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Pelagic food web structure in high nutrient low chlorophyll (HNLC) and naturally iron fertilized waters in the Kerguelen Islands region, Southern Ocean

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Abstract :

The Kerguelen Plateau is a region of natural iron fertilization in the Southern Ocean, within the typically iron limited High Nutrient Low Chlorophyll (HNLC) waters of the eastward flowing Antarctic Circumpolar Current. Between 26 February and 19 March 2018, the MOBYDICK expedition investigated pelagic ecosystem dynamics in the Kerguelen Island region on the northern plateau during the post-phytoplankton bloom period. The survey specifically targeted sampling at two stations in the HNLC waters to the west of the Kerguelen Plateau, and in the iron enriched waters on and to the east of the plateau. A combination of WP2, WP3 and Mesopelagos midwater trawl were used to sample the mesozooplankton (125 µm to ≤10 mm), macrozooplankton (10–30 mm) and micronekton (> 30 to 200 mm) communities. Stable carbon (δ13C) and nitrogen (δ15N) isotopes were measured across representative samples of taxa from all stations. Trophic positions was estimated using a Bayesian isotope mixing model (tRophicPosition), that uses both the δ13C and δ15N values of baselines and consumers. Meso/macrozooplankton trophic positions (TPs) were ~2.4 on the plateau and >0.6 TPs higher at the upstream HNLC stations. This provides empirical evidence for shorter food chains on the diatom dominated plateau and longer food chains in the HNLC region dominated by pico and nanophytoplankton. Meso/macrozooplankton TPs were also elevated downstream of the plateau, though to a lesser extent than in the upstream region, likely reflecting the effect of downstream iron transport, supporting elevated phytoplankton production in the downstream region. There was a consistent increase in trophic position between

meso/macrozooplankton and micronekton of ~ 0.6 at all stations. Food chain length was therefore determined by the composition of the trophic levels below micronekton. Similar trophic structure at the two upstream HNLC stations, on either side of the northern branch of the Polar Front, indicated that food chain length was robust to the differences in community composition, and supports the critical role of size structured trophic interactions in determining food web properties.

Highlights

► This study used stable isotopes to analyse the pelagic food web structure in the Kerguelen Island region, comparing HNLC areas upstream of the plateau and the iron enriched area on the plateau that experiences a large seasonal phytoplankton bloom. ► Trophic position was calculated for mesozooplankton, macrozooplankton, and micronekton using a Bayesian isotope mixing model approach. ► Meso and macrozooplankton trophic positions were similar but ~ 0.6 trophic positions lower than micronekton within areas. ► Median meso/macrozooplankton trophic position was ~ 2.4 on the plateau and 0.6 trophic positions higher in HNLC areas. ► Food chains were longer in HNLC regions, and food chain length was determined by the composition of the trophic levels below micronekton. ► Size based food web structure was robust to differences in community taxonomic composition.

Keywords : zooplankton, micronekton, food web, size structure, stable isotopes, Southern Ocean, Kerguelen Plateau.

Introduction

The Southern Ocean is broadly characterized as a High Nutrient Low Chlorophyll (HNLC) region where productivity is limited by the micronutrient iron (de Baar et al. 1995). Regions of naturally occurring iron enrichment occur in the vicinity of shallow bathymetry (e.g., continental shelf environments and island shelves) or in association with sea ice melt. Such regions are associated with enhanced phytoplankton productivity and can support a high biomass of consumers, for example, South Georgia (Korb et al. 2008) and Kerguelen Islands (Blain et al. 2007). While the physiological mechanisms behind the importance of iron to increased primary productivity are well understood (Olson et al. 2000), and the link to increased food web biomass established (Hindell et al. 2011), few studies have examined the

effect of iron enrichment on the structure and function of Southern Ocean food webs

(Stowasser et al. 2012, Tarling et al. 2012).

Pelagic food webs begin with phytoplankton, and here iron availability can play a key role in setting the food web base, impacting the species composition of phytoplankton communities (Hoffmann et al. 2006), their biochemical composition (Hoffmann et al. 2007), and their size structure (Sunda & Huntsman 1997). Size is a particularly important parameter in food web interactions. Synthetic analyses of predator-prey size relationships across terrestrial to marine ecosystems has demonstrated that in > 90% of feeding linkages predators are larger than their prey (Cohen et al. 1993, Barnes et al. 2010). Indeed, in marine ecosystems, body size can be a better predictor of trophic position than an organism's taxonomy (Sheldon et al. 1972, Jennings et al. 2001, Andersen et al. 2016). In the Southern Ocean, phytoplankton in HNLC regions are dominated by cells < 20 μm in size (nano and picophytoplankton) and their annual production can exceed that of larger (> 20 μm) microphytoplankton by a factor of 2.7 (Uitz et al. 2010). By contrast, in regions of natural iron enrichment microphytoplankton dominate and can form extensive and persistent blooms (Blain et al. 2007). Given the fundamental role of size in determining food web interactions, epitomized by the interaction of size and the grazing dynamics of zooplankton functional groups (Fortier et al. 1994), iron driven shifts in phytoplankton size composition may have a significant bearing on Southern Ocean pelagic food web properties.

Based on size structured feeding dynamics, as outlined above, the smaller phytoplankton size classes found in HNLC regions of the Southern Ocean are expected to favour longer food chains than iron enriched areas where large phytoplankton size classes dominate the summer high productivity period. Assuming consistent transfer efficiency between trophic levels, short food chains in iron enriched areas are therefore expected to transfer biomass more efficiently to higher trophic levels than longer food chains in HNLC regions. Food chain length is also influenced by the ratio of predator to prey size (Cousins 1987), with smaller predator-prey mass ratios (PPMR) yielding longer food chains, and vice versa. Furthermore, smaller PPMRs and longer food chains have been found to be associated with more stable environments (Jennings & Warr 2003) and this has potential implications for Southern Ocean food webs. The seasonal amplitude of primary production in HNLC vs naturally iron fertilized regions can vary by an order of magnitude (Blain et al. 2007). We can therefore hypothesise that HNLC regions, with more stable production cycles, would have smaller PPMRs than iron enriched areas, further contributing to elongation of the food chain in these regions.

Stable isotopes provide a valuable tool for empirical analysis of food web properties, including basal resources, organic matter transfer, and food chain length. Carbon and nitrogen are the two elements most commonly measured in food web isotope analyses. Carbon isotopes ($\delta^{13}\text{C}$) are typically enriched by $\sim 0.5\text{-}1\text{ ‰}$ and nitrogen isotopes ($\delta^{15}\text{N}$) by $\sim 3.4\text{ ‰}$ per trophic level in any given food web. The signature of a food web's basal resources (primary producers) are therefore more strongly conserved in consumer $\delta^{13}\text{C}$ values, and carbon isotopes have most frequently been used to trace and identify these food web sources (Minagawa & Wada 1984,

Vander Zanden & Rasmussen 2001, Post 2002, McCutchan et al. 2003). Conversely, nitrogen isotopes are most frequently used to estimate the trophic position of food web components, which can be further extrapolated to food chain length. The calculation of trophic position and food chain length from isotope values is complicated by variability / uncertainty in key values, including trophic enrichment factors, baselines, and consumers, and the associated error propagation through estimates. However, the development of Bayesian approaches have overcome some of these limitations, while also taking into account both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ in trophic position estimates (Phillips et al. 2014, Quezada-Ron egia Ili et al. 2018).

The Kerguelen plateau represents an ideal region to investigate the role of natural iron fertilization in structuring pelagic food webs in the Southern Ocean. The plateau extends southeast of the Kerguelen Islands, and has a shallow bathymetry (< 700 m) that represents a major obstacle to the Antarctic Circumpolar Current (ACC) (Park et al. 2008). Circulation interactions with the shallow topography transport iron to the surface waters of the plateau where a bloom routinely forms between November and February, with an areal extent of up to 45,000 km² (Blain et al. 2007, Mongin et al. 2008). From February to March 2018, the “Marine Ecosystem Biodiversity and Dynamics of Carbon around Kerguelen” program, hereafter referred to as MOBYDICK, set out to research contrasting productivity regimes in the Kerguelen region during the post-bloom period. The MOBYDICK expedition builds upon three previous programs in the area: the Kerguelen Ocean and Plateau compared Study (KEOPS) in summer 2005, KEOPS2 in spring 2011, and the “Myctophid assessment in relation to Oceanographic conditions: a three Dimension Density Distribution approach combining Modelling, Acoustic-and Predators” data

(MyctO-3D-MAP) in summer 2014. The MOBYDICK station positions were selected to coincide with areas sampled during these previous surveys and included one station on the plateau (iron enriched), two stations to the west of the plateau (iron limited), and one station to the east of the plateau (iron enriched).

According to size structured feeding dynamics and stability driven decreases in PPMR, we predict that iron rich regions, dominated by large microphytoplankton and large amplitude blooms, would have shorter food chains than iron limited region dominated by smaller phytoplankton size classes and lower amplitude blooms. To test this prediction, we used 1) stable isotope data collected during the MOBYDICK expedition to estimate the trophic position of the mesozooplankton, macrozooplankton and micronekton components of the pelagic food web; 2) and specifically compared trophic position estimates of these food web components between iron enriched and iron limited regions in the vicinity of the Kerguelen Plateau. These measurements are expected to provide new insights into the role of iron supply in pelagic food web structure and function in the Southern Ocean, with implications for materials flux.

Methods

Sampling during the MOBYDICK expedition was completed aboard the *Marion Dufresne II* from 26 February to 19 March 2018. The location of the four stations sampled is indicated in Figure 1. Station M2 was located on the plateau, and was the “bloom” reference station A3 in KEOPS and KEOPS2. Station M3, located to the west of the plateau, corresponded to the KERFIX

station of KEOPS, and M4, also west of the plateau, was located roughly 100 nautical miles south of the HNLC reference stations of KEOPS2 (Blain et al., 2015). Repeat sampling was conducted at stations M2, M3, and M4 (Table 1). Station M1 was located east of Kerguelen, in the core foraging area of the Kerguelen king penguin population (Scheffer et al., 2016), and was only sampled once.

Sample collection

Particulate organic matter (POM) was sampled in conjunction with each WP2+3 zooplankton net deployment, collected from the ship-board seawater line from a depth of ~ 10 m. Approximately 2 L of water was filtered onto a 25 mm pre-combusted GF/F filter by vacuum filtration. Mesozooplankton were sampled with a WP2 net (2.5 m long, 57 cm diameter) fitted with 200 μm mesh and deployed to a depth of 200 m. Macrozooplankton were collected with a WP3 net (2 m long, 1.13 m diameter) fitted with 1000 μm mesh. The WP2 and WP3 were deployed once during the night at station M1, and once each during the day and night on each visit to stations M2-M4. Three daytime and three nighttime trawls were conducted on each visit to all stations using a Mesopelagos trawl designed by Ifremer (Fisheries Biology and Technology Laboratory, LTBH, Lorient, France) (Meillat 2012). This non-closing trawl has vertical and horizontal openings which vary between 5 and 6 m and 10 and 12 m, respectively. The trawl has a mesh size of 40 mm in the wings, reducing to 5 mm in the codend. A Scanmar acoustic device (Åsgårdstrand, Norway) was attached to the net for real time monitoring of trawl depth simultaneously with acoustic measurements (Williams & Koslow 1997). Sampling depths were

dependent on acoustic backscatter, with trawls targeting areas of high backscattering signals in shallow, middle and deep layers (Table 1). During trawling, the vessel maintained a speed of 1.5 m s⁻¹ while fishing the target depth for ~30 minutes. The trawl was deployed and retrieved as fast as possible to ensure that organisms were mainly caught at the targeted depth.

Sample processing

The WP2 net samples were size fractionated into 6 size classes with a sieve column: 125-250, 250-500, 500-1000, 1000-2000, 2000-4000, and > 4000 µm. The size fractions < 4000 µm were filtered onto 47 mm pre-combusted and pre-weighed GF/F filters by vacuum filtration and then oven dried at 50°C for 48 hours onboard the vessel. The > 4000 µm size fraction was separated into species, and specimens measured, grouped into logarithmic size bins, and dried at 50°C for 48 hours onboard the vessel. Macrozooplankton and micronekton were sampled from the midwater trawls with the aim to have representation of major taxa and size classes from all trawls. All individuals were measured to the nearest 1 mm. In the case of smaller animals (< 50 mm), the entire animal was collected. In the case of larger animals, a muscle tissue sample (~ 20 mg) was collected. All samples were oven dried for 48 hours at 50°C onboard the vessel. Once returned to the laboratory all samples were weighed to the nearest 0.01 mg.

Isotope analysis

Samples were not treated to remove lipids or inorganic carbon prior to stable isotope analysis. Stable carbon and nitrogen elemental and isotopic compositions for all organisms were measured at the University of Victoria Isotope Facility using a Costech 4010 elemental analyzer (Costech, Florence) coupled via continuous flow to a Thermo Finnegan Delta Advantage isotope ratio mass spectrometer (Thermo-Finnigan, Bremen, Germany). Stable isotope values were expressed in standard δ (‰) notation and a two-point calibration anchored with internal reference materials ('Caffeine' and 'Dorm', themselves determined using IAEA N1, N2, and CH-6 as well as NBS-22; ESM Table S1) was used to calibrate $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ relative to Pee Dee Belemnite for carbon and Air for nitrogen. Standard deviations for calibration standards for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, respectively, were ± 0.3 ‰ and ± 0.2 ‰ for Caffeine ($n=3$) and ± 0.3 ‰ and ± 0.2 ‰ for Dorm ($n=5$). Carbon isotope values were lipid corrected according to individual organism C:N ratios, using the equation from (Hoffman & Sutton 2010) for fish and from (Smyntek et al. 2007) for invertebrates.

Trophic position calculation

When applying bulk isotopes to estimate food web properties an isotope baseline is required as an anchor point. The isotopic value of POM has frequently been used as representative of phytoplankton. However, this value can have high temporal variability, showing rapid response (days) to fluctuations in nutrients and phytoplankton growth rates (Lorrain et al. 2015), while also being influenced by non-phytoplankton particulates, including faecal pellets (Checkley & Entzeroth 1985) and microzooplankton. An alternative approach to establishing a trophic

baseline is to use the isotopic value of a primary consumer. Salps (tunicates) have been used in a number of studies (Post 2002, Cherel et al. 2010, Stowasser et al. 2012). However, recent analysis has identified salps as unreliable baseline measures due to their unique feeding biology (Pakhomov et al. 2019). For this study we have used the isotope values of the zooplankton size classes $\leq 1000 \mu\text{m}$ as the trophic baseline. Since no zooplankton can be considered to be truly herbivorous, we assigned a trophic level of 2.25 to this group to take into account a degree of omnivory.

The species sampled, and the number of animals sampled per species, varied between stations. To optimize the comparability of sample sets among stations for trophic position calculation, the data set was filtered to include only taxa that were common to all stations (see Figure 4). *Salpa thompsoni* were not included in the trophic position analysis due to uneven sample size, and their unique trophic ecology, and they are rather discussed separately. Common taxa, excluding *S. thompsoni*, were grouped into the length-based groups of mesozooplankton (125 μm to ≤ 10 mm), macrozooplankton (10–30 mm), micronekton (> 30 to 200 mm) (see Table s1 for sample sizes). Trophic positions were calculated using a one-source, two-isotope (carbon and nitrogen) Bayesian isotope mixing model implemented with the tRophicPosition package (version 0.7.7, Quezada-Romegialli et al., 2018) in R (R Core Team 2020). The benefits of this approach are that it explicitly includes individual variability and propagation of sampling error (trophic enrichment factors, and measurements of baselines and consumers) in the modelling approach and posterior estimates of parameters. We used Post's (2002) trophic enrichment factor values of 3.4 ± 0.98 (mean \pm standard deviations) for $\delta^{15}\text{N}$ and 0.39 ± 1.3 for $\delta^{13}\text{C}$. The Bayesian model

ran 20,000 iterations for the adaptive phase, 20,000 iterations as burnin (iterations discarded at the beginning of posterior sampling) and 20,000 actual iterations. The model used five parallel Markov Chain Monte Carlo (MCMC) simulations using the JAGS (ver. 4.3.0) Gibbs sampler (Plummer 2003). The median posterior trophic positions are presented showing the 95% credibility intervals. We then conducted pairwise comparisons of the posterior distributions among classes with a logical test that one was greater ($>$) than the other, randomly sampling posterior distributions until all posterior estimates were compared. The probability that one class had a higher trophic level than the other increased as the value approached 1. Finally, the Bhattacharyya coefficient was used to calculate the probability of overlap between two distributions, with the probability increasing towards 1 (Quezada-Romegialli et al. 2018).

Results

Water mass tracking, described in detail in Henschke et al. (2021), estimated retention times of ≥ 60 days at all sites (Table 2). This suggests that the same water masses were sampled during each repeat visit to stations M2, M3 and M4. Mixed layer depth was lowest at M1 (27m) and ranged between 50 and 90 m on repeat visits to the other stations. Mean MLD phytoplankton biomass was highest at station M2-3 ($0.58 \mu\text{g L}^{-1}$), but was generally $< 0.3 \mu\text{g L}^{-1}$. A large phytoplankton bloom was observed at Stations M1 and M2 in December – January, two to three months prior to the MOBYDICK expedition, where phytoplankton biomass exceeded $2 \mu\text{g L}^{-1}$ (Figure s1). Elevated phytoplankton biomass was also observed at the upstream stations M3 and M4 at this time, though levels were substantially lower ($\sim 0.5 \mu\text{g L}^{-1}$). Mixed layer water

temperature was highest at station M3. The absence of a temperature minimum at 200 m at M3 confirmed that it was conducted north of the APF, whereas M1, M2 and M4 stations were located south of the APF in the Antarctic Zone (Henschke et al. 2021). Zooplankton biomass was lowest at stations M1 and M3-1 ($\sim 5 \text{ mgC m}^{-3}$) and averaged 10.84 mgC m^{-3} across all other stations with a maximum of 13.65 mgC m^{-3} at Station M4-1. Highest values of crustacean biomass were observed at M2, while the highest fish biomass was observed at M3.

Trophic positions were calculated using station specific $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of zooplankton size classes $\leq 1000 \mu\text{m}$ as the trophic baseline. Comparison with other baseline proxies showed that zooplankton had lower within site variability than either POM or *Salpa thompsoni*, and that $\delta^{15}\text{N}$ values had the least variability within stations (Figure 2). Carbon isotope values (Figure 2A) were highest at M2 (average = -23.4 ‰) and lowest at M3 (ave. = -25.3 ‰) and M4 (ave. = -26.2 ‰). A decreasing trend over time in zooplankton $\delta^{13}\text{C}$ values was evident at M2 and M3. The $\delta^{13}\text{C}$ values of *S. thompsoni* generally followed the same pattern as POM. Nitrogen isotope values of zooplankton (Figure 2B) were highest at M2 (ave. = 2.3 ‰), followed by M1 (ave. = 0.9 ‰), and were lowest at M3 (ave. = -0.1 ‰) and M4 (ave. = 0.1 ‰). The $\delta^{15}\text{N}$ values of zooplankton decreased slightly over time at M2. A much stronger decreasing trend was apparent for POM and *S. thompsoni* at M2, likely reflecting the higher tissue turnover rates of these groups relative to zooplankton.

The mesozooplankton size class largely comprised zooplankton from the WP2 net size fractions $\leq 2000 \mu\text{m}$ and copepods (Figure 3A). Both the macrozooplankton and micronekton included a

mix of gelatinous and non-gelatinous taxa. Macrozooplankton included pteropods (gymnosomes and thecosomes), siphonophores, hydrozoans, gammarids, mysids, and euphausiids. Micronekton included squid, decapods, scyphozoans, tunicates, fish, chaetognaths and ctenophores. Nitrogen isotopes values typically increase with trophic level which is expected to correlate with organism size. This was largely the case in this study, and fish, which included the largest individuals, had the highest median $\delta^{15}\text{N}$ values (Figure 3B). However, the large gelatinous taxa of tunicates, chaetognaths and ctenophores all had low $\delta^{15}\text{N}$ values relative to their size. This was at least expected for the small particle grazing tunicates, which in this case comprised entirely of *Salpa thompsoni* (Figure 2). The widest range of $\delta^{15}\text{N}$ values was observed for copepods, indicating diverse foraging strategies in this group.

Mean $\delta^{15}\text{N}$ values are presented for the 20 taxa used in the posterior trophic position analysis, as well as the tunicate *S. thompsoni* (Figure 4). Overall, $\delta^{15}\text{N}$ values increased with organism size for all taxa. *Salpa thompsoni* consistently had the lowest $\delta^{15}\text{N}$ values, and interestingly the hyperiid amphipods *Cylopus magellanicus* and *Vibilia antarctica* both had low $\delta^{15}\text{N}$ values, suggesting predation on *S. thompsoni*. The myctophid *Electrona antarctica* had the highest $\delta^{15}\text{N}$ values, indicating that this species had the highest trophic level. Values of $\delta^{13}\text{C}$ also generally increased with size, although this trend was not as apparent as for $\delta^{15}\text{N}$.

The three organism size classes of mesozooplankton, macrozooplankton, and micronekton showed a consistent pattern of posterior trophic positions (PTP). Mesozooplankton and macrozooplankton were always similar with the greatest difference in trophic position between

these classes being 0.16 at station M4 (Figure 5; Table s2). The median PTP of meso / macrozooplankton was 0.74 PTPs higher at M3 than M2, 0.62 PTPs higher at M4 than M2, and 0.41 PTPs higher at M1 than M2. Micronekton were on average 0.57 trophic positions higher than meso / macrozooplankton and micronekton, with the greatest difference being 0.81 trophic positions at station M4. PTPs were lowest at M2, with an average of 2.4 for meso and macrozooplankton. Pairwise comparison of PTPs found high probability (0.9-1) that PTPs were greater at stations M1, M3 and M4 than M2 (Table 3), and a high probability that PTPs were greater at M3 and M4 than M1. The probability of overlap of PTP distributions was low between M2 and stations M3 and M4, being < 0.3 except for M4 mesozooplankton vs M2 meso / macrozooplankton (~ 0.4 ; Table 4). This provided further support for the distinct, higher, PTPs in the upstream vs plateau stations. The probability of overlap between station M1 and all other stations was generally > 0.55 , demonstrating that the downstream region was intermediate in PTP characteristics between the plateau and upstream region.

Discussion

The Kerguelen Islands and plateau are a region of natural iron fertilization within the typically iron limited High Nutrient Low Chlorophyll (HNLC) waters of the eastward flowing Antarctic Circumpolar Current. The MOBYDICK expedition provided an opportunity to investigate the effect of iron enhanced productivity on the structure and function of the lower trophic levels of the Southern Ocean food web, at the end of the summer phytoplankton bloom. This study sampled the mesozooplankton, macrozooplankton, and micronekton food web in the HNLC

waters to the west of the Kerguelen Plateau, in the iron enriched waters on the plateau, and the waters east of the plateau which are iron enriched by off-plateau advection (d'Ovidio et al. 2015). Using bulk carbon and nitrogen stable isotope we demonstrated that the trophic positions of meso and macrozooplankton were > 0.6 trophic levels higher at the upstream HNLC stations than on the plateau, and were also elevated downstream of the plateau, though to a lesser extent. This supported the prediction that the iron rich plateau would have a shorter food chain than the iron limited upstream region, and that the downstream region had enhanced production due to iron enrichment from the plateau. Below we discuss our findings in the context of the Kerguelen region pelagic ecosystem dynamics, and the implications for energy flow in the contrasting productivity regimes.

The MOBYDICK expedition took place in the post-bloom period of the seasonal production cycle and phytoplankton biomass was low in all sampled areas ($< 0.6 \mu\text{gL}^{-1}$), but lowest upstream of the plateau ($< 0.22 \mu\text{gL}^{-1}$). However, satellite observations showed that in December / January prior to the expedition phytoplankton biomass reached a peak of $> 2 \mu\text{gL}^{-1}$ over the plateau region where station M2 was located, and downstream (east) of the plateau, highlighting the effect of natural iron fertilization. The seasonal bloom cycle and spatial extent observed in 2018 were consistent with observations in previous years, indicating that this is a recurring feature of the region (Blain et al. 2007, Laurenceau-Cornec et al. 2015). The HNLC region upstream of the Kerguelen Plateau is broadly representative of the Southern Ocean and is dominated by small phytoplankton, with picophytoplankton comprising up to 50% and nanophytoplankton $> 20\%$ (mostly *Phaeocystis* and small diatoms, e.g., *Fragilariopsis*) of the total biomass (Fiala et al.

1998, Lasbleiz et al. 2016). Conversely, blooms over the plateau are dominated by large diatoms and smaller chain forming diatoms such as *Pseudonitzschia* and *Chaetoceros* (Lasbleiz et al. 2016). In the post-bloom period sampled during MOBYDICK, Prymnesiophytes, dominated by *Phaeocystis antarctica* (3 μm cell size), were the most abundant phytoplankton group, representing up to 53% and 70% of the Chl-a on and off the plateau, respectively (Irion et al. 2020). Small diatoms, e.g., *Fragilariopsis* and *Chaetoceros*, were abundant at stations M4 and M1, while M3 had the lowest contribution of diatoms (25% of Chl-a), likely due to silicate and iron co-limitation. On the plateau, large diatoms, e.g., *Corethron* dominated in low silicate post-bloom conditions in the mixed layer. Below the mixed layer, large and heavily silicified diatoms were abundant (e.g., *Eucampia* and *Odontella*).

Preliminary analysis of the zooplankton community composition during MOBYDICK found greatest similarity between stations M1 and M4, and that both of these stations overlapped in composition with M2 (Figure s4 and s5), i.e., there was not a distinct separation of the plateau zooplankton community from that upstream (M4) and downstream (M1). Station M3 showed the greatest dissimilarity to other stations, reflecting differences in the community composition north and south of the northern branch of the Polar Front (Figure 1). Copepods, particularly *Oithona* and Calanoids, dominated abundance at all stations (Figure s2 and s3), in agreement with the KEOPS surveys where copepods contributed > 80 % to total mesozooplankton abundance (Carlotti et al. 2008, Carlotti et al. 2015). Despite similarities in zooplankton community composition to M1 and M4, plateau station M2 did record the highest crustacean and gelatinous zooplankton biomass, indicating a positive response to higher phytoplankton

productivity in this region. However, there was substantial variation in zooplankton biomass among stations during MOBYDICK. While areal mesozooplankton biomass during MOBYDICK was an order of magnitude higher ($1\text{--}2.7\text{ gC m}^{-2}$) than recorded in November ($0.25\text{--}0.49\text{ gCm}^{-2}$) during the KEOPS2 survey (Carlotti et al. 2015), it was up to an order of magnitude lower than during the January / February KEOPS survey ($3.44\text{--}19.26\text{ gCm}^{-2}$) (Carlotti et al. 2008). Although the February / March sampling dates of the MOBYDICK expedition coincided with the period of peak zooplankton biomass observed during the YugNIRO times-series in 1987 / 88 (Hunt et al. 2011), seasonal Continuous Plankton Recorder data collected across the Sub-Antarctic and Polar Frontal zones in 2001 / 2002 found a peak of zooplankton abundance in February (Hunt & Hosie 2006). The seasonal timing of the zooplankton peak biomass likely varies interannually, however, based on the December / January phytoplankton bloom timing in 2018 (Figure s1), zooplankton biomass observed during MOBYDICK was probably declining after the summer peak (Semelkina 1993, Razouls et al. 2006, Hunt et al. 2011).

The combined sampling method, conducted during MOBYDICK provided an opportunity to collect stable isotope data for the mesozooplankton, macrozooplankton and micronekton components of HNLC and naturally iron fertilized water masses in the Kerguelen plateau region. An important consideration when evaluating these data was whether the stable isotope ratios measured were representative of the food web history within the sampled water mass. This would depend on the water mass retention time in the areas sampled and the tissue turnover rates of the organisms. Water mass tracking estimated water mass residence times of at least 60 days in each of the areas sampled by the four stations (Henschke et al. 2021). Organism

tissue turnover rates scale with organism size and growth rate (Fry & Arnold 1982, Hesslein et al. 1993). The highest turnover rates, and most rapid response to changing food web conditions (e.g., nitrate supply), are therefore expected to occur in particulate organic matter (POM).

Laboratory studies indicate that diatom isotope replacement rates can be on the order of days (Montoya & McCarthy 1995). Few mesocosm studies have been conducted for Southern Ocean zooplankton, with one study reporting tissue turnover rates for *Euphausia superba* of 54% for nitrogen after 30 days (Schmidt et al. 2003). However, it is expected that warmer water Sub-Antarctic and Antarctic Zone zooplankton during the growing season would have higher turnover rates. Fry and Arnold (1982) reported turnover rates for shrimp and brine shrimp of 4-19 days. In our study, there was evidence for a temporal decline in stable isotope values of POM, *S. thompsoni* and < 1000 μm zooplankton over the three visits to M2, which spanned three weeks. In the case of micronekton, turnover rates are expected to be weeks to months (Hesslein et al. 1993, Colborne et al. 2017). Given the estimated water mass residence times for the study area, it is reasonable to expect that the measured stable isotope ratios of most taxa were representative of the food web history within the water mass in which they were sampled, including the pre-voyage bloom conditions.

The elevated on-plateau $\delta^{15}\text{N}$ values of the stable isotope baseline used in this study, zooplankton < 1000 μm , reflected the higher productivity in this region. Phytoplankton $\delta^{15}\text{N}$ values increase in response to nitrate competition during high productivity periods (Altabet & Francois 2001), and these elevated $\delta^{15}\text{N}$ values were transmitted to the zooplankton on the plateau. Using a Bayesian modelling approach, that applied both carbon and nitrogen isotopes

values of baselines and consumers, we estimated that the trophic positions of meso and macrozooplankton were > 0.6 higher at the upstream HNLC stations than on the plateau. This result provided empirical support for the prediction that iron rich regions, dominated by large microphytoplankton and large amplitude blooms, would have shorter food chains than iron limited regions dominated by smaller phytoplankton and lower amplitude blooms. The dominance of pico and nanophytoplankton in the HNLC upstream region favours a prominent role of heterotrophic nanoflagellates and ciliates as trophic intermediaries between phytoplankton and zooplankton (Sommer et al. 2002), and a longer food chain. The lower amplitude seasonal phytoplankton bloom, and hence more stable production cycle in HNLC regions may also contribute to elongation of the food chain through favouring smaller predator-prey mass ratios (Jennings & Warner 2005). Conversely, the iron enriched plateau favours diatom blooms and direct consumption of phytoplankton by zooplankton. Indeed, a higher degree of herbivory was observed for meso and macrozooplankton on the plateau than off the plateau. The shorter plateau food chain promotes more efficient biomass transfer to higher trophic levels, and augmented by higher primary productivity is expected to yield enhanced on-plateau biomass accumulation. The region downstream of the plateau had meso and macrozooplankton trophic positions intermediate between the upstream and plateau region, reflecting the effect of downstream iron transport which fuels the elevated phytoplankton production in the downstream region (Blain et al. 2007).

The trophic position analysis identified two additional important features of the trophic structure in the study area. Firstly, the meso and macrozooplankton components of the food

chain had a high degree of the trophic position overlap that was independent of whether the community was in an HNLC or productive plateau region. The mean trophic position of meso and macrozooplankton on the plateau was 2.4, indicating a similar contribution of herbivory and carnivory in these size classes. The presence of large macrozooplankton grazers, such as euphausiids, may be a key factor in the effective grazing and transfer of large diatom biomass to higher trophic levels when blooms occur. There is evidence that diatoms are selected against by some zooplankton grazers, while their silicified cell walls limit the number of grazers that are capable of consuming and assimilating them (Verity & Smetacek 1996, Liu et al. 2016). The higher trophic position (~ 3) of both meso and macrozooplankton in the off-plateau areas indicated that these taxa switched to increased carnivory when large diatoms were less abundant. Secondly, there was a consistent increase in trophic position between meso / macrozooplankton and micronekton of ~ 0.6 at all stations. Food chain length was therefore determined by the composition of the trophic levels below micronekton. The similarity of trophic structure at the two upstream stations, M3 and M4, indicated that food chain length was robust to the differences in community composition that occurred across the northern branch of the Polar Front. This supports the critical role of size structured trophic interactions in determining food web properties.

While size was clearly related to trophic level in our study, with the smallest organisms (mesozooplankton) having the lowest trophic level and the largest organisms (fish) the highest, wide isotopic ranges for some groups reflected diverse within group feeding ecologies. For example, copepods spanned approximately two trophic levels, including the carnivorous genus

Paraeuchaeta and herbivorous species such as *Rhincalanus gigas*. Wide isotopic ranges were also observed for other groups, including Hydrozoans, Decapods, Chaetognaths and Hyperiid. The trophic diversity of taxa within the lower levels of pelagic food webs is an important consideration when parameterizing food web models, where the lower trophic levels are frequently aggregated into a few groups with constrained functional roles. *Salpa thompsoni* was a major exception to the size rule, being one of the largest taxa but with $\delta^{15}\text{N}$ values lower than the mesozooplankton, indicating a lower trophic level. Size fractionated particulate organic matter (POM) analyses show that, as with the rest of the food web, POM stable isotope values scale with size, and that this is particularly evident in $\delta^{15}\text{N}$ values (Trull et al. 2015). *Salpa thompsoni*'s low $\delta^{15}\text{N}$ values in our study underscore their unique functional role as large bodied small particle grazers. Due to salps ability to effectively filter extremely small particles (< 1 μm) they can separate from the classical food web (Vargas & Madin 2004, Pakhomov et al. 2019). Notably, the hyperiid amphipod, *Cylopus magellanicus* and *Vibilia antarctica* both had low $\delta^{15}\text{N}$ values, particularly at station M1 where *S. thompsoni* was most abundant (Henschke et al. 2021). Both species are known commensals of *S. thompsoni* and therefore it is highly likely that these hyperiids preyed on *S. thompsoni*, providing an indirect pathway for *S. thompsoni* biomass into the pelagic food web.

Summary

Shifts in the size structure of phytoplankton at the food web base determine trophic pathways, and the number of trophic steps leading to mesozooplankton and macrozooplankton, and by

extension the efficiency of biomass transfer to higher trophic levels. We found that HNLC waters upstream of the Kerguelen plateau, dominated by pico and nanophytoplankton, had mesozooplankton / macrozooplankton communities that were > 0.6 trophic positions higher than on the naturally iron enriched plateau which supports large summer diatom blooms. Mesozooplankton / macrozooplankton in the region downstream of the plateau were intermediate in trophic position between the upstream and plateau regions, indicating the effect of iron enrichment though downstream transport from the plateau. Food chain length was determined by the composition of interactions between trophic levels below the micronekton, which were consistently ~ 0.6 trophic positions above meso / macrozooplankton at all stations. We suggest that the efficiency of biomass accumulation in iron enriched regions depends on the presence of large zooplankton grazers that are able to effectively consume and assimilate diatoms, e.g., euphausiids. We also note the high trophic diversity of lower trophic level taxa, which supports a more nuanced approach to structuring food web models, and more effectively represent the diverse functional roles of these taxa.

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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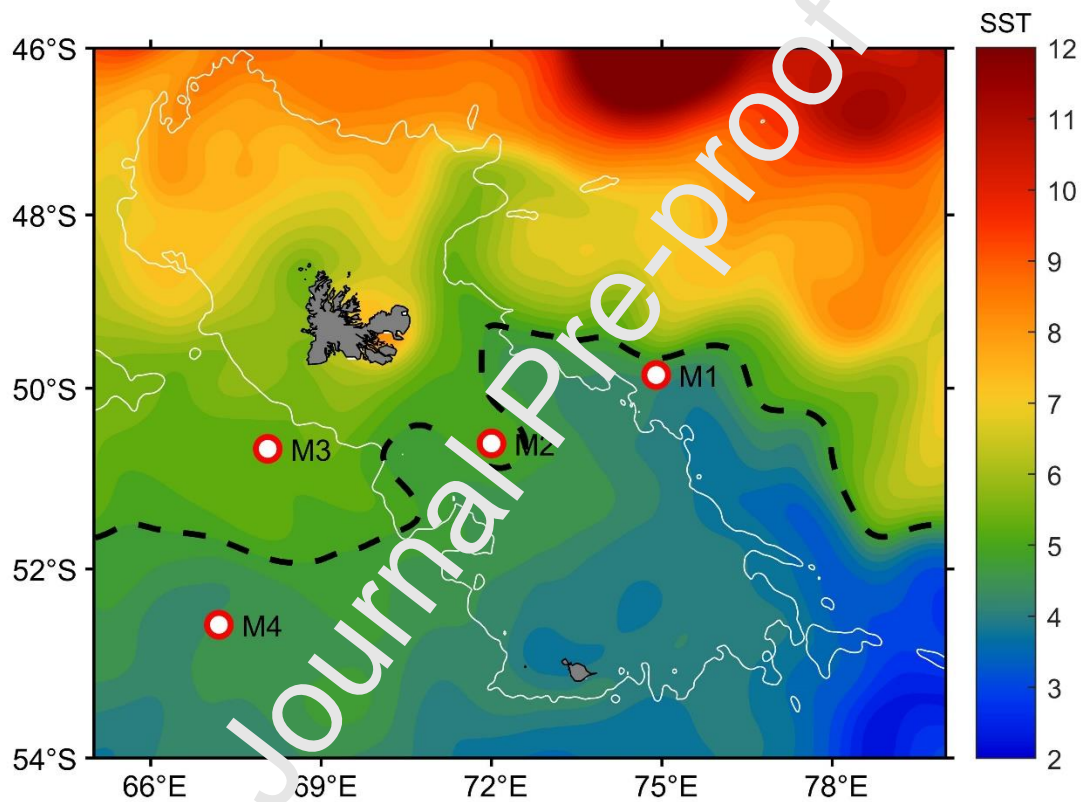


Figure 1. Map of stations sampled during the MOBYDICK expedition, from 26 February to 19 March 2018. The black dashed line indicates the position of the surface 5°C isotherm, the approximate position of the northern branch of the Polar Front (Cotte et al., this issue). SST data were retrieved using the Copernicus platform (<http://marine.copernicus.eu>). Data were produced by running the OSTIA system, a merged, multi-sensor L4 Foundation SST product,

with a spatial resolution of 0.05 degrees (Donlon et al. 2012). The continental slope (continuous white line, 1000 m isobath) and the Polar Front (dashed black line, 5°C SST) are represented.

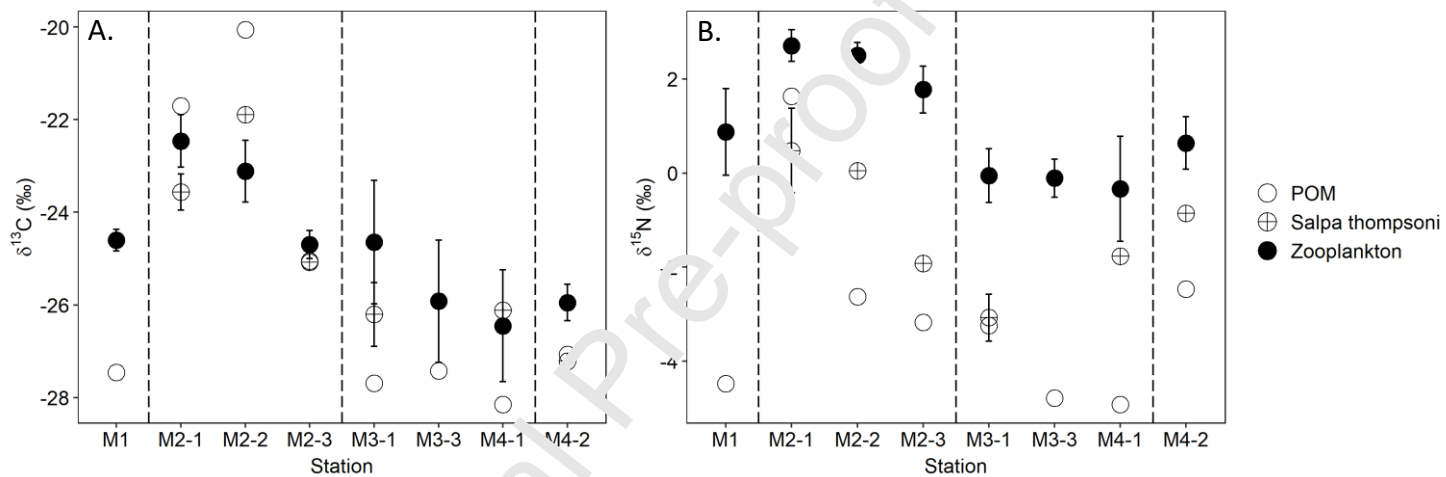


Figure 2. Values of $\delta^{13}\text{C}$ (A) and $\delta^{15}\text{N}$ (B) for trophic baseline indicators on each visit to MOBYDICK stations M1, M2, M3 and M4, including Particulate Organic Matter (POM), the tunicate *Salpa thompsoni*, and zooplankton size class $\leq 1000 \mu\text{m}$. Error bars are standard deviations.

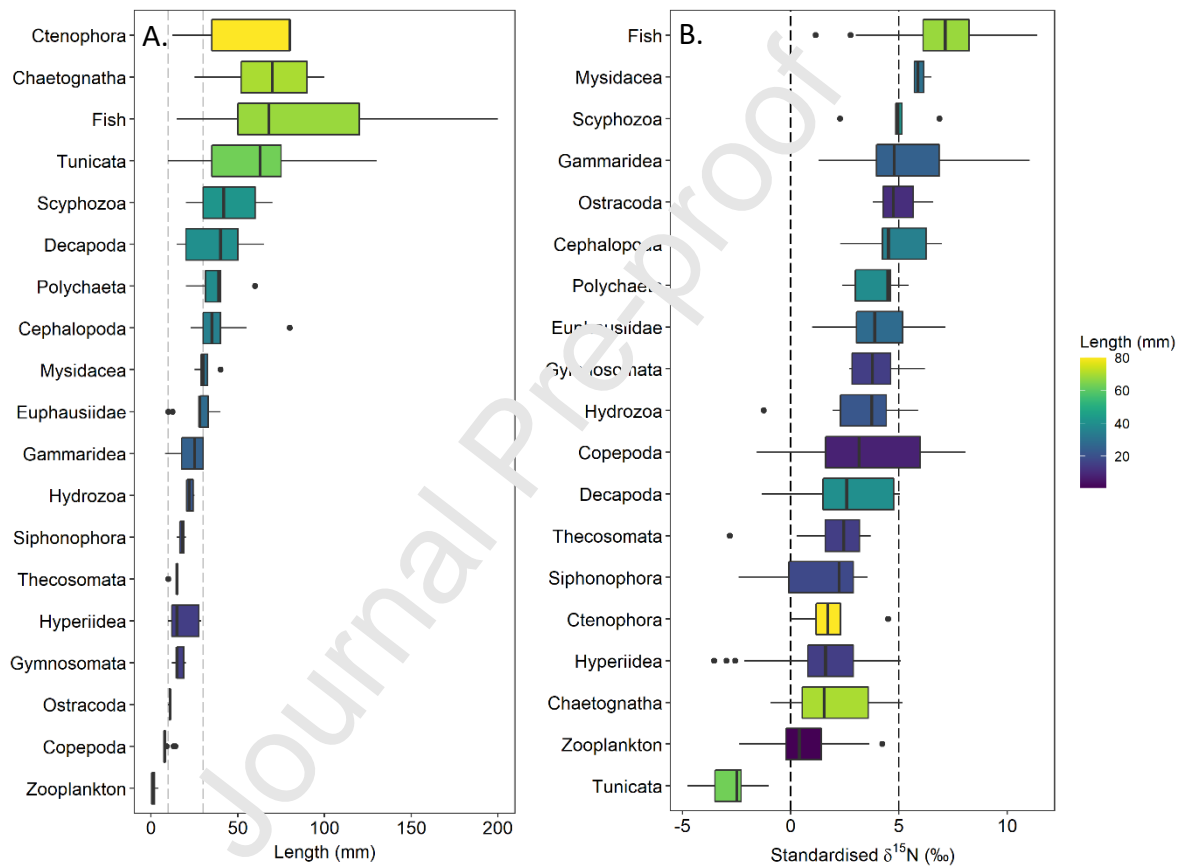


Figure 3. Length (mm) (A) and standardized $\delta^{15}\text{N}$ (B) of major taxonomic groups across all stations sampled (M1, M2, M3, M4) during MOBYDICK. The vertical lines in Figure 3A indicate the separation of pelagic organisms into mesozooplankton (125 μm to ≤ 10 mm), macrozooplankton (10 - 30 mm), and micronekton (> 30 to 200 mm). Standardized $\delta^{15}\text{N}$ was calculated by setting the baseline (mean $\delta^{15}\text{N}$ of < 1000 μm zooplankton size fractions) at each

station to 0 ‰, and adjusting all individual organism values within a station to this baseline.

Zooplankton in the figure refers to mesozooplankton size classes $\leq 2000 \mu\text{m}$. The largest zooplankton size fraction ($\geq 4000 \mu\text{m}$) was separated to species during processing, and sorted to major taxonomic groups in this figure. Tunicata were *Salpa thompsoni*. Vertical bars in the boxplots indicate median proportional biomass. The upper and lower edges of the box denote the approximate 1st and 3rd quartiles, respectively. The vertical error bars extend to the lowest and highest data value inside a range of 1.5 times the inter-quartile range, respectively. Points indicate extreme values.

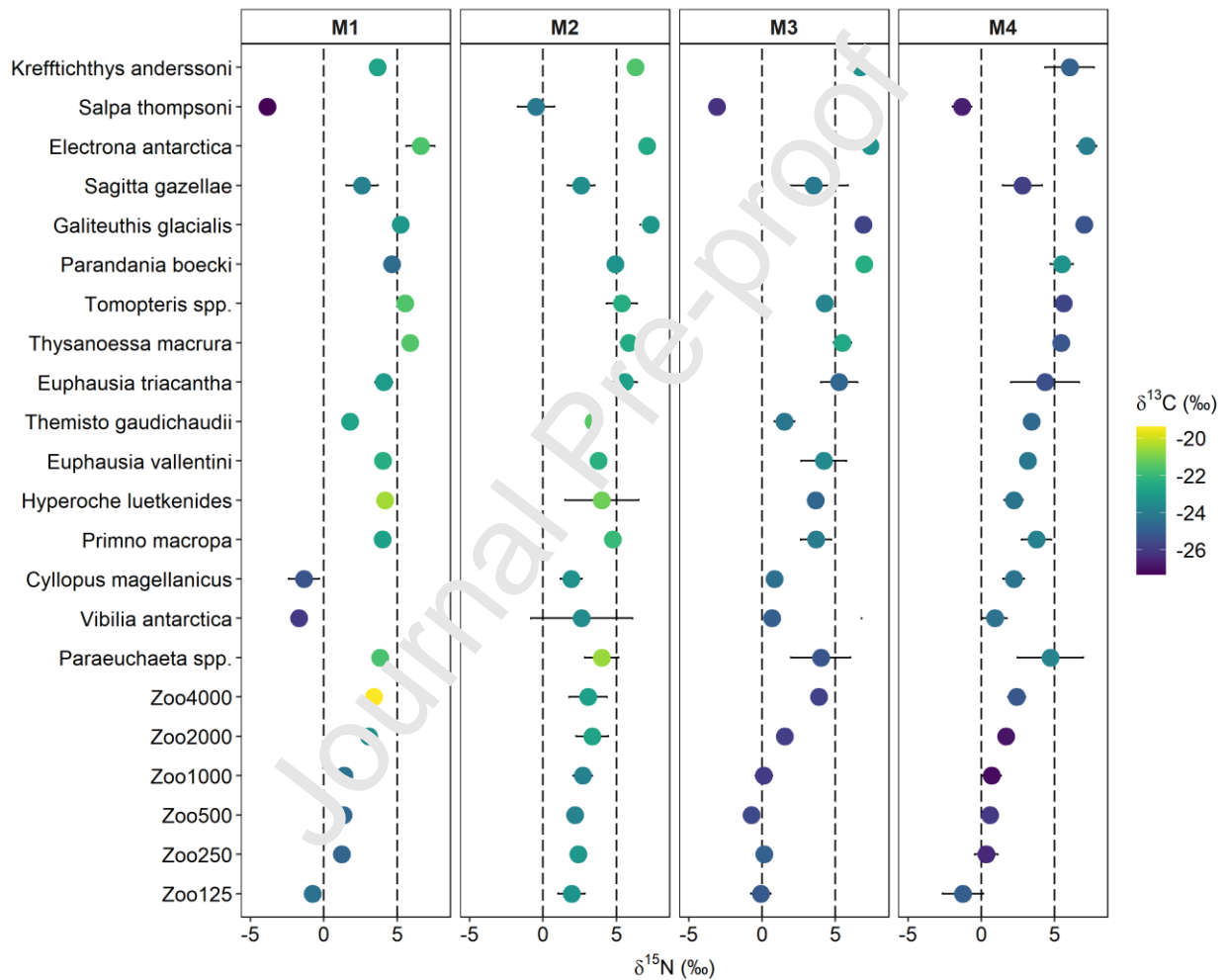


Figure 4. Mean $\delta^{15}\text{N}$ values (\pm standard deviations) at stations M1, M2, M3 and M4 for taxonomic groups that were common between all stations sampled, including *Salpa thompsoni*.

Taxa are arranged on the y-axis in increasing mean size. Points are colour scaled by $\delta^{13}\text{C}$ and the dashed lines indicates $\delta^{13}\text{C}$ values of 0 and 5 ‰.

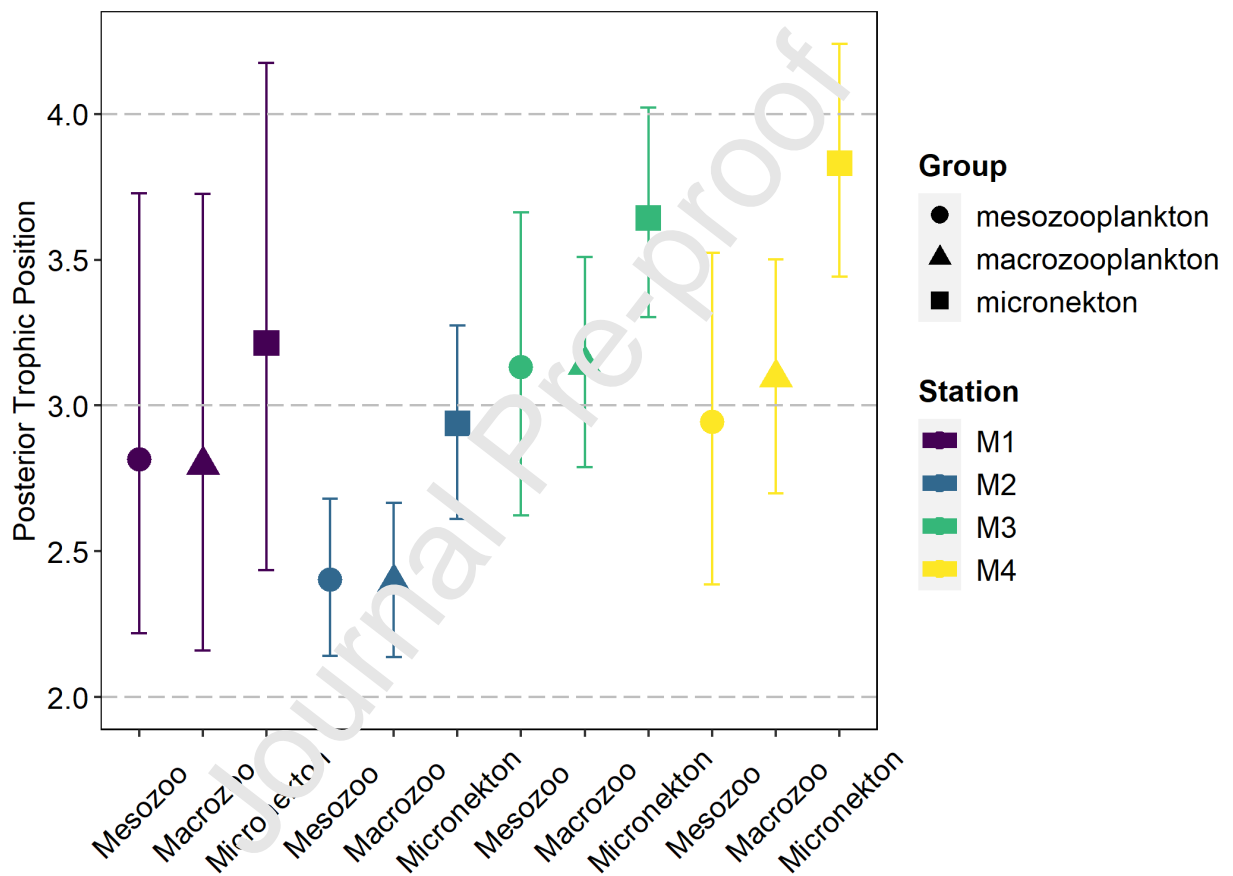


Figure 5. Posterior trophic positions (median \pm 95% credibility intervals) of mesozooplankton (125 μm to 10 mm), macrozooplankton (10 - 30 mm), micronekton (> 30 mm) sampled at stations M1-M4 during the MOBYDICK survey.

Table 1. Stations sampled, with dates, depths and gears deployed. ND – No Data

Station	Date	POM	WP2+3	n _{wp2+3}	Trawl	n _{trawl}	Trawl Depths
M1	8 Mar	1	Yes	1	Yes	5	50, 400, 632, 617, 50-632
M1	9 Mar	1	No	0	Yes	1	35, 180, 340, 350, 350-0
M2-1	26 Feb	1	Yes	1	Yes	4	50, 160, 170, 310
M2-2	7 Mar	1	Yes	1	Yes	5	70, 200, 350, 70-350
M2-3	16 Mar	1	Yes	1	Yes	3	360, 400, 460, 610
M3-1	15 Mar	1	Yes	1	Yes	4	650, 800, 810, 55-460
M3-2	18 Mar	1	Yes	1	Yes	4	ND
M3-3	19 Mar	1	Yes	1	No	0	75, 380, 560, 575
M4-1	2 Mar	1	Yes	1	Yes	4	80, 550, 600
M4-2	14 Mar	1	Yes	1	Yes	3	50, 400, 632, 617

Table 2. Mean conditions at each station occupation during the 2018 MOBYDICK expedition. Zooplankton biomass is an average of the night and day WP2 net sample (200 m depth) at each station. Zooplankton carbon content was derived from dry weight, using a mean proportional carbon content from all size fractions of 35 %, measured during isotope analysis. Crustacean, gelatinous and fish biomass are from targeted Mesopelagos trawls, fully described in Cotté et al (in review).

Station	Date	Summer bloom	Residence time (days) ^a	MLD (m) ^b	Chl _{MLDmean} ($\mu\text{g L}^{-1}$) ^b	T ^{°C} ^b	Zooplan/ton biomass (mgC m^{-3})	Crustacean biomass (mgC m^{-3}) ^c	Gelatinous biomass (mgC m^{-3}) ^c	Fish biomass (mgC m^{-3}) ^c
M1	08 Mar	Yes	~ 60	27	0.35	4.99	5.17	0.267	0.463	0.396
M2-1	26 Feb	Yes	> 60	62	0.27	5.11	12.15	0.252	0.755	0.066
M2-2	07 Mar	Yes	> 60	61	0.30	5.24	9.62	1.657	0.612	0.239
M2-3	16 Mar	Yes	> 60	68	0.58	4.99	12.07	0.143	0.229	0.029
M3-1	15 Mar	No	> 60	65	0.20	5.60	4.97	0.218	0.184	0.119
M3-2	18 Mar	No	> 60	79	0.14	5.31	10.43	0.280	0.012	0.014
M4-1	02 Mar	No	> 60	69	0.18	4.45	13.65	0.166	0.208	0.222
M4-2	14 Mar	No	> 60	81	0.21	4.46	7.12	0.021	0.305	0.002

a. Henschke et al. (2021); b. Irion et al (2020); c. Cotté et al. (in review - this issue) .

Table 3. Probability that a random sample of posterior distribution of trophic position is greater (>) in a row than in a column, and vice versa. Pairwise comparison are between mesozooplankton (125 μ m to \leq 10 mm; Meso), macrozooplankton (10 - 30 mm; Macr), and micronekton (> 30 to 200 mm; Mnek) at stations M1, M2, M3, and M4. Blue shading indicates increasing (highest = 1) and yellow shading decreasing (lowest = 0) likelihood of difference in comparisons.

M1- Meso	M1- Macr	M1- Mnek	M2- Meso	M2- Macr	M2- Mnek	M3- Meso	M3- Macr	M3- Mnek	M4- Meso	M4- Macr	M4- Mnek
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M1-Meso	0	0.51	0.21	0.91	0.91	0.35	0.22	0.18	0.03	0.38	0.22	0.02
M1-Macr	0.49	0	0.22	0.87	0.87	0.36	0.23	0.20	0.04	0.38	0.24	0.02
M1-Mnek	0.79	0.78	0	0.97	0.98	0.76	0.58	0.57	0.15	0.72	0.61	0.08
M2-Meso	0.09	0.13	0.03	0	0.52	0.01	0.01	0.00	0.00	0.04	0.00	0.00
M2-Macr	0.09	0.13	0.02	0.49	0	0.01	0.01	0.00	0.00	0.04	0.00	0.00
M2-Mnek	0.65	0.64	0.24	0.99	0.99	0	0.25	0.20	0.00	0.50	0.28	0.00
M3-Meso	0.79	0.77	0.42	0.99	0.99	0.75	0	0.49	0.05	0.70	0.55	0.02
M3-Macr	0.82	0.80	0.43	1.00	1.00	0.80	0.51	0	0.02	0.72	0.57	0.01
M3-Mnek	0.97	0.96	0.86	1.00	1.00	1.00	0.95	0.98	0	0.98	0.98	0.25
M4-Meso	0.62	0.62	0.28	0.96	0.96	0.50	0.30	0.28	0.02	0	0.33	0.01
M4-Macr	0.78	0.76	0.39	1.00	1.00	0.73	0.45	0.43	0.02	0.67	0	0.01
M4-Mnek	0.98	0.98	0.92	1.00	1.00	1.00	0.98	0.99	0.75	0.99	0.99	0

Table 4. The probability of overlap of posterior distributions of trophic position (Bhattacharya coefficient) in pairwise comparison between mesozooplankton (125 μm to ≤ 10 mm; Meso), macrozooplankton (10 - 30 mm; Macr), and micronekton (> 30 to 200 mm; Mnek) at stations M1, M2, M3, and M4. Blue shading indicate increasing (highest = 1) and yellow shading decreasing likelihood (lowest = 0) of overlap in comparisons.

	M1- Meso	M1- Macr	M1- Mnek	M2- Meso	M2- Macr	M2- Mnek	M3- Meso	M3- Macr	M3- Mnek	M4- Meso	M4- Macr	M4- Mnek
M1- Meso	0.00	0.99	0.84	0.57	0.55	0.88	0.83	0.73	0.29	0.96	0.81	0.23
M1- Macr	0.99	0.00	0.86	0.63	0.61	0.85	0.84	0.73	0.33	0.96	0.80	0.26
M1- Mnek	0.84	0.86	0.00	0.30	0.28	0.77	0.96	0.89	0.65	0.90	0.91	0.52
M2- Meso	0.57	0.63	0.30	0.00	1.00	0.24	0.23	0.09	0.00	0.43	0.17	0.00
M2- Macr	0.55	0.61	0.28	1.00	0.00	0.21	0.20	0.07	0.00	0.40	0.14	0.00
M2- Mnek	0.88	0.85	0.77	0.24	0.21	0.00	0.86	0.84	0.15	0.94	0.90	0.07
M3- Meso	0.83	0.84	0.96	0.23	0.20	0.86	0.00	0.97	0.50	0.93	0.98	0.34
M3- Macr	0.73	0.73	0.89	0.09	0.07	0.84	0.97	0.00	0.38	0.87	0.99	0.21
M3- Mnek	0.29	0.33	0.65	0.00	0.00	0.15	0.50	0.38	0.00	0.34	0.36	0.89

Mnek												
M4-												
Meso	0.96	0.96	0.90	0.43	0.40	0.94	0.93	0.87	0.34	0.00	0.93	0.23
M4-												
Macr	0.81	0.80	0.91	0.17	0.14	0.90	0.98	0.99	0.36	0.93	0.00	0.21
M4-												
Mnek	0.23	0.26	0.52	0.00	0.00	0.07	0.34	0.21	0.89	0.23	0.21	0.00

Highlights

- This study used stable isotopes to analyse the pelagic food web structure in the Kerguelen Island region, comparing HNLC areas upstream of the plateau and the iron enriched area on the plateau that experiences a large seasonal phytoplankton bloom.
- Trophic position was calculated for mesozooplankton, macrozooplankton, and micronekton using a Bayesian isotope mixing model approach.
- Meso and macrozooplankton trophic positions were similar but ~ 0.6 trophic positions lower than micronekton within areas.
- Median meso / macrozooplankton trophic position was ~ 2.4 on the plateau and 0.6 trophic positions higher in HNLC areas.
- Food chains were longer in HNLC regions, and food chain length was determined by the composition of the trophic levels below micronekton.
- Size based food web structure was robust to differences in community taxonomic composition.

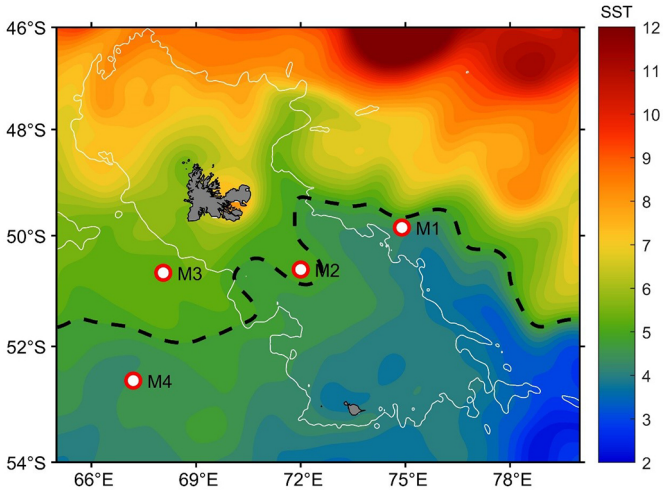


Figure 1

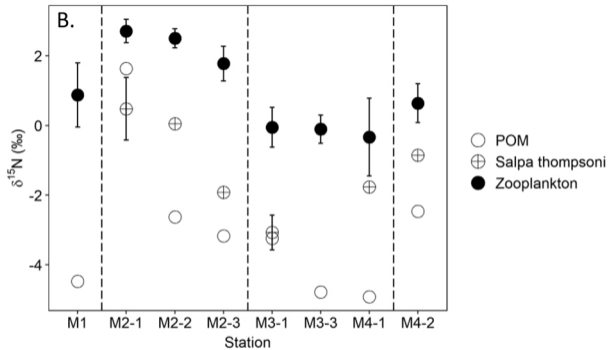
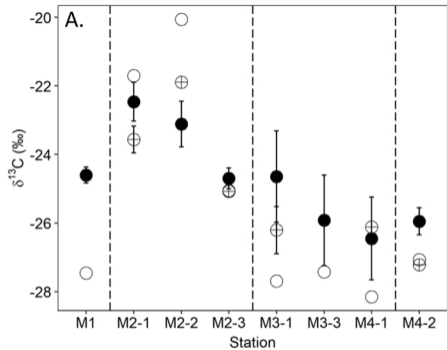


Figure 2

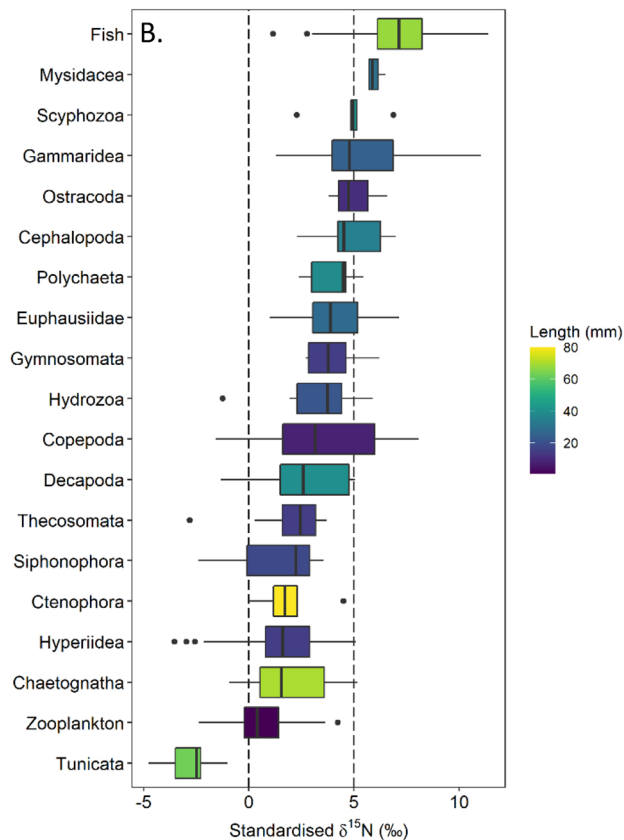
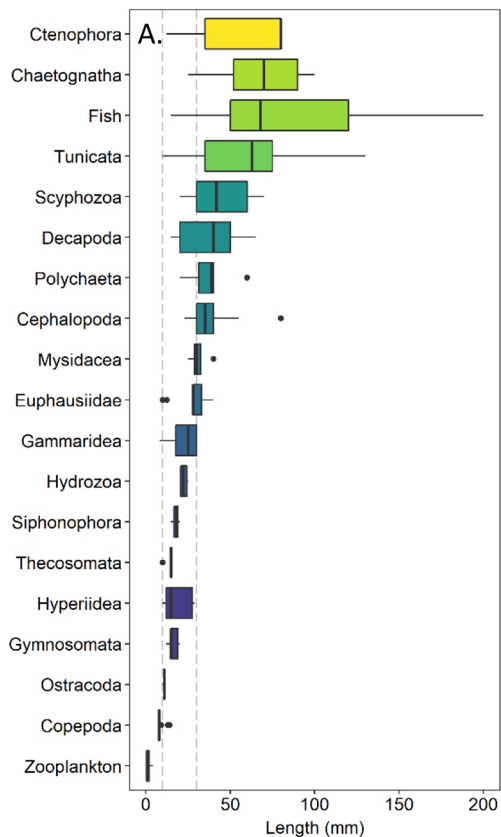


Figure 3

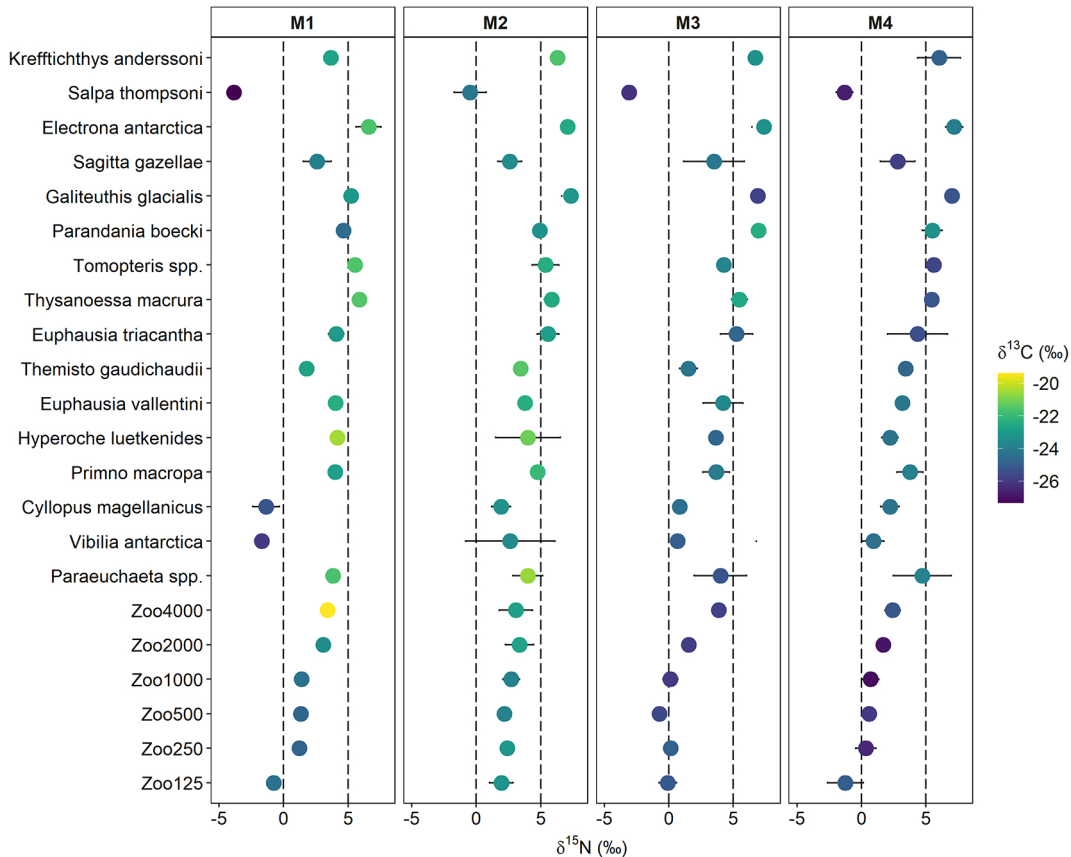


Figure 4

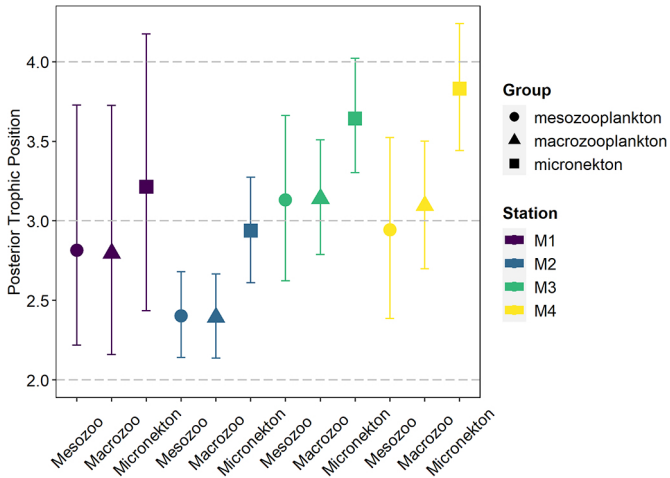


Figure 5