J. Environ. Psychol.* **2017**, *53*, 63–72. [\[CrossRef\]](http://doi.org/10.1016/j.jenvp.2017.06.008)
- 6. Ulrich, R.S. View through a Window May Influence Recovery from Surgery. *Science* **1984**, *224*, 420–421. [\[CrossRef\]](http://doi.org/10.1126/science.6143402) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/6143402)
- 7. Zhang, X.; Lian, Z.; Ding, Q. Investigation Variance in Human Psychological Responses to Wooden Indoor Environments. *Build. Environ.* **2016**, *109*, 58–67. [\[CrossRef\]](http://doi.org/10.1016/j.buildenv.2016.09.014)
- 8. Kotradyova, V.; Vavrinsky, E.; Kalinakova, B.; Petro, D.; Jansakova, K.; Boles, M.; Svobodova, H. Wood and Its Impact on Humans and Environment Quality in Healthcare Facilities. *Int. J. Environ. Res. Public Health* **2019**, *16*, 3496. [\[CrossRef\]](http://doi.org/10.3390/ijerph16183496) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31546873)
- 9. Munir, M.T.; Pailhories, H.; Eveillard, M.; Aviat, F.; Lepelletier, D.; Belloncle, C.; Federighi, M. Antimicrobial Characteristics of Untreated Wood: Towards a Hygienic Environment. *Health* **2019**, *11*, 152–170. [\[CrossRef\]](http://doi.org/10.4236/health.2019.112014)
- 10. Pailhories, H.; Munir, M.T.; Aviat, F.; Federighi, M.; Belloncle, C.; Eveillard, M. Oak in Hospitals, the Worst Enemy of *Staphylococcus aureus*? *Infect. Control Hosp. Epidemiol.* **2017**, *38*, 382–384. [\[CrossRef\]](http://doi.org/10.1017/ice.2016.304)
- 11. Chatterjee, S.; Murallidharan, J.S.; Agrawal, A.; Bhardwaj, R. Why Coronavirus Survives Longer on Impermeable than Porous Surfaces. *Phys. Fluids* **2021**, *33*, 021701. [\[CrossRef\]](http://doi.org/10.1063/5.0037924) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33746485)
- 12. Chen, J.-C.; Munir, M.T.; Aviat, F.; Lepelletier, D.; Le Pape, P.; Dubreil, L.; Irle, M.; Federighi, M.; Belloncle, C.; Eveillard, M.; et al. Survival of Bacterial Strains on Wood (*Quercus petraea*) Compared to Polycarbonate, Aluminum and Stainless Steel. *Antibiotics* **2020**, *9*, 804. [\[CrossRef\]](http://doi.org/10.3390/antibiotics9110804)
- 13. Corpet, D.E. Why Does SARS-CoV-2 Survive Longer on Plastic than on Paper? *Med. Hypotheses* **2021**, *146*, 110429. [\[CrossRef\]](http://doi.org/10.1016/j.mehy.2020.110429) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/33277105)
- 14. Alapieti, T.; Mikkola, R.; Pasanen, P.; Salonen, H. The Influence of Wooden Interior Materials on Indoor Environment: A Review. *Eur. J. Wood Wood Prod.* **2020**, *78*, 617–634. [\[CrossRef\]](http://doi.org/10.1007/s00107-020-01532-x)
- 15. Hristozov, D.; Pizzol, L.; Basei, G.; Zabeo, A.; Mackevica, A.; Hansen, S.F.; Gosens, I.; Cassee, F.R.; De Jong, W.; Koivisto, A.J.; et al. Quantitative Human Health Risk Assessment along the Lifecycle of Nano-Scale Copper-Based Wood Preservatives. *Nanotoxicology* **2018**, *12*, 747–765. [\[CrossRef\]](http://doi.org/10.1080/17435390.2018.1472314)
- 16. Hedge, A. Survival of *Escherichia coli*, *Pseudomona aeruginosa*, *Staphylococcus aureus* on Wood and Plastic Surfaces. *J. Microb. Biochem. Technol.* **2015**, *7*. [\[CrossRef\]](http://doi.org/10.4172/1948-5948.1000207)
- 17. Wang, Y.; Zhang, Y.; Li, X.; Sun, M.; Wei, Z.; Wang, Y.; Gao, A.; Chen, D.; Zhao, X.; Feng, X. Exploring the Effects of Different Types of Surfactants on Zebrafish Embryos and Larvae. *Sci. Rep.* **2015**, *5*, 10107. [\[CrossRef\]](http://doi.org/10.1038/srep10107)
- 18. EPA. Overview of Wood Preservative Chemicals. Available online: [https://www.epa.gov/ingredients-used-pesticide-products/](https://www.epa.gov/ingredients-used-pesticide-products/overview-wood-preservative-chemicals) [overview-wood-preservative-chemicals](https://www.epa.gov/ingredients-used-pesticide-products/overview-wood-preservative-chemicals) (accessed on 29 July 2020).
- 19. Kandelbauer, A.; Widsten, P. Antibacterial Melamine Resin Surfaces for Wood-Based Furniture and Flooring. *Prog. Org. Coat.* **2009**, *65*, 305–313. [\[CrossRef\]](http://doi.org/10.1016/j.porgcoat.2008.12.003)
- 20. Papadopoulos, A.N.; Taghiyari, H.R. Innovative Wood Surface Treatments Based on Nanotechnology. *Coatings* **2019**, *9*, 866. [\[CrossRef\]](http://doi.org/10.3390/coatings9120866)
- 21. WHO. *Report on the Burden of Endemic Health Care-Associated Infection Worldwide*; WHO: Geneva, Switzerland, 2011.
- 22. Khan, H.A.; Baig, F.K.; Mehboob, R. Nosocomial Infections: Epidemiology, Prevention, Control and Surveillance. *Asian Pac. J. Trop. Biomed.* **2017**, *7*, 478–482. [\[CrossRef\]](http://doi.org/10.1016/j.apjtb.2017.01.019)
- 23. Cassini, A.; Plachouras, D.; Eckmanns, T.; Abu Sin, M.; Blank, H.-P.; Ducomble, T.; Haller, S.; Harder, T.; Klingeberg, A.; Sixtensson, M.; et al. Burden of Six Healthcare-Associated Infections on European Population Health: Estimating Incidence-Based Disability-Adjusted Life Years through a Population Prevalence-Based Modelling Study. *PLoS Med.* **2016**, *13*, e1002150. [\[CrossRef\]](http://doi.org/10.1371/journal.pmed.1002150)
- 24. Benenson, S.; Cohen, M.J.; Schwartz, C.; Revva, M.; Moses, A.E.; Levin, P.D. Is It Financially Beneficial for Hospitals to Prevent Nosocomial Infections? *BMC Health Serv. Res.* **2020**, *20*, 653. [\[CrossRef\]](http://doi.org/10.1186/s12913-020-05428-7)
- 25. CDC. Health Topics—HAI—POLARIS. Available online: <https://www.cdc.gov/policy/polaris/healthtopics/hai.html> (accessed on 28 July 2020).
- 26. Haque, M.; Sartelli, M.; McKimm, J.; Abu Bakar, M. Health Care-Associated Infections—An Overview. *Infect. Drug Resist.* **2018**, *11*, 2321–2333. [\[CrossRef\]](http://doi.org/10.2147/IDR.S177247) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/30532565)
- 27. Suleyman, G.; Alangaden, G.; Bardossy, A.C. The Role of Environmental Contamination in the Transmission of Nosocomial Pathogens and Healthcare-Associated Infections. *Curr. Infect. Dis. Rep.* **2018**, *20*, 12. [\[CrossRef\]](http://doi.org/10.1007/s11908-018-0620-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29704133)
- 28. Islam, M.S.; Hossain, M.A.; Khan, S.I.; Khan, M.N.; Sack, R.B.; Albert, M.J.; Huq, A.; Colwell, R.R. Survival of *Shigella dysenteriae* Type 1 on Fomites. *J. Health Popul. Nutr.* **2001**, *19*, 177–182. [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/11761771)
- 29. Choi, H.; Chatterjee, P.; Coppin, J.D.; Martel, J.A.; Hwang, M.; Jinadatha, C.; Sharma, V.K. Current Understanding of the Surface Contamination and Contact Transmission of SARS-CoV-2 in Healthcare Settings. *Environ. Chem. Lett.* **2021**. [\[CrossRef\]](http://doi.org/10.1007/s10311-021-01186-y)
- 30. Schreck, J.H.; Lashaki, M.J.; Hashemi, J.; Dhanak, M.; Verma, S. Aerosol Generation in Public Restrooms. *Phys. Fluids* **2021**, *33*, 033320. [\[CrossRef\]](http://doi.org/10.1063/5.0040310)
- 31. Kramer, A.; Assadian, O. Survival of microorganisms on inanimate surfaces. In *Use of Biocidal Surfaces for Reduction of Healthcare Acquired Infections*; Borkow, G., Ed.; Springer International Publishing: Cham, Switzerland, 2014; pp. 7–26. ISBN 978-3-319-08056-7.
- 32. Otter, J.A.; Yezli, S.; Salkeld, J.A.G.; French, G.L. Evidence That Contaminated Surfaces Contribute to the Transmission of Hospital Pathogens and an Overview of Strategies to Address Contaminated Surfaces in Hospital Settings. *Am. J. Infect. Control* **2013**, *41*, S6–S11. [\[CrossRef\]](http://doi.org/10.1016/j.ajic.2012.12.004)
- 33. Lei, H.; Jones, R.M.; Li, Y. Exploring Surface Cleaning Strategies in Hospital to Prevent Contact Transmission of Methicillin-Resistant *Staphylococcus aureus*. *BMC Infect. Dis.* **2017**, *17*, 85. [\[CrossRef\]](http://doi.org/10.1186/s12879-016-2120-z)
- 34. Park, J.H.; Ryu, S.H.; Lee, J.Y.; Kim, H.J.; Kwak, S.H.; Jung, J.; Lee, J.; Sung, H.; Kim, S.-H. Airborne Fungal Spores and Invasive Aspergillosis in Hematologic Units in a Tertiary Hospital during Construction: A Prospective Cohort Study. *Antimicrob. Resist. Infect. Control* **2019**, *8*, 88. [\[CrossRef\]](http://doi.org/10.1186/s13756-019-0543-1)
- 35. Kanamori, H.; Rutala, W.A.; Sickbert-Bennett, E.E.; Weber, D.J. Review of Fungal Outbreaks and Infection Prevention in Healthcare Settings During Construction and Renovation. *Clin. Infect. Dis.* **2015**, *61*, 433–444. [\[CrossRef\]](http://doi.org/10.1093/cid/civ297) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/25870328)
- 36. Gupta, M.; Bisesi, M.; Lee, J. Comparison of Survivability of *Staphylococcus aureus* and Spores of *Aspergillus niger* on Commonly Used Floor Materials. *Am. J. Infect. Control* **2017**, *45*, 717–722. [\[CrossRef\]](http://doi.org/10.1016/j.ajic.2017.02.014) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28318645)
- 37. Mensah-Attipoe, J.; Reponen, T.; Veijalainen, A.-M.; Rintala, H.; Täubel, M.; Rantakokko, P.; Ying, J.; Hyvärinen, A.; Pasanen, P. Comparison of Methods for Assessing Temporal Variation of Growth of Fungi on Building Materials. *Microbiology* **2016**, *162*, 1895–1903. [\[CrossRef\]](http://doi.org/10.1099/mic.0.000372)
- 38. van Doremalen, N.; Bushmaker, T.; Morris, D.; Holbrook, M.; Gamble, A.; Williamson, B.; Tamin, A.; Harcourt, J.; Thornburg, N.; Gerber, S.; et al. Aerosol and Surface Stability of HCoV-19 (SARS-CoV-2) Compared to SARS-CoV-1. *N. Engl. J. Med.* **2020**, *382*, 1564–1567. [\[CrossRef\]](http://doi.org/10.1056/NEJMc2004973) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32182409)
- 39. Chin, A.W.H.; Chu, J.T.S.; Perera, M.R.A.; Hui, K.P.Y.; Yen, H.-L.; Chan, M.C.W.; Peiris, M.; Poon, L.L.M. Stability of SARS-CoV-2 in Different Environmental Conditions. *Lancet Microbe* **2020**, *1*, e10. [\[CrossRef\]](http://doi.org/10.1016/S2666-5247(20)30003-3)
- 40. Duan, S.-M.; Zhao, X.-S.; Wen, R.-F.; Huang, J.-J.; Pi, G.-H.; Zhang, S.-X.; Han, J.; Bi, S.-L.; Ruan, L.; Dong, X.-P.; et al. Stability of SARS Coronavirus in Human Specimens and Environment and Its Sensitivity to Heating and UV Irradiation. *Biomed. Environ. Sci.* **2003**, *16*, 246–255.
- 41. Taylor, J.; Davies, M.; Canales, M.; Lai, K.M. The Persistence of Flood-Borne Pathogens on Building Surfaces under Drying Conditions. *Int. J. Hyg. Environ. Health* **2013**, *216*, 91–99. [\[CrossRef\]](http://doi.org/10.1016/j.ijheh.2012.03.010)
- 42. Aviat, F.; Le Bayon, I.; Federighi, M.; Montibus, M. Comparative Study of Microbiological Transfer from Four Materials Used in Direct Contact with Apples. *Int. J. Food Microbiol.* **2020**, *333*, 108780. [\[CrossRef\]](http://doi.org/10.1016/j.ijfoodmicro.2020.108780)
- 43. Williams, A.P.; Avery, L.M.; Killham, K.; Jones, D.L. Persistence of *Escherichia coli* O157 on Farm Surfaces under Different Environmental Conditions. *J. Appl. Microbiol.* **2005**, *98*, 1075–1083. [\[CrossRef\]](http://doi.org/10.1111/j.1365-2672.2004.02530.x)
- 44. Tiwari, A.; Patnayak, D.P.; Chander, Y.; Parsad, M.; Goyal, S.M. Survival of Two Avian Respiratory Viruses on Porous and Nonporous Surfaces. *Avian Dis.* **2006**, *50*, 284–287. [\[CrossRef\]](http://doi.org/10.1637/7453-101205R.1)
- 45. Sakaguchi, H.; Wada, K.; Kajioka, J.; Watanabe, M.; Nakano, R.; Hirose, T.; Ohta, H.; Aizawa, Y. Maintenance of Influenza Virus Infectivity on the Surfaces of Personal Protective Equipment and Clothing Used in Healthcare Settings. *Environ. Health Prev. Med.* **2010**, *15*, 344–349. [\[CrossRef\]](http://doi.org/10.1007/s12199-010-0149-y) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21432565)
- 46. Oxford, J.; Berezin, E.N.; Courvalin, P.; Dwyer, D.E.; Exner, M.; Jana, L.A.; Kaku, M.; Lee, C.; Letlape, K.; Low, D.E.; et al. The Survival of Influenza A(H1N1) Pdm09 Virus on 4 Household Surfaces. *Am. J. Infect. Control* **2014**, *42*, 423–425. [\[CrossRef\]](http://doi.org/10.1016/j.ajic.2013.10.016) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/24679569)
- 47. Guan, J.; Chan, M.; VanderZaag, A. Inactivation of Avian Influenza Viruses on Porous and Non-porous Surfaces Is Enhanced by Elevating Absolute Humidity. *Transbound. Emerg. Dis.* **2017**, *64*, 1254–1261. [\[CrossRef\]](http://doi.org/10.1111/tbed.12499) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27059695)
- 48. Cosenza, B.J.; McCreary, M.; Buck, J.D.; Shigo, A.L. Bacteria Associated with Discolored and Decayed Tissues in Beech, Birch, and Maple. *Phytopathology* **1970**, *60*, 1547. [\[CrossRef\]](http://doi.org/10.1094/Phyto-60-1547)
- 49. Dutkiewicz, J.; Sorenson, W.G.; Lewis, D.M.; Olenchock, S.A. Levels of Bacteria, Fungi and Endotoxin in Stored Timber. *Int. Biodeterior. Biodegrad.* **1992**, *30*, 29–46. [\[CrossRef\]](http://doi.org/10.1016/0964-8305(92)90022-G)
- 50. Górny, R.L.; Gołofit-Szymczak, M.; Cyprowski, M.; Stobnicka-Kupiec, A. Nasal Lavage as Analytical Tool in Assessment of Exposure to Particulate and Microbial Aerosols in Wood Pellet Production Facilities. *Sci. Total Environ.* **2019**, *697*, 134018. [\[CrossRef\]](http://doi.org/10.1016/j.scitotenv.2019.134018) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31479905)
- 51. Munir, M.T.; Pailhories, H.; Eveillard, M.; Irle, M.; Aviat, F.; Dubreil, L.; Federighi, M.; Belloncle, C. Testing the Antimicrobial Characteristics of Wood Materials: A Review of Methods. *Antibiotics* **2020**, *9*, 225. [\[CrossRef\]](http://doi.org/10.3390/antibiotics9050225) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32370037)
- 52. Munir, M.T.; Aviat, F.; Pailhories, H.; Eveillard, M.; Irle, M.; Federighi, M.; Belloncle, C. Direct Screening Method to Assess Antimicrobial Behavior of Untreated Wood. *Eur. J. Wood Wood Prod.* **2019**, *77*, 319–322. [\[CrossRef\]](http://doi.org/10.1007/s00107-019-01390-2)
- 53. Ateş, S.; Gür, M.; Özkan, O.E.; Akça, M.; Olgun, Ç.; Güder, A. Chemical Contents and Antifungal Activity of Some Durable Wood Extractives vs. *Pleurotus ostreatus. BioResources* **2015**, *10*, 2433–2443. [\[CrossRef\]](http://doi.org/10.15376/biores.10.2.2433-2443)
- 54. Prida, A.; Puech, J.-L. Influence of Geographical Origin and Botanical Species on the Content of Extractives in American, French, and East European Oak Woods. *J. Agric. Food Chem.* **2006**, *54*, 8115–8126. [\[CrossRef\]](http://doi.org/10.1021/jf0616098)
- 55. Prida, A.; Ducousso, A.; Petit, R.J.; Nepveu, G.; Puech, J.-L. Variation in Wood Volatile Compounds in a Mixed Oak Stand: Strong Species and Spatial Differentiation in Whisky-Lactone Content. *Ann. For. Sci.* **2007**, *64*, 313–320. [\[CrossRef\]](http://doi.org/10.1051/forest:2007008)
- 56. Munir, M.T.; Pailhories, H.; Eveillard, M.; Irle, M.; Aviat, F.; Federighi, M.; Belloncle, C. Experimental Parameters Influence the Observed Antimicrobial Response of Oak Wood (*Quercus Petraea*). *Antibiotics* **2020**, *9*, 535. [\[CrossRef\]](http://doi.org/10.3390/antibiotics9090535) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32847132)
- 57. Laireiter, C.M.; Schnabel, T.; Köck, A.; Stalzer, P.; Petutschnigg, A.; Oostingh, G.J.; Hell, M. Active Anti-Microbial Effects of Larch and Pine Wood on Four Bacterial Strains. *BioResources* **2013**, *9*, 273–281. [\[CrossRef\]](http://doi.org/10.15376/biores.9.1.273-281)
- 58. Milling, A.; Kehr, R.; Wulf, A.; Smalla, K. Survival of Bacteria on Wood and Plastic Particles: Dependence on Wood Species and Environmental Conditions. *Holzforschung* **2005**, *59*, 72–81. [\[CrossRef\]](http://doi.org/10.1515/HF.2005.012)
- 59. Koch, A.P.; Kofod, C.J.; Konova, D.; Kvist, K.E.; Lindergaard, B. *Wood, Plastic and Steel—A Comparason of Hygenic Properties*; Project P 99095 "wood in the food"; The Danish Technological Institute: Taastrup, Denmark, 2002.
- 60. Milling, A.; Smalla, K.; Kehr, R.; Wulf, A. The Use of Wood in Practice—A Hygienic Risk? *Holz Roh. Werkst.* **2005**, *63*, 463–472. [\[CrossRef\]](http://doi.org/10.1007/s00107-005-0064-x)
- 61. Miller, A.J.; Brown, T.; Call, J.E. Comparison of Wooden and Polyethylene Cutting Boards: Potential for the Attachment and Removal of Bacteria from Ground Beef. *J. Food Prot.* **1996**, *59*, 854–858. [\[CrossRef\]](http://doi.org/10.4315/0362-028X-59.8.854) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31159115)
- 62. Schönwälder, A.; Kehr, R.; Wulf, A.; Smalla, K. Wooden Boards Affecting the Survival of Bacteria? *Holz Roh. Werkst.* **2002**, *60*, 249–257. [\[CrossRef\]](http://doi.org/10.1007/s00107-002-0300-6)
- 63. Munir, M.T.; Belloncle, C.; Pailhories, H.; Aviat, F.; Federighi, M.; Eveillard, M. *Wood as a Safe Material for Indoor Surface Construction of Hygienically Sensitive Places*; Northwestern Polytechnical University: Xi'an, China, 2017; pp. 79–80.
- 64. Vainio-Kaila, T.; Rautkari, L.; Nordström, K.; Närhi, M.; Natri, O.; Kairi, M. Effect of Extractives and Thermal Modification on Antibacterial Properties of Scots Pine and Norway Spruce. *Int. Wood Prod. J.* **2013**, *4*, 248–252. [\[CrossRef\]](http://doi.org/10.1179/2042645313Y.0000000038)
- 65. Vainio-Kaila, T.; Kyyhkynen, A.; Rautkari, L.; Siitonen, A. Antibacterial Effects of Extracts of *Pinus sylvestris* and *Picea abies* against *Staphylococcus aureus*, *Enterococcus faecalis*, *Escherichia coli*, and *Streptococcus pneumoniae*. *BioResources* **2015**, *10*, 7763–7771. [\[CrossRef\]](http://doi.org/10.15376/biores.10.4.7763-7771)
- 66. Vainio-Kaila, T.; Zhang, X.; Hänninen, T.; Kyyhkynen, A.; Johansson, L.-S.; Willför, S.; Österberg, M.; Siitonen, A.; Rautkari, L. Antibacterial Effects of Wood Structural Components and Extractives from *Pinus sylvestris* and *Picea abies* on Methicillin-Resistant *Staphylococcus aureus* and *Escherichia coli* O157:H7. *BioResources* **2017**, *12*, 7601–7614. [\[CrossRef\]](http://doi.org/10.15376/biores.12.4.7601-7614)
- 67. Tomiˇci´c, R.; Tomiˇci´c, Z.; Thaler, N.; Humar, M.; Raspor, P. Factors Influencing Adhesion of Bacteria *Escherichia coli*, *Pseudomonas aeruginosa*, *Staphylococcus aureus* and Yeast *Pichia membranifaciens* to Wooden Surfaces. *Wood Sci. Technol.* **2020**, *54*, 1663–1676. [\[CrossRef\]](http://doi.org/10.1007/s00226-020-01222-0)
- 68. Báder, M.; Németh, R. Moisture-Dependent Mechanical Properties of Longitudinally Compressed Wood. *Eur. J. Wood Prod.* **2019**, *77*, 1009–1019. [\[CrossRef\]](http://doi.org/10.1007/s00107-019-01448-1)
- 69. Vainio-Kaila, T.; Hänninen, T.; Kyyhkynen, A.; Ohlmeyer, M.; Siitonen, A.; Rautkari, L. Effect of Volatile Organic Compounds from *Pinus sylvestris* and *Picea abies* on *Staphylococcus aureus*, *Escherichia coli*, *Streptococcus pneumoniae* and *Salmonella enterica* Serovar Typhimurium. *Holzforschung* **2017**, *71*, 905–912. [\[CrossRef\]](http://doi.org/10.1515/hf-2017-0007)
- 70. Anouhe, J.-B.S.; Niamké, F.B.; Faustin, M.; Virieux, D.; Pirat, J.-L.; Adima, A.A.; Kati-Coulibaly, S.; Amusant, N. The Role of Extractives in the Natural Durability of the Heartwood of Dicorynia Guianensis Amsh: New Insights in Antioxydant and Antifungal Properties. *Ann. For. Sci.* **2018**, *75*, 15. [\[CrossRef\]](http://doi.org/10.1007/s13595-018-0691-0)
- 71. Shebani, A.N.; van Reenen, A.J.; Meincken, M. The Effect of Wood Extractives on the Thermal Stability of Different Wood Species. *Thermochim. Acta* **2008**, *471*, 43–50. [\[CrossRef\]](http://doi.org/10.1016/j.tca.2008.02.020)
- 72. Geffert, A.; Geffertova, J.; Dudiak, M. Direct Method of Measuring the PH Value of Wood. *Forests* **2019**, *10*, 852. [\[CrossRef\]](http://doi.org/10.3390/f10100852)
- 73. Vainio-Kaila, T.; Kyyhkynen, A.; Viitaniemi, P.; Siitonen, A. Pine Heartwood and Glass Surfaces: Easy Method to Test the Fate of Bacterial Contamination. *Eur. J. Wood Wood Prod.* **2011**, *69*, 391–395. [\[CrossRef\]](http://doi.org/10.1007/s00107-010-0453-7)
- 74. Acuff, G.R.; Vanderzant, C.; Hanna, M.O.; Ehlers, J.G.; Gardner, F.A. Effects of Handling and Preparation of Turkey Products on the Survival of *Campylobacter jejuni*. *J. Food Prot.* **1986**, *49*, 627–631. [\[CrossRef\]](http://doi.org/10.4315/0362-028X-49.8.627)
- 75. Ak, N.O.; Cliver, D.O.; Kaspar, C.W. Cutting Boards of Plastic and Wood Contaminated Experimentally with Bacteria. *J. Food Prot.* **1994**, *57*, 16–22. [\[CrossRef\]](http://doi.org/10.4315/0362-028X-57.1.16)
- 76. Ak, N.O.; Cliver, D.O.; Kaspar, C.W. Decontamination of Plastic and Wooden Cutting Boards for Kitchen Use. *J. Food Prot.* **1994**, *57*, 23–30. [\[CrossRef\]](http://doi.org/10.4315/0362-028X-57.1.23)
- 77. Zangerl, P.; Matlschweiger, C.; Dillinger, K.; Eliskases-Lechner, F. Survival of *Listeria monocytogenes* after Cleaning and Sanitation of Wooden Shelves Used for Cheese Ripening. *Eur. J. Wood Wood Prod.* **2010**, *68*, 415–419. [\[CrossRef\]](http://doi.org/10.1007/s00107-009-0381-6)
- 78. Lucke, F.-K.; Skowyrska, A. Hygienic Aspects of Using Wooden and Plastic Cutting Boards, Assessed in Laboratory and Small Gastronomy Units. *J. Verbr. Lebensm.* **2015**, *10*, 317–322. [\[CrossRef\]](http://doi.org/10.1007/s00003-015-0949-5)
- 79. Imhof, R.; Schwendimann, L.; Scettrini, P.R. Sanitising Wooden Boards Used for Cheese Maturation by Means of a Steam-Mediated Heating Process. *J. Consum. Prot. Food Saf.* **2017**, *12*, 255–263. [\[CrossRef\]](http://doi.org/10.1007/s00003-017-1114-0)
- 80. Cruciata, M.; Gaglio, R.; Scatassa, M.L.; Sala, G.; Cardamone, C.; Palmeri, M.; Moschetti, G.; Mantia, T.L.; Settanni, L. Formation and Characterization of Early Bacterial Biofilms on Different Wood Typologies Applied in Dairy Production. *Appl. Environ. Microbiol.* **2018**, *84*, e02107-17. [\[CrossRef\]](http://doi.org/10.1128/AEM.02107-17) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/29180375)
- 81. Deza, M.A.; Araujo, M.; Garrido, M.J. Efficacy of Neutral Electrolyzed Water to Inactivate *Escherichia coli*, *Listeria monocytogenes*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* on Plastic and Wooden Kitchen Cutting Boards. *J. Food Prot.* **2007**, *70*, 102–108. [\[CrossRef\]](http://doi.org/10.4315/0362-028X-70.1.102) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/17265867)
- 82. DeVere, E.; Purchase, D. Effectiveness of Domestic Antibacterial Products in Decontaminating Food Contact Surfaces. *Food Microbiol.* **2007**, *24*, 425–430. [\[CrossRef\]](http://doi.org/10.1016/j.fm.2006.07.013)
- 83. Thormar, H.; Hilmarsson, H. Killing of *Campylobacter* on Contaminated Plastic and Wooden Cutting Boards by Glycerol Monocaprate (Monocaprin). *Lett. Appl. Microbiol.* **2010**, *51*, 319–324. [\[CrossRef\]](http://doi.org/10.1111/j.1472-765X.2010.02898.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20666986)
- 84. Al-Qadiri, H.M.; Ovissipour, M.; Al-Alami, N.; Govindan, B.N.; Shiroodi, S.G.; Rasco, B. Efficacy of neutral electrolyzed water, quaternary ammonium and lactic acid-based solutions in controlling microbial contamination of food cutting boards using a manual spraying technique. *J. Food Sci.* **2016**, *81*, M1177–M1183. [\[CrossRef\]](http://doi.org/10.1111/1750-3841.13275) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/27027449)
- 85. Gehrig, M.; Schnell, G.; Zürcher, E.; Kucera, L.J. Hygienische Eigenschaften von Holz-und Kunststoffbrettern in der Nahrungsmittelverarbeitung und-präsentation: Ein Vergleich. *Holz Roh. Werkst.* **2000**, *58*, 265–269. [\[CrossRef\]](http://doi.org/10.1007/s001070050423)
- 86. Chiu, T.-H.; Duan, J.; Liu, C.; Su, Y.-C. Efficacy of Electrolysed Oxidizing Water in Inactivating *Vibrio parahaemolyticus* on Kitchen Cutting Boards and Food Contact Surfaces. *Lett. Appl. Microbiol.* **2006**, *43*, 666–672. [\[CrossRef\]](http://doi.org/10.1111/j.1472-765X.2006.02006.x)
- 87. Shi, Y.; Sun, R.; An, D.; Lu, W.; Zhang, C.; Wang, L.; Liu, Y.; Wang, Q. Mathematical Quantification of Inactivation of *Vibrio parahaemolyticus* on Two Types of Surface Soiled with Different Substrates. *Food Control* **2017**, *74*, 1–8. [\[CrossRef\]](http://doi.org/10.1016/j.foodcont.2016.11.025)
- 88. Prechter, S.; Betz, M.; Cerny, G.; Wegener, G.; Windeisen, E. Hygienische Aspekte von Schneidebrettern aus Holz bzw. Kunststoff. *Holz Roh. Werkst.* **2002**, *60*, 239–248. [\[CrossRef\]](http://doi.org/10.1007/s00107-002-0301-5)
- 89. Dubreil, L.; Aviat, F.; Anthoine, V.; Ismail, R.; Rossero, A.; Federighi, M. Confocal Spectral Microscopy—An Innovative Tool for Tracking of Pathogen Agents on Contaminated Wooden Surfaces. *Eur. J. Wood Wood Prod.* **2018**, *76*, 1083–1085. [\[CrossRef\]](http://doi.org/10.1007/s00107-018-1294-z)
- 90. Carpentier, B. Sanitary Quality of Meat Chopping Board Surfaces: A Bibliographical Study. *Food Microbiol.* **1997**, *14*, 31–37. [\[CrossRef\]](http://doi.org/10.1006/fmic.1996.0061)
- 91. Ismail, R.; Aviat, F.; Gay-Perret, P.; Le Bayon, I.; Federighi, M.; Michel, V. An Assessment of *L. monocytogenes* Transfer from Wooden Ripening Shelves to Cheeses: Comparison with Glass and Plastic Surfaces. *Food Control* **2017**, *73*, 273–280. [\[CrossRef\]](http://doi.org/10.1016/j.foodcont.2016.08.014)
- 92. Montibus, M.; Ismaïl, R.; Michel, V.; Federighi, M.; Aviat, F.; Le Bayon, I. Assessment of *Penicillium expansum* and *Escherichia coli* Transfer from Poplar Crates to Apples. *Food Control* **2016**, *60*, 95–102. [\[CrossRef\]](http://doi.org/10.1016/j.foodcont.2015.07.025)
- 93. Dantas, S.T.A.; Rossi, B.F.; Bonsaglia, E.C.R.; Castilho, I.G.; Hernandes, R.T.; Fernandes, A.; Rall, V.L.M. Cross-Contamination and Biofilm Formation by *Salmonella enterica* Serovar Enteritidis on Various Cutting Boards. *Foodborne Pathog. Dis.* **2018**, *15*, 81–85. [\[CrossRef\]](http://doi.org/10.1089/fpd.2017.2341)
- 94. Munir, M.T.; Belloncle, C.; Irle, M.; Federighi, M. Wood-Based Bedding in Poultry Production: A Review. *World Poult. Sci. J.* **2019**, *75*, 5–16. [\[CrossRef\]](http://doi.org/10.1017/S0043933918000909)
- 95. Cimiotti, W.; Glunder, G.; Hinz, K.H. Survival of the Bacterial Turkey Coryza Agent. *Vet. Rec.* **1982**, *110*, 304–306. [\[CrossRef\]](http://doi.org/10.1136/vr.110.13.304)
- 96. Coughenour, C.; Stevens, V.; Stetzenbach, L.D. An Evaluation of Methicillin-Resistant *Staphylococcus aureus* Survival on Five Environmental Surfaces. *Microb. Drug Resist.* **2011**, *17*, 457–461. [\[CrossRef\]](http://doi.org/10.1089/mdr.2011.0007)
- 97. Xi, L.; Qin, D.; An, X.; Wang, G. Resistance of Natural Bamboo Fiber to Microorganisms and Factors That May Affect Such Resistance. *BioResources* **2013**, *8*, 6501–6509. [\[CrossRef\]](http://doi.org/10.15376/biores.8.4.6501-6509)
- 98. Tomičić, R.; Tomičić, Z.; Raspor, P. Adhesion of *Candida* Spp. and *Pichia* Spp. to Wooden Surfaces. Food Technol. Biotechnol. 2017, *55*, 138–142. [\[CrossRef\]](http://doi.org/10.17113/ftb.55.01.17.4514) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28559743)
- 99. Munir, M.T. Wood and Hospital Hygiene: Investigating the Hygienic Safety and Antimicrobial Properties of Wood Materials. Ph.D. Thesis, Ecole Centrale de Nantes, Nantes, France, 2021.