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May the *Oxa*-Michael Reaction of 2-Trifluoromethacrylic Acid Lead to Original Fluorinated Polyester?

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Abstract

2-Trifluoromethacrylic acid (MAF) is a peculiar fluorinated functional monomer. Though it is known to polymerise via C=C bond cleavage under anionic initiation, its *oxa*-Michael addition polymerisation afforded polyesters bearing trifluoromethyl side groups. This *oxa*-Michael addition was attempted in the presence of radical (peroxides), basic (amines, phosphines) and acidic (AICl₃, triflic acid, *p*-toluenesulfonic acid) reactants and catalysts which led to oligomers in various yields (up to 57 %) and molar masses (up to 2200 g.mol⁻¹). The obtained polyestes were characterised by GC/MS, IR, ¹⁹F, ¹H and ¹³C NMR spectroscopies, MALDI-TOF and their thermal properties were determined by thermogravimetric analysis and differentiel scanning calorimetry. Triphenylphosphine and piperidine led to the highest degree of polymerization DP_n (up to 16th adduct) with a degradation temperature of about 200 °C. The glass transition temperatures (T_g) were -59 and -46 °C for MAF bisadduct (diMAF) and oligoMAF, respectively. Such oligomers were hydrophobic, evidenced by water contact angle measurement of 107 ° showing that the contribution of trifluoromethyl group was higher than that of the ester function.

1. Introduction

Polyesters are considered as a solution for the sustainability problems resulting from the plastics waste in the environnement because their production requires lower energy than other materials such as polyamides. Additionnally, they have a good biodegrability thanks to the labile ester bonds as target sites for enzymatic or catalytic chemical attack, releasing carboxylic acid and hydroxyl residues.¹ Various polyesters and co-polyesters have been industrially produced, essentially by esterification of diols and diacids, or ring-opening of lactone-type monomers,^{2–4} but rarely by oxa-Michael addition from unsaturated carboxylic acids.

Oxa-Michael addition is an addition of a nucleophilic oxygenated group, usually an alcohol as Michael donnor, onto a compound bearing a conjugated system, called « Michael acceptor ».⁵ Alcoolates, the most known Michael donor, are good nucleophiles whereas carboxylates are moderate ones. This is explained by negative charge centered on the oxygen atom of alcoolate but delocalised by resonance

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on both oxygen atoms of carboxylate, making the latter less nucleophilic. Using carboxylate as Michael donor requires a good Michael acceptor, under catalytic or harsh conditions. *Oxa*-Michael addition using conjugated carboxilic acid, whether as Michael donor^{6,7} or acceptor,⁸ was reported in few articles in organic synthesis field.

Polymerisation of monomer bearing vinyl functions can be achieved either under radical or ionic initiations. More scarcely, unsatured carboxylic acids and derivatives might also be polymerised *via oxa*-Michael process. One typical example is acrylic acid (AA) that easily polymerises under radical conditions by π orbital cleavage in C=C bond, but has also been involved in *oxa*-Michael addition, affording polyesters in low molar masses (Scheme 1).⁹⁻¹³ In addition, dimer of AA was firstly observed under its storage conditions.^{13,14}



Scheme 1 : Oxa-Michael addition homopolymerisation of acrylic acid

According to Matsuoka's group,¹⁵ Lewis bases such as phosphines and *N*-heterocyclic carbene (NHC) underwent Michael addition to AA, followed by proton transfer as step-growth polymerisation, which produces a variety of polymers (random and telechelic ones). On the other hand, strong Brønsted acid catalysts (especially sulfonic acids) activate the carbonyl group of AA, affording high molar mass polyesters.⁹ However, the use of Ti(OⁱPr)₄, 4-MeOC₆H₄OH, hydroquinone and CuCl at 140 °C led to oligomer with DP_n close to 2.^{10,11}

As a matter of fact, 2-trifluoromethacrylic acid (MAF) is an attractive synthon¹⁶ and precursor of functionnal monomers.^{17,18} One of our research topics is exploring the reactivity of MAF and applying it in different polymerisation procedures, such as *co*- and *ter*polymerisations. According to the literature, MAF cannot be polymerised from radical initiation^{19,20} but both its electron-withdrawing trifluoromethyl and carboxylic acid enable it to be polymerised under anionic conditions.^{21–23} Hence, it can be copolymerised with electron-donating monomers (e.g., α -alkenes,²⁴ norbornene,^{25,26} vinyl ethers²⁷) and also with vinylidene fluoride (VDF).^{18,28}

However, contrarily to such reactions, to the best of our knowledge, the *oxa*-Michael addition of MAF was mentioned in only two reports. First, in 2003, a Japanese patent claimed the polymerisation of MAF *via* an *oxa*-Michael poly(addition) leading to adducts of low molar masses in the presence of triethylamine but this report lacks of many clues and evidences to really identify the expected structures of such resulting oligomers.²⁹ Second, in 2009, an analytical article reported the same polymerisation which occurred under UV radiation (365 nm) at 4 °C for 15 h, then at 100 °C for 1 h.

During the ephedrine titration in the presence of MAF in CDCl₃, MAF polymers were also observed, as undesired products. The mixture was then characterized by ¹H NMR spectroscopy. Despite the given assignments, MAF polymers were not isolated and no other analyses were supplied.³⁰ Thus, it has been of interest to deeper investigate a similar reaction, taking into account the experimental conditions of the reaction involving acrylic acid, and considering that MAF contains a specific trifluoromethyl group that may greatly influence the electronic effects and thus the reactivity of this monomer. Hence, the objective of this article is to synthesise and characterise fluorinated polyesters based on MAF by *oxa*-Michael addition.

2. Materials and Methods

Chemicals. 2-trifluoromethacrylic acid (MAF) and *tert*-butylperoxypivalate (*t*BuOOC(O)*t*Bu, TBPPi) (75%) were kindly offered by Tosoh Fine Chemicals Company (Shunan, Japan) and Akzo Nobel (Chalons sur Marne, France), respectively. KOH, 2,2,6,6-tetramethylpiperidine, piperidine, 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), *N*,*N*-diisopropylethylamine (DIPEA), P(*n*Bu₃)₃, PPh₃, concentrated HCl, FeSO₄.7H₂O, B(C₆F₅)₃, AlCl₃, MgSO₄, *p*-toluenesulfonic acid monohydrate (PTSA.H₂O), triflic acid, tricyclohexylphosphine P(C₆H₁₁)₃, racemic (2,2'-bis(diphenylphosphino)-1,1'-binaphthyl) (BINAP) and Amberlyst 15 were purchased from Aldrich Sigma, Fischer Scientific, Acros Organics and FluoroChem and used without further purification.

Gas chromatography coupled with mass spectrometry (GC/MS). GC model was GC-2010 Plus Shimadzu with injection port SPL1 (temperature : 250° C ; flow 26.6 mL/min ; purge flow 3 mL/min, inlet pressure 100 kPa) and column Zebron ZB-5MS (total flow 26.6 mL/min ; colum flow 0.76 mL/min ; linear velocity 38.2 cm/sec ; purge flow 3mL/ min). GC Column dimensions were 20 m of length x 0.18 mm of inner diameter x 0.18 µm of film thickness. The carrier gas was He and the column oven temperature was programmed from 30 °C isothermal for 5 min then 22 °C/min to 280 °C and isothermal for 5 min. MS model was GCMS-QP2010SE which is a quadrupole mass spectrometer equipped with electron ionisation at 200 °C. The measurements were performed in full-scan mode (m/z 2.0–800).

Nuclear magnetic resonance (NMR) spectroscopy. The ¹³C, ¹H and ¹⁹F NMR spectra (Figures 1-2) were recorded on Bruker Avance 400 instruments, using CDCl₃ as deuteurated solvent, signal of which was calibrated at 7.26 ppm. Hexafluorobenzene C_6F_6 was used as internal standard for ¹⁹F NMR analysises, its signal was calibrated at -164.9 ppm. Coupling constants and chemical shifts are given in Hertz (Hz) and parts per million (ppm), respectively. The experimental conditions for recording ¹H [or ¹⁹F] NMR spectra were as follows: angle 90° [or 30°], acquisition time 4.5 s [or 0.7 s], pulse delay 2 s [or 5 s], number of scans 36 [or 64], and a pulse width of 5 µs for ¹⁹F NMR. ¹H

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decoupling is applied on all ¹³C NMR analyses and almost on Attached Proton Test mode (¹³C{¹H}-APT). In the details of NMR characterisation, s, d, t, q, and m stand for singlet, doublet, triplet, quintet, and multiplet, respectively.

For 2D NMR spectroscopy, HMQC spectrum (Figure 3) was obtained with 2048 points in the F1 (13 C) dimension and 8192 points in F2 (1 H) one, a spectral width of 25.2 kHz in F1 and 8.0 kHz in F2. Acquisition times were 0.04 s in F1 and 0.5 s in F2. Two scans were applied to lead to a suitable signal/noise ratio and to respect phase of the pulse program sequence. A relaxation delay of 1.5 s was chosen.

HeteroCOSY ${}^{1}H{}^{19}F$ spectrum (Figure S16) was obtained with 1400 points in the F1 (${}^{1}H$) dimension and 15058 points in F2 (${}^{19}F$) one, a spectral width of 4.0 kHz in F1 and 7.5 kHz in F2. Acquisition times were 0.17 s in F1 and 1.0 s in F2. Eight scans and relaxation delay of 1 s were applied.

gHSQC spectrum (Figure S17) was obtained with 1024 points in the F1 (13 C) dimension and 7528 points in F2 (19 F) one, with a spectral width of 20.1 kHz in F1 and 3.8 kHz in F2. Acquisition times were 0.03 s in F1 and 1.0 s in F2. Eight scans and relaxation delay of 1 s were applied. ¹H decoupling was applied during the whole sequence.

Matrix-Assisted Laser Desorption Ionisation (MALDI-TOF) spectrometry. MALDI-TOF mass spectra were recorded using a Bruker Rapiflex time-of-flight mass spectrometer using a nitrogen laser for MALDI (λ 337 nm). The measurements in positive ion mode were performed with a voltage and reflector lens potential of 25 and 20 kV, respectively. Mixtures of peptides were used for external calibration. The matrix and cationising agent were *trans*-2-[3-(4-tertbutylphenyl)-2-methylprop-2-enylidene]malononitrile (DCTB, 10 mg.mL⁻¹ in CHCl₃) and sodium iodide (NaI, 6 mg.mL⁻¹ in methanol), respectively. The polymer concentration was 20 mg.mL⁻¹ in dichloromethane. The polymer and matrix were mixed in a 5 : 10 volume ratio. NaI was first deposited on the target. After evaporation of the solvent, the mixture, composed of the polymer and matrix, was placed on the MALDI target. The dry droplet sample preparation method was used.

Thermogravimetric analysis (TGA). The thermogravimetric analysis of the polymers (*ca.* 10 mg) was carried out under N₂ with a TA Instruments TGA 51 apparatus. The polymer samples were heated from 20 to 400 °C at a heating rate of 20 °C min⁻¹.

Fourier transform infrared (FTIR) spectroscopy. FTIR analyses of the copolymers were performed using a PerkinElmer Spectrum 1000 in ATR mode, with an accuracy of ± 2 cm⁻¹.

Differential scanning calorimetry (DSC). DSC analysis, under N₂ atmosphere, of the purified oligoMAF (*ca.* 10 mg) was carried out using a TA instruments Q100 instrument equipped with DSC

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200 F3 software. The instrument was calibrated with noble metals and checked before analysis with an indium sample ($T_m = 156.6$ °C). After its insertion into the DSC apparatus, the sample was cooled down from 20 °C to -120 °C and stabilized at latter temperature for 12 min. Then, the first scan was performed at a heating rate of 10 °C min⁻¹ up to 30 °C. The cycle cooling to -120 °C, stabilizing and warming up to 30 °C was repeated two more times. T_g was evaluated from three heatings, taken at the half-height of the heat capacity jump of the glass transition. The sample was amorphous and the glass transition temperature was determined by the inflection point in the heat capacity jump of the third scan.

Water contact angle (WCA). WCA measurements were carried out on Contact Angle System OCA-Data Physics. The water sessile drop method was used for the statics contact angle (CA) measurements at ambient temperature. The probe liquid was water (θ_{H2O}) and the average CA value was determined on five different drops of *ca.* 4.0 µL deposited on the same sample. OligoMAF coating was prepared by casting onto a glass slide from oligoMAF dissolved in dichloromethane.

Polymerisation procedure. In a reaction tube (Figure S1a) equipped with a magnetic stirrer, were introduced MAF and additive. The tube was purged under N₂ before being sealed and immersed into an oil bath heated at the requested temperature. The polymerisation was stopped after 24 hours by cooling the tube to room temperature. Immersion of the tube enabled to prevent the sublimation of MAF. Compared to round bottom flasks, the tubes allow to easily proceed simultaenous several (4 in the case of our experiments) reactions in 1.4 gram scale (Figure S1b). For reactions with amines, a filtration over Amberlyst 15 was nessecary to remove the produced ammonium (Scheme S1 & Figure S2). A subsequent sublimation at 60 - 70 °C under reduced pressure was achieved to remove the unreacted MAF and afforded oligoMAF as a yellowish and translucient wax. A more detailed description is supplied in the electronic Supplementary Information.

3. Results and discussion

To the best of our knowledge, the oligomerisation of MAF by *oxa*-Michael addition leading to low molar mass-oligomers was claimed in Japanese patent²⁹ and an analytical publication.³⁰ However, only the descriptions from ¹H and ¹³C NMR spectra of the dimer and trimer were reported. Thus, it was worth to confirm this formation by means of complementary techniques as ¹⁹F, 2D NMR and IR spectrocopies, GC/MS and MALDI-TOF analyses, as well as to attempt increasing the average degree of polymerisation (DP_n).

Based on the oligomerisation acrylic acid, polymerisation of MAF was then studied. Actually, CF_3 substituent makes MAF much more acidic than AA (pKa(MAF) = -2.1;³¹ pKa(AA) = 4.25)³² and might lead to significant change in term of reactivity under polymerisation conditions (Scheme 2). This

section is composed of two parts : i) first, the characterisations of oligoMAF are supplied and then ii) the influences of the experimental (radical, basic or acidic) conditions on the MAF conversion and DP_n are studied.



Scheme 2: Oligomerisation of MAF by oxa-Michael addition yielding fluorinated oligo(ester)s

3.1.Characterisation of oligo(MAF)s

Nuclear magnetic resonance (NMR) spectroscopy

After purification and drying, the oligo(MAF)s were analysed by NMR, GC/MS, MALDI and IR. The characterisation of oligoMAF was achieved by comparing those of MAF, diMAF and triMAF.

MAF. The NMR spectrum of MAF (Figure S4) shows two quartets centered at 6.88 ppm (${}^{4}J_{HFtrans} = 1.8$ Hz) and 6.60 ppm (${}^{4}J_{HFcis} = 1.3$ Hz) assigned to ethylenic CH₂= of MAF and a doublet (${}^{4}J_{FH} = 1.7$ Hz) of doublets (${}^{4}J_{FH} = 1.3$ Hz) at -68.81 ppm, attributed to the CF₃. The assignments are consistent with previous literature report.³³ Its ¹³C NMR spectrum (Figure S5) displays a singlet at 167.28 ppm, attributed to the carbonyl group in *c*. The quartets at 135.50 ppm (${}^{3}J_{CF} = 4.9$ Hz), 130.99 ppm (${}^{2}J_{CF} = 32.6$ Hz) and 121.18 ppm (${}^{1}J_{CF} = 272.3$ Hz) are assigned to CH₂ *d*, *C*(CF₃)(COOH) *b* and CF₃ *a*, respectively. Similar signals were observed in D₂O with the supplementary coupling ${}^{2}J_{HH} = 0.5$ Hz of the geminal protons of vinyl CH₂= (Figures S6-7).

DiMAF. The identification of diMAF (2m) was performed by analysing the mixture of MAF (84 %) and diMAF (16 %), with trace of triMAF, obtained from the reaction without any additive (Entry 1-Table 1), before MAF removal. The ¹H NMR spectrum (Figures S10-top) displays two quartets assigned to ethylenic hydrogens $CH_2 = d^1$ at 6.76 ppm (${}^4J_{HF} = 1.7$ Hz) and at 6.51 ppm (${}^4J_{HF} = 1.2$ Hz). The doublet (${}^3J_{HH} = 5.9$ Hz) at 4.71 ppm and the quartet (${}^3J_{HF} = 8.0$ Hz) of triplets (${}^3J_{HH} = 5.9$ Hz) at 3.64 ppm, are assigned to $CH_2 d^2$ and $CH b^2$, respectively.²⁹ CF₃ group in C=C(*CF*₃)(COOH) a^1 gives a doublet (${}^4J_{FH} = 1.7$ Hz) of doublets (${}^4J_{FH} = 1.2$ Hz) at -68.91 ppm, while that in CH₂-C(*CF*₃)(COOH) a^2 affords a doublet

 $(^{3}J_{FH} = 8.1 \text{ Hz})$, because of its coupling with geminal hydrogens, at -69.51 ppm (Figures S10 – bottom).

On the ¹³C NMR spectrum (Figure S11), the -COO- c^1 and c^2 lead to signals at 170.41 and 160.77 ppm. The quartets at 134.46 ppm (${}^{3}J_{CF} = 5.0 \text{ Hz}$) and 130.69 ppm (${}^{2}J_{CF} = 32.7 \text{ Hz}$) are assigned to CH₂= d^1 and CH₂= $c(CF_3)(COO)$ b^1 , respectively. The CH₂= $c(CF_3)(COO)$ a^1 is represented by a quartet (${}^{1}J_{CF} = 272.3 \text{ Hz}$) at 121.18 ppm which is totally overlapped with the corresponding carbon atom of the MAF monomer. The CH₂-CH(CF_3)(COO) a^2 gives a quartet at 123.29 (${}^{1}J_{CF} = 280.2 \text{ Hz}$). The quartets located at 60.23 ppm (${}^{3}J_{CF} = 2.9 \text{ Hz}$) and 49.35 ppm (${}^{2}J_{CF} = 28.6 \text{ Hz}$) are attributed to CH₂ d^2 and CH b^2 , respectively.²⁹ These assignments are confirmed below in the analysis of the diMAF/triMAF mixture (Figures S12-14).

TriMAF. Within the same reaction (Entry 1-Table 1), after MAF removal by sublimation, a mixture of diMAF (67 %) and triMAF (33 %) was analysed. The spectra of triMAF exhibit all signals characteristic of the diMAF, and therefore the CH₂CH(CF₃)COOH moiety is described hereafter. The ¹H NMR spectrum (Figure S12-top) displays a doublet (${}^{4}J_{HF} = 5.9$ Hz) of multiplets at 4.67 ppm assigned to CH₂ d^{3} and a quartet (${}^{3}J_{HF} = 8.0$ Hz) of triplets (${}^{3}J_{HH} = 5.9$ Hz) at 3.60 ppm attributed to CH b^{3} . The multiplicity of the latter signal was determined thanks to ${}^{1}H{}^{19}F{}$ NMR spectrum (Figure S12-bottom).



Figure 1 : ¹H (top), ¹⁹F (middle) and ¹⁹F{¹H} (bottom) NMR spectra, recorded in CDCl₃, of oligoMAF obtained without additive, 105 °C, 63 h, after MAF removal (Entry 2 – Table 1). $DP_n = 2.7$ was dertermined with both of ¹H and ¹⁹F NMR.

The ¹⁹F NMR spectrum (Figure S13 - bottom) shows the signal of CF₃ end unit, CH₂=C(*CF*₃)(COO) *a*¹ in triMAF, at -68.89 ppm as a doublet (⁴ $J_{FH} = 1.7$ Hz) of doublets (⁴ $J_{FH} = 1.2$ Hz) of doublets ($J_{FF} = 1.8$ Hz), distinguished from the corresponding CF₃ of diMAF. The CF₃ of unit 3 gives a doublet (³ $J_{FH} = 8.1$ Hz) of doublets ($J_{FF} = 3.2$ Hz) at -69.56 ppm. Two signals involved J_{FF} were observed with weak intensities and will be discussed later. These assignments were achieved thanks to decoupled ¹⁹F{¹H} and ¹⁹F{¹H}{¹³C} NMR spectra (Figure S13 - top) which gave the same profile. It is worth noting that the elementary doublets ($J_{FF} = 1.8$ Hz and $J_{FF} = 3.2$ Hz), due to the through-space coupling that is explained further, were not present in diMAF.

Distinct signals were also observed in the ¹³C NMR spectrum (Figure S14), as the CH₂= d^1 and CH₂=c(CF₃)(COO) b^1 of triMAF are represented by two quartets at 134.60 (${}^{3}J_{CF}$ = 5.0 Hz) and 130.55 ppm (${}^{2}J_{CF}$ = 32.9 Hz), respectively, slightly shifted from the corresponding signals of diMAF. The MAF unit 3 gives two peaks at 170.03 and 164.15 ppm, attributed to COO c^3 , a quartet at 123.16 ppm (${}^{1}J_{CF}$ = 286.2 Hz) to CH₂=C(*CF*₃)(COO) a^3 , quartet (${}^{3}J_{CF}$ = 2.4 Hz) at 60.63 ppm to CH₂ d^3 and two quartets of doublets at 49.32 ppm (${}^{2}J_{CF}$ = 28.5 Hz, J_{FF} = 2.2 Hz) and 49.17 ppm (${}^{2}J_{CF}$ = 28.7 Hz, J_{FF} = 2.9 Hz) assigned to CH b^3 .²⁹

The coupling atoms of previously mentionned J_{FF} was confirmed because the corresponding doublets were observed on both ¹⁹F{¹H} and ¹⁹F{¹H}{¹³C} NMR spectra (Figure S13-top), which have the same profile. Similar coupling constants are also noted in the ¹³C NMR spectrum (Figure S14). It indicates likely two fluorine atoms from two consecutive CF₃ groups of oligoMAF, separated by 8 chemical bonds. Indeed, according to the literature, ³⁴ the values 2.2 and 2.9 Hz should correspond to the coupling constant of ⁿJ_{FF} with n = 5 – 7, which are not found in the oligoMAF structure. As reported by Foris and Roche, ^{35,36} direct through bond ⁸J_{FF} coupling can be eliminated as these authors found value up to only 2 and because of the presence of the O atoms between two CF₃ groups.^{37,38} Indirect through space coupling seems therefore to be a reliable explanation.

OligoMAF. The oligoMAF obtained from the reaction without additive, heated at 105 °C for 63 h (Entry 2- Table 1) after MAF removal was characterised by NMR spectroscopy. The ¹H NMR spectrum (Figure 1 - top) shows multiplet centered at 6.76 ppm and 6.51 ppm attributed to ethylenic protons $CH_2 = d^1$. The complex multiplets ranging from 4.72 to 4.57 ppm and from 3.69 to 3.59 ppm are assigned to $CH_2 d^2...d^n$ and $CH(CF_3)COOH b^2...b^n$ moities of the oligomers, respectively. As the adjacent $CH(CF_3)$ group bears an asymetric carbon atom, both protons in such a methylene group are non-equivalent. The ¹⁹F and ¹⁹F{¹H} NMR spectra (Figure 1 – middle and bottom) display $C=C(CF_3) a^1$ multiplet centered at -68.91 ppm and $-CH(CF_3)COO-a^2...a^n$ signal centered at -69.51 ppm.

Concerning the ¹³C NMR spectroscopy (Figure 2), the singlets found from 160.7 to 169.7 ppm are attributed to COO $c^1...n^n$ while those from 134.3 to 135.5 ppm are assigned to CH₂ d^1 . Quaternary carbon in CH₂=c(CF₃)(COO) b^1 gives quartets (² $J_{CF} \approx 33$ Hz) centered at 130.5 ppm. Quartet (¹ $J_{CF} = 272.4$ Hz) at 121.14 ppm is attributed to CF₃ a^1 while the quartet (¹ $J_{CF} = 280.2$ Hz) of multiplets at *ca*. 123 ppm is assigned to CF₃ $a^2...a^n$. Signals from 60.8 to 60.0 ppm and from 49.8 to 48.5 ppm are attributed to $d^2...d^n$ and $b^2...b^n$, respectively.



Figure 2 : ${}^{13}C{}^{1}H$ -APT NMR spectrum, recorded in CDCl₃, of oligoMAF (obtained without additive, 105 °C, 63 h, after MAF removal) (Entry 2-Table 1)

The 2D ¹H-¹³C HMQC NMR spectrum (Figure 3) confirms the assignments. Both protons in the methylene group d^1 signals, centered at 6.76 and 6.51 ppm in the ¹H NMR spectrum, are connected to the corresponding signal of the carbon atom, found at 134.6 ppm in the ¹³C NMR spectrum. Multiplets at 4.75-4.61 ppm and 3.69-3.52 ppm, are connected to those at 60.81-59.99 ppm and 49.76-48.57 ppm, showing the correlations for CH₂ $d^2...d^n$ and CH $b^2...b^n$ moities, respectively. These clues evidence that the *oxa*-Michael reaction occured instead of a polymerisation by the opening of the C=C π bond.

The 2D ¹H-¹⁹F and ¹³C-¹⁹F NMR spectra were also recorded as evidences of these assignments. The HeteroCOSY ¹H-¹⁹F spectrum (Figure S16) shows the correlations of F atoms with the nearest H ones, *i.e.* CH₂= d^1 /CF₃ a^1 and CH $b^2...b^n$ / CF₃ $a^1...a^n$ The signals of these protons are intense on the one dimension spectrum (¹H). Signal at 4.6 ppm of further protons as CH₂ $d^2...d^n$, compared to CH $b^2...b^n$, have weak intensity resulting from weak correlation. The gHSQC spectrum (Figure S17) highlights

only ${}^{1}J_{CF}$, corresponding to CF_{3} a^{1} and $a^{2}...a^{n}$. ${}^{1}H$ decoupling is applied during the whole sequence and, therefore, only the signals of the C atoms that have ${}^{1}J_{CF}$ are displayed on the one dimension spectrum (${}^{13}C$).



Figure 3 : 2D ${}^{1}H^{-13}C$ HMQC NMR spectrum, recorded in CDCl₃, of oligoMAF (obtained without additive, 105 °C, 63 h) after MAF removal (Entry 2 – Table 1). The one-dimensional spectra are plotted at the top (${}^{1}H$) and left (${}^{13}C{}^{1}H$)-APT) of the 2D plots

Based on these above NMR characterisations, the formulas for MAF conversions and average degrees of oligomerisation, DP_n , can be proposed. As each unit in oligomers has distinguished CH (vinyl =C*H* or -C*H*(CF₃)COOH moities) and CF₃ (C=C(*CF₃*) or -CH(*CF₃*)COO-), MAF conversion can be calculated with the formulas (1) or (2) using integrals from the characteristic signals in ¹H or ¹⁹F NMR spectra of the reaction mixtures before work-up, respectively (Figure 4 - top). For phosphines-catalysed reactions, see ESI.

$$\operatorname{Conv.(MAF)(\%)} = \frac{\int_{6.7}^{6.8} CH(oligomers) + \int_{3.6}^{3.7} CH(oligomers)}{\int_{6.7}^{6.8} CH(oligomers) + \int_{3.6}^{3.7} CH(oligomers) + \int_{6.8}^{6.9} CH(unreacted MAF)} x \ 100$$
(1)

 $Conv.(MAF) (\%) = \frac{\int_{-69.0}^{-68.9} CF_3(oligomers) + \int_{-69.6}^{-69.5} CF_3(oligomers)}{\int_{-69.0}^{-68.9} CF_3(oligomers) + \int_{-69.6}^{-69.5} CF_3(oligomers) + \int_{-69.1}^{-69.0} CF_3(unreacted MAF)} x100$ (2)

For the same reason, DP_n can be also determined in two ways (equations 3 and 4) :

$$DP_{n} = \frac{\int_{6.7}^{6.8} CH(oligomers) + \int_{3.6}^{3.7} CH(oligomers)}{\int_{6.7}^{6.8} CH(oligomers)} = 1 + \frac{\int_{3.6}^{3.7} CH(oligomers)}{\int_{6.7}^{6.8} CH(oligomers)}$$
(3)

$$\mathsf{DP}_{\mathsf{n}} = \frac{\int_{-69.0}^{-68.9} CF_3(oligomers) + \int_{-69.1}^{-69.0} CF_3(oligomers)}{\int_{-69.0}^{-68.9} CF_3(oligomers)} = 1 + \frac{\int_{-69.1}^{-69.0} CF_3(oligomers)}{\int_{-69.0}^{-68.9} CF_3(oligomers)}$$
(4)



Figure 4 : ¹H (left) and ¹⁹F (right) NMR spectra, recorded in CDCl₃, of the product resulting from the oligomerisation of MAF by oxa-Michael addition in case of $DP_n = 2.3$ (top) and $DP_n = 3.7$ (bottom) (Entries 1 & 6 - Table 1); starred signals represent unreacted MAF

Fourier transform infrared (FTIR) spectroscopy

IR spectrum of MAF (Figure S9) displays a large band centered at 3500 cm⁻¹, corresponding to O-H (in COOH) and C(sp²)-H of ethylenic CH₂= end-group. C=O (in COOH) frequency at 1720 cm⁻¹ and that of C-F at 1145 and 1102 cm⁻¹ have high intensities because of the high polarity of the bonds. C=C bands with medium intensities were also observed at 1635 and 1407 cm⁻¹.

The IR spectrum of oligoMAF (Figure S20 – red spectrum) exhibits a wide band with weak intensity centered at 2977 cm⁻¹, corresponding to the vanishing of O-H(COOH) and C(sp²)-H and also the presence of C(sp³)-H. The C=O(ester) band at 1742 cm⁻¹ is slightly shifted compared to that of C=O(COOH) of MAF. Weaker intensity was observed at the C=C bands at 1635 and 1407 cm⁻¹. Stronger signals, compared to those in IR spectrum of MAF, observed in 1300 – 900 cm⁻¹ window, are assigned to formed C-C and C-O-C=O bonds. All the described frequencies are in stretching mode.

Gas chromatography coupled with mass spectrometry (GC/MS)

The GC spectrum of MAF shows a peak at a retention time (t_R) of 6.77 min (Figure S8). The GC spectrum of example 2 – Table 1 (Figure 5 - top) exhibits two peaks: that at t_R = 10.18 min correponds

to diMAF while the one at $t_R = 11.95$ min to triMAF and longer oligoMAFs, confirmed by GC analysis of diMAF/triMAF mixture (Figure S15). The mass spectroscopy spectra of diMAF and triMAF/oligoMAF (Figure 5 – two middle spectra) display m/z = 123, corresponding to $CH_2=C(CF_3)COO^{\bullet}$ fragment, as the base peak.

Based on the obtained m/z values, two decomposition mechanisms of diMAF can be proposed (Scheme 3). Pathway 1 starts with the cleavage of the ester C-O bond, giving $C_4H_4F_3O_3^{\bullet}$ (m/z = 157) fragment. The subsequent fragmentations of O atom and -COOH group afford $C_4H_4F_3O_2^{\bullet}$ (m/z = 141) and CHO_2^{\bullet} (m/z = 45), respectively. The second pathway separates 2 MAF units and leads to $C_4H_2F_3O_2^{\bullet}$ (m/z = 139). The ester C-O bond is then broken, releasing $CH_2=C(CF_3)CO^{\bullet}$ (m/z = 123). Two final fragmentations give $C_3H_2F_3^{\bullet}$ (m/z =95) and CF_3^{\bullet} (m/z = 69). In addition, decarboxylation of M leads to m/z = 235 (decomposition 3).



Figure 5 : GC/MS spectra of oligoMAF mixture (Entry 2 - Table 1)



Scheme 3 : Proposed fragmentation pathways based on the m/z values given by GC/MS (Entry 2 - Table 1)

OligoMAF, obtained in presence of diisopropylethylamine (DIPEA) (Entry 9-Table 1), was analysed by MALDI-TOF spectrometry. The positive reflectron spectrum (Figure 6) exhibits two signal series with unsymmetric Gaussian shape distribution. The first series, marked in green is assigned to [OligoMAF-Na]⁺ with m/z = 140y + 23 and the mass difference corresponding to a MAF unit (m/z = 140), ranging between m/z = 583 (DP₄) and m/z = 2263 (DP₁₆). The distribution was centered on a DP₆ chain at m/z = 863, the formula of which was determined as CH₂=C(CF₃)COOCH₂[CH(CF₃)COOCH₂]₄CH(CF₃)COOH from isotopic distribution (Figure S21). M_n and M_w are 975 and 1083, respectively, leading to a dispersity of 1.1. The MS-MS analysis of 723 m/z fragment gave similar profile as in case of oligoMAF obtained without additive (Entry 9-Table 1 and Figure S22).





Figure 6 : MALDI-TOF mass positive reflectron spectrum with expansion from 850 and 1090 m/z with NaI and trans-2-[3-(4-tertbutylphenyl)-2-methylprop-2-enylidene]malononitrile (DCTB) as the matrix (Entry 9-Table 1)

In the second series, marked in orange (Figure 6), all signals display quasi the same intensity as those of the green series. This probably indicated that all signals of the green distribution have lost some fragment. Indeed, after loosing a sodium atom, further CO_2H end-group was also removed (Figure 7), probably resulted from the presence of DIPEA trace in the sample. The green series is then attributed to [OligoMAF-COOH]⁺, with the formula : m/z = 140y - 45. Its distribution is centered on a m/z = 975 (DP₆) and ranging between m/z = 655 (DP₄) and m/z = 2195 (DP₁₆). Average M_n and M_w are 1006 and 1148, respectively, affording a dispersity of 1.1. MS-MS analysis of 935 m/z (DP₇) fragment (Figure S23) shows a cascade of MAF units (140) and, its isotopic distibution was also supplied.



Figure 7: OligoMAF - COOH corresponding to (140y -45) series, where y is the MAF number

Another series of signals of small intensity next to the former one with m/z = 140y + 23 + 4 is also observed but no explaination could be provided so far. OligoMAF obtained without additive (Entry 2-Table 1) was also analysed and gave coherent results which are described in ESI (Figures S18-S19, Table S1).

Thermogravimetric analysis (TGA)



Figure 8 : TGA thermogram and its derivative of oligo(MAF) under nitrogen (Entry 9-Table 1)

The thermal stability of oligoMAF was studied by thermogravimetric analysis (TGA) under N₂.

Thermogram of oligoMAF (Figure 8) exhibits one step-degradation from 96 °C (1.5 % loss) and 50 % loss at 176 °C. This degradation temperature is lower than that of non-fluorinated esters from acrylic acid that occurs in 250 - 300 °C range,⁹ or those from diols and diacids in 220 – 230 °C range.¹ The electron-withdrawing CF₃ group may induce some electronic effect to favor a faster degradation in addition to low molar mass of oligoMAF. Furthermore, it is known that depolymerisation occurs by unzipping from the COOH end group that first undergoes a decarboxylation.³⁹

Differential scanning calorimetry (DSC)

The glass transition temperatures (T_g) of oligoMAF was determined by DSC. The thermograms of which (Figures S29 – S30) exhibit a single T_g : that of low molar mass diMAF (Entry 1 – Table 1) was - 59 °C while oligoMAF obtained with DIPEA (Entry 9 –Table 1) has a T_g of -46 °C.

Water contact angle (WCA)

The surface properties of oligoMAF were assessed by means of water contact angle measurements (Figure 9 and Table S2). A glass substrate was coated by casting with oligoMAF (Entry 11–Table 1) that was first dissolved in dichloromethane and spread onto glass. A hydrophobic character was noted in the first 12 seconds. As expected, in contrast to polyacrylates bearing C_8F_{17} side groups,^{40,41} the oligoMAF does not display so high hydrophobic properties, probably limited by the presence of hydrophilic ester and carboxylic end groups.



Figure 9 : Water contact angles on oligoMAF obtained with $P(C_6H_{11})_3$ (Entry 11 – Table 1) coated onto a glass substrate at different intervals of time.

3.2.Additives screening

The second part of this section aims at optimising such a reaction according to radical, basic or acidic conditions and the results are listed in Table 1. First, without any additive, heating MAF at 95 °C for 24 h afforded oligoMAF in 15 % yield in low molar mass ($DP_n = 2.3$) (Entry 1-Table 1). MAF conversion (66 %) and propagation ($DP_n = 2.6$) increased with reaction temperature (105 °C) and time (63 h) (Entry 2 – Table 1). However, using toluene as solvent did not enhance these features (Entry 3 – Table 1). Then, various additives were used and our interest was to study the influence of radical,

basic or acidic species on the MAF conversion and $\mathsf{DP}_{\mathsf{n}}.$

Table 1 : Additives screening and experime	ntal conditions for the oliaomerisation	of MAF by oxa-Michael addition
		-,

Entry	Additive type	Conditions ^a	MAF Conv. (%) ^{b, c}	DPn ^{b, d} (DPn ^{b, e})	OligoMAF yield (%) ^f
1	No additive	-	11	2.3 (2.2)	15
2	No additive	105 °C, 63 h	66	2.6	-
3	No additive	Toluene, 105 °C, 63 h	12	2.3	-
4	Radical	TBPPi (3.5 %)	15 ^{<i>j</i>}	2.5 (2.1)	-
5	Base	KOH (5 %)	73	3.2 (3.1)	-
6	Base	2,2,6,6- Tetramethylpiperidine (5 %)	71	3.0 (2.6)	57
7	Base	Piperidine (5 %)	76	3.5 (3.0)	-
8	Base	DBU (5 %)	76	3.1 (3.0)	49
9	Base	DIPEA (5 %)	78	3.1 (3.4)	-
10	Base	P([″] Bu)₃ (5 %)	66	3.2 (2.9)	-
11	Base	P(C ₆ H ₁₁) ₃ (5 %)	73	2.9 (3.2)	-
12	Base	racemic BINAP (2.5 %)	76	2.7 (3.1)	32
13	Base	PPh₃ (0.5 %)	83	3.5 (3.3)	-
14	Base	PPh3 (5 %)	71	3.3 (3.0)	-
14A	Base	PPh₃ (5 %) MgSO₄ (5 %), 17 h	71	3.1 (2.8)	-
14B	Base	PPh₃ (5 %) MgSO₄ (5 %)	71	2.9 (2.9)	-
14C	Base	PPh₃ (5 %) MgSO₄ (5 %), 41 h	66	3.2 (2.7)	-
15	Base	PPh₃ (5 %), 60 °C	73	3.5 (3.8)	-
16	Base	PPh₃ (5 %), 80 °C	87	3.5	-
17	Base	PPh₃ (5 %), 120 °C	- ^g	-	-
18	Base	PPh ₃ (10 %)	79	2.8 (2.6)	-
19	Base	PPh₃ (20 %), 10 min	_ ^g	-	-
20	Acid	HCl _{aq 35 %} (8 %)	26	2.9 (2.1)	-
21	Acid	FeSO ₄ .7H ₂ O (5 %)	14	1.9 (1.7)	-
22	Acid	B(C ₆ F ₅) ₃ (5 %)	10	2.2 (2.1)	-
23	Acid	AICl ₃ (5 %)	72	3.5 (3.5)	-
24	Acid	MgSO ₄ (5 %)	13	2.2 (2.1)	-
25	Acid	PTSA.H₂O (5 %)	43	2.3 (2.5)	-
26	Acid	Triflic acid (5 %)	67	2.6 (2.6)	-
27	Acid/Base	PPh ₃ (5 %)	81	3.3	-

		AlCl₃ (5 %), 60 °C			
28	Acid/Base	PPh ₃ (5 %), AlCl ₃ (5 %),	69	3.1 (3.2)	-
		Toluene, 60 °C, 63h			

^{*a*} 95 °C, 24h, under N₂, otherwise stated in Table; ^{*b*} Determined from reaction mixtures; ^{*c*} Determined from ¹H NMR spectra and equation (1) or (1'); ^{*d*} Determined from ¹H NMR spectra and equation (3) or (3'); ^{*e*} Determined from ¹⁹F NMR spectra and equation (4); ^{*f*} Determined from ¹H NMR spectra of reaction mixtures after work-up (ammonium elimination and MAF sublimation); ^{*g*} with other unidentified product(s).

TBPPi, DBU, DIPEA and PTSA stand for *tert*-butylperoxypivalate, 1,8-diazabicyclo[5.4.0]undec-7-ene, *N*,*N*-diisopropylethylamine and *p*-toluenesulfonic acid, respectively.

Radical additives

Though MAF is known not to homopolymerise under radical initiation,^{19,20} Aglietto *et al.*²⁴ reported that *tert*-butylcyclohexyl peroxydicarbonate led to low molar mass oligomers in poor yield. *tert*-Butylperoxypivalate (TBPPi) (3.5 mol%) was attempted as initiator for a radical trial, but did not lead to any remarkable change of MAF conversion (Entry 4 - Table 1).

Basic additives

Inorganic base as KOH led to high MAF conversion (76%) and a DP_n of 3.2 (Entry 5 - Table 1). Hindered secondary and tertiary amines such as piperidine, 2,2,6,6-tetramethylpiperidine, *N*,*N*-diisopropylethylamine (DIPEA) and 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) afforded the oligomers with almost the same results (MAF conversion of 71-78 % and DP_n = 3.1-3.7) (Entries 6-9 - Table 1).

Different aliphatic and aromatic phosphines such as $P(^{n}Bu)_{3}$, $P(C_{6}H_{11})_{3}$, PPh_{3} (5 mol%) and racemic (2,2'-bis(diphenylphosphino)-1,1'-binaphthyl), BINAP, (2.5 mol%), led to almost good MAF conversions (66-76 %) and DP_{n} (2.7-3.3) (Entries 10 - 12, 14 - Table 1). Highest DP_{n} value was obtained when the reaction was catalyzed by PPh_{3} , the loading of which was then considered.

Lower PPh₃ loading (0.5 mol%) or temperatures (60 and 80 °C) (Entries 13, 15, 16 – Table 1) led to same DP_n (3.5) and higher MAF conversions (73-87 %). But higher PPh₃ loading (10 and 20 mol%) or temperature (120 °C) (Entries 17-19, Table 1) gave lower DP_n (2.8) or undesired side-products which were not identified.

Acidic additives

HCl simultaneously led to oligoMAF and unexpected ClCH₂-CH(CF₃)COOH (MAF-Cl), resulted from electrophilic addition of HCl onto C=C bond of MAF (Entry 20 – Table 1). MAF-Cl was intentionnally formed by adding HCl_{aq} 35% to MAF (Scheme S3, Figures S27-S28), including NMR evidence. Lewis acids such as FeSO₄.7H₂O, B(C₆F₅)₃ almost did not improve the reaction (Entries 21, 22 - Table 1). AlCl₃

yielded satisfactory MAF conversion into oligomers with DP_{4.5} but also to CICH₂-CH(CF₃)COOH (13 %) (Entry 23 - Table 1, Figure S19). It is worth to note that AlCl₃ is known as a Lewis-acid catalyst for electrophilic chlorination of aromatic rings or of alcohols.⁴² Indeed, vinylic chlorination with AlCl₃ requires benzeneseleninyl chloride.⁴³ In addition, chlorination of fluorinated telomers was also succesfully achevied in presence of AlCl₃.⁴⁴ To the best of our knowledge, the use of AlCl₃ as a chlorine source for chlorination of fluorinated olefin has not been reported so far.

Organic Brønsted acid such as *p*-toluenesulfonic acid monohydrate (PTSA.H₂O) and triflic acid increased the MAF conversion to 43 and 67%, respectively, but with low DP_n were produced (2.3 and 2.6) (Entries 25-26 - Table 1).

Other parameters

Low MAF conversion might be due to the presence of water molecules, as in cases of PTSA.H₂O and FeSO₄.7H₂O (Entries 21 and 25 - Table 1). To verify it, MgSO₄ that traps water was used and slightly increased the MAF conversion (Entry 24 - Table 1) but did not lead to any significative change in the presence of PPh₃ (Entry 14B – Table 1).

From different reaction times (17, 24 and 41 h), slight differences of DP_n (2.9-3.2) and MAF conversions (66-71 %) were observed (Entries 14A - 14C - Table 1).

Screening of additive combinations

As using PPh₃ and AlCl₃ separately led to good results, our next interest was exploiting the reactivity of their combination, as a Lewis acid base couple. It was barely possible to perform the reaction catalysed by both of AlCl₃ and PPh₃ at 95 °C because the additives mixture turned to red then to black in a few seconds, indicating its burning before MAF melting (60 °C). In other cases, MAF played the role of solvent. Lower temperature (60 °C) (Entry 27 – Table 1) was then attempted and the same DP_n (3.3) was observed with 81 % MAF conversion, which is slightly better than the result obtained with only PPh₃ at 95 °C (Entry 14 – Table 1). Toluene was also used as the solvent to « homogenise » the mixture. Similar to the no-additive procedure mentionned above (Entries 2-3 - Table 1), the presence of toluene led to lower MAF conversion (69 %) and DP_n (3.1) (Entry 28 - Table 1). In these two attempts, ClCH₂-CH(CF₃)COOH was also obtained in low yields (13%).

Finally, the best conditions for high MAF conversion (83-87 %) were achieved in presence of PPh₃ (0.5 % at 95 °C or 5 % at 80 °C) for DP_n of 3.5. As a matter of fact, a slight difference of DP_n values was observed according to the analysis but the results are still consistent when supplied from the same method. OligoMAF obtained without additive (Entry 2 – Table 1) had DP_{2.6} from ¹H NMR and DP₅ from MALDI-TOF, while the one obtained with DIPEA (Entry 9 – Table 1) had DP_{3.1} from ¹H NMR and

DP₆ from MALDI-TOF). MAF removal was applied on some of reactions and showed relative coherence with calculated MAF conversions. However, retro oxa-Michael was observed during the sublimation of oligoMAF phosphonium (Scheme S2).

3.3. Mechanisms of the dimerisation of MAF by oxa-Michael addition

In order to explain these obtained results, the oligomerisation mechanisms, classified by additive types, were considered. To simplify the discussion, only dimerisation mechanisms are supplied.

Without additive

Without any additive, two molecules of MAF could undergo an intermolecular *oxa*-Michael addition, affording diMAF *via* a typical six-membered transition state⁴⁵ **Int1**, eventually disfavored by the poor electron C=C bond (Scheme 4 – first mechanism).



Figure 10 : Possible rearrangment by intermolecular hydrogen bond of MAF

Furthermore, the rearrangment by intermolecular hydrogen bond of MAF **Int1'** (Figure 10), corresponding to a eight membered transition state that dominates the desired reaction and led to low MAF conversion. MAF can also be considered as a Brønsted acid that catalyses the reaction, the mechanism of which is described below (Scheme 4 – fourth mechanism).

Without additive



Scheme 4 : Mechanisms of dimerisation of MAF by oxa-Michael addition under different acidic and basic conditions

Basic additives

In the presence of a Brønsted base (such as KOH or secondary and tertiary amines), MAF would be deprotonated to generate carboxylate **Int2** that attacks, as a nucleophile, an second MAF molecule. The resulting carboxylate **Int3** is then protonated to afford diMAF (Scheme 4 – second mechanism).⁴⁶

By using Lewis base (such as tertiary phosphines PR₃), the nucleophilic conjugate addition of the Lewis base on the vinyl group of MAF takes place to form the zwitterion **Int4**, known as the activated specy of Morita-Baylis-Hilman or Rauhut–Currier reactions.^{47,48} **Int4** can undergo a rearrangment to yield **Int5**, that propagates to form **Int6**. **Int4** might also undergo a proton migration to give carboxylate **Int5'** that is then added onto the vinyl group of a second MAF molecule to yield

intermediate **Int6'**. According to the ³¹P NMR spectrum (Figure S26), the Lewis base was not eliminated at the end of the reactions. The desired diMAF was then obtained with its phosphinated derivates (Scheme 4 – third mechanism).⁴⁶

Tertiary amines act mostly like a Brønsted base in this reaction because of the domination of acidbase reaction vis-a-vis the conjugated addition. This was proved by the formation of the organic ammonium salt, $(RCOO)^{-}(R'_2NH_2)^{+}$, when mixing equimolar 2,2,6,6-tetramethylpiperidine and MAF (Scheme 5). The vanishing of COOH signal was observed in ¹H NMR spectrum (Figure S25). The electrophilic addition adduct, of amine onto MAF C=C bond, was not observed.¹⁶ With the steric hindrance, the ammonium cation should prevent from the propagation of the attached carboxylates.



Scheme 5: Formation of the organic ammonium salt from MAF and 2,2,6,6-tetramethylpiperidine

 PR_3 phosphines are less basic but more nucleophilic than similarly substituted amines NR_3 and led clearly to third mechanism.^{49,50} Additionnally, trialkylated phosphines are more nucleophilic than PPh_3 ,^{49,50} and therefore generate more **Int6'** species than PPh_3 does, affording shorter oligomer chains. The availability of carboxylate group of phosphonium should promote the propagation and explain higher DP_n of oligoMAF.

Acidic additives

The Brønsted acid (such as PTSA.H₂O and triflic acid) activates the carboxylic acid function of MAF, triggering the conjugate addition in a six-membered transition state, affording **Int9**. A subsequent hydrogen shift gives diMAF (Scheme 4 – fourth mechanism).^{9,13}

The Lewis acid (such as $FeSO_4.7H_2O$ and $B(C_6F_5)_3$) did not promote the oligomerisation, giving low MAF conversions and DP_n. It is worth to know that heating the equimolar MAF and MAF-Cl mixture at 95 °C for 1 day did not lead to any conversion (Scheme S4). No reasonnable explanation of high DP_n when using AlCl₃ can be given.

No matter the pathway, the key **Int3**, **Int6**, **Int6'** and **Int9** intermediates are able to undergo further Michael addition to propagate the oligomer chains.

4. MAF polymerisation pathways

MAF can undergo homopolymerisation with two different monomer units: $-[CH_2-C(CF_3)(COOH)]$ - unit formed via C=C π bond cleavage, under basic conditions ^{21–23} and $-[CH_2-CH(CF_3)-COO]$ - unit formed from the oxa-Michael addition (Scheme 6). The latter reaction was underdevelopped in the literature^{29,30} and therefore can be highlighted in this present study. The $-[C(CF_3)(COOH)-CH_2]$ - unit is simplified to (-C^oC-), as the fragment starts and ends by C atoms. The $-[CH_2-CH(CF_3)-COO]$ - unit is symbolised by (-C^oO-), as the fragment starts from a C atom and ends with an O atom.



Scheme 6: $-[C(CF_3)(COOH)-CH_2]-(-C^{\cap}C-)$ unit formed via C=C bond cleavage of MAF and $-[CH_2-CH(CF_3)-COO]-(-C^{\cap}O-)$ monomer unit generated via oxa-Michael addition.

The (-C^oC-) and (-C^oO-) units of polyMAF can be also differenciated by ¹³C NMR spectroscopy. Concerning (-C^oC-) unit, a previous article⁵¹ reports the alternating copolymerisation of vinyl acetate (VAc) with *tert*-butyl-2-trifluoromethacrylate (MAFTBE). The resulting poly(VAc-*alt*-MAFTBE) copolymers were hydrolyzed into NC-CH₂-[CH₂-CH(OH)-CH₂-C(CF₃)(COOH)]_n-SH. As a comparison, the ¹³C NMR spectrum (Figure 11C) displays an expected quartet (¹ J_{CF} = 280 Hz) centered at 127 ppm assigned to CF₃ a^3 , slightly low shield shifted compared to a, a^1 and a^2 . The quartet at 54.74 ppm (² J_{CF} = 30 Hz) is attributed to the quaternary carbon atom b^3 while COO c^3 is represented by a signal at 176.16 ppm. The broad signal centered in the 35.8–45.4 ppm range is assigned to -CH₂- d^3 and CH₂ in vinyl alcohol unit.



Figure 11 : ¹³C NMR spectra of MAF (top-recorded in CDCl₃) ; of diMAF (APT spectrum- middle-recorded in CDCl₃) and NC-CH₂-[CH₂-CH(OH)-CH₂-C(CF₃)(COOH)]_n-SH copolymer (bottom-recorded in D₂O, extracted from Polym. Chem. 2021, 12, 277).

The most important clue of the (-C^oO-) unit is the tertiary carbon -CH- b^2 at 49.35 ppm which is neither present in MAF nor in (-C^oC-) unit structures. The APT mode of ¹³C NMR spectrum allows to evidence -CH- signal, found in the opposite side of secondary and quaternary carbons signals. Additionnally, directly attached to a CF₃ group, it displays a quartet with ²J_{CF} \approx 30 Hz (Figure 11B). Indeed, the absence of signal at 55 ppm attributed to -CH₂-**C**(CF₃)(COOH)-CH₂- of the copolymer evidences the absence of radical homopolymerisation (Figure 11C).

5. Conclusion

This study reports the *oxa*-Michael oligomerisation of MAF in various conditions : in the absence of catalyst, in the presence of Brønsted acids, Brønsted and Lewis bases. The additives screening showed that the oligomerisation of MAF was not promoted in the radical way and confirms previous studies.^{19,20} Brønsted acids and bases led to oligomers with good MAF conversions but in low DP_n. Et₃N, in the Japanese patent,²⁹ does not seem to be the optimal reactant to get high DP_n. The reaction can be also assisted by triflic acid as Brønsted acid, which gives good MAF conversion (67%) and average DP_n (2.6). Mostly, Lewis acids do not favor the desired reaction. Particularly, the use of AlCl₃ allowed to reach a DP_n of 4.1 but simultaneously gave ClCH₂-CH(CF₃)COOH as a by-product. Brønsted bases such as hindered tertiary amines and phosphines led to good MAF conversions. The best results of this screening were obtained with PPh₃ and piperidine, similarly to the oxa-Michael

addition polymerisation of acrylic acid. The use of PPh₃ ((0.5 % at 95 °C or 5 % at 80 °C) yielded good MAF conversions (83-87 %) and DP_n (3.5) but the purification of phosphoniums is delicate because MAF removal process favors led to retro oxa-Michael reaction. The piperidine lead also to correct MAF conversion (76 %) and DP_n (3.5). This study might be developped in copolymerisation processes with diols, vinylsulfones or sulfonate derivatives.^{9,52,53}

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Conflict of Interets

The authors declare there is no conflict of interest.

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