

Disentangling tropicalization and deborealization in marine ecosystems under climate change

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1 Disentangling tropicalization and deborealization in marine 2 ecosystems under climate change

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38 As climate change accelerates, species are shifting poleward and subtropical and tropical species are appearing in temperate environments¹⁻³. A popular approach for characterizing 39 40 such responses is the community temperature index (CTI), which tracks the mean thermal affinity of a community. Studies in marine⁴, freshwater⁵, and terrestrial⁶ ecosystems have 41 42 documented increasing CTI under global warming. However, most studies have only linked 43 increasing CTI to increases in warm-affinity species. Here, using long-term monitoring of 44 marine fishes across the Northern Hemisphere, we decomposed CTI changes into four 45 underlying processes – tropicalization (increasing warm-affinity), deborealization (decreasing 46 cold-affinity), borealization (increasing cold-affinity), and detropicalization (decreasing 47 warm-affinity) - for which we examined spatial variability and drivers. CTI closely tracked changes in sea surface temperature, increasing in 72% of locations. However, 31% of these 48 49 increases were primarily attributable to decreases in cold-affinity species, i.e., deborealization. 50 Thus, increases in warm-affinity species were prevalent, but not ubiquitous. Tropicalization 51 was stronger in areas that were initially warmer, experienced greater warming, or were 52 deeper, while deborealization was stronger in areas that were closer to human population 53 centers or that had higher community thermal diversity. When CTI (and temperature) 54 increased, species that decreased were more likely to be living closer to their upper thermal 55 limits or to be commercially fished. Additionally, warm-affinity species that increased had 56 smaller body sizes than those that decreased. Our results show that CTI changes arise from a 57 variety of underlying community responses that are linked to environmental conditions, 58 human impacts, community structure, and species characteristics.

59

60 Results and Discussion

Fish communities worldwide are responding to global warming through shifts in mean thermal affinity, which can be represented by the community temperature index (CTI)^{4,7–9}. An increase in CTI necessarily implies an increase in the relative abundance of warm-affinity species. However, a key question is whether this is primarily due to increases in the total abundance of warm-affinity species or to decreases in the total abundance of cold-affinity species. To resolve this, we decomposed CTI changes into four underlying processes:

- 'tropicalization' (increasing abundance of warm-affinity species)
- 'deborealization' (decreasing abundance of cold-affinity species)
- 'borealization' (increasing abundance of cold-affinity species)
- 'detropicalization' (decreasing abundance of warm-affinity species)

Here, we define warm-affinity and cold-affinity species locally within each community: 71 72 warm-affinity species are those whose thermal affinity is higher than the mean of the 73 community and cold-affinity species are those whose thermal affinity is lower than the mean. Additionally, whereas past literature has used the term 'tropicalization' to describe increasing 74 CTI^7 or poleward distribution shifts^{3,10–12}, we explicitly use this term to refer to an increase in 75 76 warm-affinity species. We applied this approach to fish communities using scientific 77 monitoring data from 558 grid cells covering 12 marine regions across the Northern 78 Hemisphere that showed contrasting changes in sea surface temperatures (SST) over the 79 period 1990 to 2015. We calculated the relative strength of each underlying processes in each 80 grid cell and identified which process was strongest when CTI increased or decreased. 81 Finally, we examined the potential influences of environmental conditions, human impacts, 82 and community structure on differences in the strength of the underlying processes and examined differences between species contributing to opposite processes (e.g., borealization
vs. deborealization).

85 Mean-annual SST increased in 72.4% (404) of grid cells between 1990 and 2015 with a mean of 0.23 ± 0.007 °C decade⁻¹ (mean \pm standard error), while it decreased in 27.6% of 86 cells (154) with a mean of -0.10 ± 0.008 °C decade⁻¹ (Figure 2A). CTI closely mirrored SST 87 88 (Pearson's correlation: 0.47), increasing in 71.3% (398) of cells, with a mean of 0.28 \pm 0.013° C decade⁻¹ (Figure 2B), and decreasing in 28.7% (160), with a mean of $-0.14 \pm 0.014^{\circ}$ C 89 decade⁻¹ (Figure 2B). Increases in CTI occurred primarily along the northeast coast of the 90 91 United States, in the Scottish Seas, the North Sea, the Baltic Sea, the Barents Sea, and around 92 the Aleutian Islands, while decreases were more prominent along the west and southeast 93 coasts of the United States and in the Bering Sea (Figure 2B).

94 We next decomposed changes in CTI and quantified the strength of each underlying 95 processes within each grid cell. Across all grid cells, tropicalization was the strongest process 96 on average being dominant in 47% of cells, while detropicalization was the weakest, being 97 dominant in only 7% of cells (Figure S1). Among the grid cells where CTI increased, 98 tropicalization was stronger than deborealization in 68.6% (while deborealization was 99 stronger in 31.4%) (Figure 2C). Hence, while past literature has focused extensively on increases in warm-affinity species and poleward distribution shifts^{3,7,11,13}, over one third of 100 101 CTI increases were attributable to decreases in cold-affinity species. Among the grid cells 102 where CTI decreased, borealization was stronger than detropicalization in 75% (Figure 2D). 103 These patterns were clearly spatially structured, as tropicalization was stronger than 104 deborealization along the east coast of the United States, in the Scottish Seas, the North Sea, 105 the Baltic Sea, along the west coast of Norway, in the western Barents Sea, and around the 106 Aleutian Islands. Deborealization was stronger in the Bering Sea, the Gulf of Mexico, and the 107 eastern Barents Sea (Figure 2C). Borealization was stronger than detropicalization in nearly 108 every region where CTI decreased, especially in the Bering Sea and along the west coast of109 the United States (Figure 2D).

110 To identify the biotic or abiotic conditions associated with each process, we next 111 modelled the difference in the strength of (i) tropicalization vs. deborealization when CTI 112 increased, and (ii) borealization vs. detropicalization when CTI decreased. Thus, the 113 difference in the strength of the processes was the response variable (i.e., tropicalization 114 minus deborealization; borealization minus deborealization). Explanatory variables were the 115 rate of change in SST, initial (i.e., baseline) SST, mean-annual SST variation, depth, distance 116 to the nearest human population center, mean maximum body size, community thermal 117 diversity (CTDIV), and community thermal range (CTR) (see STAR methods and Table S2 118 for details). We used linear mixed effects models with Gaussian likelihood distributions 119 where grid cells were the unit of observation and survey campaign was included as a random 120 effect (i.e., varying intercept). When CTI increased, tropicalization was stronger than 121 deborealization in cells that were initially warmer (effect size = 0.16 [0.07, 0.24; 95% CI]), 122 experienced greater warming (effect size = 0.07 [0.02, 0.13]) or were deeper (effect size = 123 0.07 [0.02, 0.11]; Figure 3A). Deborealization was stronger than tropicalization in cells that 124 were closer to human population centers (effect size = 0.07 [0.02, 0.11]) or that had greater 125 community thermal diversity (effect size = -0.05 [-0.10, -0.01]; Figure 3A). When CTI 126 decreased, borealization was stronger than detropicalization in cells that were initially warmer 127 (effect size = 0.13 [0.01, 0.25]), had greater temperature increases (effect size = 0.07 [0.01, 128 0.12]) (or lower temperature decreases since CTI decreases are mostly associated with 129 cooling), or were deeper (effect size = 0.06 [0.01, 0.11]; Figure 3B).

Theoretically, ignoring all factors other than temperature, when temperature and CTI are increasing, borealization and detropicalization should not occur, and when temperature and CTI are decreasing, tropicalization and deborealization should not occur. However, all

133 four processes occurred to some extent in nearly every grid cell (Figure S1). We therefore 134 hypothesized that there were mechanistic differences between species that explained this 135 anomaly. For instance, when CTI is increasing, species that contribute to borealization likely 136 differ in some key features from species that contribute to deborealization. We identified 137 differences between species contributing to (i) borealization vs. deborealization, and (ii) 138 tropicalization vs. detropicalization, using linear mixed effects models with binomial 139 likelihood distributions where species were the unit of observation and grid cell nested in 140 survey campaign were included as random effects (see STAR methods and Table S3 for 141 details). Thus, the binary response variable was whether a species was contributing to i) 142 deborealization (0) or borealization (1), or to ii) detropicalization (0) or tropicalization (1). In 143 grid cells where CTI increased, explanatory variables included maximum thermal limit, 144 thermal range, maximum body size, and whether species are commercially fished. In grid 145 cells where CTI decreased, the same explanatory variables were used except that minimum 146 thermal limit was used in place of maximum thermal limit. When CTI increased, species 147 contributing to borealization had higher maximum thermal limits (i.e., more tolerant of 148 warming) (effect size = 0.72 [0.53, 0.91]) while species contributing to deborealization were 149 more likely to be commercially fished (effect size = -0.34 [-0.49, -0.19]) and had wider 150 thermal ranges (effect size = -0.16 [-0.28, -0.04]; Figure 4A). Similarly, species contributing 151 to tropicalization had higher maximum thermal limits (effect size = 0.57 [0.38, 0.76]) and smaller body sizes (effect size = -0.17 [-0.24, -0.10]) and species contributing to 152 153 detropicalization had wider thermal ranges (effect size = -0.15 [-0.27, -0.03]; Figure 4A). 154 When CTI decreased, species contributing to borealization had wider thermal ranges than 155 those contributing to deborealization (effect size = 0.17 [0.04, 0.29]). Species contributing to 156 detropicalization had higher minimum thermal limits (effect size = -0.35 [-0.52, -0.17]), were more likely to be commercially fished (effect size = -0.26 [-0.44, -0.08]), and had smaller 157

body sizes (effect size = 0.09 [0.01, 0.18]; Figure 4B) than those contributing to
tropicalization.

160 Previous studies have documented large-scale changes in CTI but have not identified the underlying processes of these community thermal shifts^{3,4,6}. Unraveling these processes 161 162 has clear implications for predicting future biodiversity responses under global warming, as well as potential impacts on community trait composition^{14,15} and their consequences for 163 ecosystem structure and functioning¹⁶⁻¹⁸. For example, communities increasing in CTI due to 164 165 emigration or mortality of cold-affinity species (i.e., deborealization) could experience 166 population crashes or local extinctions under future warming and could be considered conservation priorities¹⁹⁻²¹. In contrast, communities increasing in CTI due to immigration or 167 168 population growth of warm-affinity species (i.e., tropicalization) may have increased 169 abundance and productivity despite changing composition^{8,22,23}, and could be resilient to well-170 managed fishing pressure.

While increases in CTI have been frequently linked to immigration or poleward 171 distribution shifts by warm-affinity species^{3,10,13}, we observed that over one third of CTI 172 173 increases were primarily explained by decreases in cold-affinity species (i.e., deborealization). 174 This result has major implications for understanding climate change impacts on community 175 structure, particularly as tropicalization and deborealization were spatially non-random and 176 associated with environmental variation and human impacts. Tropicalization was stronger 177 than deborealization in areas with warmer initial temperatures and areas with greater overall 178 warming. This is consistent with previous studies showing that community thermal shifts depend not only on the rate of warming, but also baseline climate. For instance, Antão et al.²⁴ 179 180 showed that in marine communities exposed to warming, species gains outpaced species losses under warmer initial conditions, and Lenoir et al.²⁵ showed that marine species track 181 182 isotherms more rapidly in initially warm waters. These results are consistent with faster

colonization and range edge expansion and slower extirpation and range edge contraction^{11,26}. 183 184 These results may also be explained by more rapid dispersal and population growth in warmer 185 environments. In marine organisms, the speed of metabolic and demographic processes increases with temperature²⁷, and both range expansion by new species and population growth 186 187 of existing species should occur more rapidly under warmer conditions. Warm species gains 188 may also dominate in warmer environments due to the latitudinal gradient in species richness, 189 as greater numbers and proportions of warm-affinity species are expected in warm, speciesrich areas 28 . 190

191 Tropicalization was generally stronger than deborealization in deeper areas, likely 192 owing to greater vertical temperature refuge for cold-affinity species⁹. For instance, 193 tropicalization was particularly strong along the east coast of the United States, in the Scottish 194 Seas, and in the western Barents Sea. These regions are situated along deep, open shelves, 195 which could enable cold-affinity species to temporarily seek refuge in cooler, deeper waters during warming episodes, preventing their loss locally²⁹. This is consistent with previous 196 197 studies showing that relatively small shifts in depth may allow species to remain within their thermal niches^{9,30} In the North Sea, a system primarily characterized by tropicalization, many 198 species have shifted to cooler, deeper waters over the last few decades³⁰. However, the North 199 Sea is a relatively shallow, semi-enclosed ecosystem and Rutterford et al.³¹ showed that North 200 201 Sea fishes will eventually be constrained by depth limitations, compressing habitat suitability 202 and potentially driving local extinction. Thus, the increase or immigration of warm-affinity 203 species could be currently out-pacing the decline or emigration of cold-affinity species, but 204 this trend could reverse in the future if cold-affinity species are unable to find thermal refuge.

Areas characterized by deborealization or detropicalization, i.e., decreasing abundance, had greater community thermal diversity than areas characterized by tropicalization or borealization. One hypothesis could be that communities with higher thermal diversity have fewer vacant niches (i.e., niche saturation) and therefore fewer opportunities for immigration and establishment by new species^{32,33}. Communities with greater thermal diversity may also contain more species living closer to their thermal limits, and thus have greater potential for species losses or population declines due to temperature rises⁹. For instance, Burrows et al.⁹ showed that communities with greater thermal diversity may have higher sensitivity to temperature changes, as species near their thermal limits can be rapidly lost or gained²⁸.

214 Tropicalization and borealization were more common than deborealization or 215 detropicalization. This suggests that habitat suitability is expanding for warm-affinity species faster than it is retracting for cold-affinity species²⁶. Hence, many cold-affinity species may be 216 217 tolerant of current warming, yet future warming could trigger major losses, potentially 218 shifting the balance between tropicalization and deborealization. Even when CTI decreased, 219 detropicalization was rarely dominant, as warm-affinity species rarely showed strong 220 decreases. While some areas did experience cooling during the study period, the average rate 221 of cooling was roughly half of the rate of warming, and all regions have experienced long-222 term temperature rises. Thus, warm-affinity species appear to be less impacted by periods of 223 mild cooling, and detropicalization should become increasingly rare under future warming.

224 Interestingly, we found that when CTI increased, some cold-affinity species increased 225 and some warm-affinity species decreased, counter to expectation. This was primarily 226 explained by thermal limits and apparent fishing pressure. Cold-affinity species that increased had higher maximum thermal limits than those that decreased, and those that decreased were 227 228 more likely to be commercially fished. Because species were compared within the same grid 229 cells, species with lower thermal maxima were living closer to their upper limits. Species 230 decreases can therefore be attributed to temperature rises surpassing thermal tolerances as 231 well as potential overfishing. Hence, both thermal tolerance and fishing pressure are shaping 232 long-term changes in marine fish communities, and future community responses will be

236 When CTI increased, warm-affinity species that increased had smaller body sizes than 237 those that decreased. Smaller-bodied species generally have faster growth rates, shorter 238 generation times, and less parental investment, enabling populations to rapidly track environmental changes^{14,35,36}. Thus, small-bodied species whose upper thermal limits were 239 240 not surpassed by temperature rises could rapidly increase in abundance following warming, 241 particularly as warming elevates metabolic and demographic rates. In contrast, large-bodied 242 species have slower growth rates and reproduce later in life, leading to slower population turnover and environmental tracking^{35,36}. Large-bodied species are also more susceptible to 243 244 human impacts³⁷. Hence, even large-bodied species that are favored by temperature rises 245 might be decreasing in abundance faster than they can reproduce, leading to population 246 declines despite warm-water affinities.

247 While limited to fish communities from 12 marine regions over a 26-year period, our 248 approach is applicable to other ecosystems and taxa and may help unravel the underlying processes of community thermal shifts at a global scale³⁸. Identifying how changes in species' 249 250 distributions and abundances are impacting overall diversity and community dynamics will be key for planning future conservation and management efforts³⁹⁻⁴². Areas with net losses of 251 252 cold-affinity species may require careful fisheries regulation, whereas areas gaining warmaffinity species may have increased productivity and exploitation opportunities^{8,23,43,44}. 253 254 Overall, we found that over one third of CTI increases were more strongly explained by 255 decreases in cold-affinity species than by increases in warm-affinity species, with significant 256 roles of environmental conditions, human impacts, and community structure. Additionally, we 257 found that species tendencies to increase or decrease in response to temperature changes were

dictated by thermal limits and commercial fishing status. Future studies should link spatial patterns in the underlying processes of CTI to changes in seasonality, ocean currents, and other abiotic factors likely to be modified by climate change, as well as changes in fishing pressure and human impacts. While past studies have documented extensive shifts in CTI, ours is the first to decompose CTI into underlying processes at a multi-continental scale, which could help in anticipating future changes in biodiversity under climate change and implementing adapted management strategies.

265

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279

280 Author contributions

- A.A., D.M. and M.M. designed the research, A.A.M., T.H., and M.M. collected and processed
- the data, M.M. analyzed the data, and all authors contributed to conceptual development and
- writing.
- 284

285 **Declaration of interests**

286 The authors declare no competing interests.

287

288 Figure legends

Figure 1. The four underlying processes contributing to changes in CTI. Increases in CTI occur when the combination of tropicalization (red) and deborealization (orange) is stronger than the combination of borealization (blue) and detropicalization (purple). CTI increases can therefore be attributed to either topicalization or deborealization, whichever process is stronger, and CTI decreases can be attributed to either borealization or detropicalization, whichever process is stronger.

295 Figure 2. Maps showing the rate of change in SST and CTI along with differences in the 296 strength of the underlying processes. Rate of change in SST (A) and CTI (B) across the 558 297 spatial sampling grid cells for the period 1990 - 2015. Differences in the strength of 298 tropicalization and deborealization in grid cells where CTI increased (C), and differences in 299 the strength of borealization and detropicalization in grid cells where CTI decreased (D). See 300 also Figure S1, which shows average relative strength of each underlying process, Figure S2, 301 which shows the area covered by each monitoring survey, Table S1, which provides details on 302 the monitoring surveys, Figure S3, which shows the method for calculating the strength of 303 each underlying process, and Figure S4, which compares the rate of change in CTI vs. 304 (topicalization + deborealization) – (borealization + detropicalization).

305 Figure 3. Results of linear mixed effects models of differences in the strength of tropicalization and deborealization in grid cells where CTI increased (A), and of 306 differences in the strength of borealization and detropicalization in grid cells where CTI 307 308 decreased (B). Grey circles represent standardized effect sizes and black horizontal bars 309 represent 95% confidence intervals. In panel A, positive effects are associated with stronger 310 tropicalization, and negative effects are associated with stronger deborealization. In panel B, positive effects area associated with stronger borealization, and negative effects are associated 311 312 with stronger detropicalization. See also Table S2, which shows the output summary for each 313 model.

Figure 4. Results of linear mixed effects models of i) the probability that a species contributed to borealization or deborealization, and ii) the probability that species 316 contributed to topicalization or detropicalization when CTI increased (A) and when CTI 317 decreased (B). Grey circles represent standardized effect sizes and black horizontal bars 318 represent 95% confidence intervals. Positive effects are associated with species that 319 contributed to borealization or tropicalization, and negative effects are associated with species 320 that contributed to deborealization or detropicalization. See also Table S3, which shows the 321 output summary for each model, and Table S4, which compares model results using different 322 subsets of species based on quantiles of abundance changes.

323

324 STAR METHODS

325

326 **RESOURCE AVAILABILITY**

- 327 Lead Contact
- 328 Further information and requests should be directed to and will be fulfilled by the lead
- 329 contact, Matthew McLean (mcleamj@gmail.com).
- 330
- 331 Materials Availability
- 332 This study did not generate new unique reagents.

333

334 Data and Code Availability

This paper analyzes existing, publicly available data. Links for the datasets are provided in the key resources table. This paper does not report original code. Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

339

340 EXPERIMENTAL MODEL AND SUBJECT DETAILS

All fish monitoring data used in this study are freely available and open access; references and
links are provided in the Key resources table and Supplemental information. No experimental
models (animals, human subjects, plants, microbe strains, cell lines, primary cell cultures)
were used in the study.

345

346 METHOD DETAILS

347 Fish community data

348 Thirteen bottom-trawl surveys from 12 marine regions across the northern hemisphere were 349 used to examine changes in the community temperature index (CTI) in fish communities over 350 a large geographic scale with substantial longitudinal, latitudinal, and depth gradients. All 351 surveys used similar sampling protocols, where bottom trawls were towed for an average of 352 30 minutes and the species composition and abundances of all captured fishes were identified 353 and recorded (see Table S1). Spatial coverage and resolution differed across surveys, and we 354 therefore aggregated trawl surveys to 1° longitude $\times 1^{\circ}$ latitude spatial grid cells. A 1° 355 longitude $\times 1^{\circ}$ latitude resolution was chosen to adequately capture both inter and intra-survey 356 variation, to reveal gradients in community responses, to maximize data availability, and to 357 match with the spatial resolution of the HadISST database (see 'Sea surface temperature' 358 below). The length of time series also differed between surveys, and we therefore examined 359 the period 1990 – 2015, which maximized temporal overlap between surveys. Following 360 Burrows et al.⁹, along the US West Coast, two surveys with overlapping spatial coverage but 361 adjacent temporal periods were combined (see Figure S1 and Table S1). The combined data 362 were inspected for discontinuities, and we verified that our main results and conclusions were 363 robust to removing these data from the analyses. Because some surveys are conducted in 364 multiple seasons, for each grid cell, we only used data for the quarter with the greatest number

365 of years surveyed. Lastly, because of spatial and temporal heterogeneity in sampling effort 366 both between and within grid cells, we performed a bootstrap sampling procedure. We 367 randomly selected four trawl surveys per grid cell, per year (four was the median number of trawls per cell, per year), recorded the resulting species' abundances, repeated this procedure 368 369 99 times, and calculated species' mean abundances across the 99 permutations. Only grid 370 cells with with maximum sampling gaps of five years or less were considered (some surveys 371 are only conducted every 3-5 years), resulting in a total of 558 cells. All survey abundance 372 data were then $log_{10}(x+1)$ transformed before analyses. While we recognize that aggregating 373 bottom trawl data to a 1° longitude $\times 1^{\circ}$ latitude scale creates species assemblages that are not 374 true locally interacting biological communities, we use the term 'community' for consistency 375 with existing literature on concepts such as the community temperature index and community 376 thermal diversity.

377

378 Sea surface temperature (SST)

379 For each grid cell, we extracted mean-annual sea surface temperature (SST) and annual SST variation. Minimum and maximum SST were also initially considered, but later dropped 380 381 because they were highly correlated with mean SST, but much less informative (i.e., never 382 had discernable effects in statistical models). SST data for each grid cell were derived from 383 the Hadley Centre for Climate Prediction and Research's freely available HadISST1 database⁴⁶. The HadISST1 database provides global monthly SST on a 1° longitude \times 1° 384 385 latitude spatial grid and is available for all years since 1870. These data were used to examine 386 temperature changes during the study period and to model the underlying processes of CTI.

387

388 Calculating community temperature index (CTI)

389 Community temperature index (CTI) is the abundance-weighted mean thermal affinity of a 390 community or assemblage, which reflects the relative abundance of warm-affinity or coldaffinity species⁵⁰. The inferred thermal affinity for each fish species in this study (1091 391 392 species total) was first calculated as the median temperature of each species' occurrences 393 across its' global range of observations for which data were available (Figure S2). Rather than 394 surface temperature or bottom temperature, we used mid-water-column temperature (i.e., 395 from the surface to 200 meters depth) because the surveys included a mixture of demersal 396 (bottom-living) and pelagic species. We used temperature climatologies from the global 397 database WOD V2 (https://www.nodc.noaa.gov/cgi-2013 398 bin/OC5/woa13/woa13.pl?parameter=t) with a spatial resolution of ¹/₄°. These climatologies 399 represent average decadal temperatures for 1955-1964, 1965-1974, 1975-1984, 1985-1994, 400 1995-2004 and 2005-2012 on 40 depth layers. These data were aggregated vertically by 401 calculating average temperature of the first 200 m depth. Species' occurrences were extracted 402 from several databases including OBIS (https://obis.org/), GBIF (https://www.gbif.org/), 403 VertNet (http://vertnet.org/) and ecoengine (https://ecoengine.berkeley.edu/). After removing 404 duplicate occurrence records, we made a spatiotemporal match-up between temperature 405 climatologies and species occurrences, considering both the geographic coordinates of 406 occurrences, as well as their corresponding decade (to control for climate trends over the past 407 58 years). We then took the median value of temperature from these records for each species. 408 Although we included both demersal and pelagic species and used mid-water-column 409 temperature to infer thermal affinities in our analyses, we tested the sensitivity of our results 410 to these choices by recalculating thermal affinities using surface temperature and bottom 411 temperature, both with and without pelagic species (see Supplementary Material). Separate 412 data sources were used to calculate species' thermal affinities and to model the underlying 413 processes of CTI because estimating species' thermal affinities required matching species'

414 occurrences with mid-water column temperatures, whereas modelling the underlying 415 processes required a standardized, continuous, temporally resolved database. Mid-water-416 column data were only available as decadal averages and did not cover the entire study 417 period. Lastly, for each grid cell, we calculated the rate of change in SST and CTI as the slope 418 of simple linear regressions of SST and CTI vs. time.

419

420 Comparison of thermal affinities with Cheung et al. 2013⁴

421 While a variety of past studies have quantified species' thermal affinities using species' distribution models⁵¹ or the midpoint of species minimum and maximum temperature 422 observations²⁸, here we inferred thermal affinities as the median temperature value across a 423 424 species' range of observations. To determine the accuracy of this approach, we compared our data with those of Cheung et al. 2013⁴ for 252 overlapping species. We found an 83% 425 correlation between our data and those of Cheung et al. 2013⁴, indicating high consistency 426 427 between the two studies. This provides strong support for our approach because Cheung et al. 2013⁴ is a landmark study investigating changes in the community temperature index in 428 429 marine fishes.

430

431 QUANTIFICATION AND STATISTICAL ANALYSIS

432 All data handling and quantitative analyses were performed using R^{45} version 4.0.0.

433

434 Decomposing CTI into the four underlying processes

CTI is a community weighted mean and therefore reflects changes in the relative abundances 435 of warm-affinity and cold-affinity species. CTI will increase when species with thermal 436 437 affinities greater than the mean of the community increase and when species with thermal 438 affinities lower than the mean of the community decrease. Conversely, CTI will decrease 439 when species with thermal affinities greater than the mean of the community decrease and 440 when species with thermal affinities lower than the mean of the community increase. Hence, 441 CTI changes can be decomposed into four underlying process – tropicalization (increasing 442 warm-affinity species), deborealization (decreasing cold-affinity species), borealization 443 (increasing cold-affinity species), and detropicalization (decreasing warm-affinity species). 444 The overall change in CTI reflects the relative strength of these processes. For instance, CTI 445 will increase when the strength of tropicalization + deborealization is greater than the strength 446 of borealization + detropicalization. To determine the strength of each underlying process, 447 species within each grid cell must first be classified as either warm-affinity or cold-affinity. 448 Because CTI (the mean thermal affinity of the community) changes every year, species may 449 be warm-affinity one year (i.e., having a thermal affinity higher than the community mean) 450 and cold-affinity the next (i.e., having a thermal affinity lower than the community mean). 451 Therefore, to classify species as either warm or cold affinity within each grid cell, we used the 452 mean CTI value across all years in the time series (i.e., mean of CTI values for 1990 to 2015 453 for each grid cell). We then separated warm and cold-affinity species into those that increased 454 in abundance and those that decreased (Figure 1). Because CTI will shift up or down based on 455 the amount of increase or decrease in species abundances along the thermal affinity axis (i.e., 456 Figure 1), the strength of each process can be thought of as the amount of "pull" that each 457 process exhibits on the overall community mean. This is determined by the degree to which 458 species contributing to each process influence the overall community mean. Species that have 459 thermal affinities much greater or much lower than the community mean will exhibit more

influence than those with thermal affinities very similar to the mean. Additionally, species 460 461 with large abundance changes will exhibit more influence than those with small abundance 462 changes. Hence, each species contribution to the change in CTI is a combination of the 463 difference between its individual thermal affinity (STI) and that of the community (CTI) and 464 its change in abundance. We therefore calculated the strength of each processes by (i) 465 calculating the difference between each species' thermal affinity and the mean of the 466 community, (ii) multiplying this value by each species' change in abundance, and (iii) taking 467 the sum of the resulting values for all species within each process (Figure S3). We assessed 468 the accuracy of this approach by comparing the value of (tropicalization + deborealization) – 469 (borealization + detropicalization) to the rate of change in CTI for each grid cell. Note, these 470 two values will never be a perfect match because, as mentioned above, some species fluctuate 471 between warm and cold-affinity over time, especially in grid cells where CTI is highly 472 variable across years. However, we found a correlation of 0.85 between the two values, 473 indicating that our metric for estimating the strength of the underlying processes accurately 474 captured changes in CTI (Figure S4).

475

476 Conditions associated with the underlying processes

To identify the biotic and abiotic conditions associated with each underlying process, we modelled the difference in the strength of tropicalization vs. deborealization (i.e., tropicalization minus deborealization) when CTI increased, and the difference in the strength of borealization vs. detropicalization (i.e., borealization minus detropicalization) when CTI decreased. We used linear mixed effects models with Gaussian likelihood distributions and included survey campaign as a random effect (i.e., varying intercept). Explanatory variables were the rate of change in SST, initial (i.e., baseline) SST, mean-annual SST variation, depth, 484 distance to the nearest human population center, mean maximum body size, community thermal diversity (CTDIV), and community thermal range (CTR). Initial SST was defined as 485 486 the mean-annual SST for each grid cell for the period 1980-1989, the ten years prior to the 487 study period. Depth was recorded during each trawl survey, and we calculated mean depth per grid cell. Distance to the nearest human population center came from Yeager at al.⁴⁷, which is 488 489 calculated as the straight-line distance to the nearest provincial capital as defined by the ESRI 490 World Cities data set. Body size data came from the open-access trait database of Beukhof et 491 al.⁴⁸. CTDIV was defined as the variation in thermal affinities in the community and was calculated as the abundance-weighted standard deviation of species' thermal affinities⁹. CTR 492 describes whether species in the community have narrow or wide thermal ranges and was 493 494 calculated as the abundance-weighted mean of species' thermal ranges⁹. Thermal ranges were defined as the difference between the 90th and 10th percentiles of species thermal affinity 495 496 observations. For CTDIV, CTR, and mean body size, we took the mean across the first 10 497 years of the study period for each grid cell to define baseline conditions in community 498 structure that may have shaped community responses to warming. All metrics were calculated 499 for the entire community sampled in each grid cell. Hence, identical predictors were used for 500 both models, rather than sub-setting predictors to only species contributing to tropicalization 501 and deborealization or to borealization and detropicalization.

502

503 Species contributing to opposite processes

To identify differences between species contributing to borealization vs. deborealization, and between species contributing to tropicalization vs. detropicalization, we used linear mixed effects models with binomial likelihood distributions and grid cell nested in survey campaign as random effects (i.e., varying intercepts). In grid cells where CTI increased, explanatory 508 variables included maximum thermal limit, thermal range, maximum body size, and whether 509 species are commercially fished. In grid cells where CTI decreased, the same explanatory 510 variables were used except that minimum thermal limit was used in place of maximum thermal limit. Maximum and minimum thermal limits were defined as the 90th and 10th 511 512 percentiles of species thermal affinity observations, respectively, and species thermal ranges were defined as the difference between the 90th and 10th percentiles. Body size again came 513 from Beukhof et al.⁴⁸. We defined whether a species was commercially fished according to 514 categories of commercial importance available from FishBase⁴⁹. Species listed as 'highly 515 516 commercial', 'commercial', or 'minor commercial', were considered commercially fished, 517 and species listed as 'of no interest', 'of potential interest', 'subsistence fisheries', or 518 'unknown' were considered not commercially fished. All models were performed using the R 519 package lme4⁵². Model quality and assumptions were verified using the R packages performance⁵³ and MuMin⁵⁴ (see Supplementary Material). Initial model inspection revealed 520 521 low predictive accuracy and explained variation for the binomial models. This was likely 522 because all species were initially included in this analysis whether they showed very slight or 523 very large changes in abundance, i.e., any cold-affinity species whose change in abundance 524 was greater than 0 was classified as contributing to borealization. All species populations 525 fluctuate naturally, and so small increases or decreases in abundance are expected that may be 526 independent of thermal affinity. Hence, including all species in this analysis could potentially 527 blur patterns. We therefore reran models using i) all species, ii) species whose abundance 528 changes were in the top 75%, iii) species whose abundance changes were in the top 50%, and iv) species whose abundance changes were in the top 25%. All approaches yielded very 529 530 similar results, but with predictive accuracy and explained variation increasing with stricter 531 species subsets. We therefore selected the model using species whose abundance changes were in the top 50% as a compromise between data deletion and model quality (at least 2000 532

observations per model and predictive accuracy over 70%), however, all model results arereported in Table S4.

535

536 Model performance

537 We assessed the performance of all models using the R package performance. For the two 538 Gaussian models, we assessed linearity (i.e., residuals vs fitted values), homogeneity of 539 variance, collinearity, the potential influence of high leverage observations, normality of 540 residuals, and normality of random effects. This was accomplished with the function 541 check_model. We also assessed predictive accuracy via the correlation between fitted values 542 and observed values and via k-fold cross validation using the function performance accuracy. 543 Because cross validation results vary between iterations, we ran the performancy accuracy 544 function 99 times and recorded the average score. Both models satisfied all assumptions, 545 including no high leverage observations and Variance Inflation Factors under 2.5 for all 546 variables. For the model of differences between the strength of tropicalization and 547 deborealization, the correlation between fitted values and observed values was 62% and the 548 average cross validation accuracy was 57%. For the model of differences between the strength 549 of borealization and deborealization, the correlation between fitted values and observed 550 values was 79% and the average cross validation accuracy was 71%.

For the four binomial models, we assessed binned residuals and predictive accuracy using the functions binned_residuals and performance_accuracy. Binned residuals are assessed by first ordering predicted probabilities from smallest to largest and calculating raw residuals. Data are then split into bins of equal numbers of observations and the average residual is plotted against the average predicted probability for each bin. The quality of the model is then evaluated based on the percentage of binned residuals that lie within confidence 557 limits/error bounds. Predictive accuracy was assessed as the area under the receiver operating 558 characteristic curve (AUC - ROC), which evaluates how accurately a binomial model predicts 559 group classification. AUC – ROC is bounded between 0 and 1, with 0 indicating 0% accuracy 560 and 1 indicating 100% accuracy. For sites where CTI increased, the model of differences 561 between species contributing to borealization and deborealization had 85% of residuals within 562 error bounds and a predictive accuracy of 73%, while the model of differences between 563 species contributing to tropicalization and detropicalization had 84% of residuals within error 564 bounds and a predictive accuracy of 73%. For sites where CTI decreased, the model of 565 differences between species contributing to borealization and deborealization had 83% of 566 residuals within error bounds and a predictive accuracy of 71%, while the model of 567 differences between species contributing to tropicalization and detropicalization had 86% of 568 residuals within error bounds and a predictive accuracy of 70%.

569 Altogether, these results show that our models did not violate assumptions, but that 570 predictive accuracy was less than desirable. This likely indicates that other drivers that we 571 were unable to assess are important in explaining variation in the strength of processes and in 572 differences between species contributing to opposite processes. Further exploration showed 573 that poor predictive accuracy may have also resulted from inconsistent relationships between 574 surveys (i.e., regions). For example, including a random slope term for survey in the binomial 575 models showed that, in sites where CTI decreased, upper thermal maximum was a strong 576 predictor of whether species underwent borealization or derealization for all surveys except the Gulf of Alaska, Gulf of Mexico, and Baltic Sea. Additionally, commercially fished status 577 578 was a strong predictor of whether species underwent borealization or deborealization in 579 regions that were closer to human population centers, but not those that were further from 580 population centers. However, models that included random slope terms did not have greater

predictive accuracy, indicating that improving model accuracy ultimately hangs on
uncovering other important drivers of process strength and species differences.

583

584 Sensitivity to pelagic species and temperature zone

585 To determine how including or excluding pelagic species influenced our results, we 586 recalculated i) the rate of change in CTI, ii) the difference in the strength of tropicalization 587 and deborealization, and ii) the difference in the strength of borealization and 588 detropicalization after removing pelagic species. Additionally, to examine the impact of 589 calculating thermal affinities with different water column zones (i.e., bottom temperature, 590 mid-water-column temperature, and sea surface temperature) we recalculated the above three 591 metrics using all three temperature zones. We did this for all possible scenarios, hence for all 592 species using bottom, mid-water-column, and surface temperature, and for demersal species 593 only using bottom, mid-water-column, and surface temperature. We then examined the 594 correlation in metrics across all six scenarios. Across the six scenarios, correlation values for 595 the rate of change in CTI ranged from 0.666 to 0.996 with a mean of 0.83, correlation values 596 for the difference in the strength of tropicalization and deborealization ranged from 0.776 to 597 0.997 with a mean of 0.873, and correlation values for the difference in the strength of 598 borealization and detropicalization ranged from 0.816 to 0.997 with a mean of 0.894, 599 altogether indicating that results were robust to including or excluding pelagic species and to 600 potential choices in thermal affinity calculation.

601

602 **References**

- 603 1. Chen, I.-C., Hill, J.K., Ohlemüller, R., Roy, D.B., and Thomas, C.D. (2011). Rapid Range
 604 Shifts of Species Associated with High Levels of Climate Warming. Science *333*, 1024.
- Pinsky, M.L., Worm, B., Fogarty, M.J., Sarmiento, J.L., and Levin, S.A. (2013). Marine
 taxa track local climate velocities. Science *341*, 1239–1242.
- Verges, A., Steinberg, P.D., Hay, M.E., Poore, A.G.B., Campbell, A.H., Ballesteros, E.,
 Heck, K.L., Booth, D.J., Coleman, M.A., Feary, D.A., et al. (2014). The tropicalization of
 temperate marine ecosystems: climate-mediated changes in herbivory and community
 phase shifts. Proc. R. Soc. B Biol. Sci. 281, 20140846–20140846.
- 611 4. Cheung, W.W., Watson, R., and Pauly, D. (2013). Signature of ocean warming in global
 612 fisheries catch. Nature 497, 365–368.
- 613 5. Comte, L., Olden, J.D., Tedesco, P.A., Ruhi, A., and Giam, X. (2021). Climate and land614 use changes interact to drive long-term reorganization of riverine fish communities
 615 globally. Proc. Natl. Acad. Sci. *118*, e2011639118.
- 6. Devictor, V., Van Swaay, C., Brereton, T., Chamberlain, D., Heliölä, J., Herrando, S.,
 Julliard, R., Kuussaari, M., Lindström, Å., and Roy, D.B. (2012). Differences in the
 climatic debts of birds and butterflies at a continental scale. Nat. Clim. Change 2, 121.
- 619 7. Cheung, W.W., Meeuwig, J.J., Feng, M., Harvey, E., Lam, V.W., Langlois, T., Slawinski,
 620 D., Sun, C., and Pauly, D. (2012). Climate-change induced tropicalisation of marine
 621 communities in Western Australia. Mar. Freshw. Res. *63*, 415–427.
- 8. Day, P.B., Stuart-Smith, R.D., Edgar, G.J., and Bates, A.E. (2018). Species' thermal
 ranges predict changes in reef fish community structure during 8 years of extreme
 temperature variation. Divers. Distrib. 24, 1036–1046.
- Burrows, M.T., Bates, A.E., Costello, M.J., Edwards, M., Edgar, G.J., Fox, C.J., Halpern,
 B.S., Hiddink, J.G., Pinsky, M.L., and Batt, R.D. (2019). Ocean community warming
 responses explained by thermal affinities and temperature gradients. Nat. Clim. Change 9,
 959–963.
- 10. Vergés, A., Doropoulos, C., Malcolm, H.A., Skye, M., Garcia-Pizá, M., Marzinelli, E.M.,
 Campbell, A.H., Ballesteros, E., Hoey, A.S., Vila-Concejo, A., et al. (2016). Long-term
 empirical evidence of ocean warming leading to tropicalization of fish communities,
 increased herbivory, and loss of kelp. Proc. Natl. Acad. Sci. *113*, 13791–13796.
- 11. Bates, A.E., Pecl, G.T., Frusher, S., Hobday, A.J., Wernberg, T., Smale, D.A., Sunday,
 J.M., Hill, N.A., Dulvy, N.K., and Colwell, R.K. (2014). Defining and observing stages of
 climate-mediated range shifts in marine systems. Glob. Environ. Change 26, 27–38.
- Horta e Costa B, Assis J, Franco G, Erzini K, Henriques M, Gonçalves EJ, and Caselle JE
 (2014). Tropicalization of fish assemblages in temperate biogeographic transition zones.
 Mar. Ecol. Prog. Ser. 504, 241–252.
- 639 13. Osland, M.J., Stevens, P.W., Lamont, M.M., Brusca, R.C., Hart, K.M., Waddle, J.H.,
 640 Langtimm, C.A., Williams, C.M., Keim, B.D., and Terando, A.J. (2021). Tropicalization
 641 of temperate ecosystems in North America: The northward range expansion of tropical
 642 organisms in response to warming winter temperatures. Glob. Change Biol.

- 643 14. Pecuchet, L., Lindegren, M., Hidalgo, M., Delgado, M., Esteban, A., Fock, H.O., Gil de
 644 Sola, L., Punzón, A., Sólmundsson, J., and Payne, M.R. (2017). From traits to life-history
 645 strategies: Deconstructing fish community composition across European seas. Glob. Ecol.
 646 Biogeogr. 26, 812–822.
- 647 15. Beukhof, E., Frelat, R., Pecuchet, L., Maureaud, A., Dencker, T.S., Sólmundsson, J.,
 648 Punzón, A., Primicerio, R., Hidalgo, M., Möllmann, C., et al. (2019). Marine fish traits
 649 follow fast-slow continuum across oceans. Sci. Rep. 9, 17878.
- 16. Duffy, J.E., Lefcheck, J.S., Stuart-Smith, R.D., Navarrete, S.A., and Edgar, G.J. (2016).
 Biodiversity enhances reef fish biomass and resistance to climate change. Proc. Natl.
 Acad. Sci. *113*, 6230–6235.
- Maureaud, A., Hodapp, D., van Denderen, P.D., Hillebrand, H., Gislason, H., Spaanheden
 Dencker, T., Beukhof, E., and Lindegren, M. (2019). Biodiversity–ecosystem functioning
 relationships in fish communities: biomass is related to evenness and the environment, not
 to species richness. Proc. R. Soc. B Biol. Sci. 286, 20191189.
- Maureaud, A., Andersen, K.H., Zhang, L., and Lindegren, M. (2020). Trait-based food
 web model reveals the underlying mechanisms of biodiversity–ecosystem functioning
 relationships. J. Anim. Ecol. 89, 1497–1510.
- 500 19. Stuhldreher Gregor, Hermann Gabriel, and Fartmann Thomas (2014). Cold-adapted
 501 species in a warming world an explorative study on the impact of high winter
 502 temperatures on a continental butterfly. Entomol. Exp. Appl. *151*, 270–279.
- 20. Pearce-Higgins James W., Eglington Sarah M., Martay Blaise, Chamberlain Dan E., and
 Both Christiaan (2015). Drivers of climate change impacts on bird communities. J. Anim.
 Ecol. 84, 943–954.
- Auber, A., Gohin, F., Goascoz, N., and Schlaich, I. (2017). Decline of cold-water fish
 species in the Bay of Somme (English Channel, France) in response to ocean warming.
 Estuar. Coast. Shelf Sci. *189*, 189–202.
- 669 22. Hiddink, J., and Ter Hofstede, R. (2008). Climate induced increases in species richness of
 670 marine fishes. Glob. Change Biol. 14, 453–460.
- Barange, M., Merino, G., Blanchard, J.L., Scholtens, J., Harle, J., Allison, E.H., Allen,
 J.I., Holt, J., and Jennings, S. (2014). Impacts of climate change on marine ecosystem
 production in societies dependent on fisheries. Nat. Clim. Change 4, 211.
- Antão, L.H., Bates, A.E., Blowes, S.A., Waldock, C., Supp, S.R., Magurran, A.E.,
 Dornelas, M., and Schipper, A.M. (2020). Temperature-related biodiversity change across
 temperate marine and terrestrial systems. Nat. Ecol. Evol. *4*, 927–933.
- 677 25. Lenoir, J., Bertrand, R., Comte, L., Bourgeaud, L., Hattab, T., Murienne, J., and
 678 Grenouillet, G. (2020). Species better track climate warming in the oceans than on land.
 679 Nat. Ecol. Evol. 4, 1044–1059.
- 680 26. Fredston-Hermann, A., Selden, R., Pinsky, M., Gaines, S.D., and Halpern, B.S. (2020).
 681 Cold range edges of marine fishes track climate change better than warm edges. Glob.
 682 Change Biol. 26, 2908–2922.

- 683 27. Clarke, A., and Johnston, N.M. (1999). Scaling of metabolic rate with body mass and
 684 temperature in teleost fish. J. Anim. Ecol. 68, 893–905.
- 585 28. Stuart-Smith, R.D., Edgar, G.J., Barrett, N.S., Kininmonth, S.J., and Bates, A.E. (2015).
 586 Thermal biases and vulnerability to warming in the world's marine fauna. Nature 528,
 587 88–92.
- 488 29. Jorda, G., Marbà, N., Bennett, S., Santana-Garcon, J., Agusti, S., and Duarte, C.M.
 (2020). Ocean warming compresses the three-dimensional habitat of marine life. Nat.
 Ecol. Evol. 4, 109–114.
- 30. Dulvy, N.K., Rogers, S.I., Jennings, S., Stelzenmller, V., Dye, S.R., and Skjoldal, H.R.
 (2008). Climate change and deepening of the North Sea fish assemblage: a biotic
 indicator of warming seas. J. Appl. Ecol. 45, 1029–1039.
- 81. Rutterford, L.A., Simpson, S.D., Jennings, S., Johnson, M.P., Blanchard, J.L., Schön, P.J., Sims, D.W., Tinker, J., and Genner, M.J. (2015). Future fish distributions constrained
 by depth in warming seas. Nat. Clim. Change 5, 569.
- 697 32. Givan, O., Parravicini, V., Kulbicki, M., and Belmaker, J. (2017). Trait structure reveals
 698 the processes underlying fish establishment in the Mediterranean. Glob. Ecol. Biogeogr.
 699 26, 142–153.
- 33. Beaugrand, G., Luczak, C., Goberville, E., and Kirby, R.R. (2018). Marine biodiversity
 and the chessboard of life. PLOS ONE *13*, e0194006.
- 34. Bowler, D., and Böhning-Gaese, K. (2017). Improving the community-temperature index
 as a climate change indicator. PLOS ONE *12*, e0184275.
- 35. McLean, M., Mouillot, D., Lindegren, M., Engelhard, G., Villéger, S., Marchal, P.,
 Brind'Amour, A., and Auber, A. (2018). A climate-driven functional inversion of
 connected marine ecosystems. Curr. Biol. 28, 3654–3660.
- 36. McLean, M., Mouillot, D., and Auber, A. (2018). Ecological and life history traits explain
 a climate-induced shift in a temperate marine fish community. Mar. Ecol. Prog. Ser. 606,
 175–186.
- 37. Mellin, C., Mouillot, D., Kulbicki, M., Mcclanahan, T.R., Vigliola, L., Bradshaw, C.,
 Brainard, R., Chabanet, P., Edgar, G., and Fordham, D. (2016). Humans and seasonal
 climate variability threaten large-bodied coral reef fish with small ranges. Nat. Commun.
 7, 1–9.
- 38. A Maureaud, A., Frelat, R., Pécuchet, L., Shackell, N., Mérigot, B., Pinsky, M.L.,
 Amador, K., Anderson, S.C., Arkhipkin, A., and Auber, A. (2021). Are we ready to track
 climate-driven shifts in marine species across international boundaries?-A global survey
 of scientific bottom trawl data. Glob. Change Biol. 27, 220–236.
- 39. Brander, K.M. (2007). Global fish production and climate change. Proc. Natl. Acad. Sci. *104*, 19709–19714.

- 40. Wernberg, T., Bennett, S., Babcock, R.C., de Bettignies, T., Cure, K., Depczynski, M.,
 Dufois, F., Fromont, J., Fulton, C.J., and Hovey, R.K. (2016). Climate-driven regime shift
 of a temperate marine ecosystem. Science *353*, 169–172.
- 41. Gaines, S.D., Costello, C., Owashi, B., Mangin, T., Bone, J., Molinos, J.G., Burden, M.,
 Dennis, H., Halpern, B.S., and Kappel, C.V. (2018). Improved fisheries management
 could offset many negative effects of climate change. Sci. Adv. *4*, eaao1378.
- 42. Goldenberg, S.U., Nagelkerken, I., Marangon, E., Bonnet, A., Ferreira, C.M., and
 Connell, S.D. (2018). Ecological complexity buffers the impacts of future climate on
 marine consumers. Nat. Clim. Change 8, 229.
- 43. Cheung, W.W., Lam, V.W., Sarmiento, J.L., Kearney, K., Watson, R., Zeller, D., and
 Pauly, D. (2010). Large-scale redistribution of maximum fisheries catch potential in the
 global ocean under climate change. Glob. Change Biol. *16*, 24–35.
- 44. Sumaila, U.R., Cheung, W.W., Lam, V.W., Pauly, D., and Herrick, S. (2011). Climate
 change impacts on the biophysics and economics of world fisheries. Nat. Clim. Change *1*,
 449–456.
- 45. Team, R.C. (2013). R: A language and environment for statistical computing.
- 46. Rayner, N., Parker, D., Folland, C., Horton, E., Alexander, L., and Rowell, D. (2003). The
 global sea-ice and sea surface temperature (HadISST) data sets. J Geophys Res *108*, 2–22.
- 47. Yeager, L.A., Marchand, P., Gill, D.A., Baum, J.K., and McPherson, J.M. (2017). Marine
 Socio-Environmental Covariates: queryable global layers of environmental and
 anthropogenic variables for marine ecosystem studies. Ecology *98*, 1976–1976.
- 48. Beukhof, E., Dencker, T.S., Palomares, M.L.D., and Maureaud, A. (2019). A trait
 collection of marine fish species from North Atlantic and Northeast Pacific continental
 shelf seas. PANGEA.
- Froese, R., and Pauly, D. (2010). FishBase (www database). Available at
 http://www.Fishbase.org
- 50. Devictor, V., Julliard, R., Couvet, D., and Jiguet, F. (2008). Birds are tracking climate
 warming, but not fast enough. Proc. R. Soc. B Biol. Sci. 275, 2743.
- 51. Flanagan, P.H., Jensen, O.P., Morley, J.W., and Pinsky, M.L. (2019). Response of marine
 communities to local temperature changes. Ecography *42*, 214–224.
- 52. Bates, D., Mächler, M., Bolker, B., and Walker, S. (2014). Fitting linear mixed-effects
 models using lme4. ArXiv Prepr. ArXiv14065823.
- 53. Lüdecke, D., Ben-Shachar, M.S., Patil, I., Waggoner, P., and Makowski, D. (2021).
 performance: An R package for assessment, comparison and testing of statistical models.
 J. Open Source Softw. 6.
- 755 54. Barton, K., and Barton, M.K. (2015). Package 'mumin.' Version 1, 439.

756



SST Rate of Change





D

С

Α

В

Borealization – Detropicalization







Increasing CTI



Decreasing CTI