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Lucie Cases, Pauline Adler, Franck Pelissier, Sébastien Diliberto, Clotilde Boulanger, et al.. New biomaterials for Ni biosorption turned into catalysts for Suzuki–Miyaura cross coupling of aryl iodides in green conditions. RSC Advances, 2021, 11 (45), pp.28085 - 28091. 10.1039/d1ra04478h. hal-03324149

HAL Id: hal-03324149

<https://hal.science/hal-03324149>

Submitted on 23 Aug 2021

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 Cite this: *RSC Adv.*, 2021, 11, 28085

New biomaterials for Ni biosorption turned into catalysts for Suzuki–Miyaura cross coupling of aryl iodides in green conditions†

 Lucie Cases,^a Pauline Adler,^a Franck Pelissier,^a Sébastien Diliberto,^b Clotilde Boulanger^b and Claude Grison^b   ^{*a}

In parallel with increasing Ni production and utilisation, Ni pollution in the soil–water continuum has become an alarming and global problem. Solutions for removing Ni from industrial effluents have been widely investigated and biosorption has emerged as an efficient, cost-effective, scalable and sustainable alternative for water treatment. However, the biosorption capacity is limited by the chemical composition of the biomaterial and the Ni-enriched biomaterials are rarely valorised. In this work, the biosorption capacity of three abundant biomaterials with different chemical properties – water hyacinth, coffee grounds and pinecones – was studied before and after functionalization, and reached a maximum biosorption capacity of 51 mg g⁻¹ of Ni(II). A bioinspired functionalization approach was investigated introducing carboxylate moieties and was conducted in green conditions. The Ni-enriched biomaterials were valorised by transformation into catalysts, which were characterised by MP-AES and XRPD. Their characterisation revealed a structure similar to nickel formate, and hence the Eco-Ni(HCOO)₂ catalysts were tested in Suzuki–Miyaura reactions. Several aryl iodides were successfully cross-coupled to phenylboronic acids using Eco-Ni(HCOO)₂ without any ligand, a mild and green base in a mixture of green solvents.

 Received 9th June 2021
 Accepted 2nd August 2021

DOI: 10.1039/d1ra04478h

rsc.li/rsc-advances

Introduction

Nickel is a transition metal, which is used in metallurgical processes, in electroplating, in the production of batteries and as a catalyst in fine chemistry. Within the last decade, its global mine production has almost doubled and reached 2.5 billion tons of nickel in 2020. The anthropogenic activities using Ni have led to the release of significant amounts of Ni and derivatives into the environment. Ni pollution of the soil–water continuum is serious since long-term exposure to high concentrations of Ni can cause dermatitis,¹ asthma,² cardiovascular and kidney diseases, lung fibrosis,³ lung and nasal cancers.⁴ Exposure to Ni pollution mainly concerns oral ingestion through water and food.⁵ Removal of Ni from industrial effluents before their discharge in the environment has become a priority. However, nickel is hardly extractable due to its strong affinity for aqueous media. Several processes have been

reported in the literature including chemical treatments, as chemical precipitation⁶ and electrocoagulation,⁷ membrane separation processes as ultrafiltration,⁸ reverse osmosis, electro dialysis⁹ and ion exchange methods.¹⁰ Although efficient, the industrialisation of the aforementioned processes is still limited, mostly due to the high cost, the secondary pollution and the generation of problematic sludge.

Biosorption has been emerging as an alternative and promising approach to water treatment. Biosorption relies on the adsorption of pollutants from wastewater using a biomaterial. This technique presents several advantages as the abundant availability in nature of its biomaterial, hence its low cost and the absence of a secondary pollution. Several types of biomaterials have been developed for the biosorption of Ni, from bacterial biomass,^{11,12} microalgae,¹³ tree bark,¹⁴ plant species¹⁵ to fruit peels.^{16,17} However, their biosorption capacity is limited by the chemical properties of the biomaterial. Moreover, as in every depollution process, the main drawback is the absence of valorisation of the depolluting membrane or biomaterial. In this paper, we propose to combine two strategies to overcome these limitations: the biosorption of Ni from wastewater using biomaterials modified in a bioinspired process and the utilisation of the Ni-enriched biomaterial as a catalyst in organic chemistry.

We have recently studied the ability of several aquatic plants to remove palladium from wastewater.¹⁸ The derived root

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† Electronic supplementary information (ESI) available: Materials and methods, IR spectra of biomaterials and Eco-Ni(HCOO)₂, description of products, XPS of Eco-Ni(HCOO)₂. See DOI: 10.1039/d1ra04478h



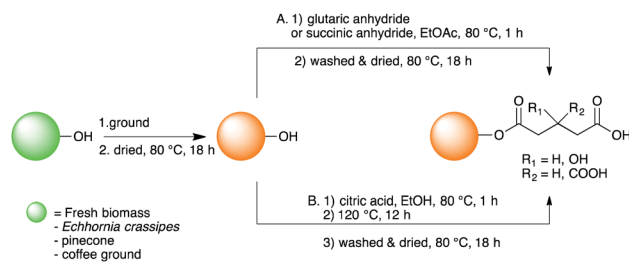
powders were able to adsorb high concentrations of Pd, following a passive process of biosorption. However, the affinity of a biomaterial to a transition metal depends on its chemical properties. Indeed, a few natural biomaterials were chemically modified to improve their biosorption capacity. Grafted copolymers introduced on orange peels and polyethylenimine crosslinked to glutaraldehyde itself crosslinked to biomass of *Penicillium chrysogenum* largely improved the maximum biosorption capacity of Ni from solution.^{11,17} However, these chemical methods of functionalization are far from being environmental-friendly processes.

In this paper, we first investigated a functionalization methodology to improve the affinity of three biomaterials, water hyacinth, pinecone and coffee ground, towards Ni salts. The functionalization methodology was designed and conducted in green conditions. We then studied their biosorption capacity to remove Ni from wastewater. And we finally studied the recovery of the Ni-enriched biomaterials and developed its utilisation as a catalyst in green and sustainable chemistry. The example of Ni-catalysed Suzuki–Miyaura reaction was chosen to illustrate the valorisation of the biomaterial.

Results and discussion

Biosorption of Ni using functionalized biomass feedstock

Biosorption of nickel from aqueous solution was studied in batch using three biomaterials with different chemical properties: roots of an aquatic plant, *Echhornia crassipes* (water hyacinth) rich in carboxylate functions, coffee grounds rich in tannins and pinecones rich in lignin. The three biomaterials were ground, dried and put into solution containing increasing concentrations of nickel sulfate (12, 40 mg L⁻¹). Although presenting high biosorption efficiency,^{13–16,19} the three biomaterials rapidly showed a limited biosorption efficiency of around 36% for a solution of 40 mg L⁻¹ of NiSO₄ (Table 1 entries 1–6).



Scheme 1 Functionalisation of biomass using (A). glutaric anhydride or succinic anhydride or using (B). citric acid for biosorption of Ni.

Functionalization of the biomaterials was hence envisaged in order to increase their Ni biosorption performances.

Our functionalization methodology was inspired from the natural mechanism of Ni storage in Ni-hyperaccumulating plant shoots, in which Ni is stored as nickel carboxylate (citrate, malate and malonate).²⁰ Two strategies of functionalization, both based on the introduction of carboxylate moieties onto the hydroxyl groups of the biomaterials, were investigated (Scheme 1). Strategy A relies on the direct esterification of hydroxyl groups using carboxylic anhydride (succinic or glutaric anhydride). Ethyl acetate was the solvent chosen for this strategy since it does not decompose the anhydrides unlike a protic solvent. Strategy B relies on the auto-catalysed esterification of hydroxyl groups by citric acid. Unlike strategy A, ethyl acetate could not be used due to the poor solubility of citric acid in organic solvents, and water would deactivate the reaction, so ethanol was chosen. Both strategies were conducted in green conditions, using biosourced reagents (the biomass and the carboxylic acid or anhydrides) in a green solvent (ethyl acetate or ethanol).

Following the functionalization, the functionalised biomaterials were purified to remove the excess of carboxylic acids and were characterised by infrared spectroscopy (Table 2 and Fig.S1†). Infrared analyses showed new bands of higher

Table 1 Properties of the new biomaterials towards biosorption of Ni in solution

Entry	Starting biomaterial	Functionalisation reagent	Name of biomaterial	Carboxylate (mmol g ⁻¹ of material)	Solution to biosorb (mg per L of metal per g of material)	Biosorption efficiency (%)
1	<i>Echhornia crassipes</i>	—	Ec	—	12 mg L ⁻¹ of NiSO ₄	62
2	Pinecone	—	Pn	—	12 mg L ⁻¹ of NiSO ₄	62
3	Coffee ground	—	Cg	—	12 mg L ⁻¹ of NiSO ₄	30
4	<i>Echhornia crassipes</i>	—	Ec	—	40 mg L ⁻¹ of NiSO ₄	36
5	Pinecone	—	Pn	—	40 mg L ⁻¹ of NiSO ₄	16
6	Coffee ground	—	Cg	—	40 mg L ⁻¹ of NiSO ₄	17
7	<i>Echhornia crassipes</i>	Succinic anhydride	Ec-SA	1.4	40 mg L ⁻¹ of NiSO ₄	99
8	Pinecone	Succinic anhydride	Pn-SA	1.1	40 mg L ⁻¹ of NiSO ₄	78
9	Coffee ground	Succinic anhydride	Cg-SA	1.4	40 mg L ⁻¹ of NiSO ₄	80
10	<i>Echhornia crassipes</i>	Glutaric anhydride	Ec-GA	0.7	40 mg L ⁻¹ of NiSO ₄	64
11	<i>Echhornia crassipes</i>	Citric acid	Ec-CA	1.4	40 mg L ⁻¹ of NiSO ₄	89
12	Pinecone	Citric acid	Pn-CA	1.6	40 mg L ⁻¹ of NiSO ₄	70
13	Coffee ground	Citric acid	Cg-CA	1.8	40 mg L ⁻¹ of NiSO ₄	99
Maximum of biosorption						
14	<i>Echhornia crassipes</i>	Succinic anhydride	Ec-SA	1.4	1024 mg L ⁻¹ of NiSO ₄	51 mg g ⁻¹



Table 2 Infrared analysis of the dried materials and the citric acid functionalised materials^a

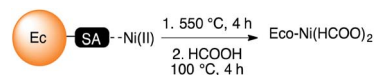
Entry	Biomaterial	C=O ester (cm ⁻¹)
1	Ec	None
2	Ec-CA	1722 (s)
3	Pn	1738 (w)
4	Pn-CA	1726 (s)
5	Cg	1730 (vw)
6	Cg-CA	1718 (m)

^a s: strong; m: moderate; w: weak; vw: very weak.

percentage of transmission in the 1720–1740 cm⁻¹ region corresponding to the stretching vibration of the ester group, which was formed during the functionalization process.

The efficacy of the functionalization strategies was compared by titrating the introduced carboxylate moieties using a solution of sodium hydroxide (Table 1, entries 7–13). Both strategies A and B led to similar molar quantities of carboxylates per gram of biomaterial, except when glutaric anhydride was used, the functionalization was clearly less efficient and was therefore not pursued (Table 1, entry 10).

The functionalised biomaterials were then used to test whether functionalization had improved their biosorption capacity of nickel in batch (Table 1, entries 7–13). They were put into solutions of 40 mg L⁻¹ of nickel sulfate, corresponding to the highest concentration at which the non-functionalised biomass was saturated with. In general, the functionalised biomass showed one-third to twice more biosorption capacity than the non-modified biomass (Table 1, entries 7–9 & 11–13), except for Ec-GA (Table 1, entry 10). Indeed the biosorption capacity of Ec-GA is similar to the one of Ec, which can be related to the poor efficiency of functionalization using glutaric acid and can be seen as a negative control of this experiment. It is noteworthy that Ec-SA and Cg-CA removed Ni in totality from the concentrated aqueous solution. The maximum capacity of biosorption of Ec-SA was hence tested using a saturating solution of 1024 mg L⁻¹ of nickel sulfate. Ec-SA was able to adsorb up to 51 mg g⁻¹ of Ni, which is higher than non-modified natural materials^{12,14–16,19} and in the same range as materials that have been modified in eco-unfriendly conditions (Table 3).^{11,17} These new biomaterials constitute novel, efficient and sustainable solutions to treat heavy metal polluted effluents.

**Scheme 2** Preparation of Eco-Ni(HCOO)₂ catalyst. Ec-SA stands for *Echhornia crassipes* biomaterial functionalised with succinic anhydride.

Preparation and analyses of the Eco-Ni(HCOO)₂ catalyst

One functionalised biomaterial, Ec-SA, was transformed into a catalyst for Suzuki–Miyaura coupling reactions (Scheme 2). Ec-SA was heated at 500 °C for 4 h to give an inorganic powder of nickel oxides that was treated with formic acid, a biosourced reducing reagent, leading to the Eco-Ni(HCOO)₂ catalyst.

The composition of Eco-Ni(HCOO)₂ catalyst was determined by Microwave Plasma Atomic Emission Spectroscopy (MP-AES). As expected from the biosorption experiments, the Eco-Ni(HCOO)₂ catalyst presented a high mass fraction of Ni, about 30 wt% (Table 4). The high concentration in Ni was promising to promote organic catalysis.

The Eco-Ni(HCOO)₂ catalyst was characterised by XRPD experiments (black curve, Fig. 1). In parallel, XRPD experiment was also conducted on synthetic and non-biosourced Ni(HCOO)₂ for comparison (blue curve, Fig. 1). Although more complex, the diffraction pattern of Eco-Ni(HCOO)₂ shows similar peaks, as the diffraction pattern of the non-biosourced Ni(HCOO)₂, that is highlighted by the green bars corresponding to the theoretical family of peaks of Ni(HCOO)₂. A second family of peaks of Eco-Ni(HCOO)₂ was also identified to CaSO₄, as shown by the red bars in comparison to database. This salt in biosourced Ni(HCOO)₂ was of plant origin.

FT-IR and XPS analyses were conducted on the ecocatalyst (Fig. S2 and S17[†]). Eco-Ni(HCOO)₂. The spectra were added to the ESI.[†] The IR spectrum shows absorption bands at 1580 cm⁻¹ and 1325 cm⁻¹ that are characteristic of the carboxylate group of Eco-Ni(HCOO)₂.

In XPS, The Ni 2p_{3/2} binding energy (BE) of 857.8 eV, indicated presence of Ni(II). In conclusion, MP-AES, XRDP, XPS and FT-IR confirmed the generation of Ni(HCOO)₂.

Suzuki–Miyaura reactions of aryl iodides

The Suzuki–Miyaura reaction (SMR) has arguably become one of the most utilised tools for the construction of C–C bonds and has found academic use and industrial applications for the

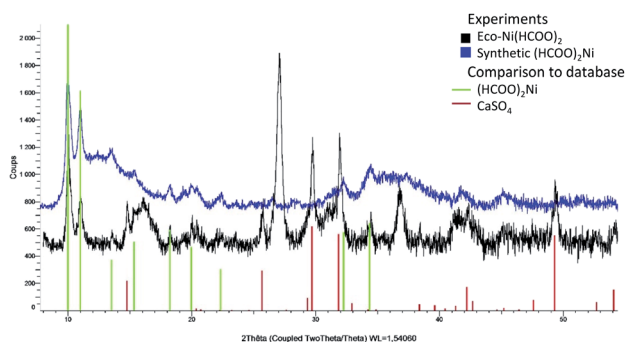
Table 3 Maximum capacity of biosorption of non-modified natural and materials modified in eco-unfriendly conditions

Biomaterial	C _{max} Ni (mg g ⁻¹)	References
Algae	13–50	14
Lichen (<i>Cladonia furcata</i>)	7.9	19
Biomass of <i>Penicillium chrysogenum</i> modified with polyethylenimine (PEI) and then crosslinked with glutaraldehyde	55	11
Aloe barbadensis Miller modified by Na ₂ CO ₃	29	15
Lemon peel modified by NaOH	36	16
Ec-SA	51	This work



Table 4 Mineral composition of the catalyst Eco-Ni(HCOO)₂ determined by MP-AES

Catalyst	Mineral composition (wt% ± rsd)			
	Ni	Na	Mg	Ca
Eco-Ni(HCOO) ₂	31.2 ± 2.05	0.88 ± 0.29	0.10 ± 0.31	2.47 ± 1.06

**Fig. 1** XPRD analyses of Eco-Ni(HCOO)₂ (black curve) and of synthetic non-biosourced Ni(HCOO)₂ (blue curve) and comparison to database (red and green bars).

production of polymers, fine chemicals, materials and pharmaceuticals. SMR is generally defined as the transition-metal-catalysed cross coupling between an organoboron reagent and an organic halide in the presence of a catalyst and a base. Diverse catalytic systems were developed for SMR, but Pd-based catalysts remained the most efficient and hence compatible with mild conditions.²¹ However, the depletion of Pd resources in parallel of its geo-economic context have risen the price of Pd to its highest records. Nickel-catalysed SMR has been recently emerging as an interesting and cost-effective alternative.²² Indeed Ni is more abundant and on average ten times cheaper than palladium. However, nickel possesses distinctive catalytic properties than palladium and nickel-catalysed SMR reactions are usually conducted in less mild conditions. Therefore nickel-catalysed SMR deserve the attention for developing greener strategies.²³

The activity of Eco-Ni(HCOO)₂ catalyst was tested in SMR using several aryl iodides and aryl boronic acids (Table 4). Following the principles of green chemistry, not only the catalyst was biosourced but every component of the reactions was considered. Since Eco-Ni(HCOO)₂ presents a degree of oxidation of Ni(II) (Fig. S17†), it should be reduced *in situ* to Ni(0) for SMR. A temperature of 380 °C is necessary to induce the thermal decomposition of formate nickel into Ni(0),²⁴ but is not compatible with SMR. A hydrogen donating solvent, glycerol, was therefore chosen.

The conditions of the coupling reaction were first determined using *p*-iodoacetophenone and phenylboronic acid (entry 1, Table 4). The reaction led to a quantitative conversion and an excellent yield, using 5 mol% of the bio-sourced Eco-Ni(HCOO)₂ catalyst without any ligand. Moreover, the reaction

was conducted in eco-friendly conditions using a mild and green base, potassium carbonate, in a mixture of green solvents, glycerol and *n*-butanol at a moderate temperature.

The scope of the SMR was expanded to phenylboronic acids bearing electron-withdrawing groups (entries 2 & 3, Table 5). The trifluoromethyl group led to similar conversion and yield while the nitro group gave poor conversion, probably due to the low stability of the *p*-nitrophenylboronic acid.

Diversely *para* substituted iodobenzene compounds were then tested (entries 4–8, Table 5). *p*-Nitroiodobenzene led to excellent conversion and yield with no trace of the reduction of the nitro group, while *p*-iodobenzonitrile mostly led to the reduction of the nitrile group into *p*-iodobenzylamine.

The presence of an electron-donating group, as a fluor or a methyl, led to reduced yields since the C–I bond was reduced instead. The use of iodobenzene with a heterocyclic compound, as 3-pyridineboronic acid, gave an excellent conversion and yield. It is noteworthy that the presence of pyridine derivative did not deactivate the Eco-Ni(HCOO)₂ catalyst, especially since the reaction was conducted without any ligand.

Although a few investigations have been made for Pd-catalysed SMR carried out in green conditions, a few examples are found for Ni-catalysed SMR, illustrating needs in this area.

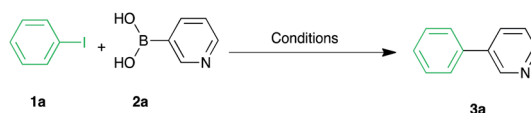
The coupling conditions between iodobenzene and 3-pyridineboronic acid were compared to Ni-catalysed SMR from literature (Table 5). For this specific reaction, the Ni catalyst is always coordinated to complex ligands, a triazine-based Ni(II) PNP pincer complex²⁵ or graphene oxide crosslinked nickel

Table 5 Scope of the Suzuki–Miyaura coupling reactions using Eco-Ni(HCOO)₂ catalyst

Entry	Product 3	Yield ^a (conv.)
1		83% (99%)
2		73% (98%)
3		13% (30%)
4		83% (89%)
5		6% (81%)
6		43% (71%)
7		18% (90%)
8		89% (99%)

^a Yields were quantified by GC FID with *p*-cymene (0.2 eq.) as an internal standard and then confirmed by ¹H NMR.



Table 6 Comparison of the coupling reaction between iodobenzene and 3-pyridineboronic acid with Eco-Ni(HCOO)₂ and literature

Entry	Conditions	Yield (conv.)
1 (ref. 25)	[Ni(II) PNP pincer complex] (2 mol%) <i>t</i> BuOK, toluene 180 °C, 16 h	85%
2 (ref. 26)	[GO/NiTAPP] (3 mol%) K ₃ PO ₄ (2 eq.), dioxane 80 °C, 1.5 h	92%
3	Eco-Ni(HCOO) ₂ (5 mol%) Na ₂ CO ₃ (2 eq.), glycerol/ <i>n</i> -BuOH 100 °C, 18 h	89% (99%)

5,10,15,20-tetrakis(aminophenyl)porphyrin complex.²⁶ However, the Eco-Ni(HCOO)₂ catalyst exhibits similar efficiency and is ligand-free. Moreover, hazardous solvents, dioxane and toluene, were used in the two cited reactions, while a mixture of green solvents was used here. In general, mild conditions should be preferred if applicable as using a mild and inorganic base, and a moderate temperature (Table 6, entries 2 & 3).

Conclusions

In this work, new eco-friendly modified biomaterials were found for removing efficiently Ni from aqueous solutions. A functionalization method for introducing carboxylate moieties was developed using biosourced reagent in green conditions. Water hyacinth and coffee ground functionalised with succinic anhydride led to the best biosorption capacity and could remove Ni totally from a 40 mg L⁻¹ solution of nickel sulfate. The maximum biosorption capacity reached 51 mg g⁻¹ of Ni(II) for water hyacinth functionalized with succinic anhydride.

The Ni-enriched biomaterials post biosorption were valorised as an efficient catalyst of Suzuki–Miyaura reactions, which were conducted in green conditions. Several biaryl compounds were obtained in good yields using a biosourced cocatalyst, no ligand, a mild base, in a mixture of green solvents at a moderate temperature.

Experimental

Materials and methods

All chemicals and solvents were purchased from commercial sources. Plants of *Eichhornia crassipes* were purchased from a specialized grower (Nymphaea company, France). Bark pine and pinecone were collected from black pine trees, *Pinus nigra*, in the region of Montpellier in autumn 2018. Coffee grounds were obtained directly from our laboratory.

The samples for MP-AES analyses were digested in 10 mL of reversed aqua regia (1 : 2 hydrochloric acid (37%); nitric acid (65%)) under a microwave-assisted digestion (Multiwave-Go Anton Paar) with the following program: 20–165 °C in 20 min and then 10 min isothermal at 165 °C. Samples were filtered and then diluted to 0.4 mg L⁻¹ in 1% aqueous nitric acid. Mineral compositions were determined by using a microwave plasma-atomic emission spectroscopy (MP-AES) 4200 (Agilent

Technologies) equipped with a concentric nebulizer and a double-pass cyclonic spray chamber. The pump speed during analysis was kept at 10 rpm and the sample introduction tube diameter was 0.89 mm. The analytical cycle consisted of 30 s rinsing with aq. 1% nitric acid followed by 25 s of sample uptake (pump speed 40 rpm) and then 20 s of equilibration before the reading at preselected integration times (pump speed 10 rpm). The integration time was set to 3 s for all elements. Unless otherwise stated, the automatic background correction mode available in the software was used. An Agilent SPS3 autosampler was used throughout the study.

Gas chromatography analyses were performed using a Thermo Scientific Trace 1300 device equipped with an EI ionization source and an ISQ-QD detector. The reactions were monitored by using *para*-cymene as internal standard and FID methods for the calibration.

IR spectra were recorded on a PerkinElmer Spectrum 100 FTIR® spectrometer in ATR (Attenuated Total Reflexion) mode. The number of scans was 32, the resolution was 1 point per cm. The acquisition was done from 650 to 4000 cm⁻¹. The detector used was a DTGS (Deuterated-TriGlycine Sulfate). The background was done in air.

NMR spectra were recorded on a Brücker Avance 400 spectrometer at room temperature. ¹H frequency was at 400 MHz and ¹³C frequency was at 100 MHz. ¹H and ¹³C NMR data match that reported in the literature.

Procedure for preparation of the biomaterial

Eichhornia crassipes roots, bark pines and pinecones were dried in an oven at 80 °C for 8 hours. They were then ground, sifted through a 1.25 mesh sieves, and washed with water (3 × 100 mL g⁻¹). They were then dried for 18 hours at 80 °C before functionalization. Coffee grounds were washed several times with hot water until the filtrate became colourless and then were dried for 18 hours at 80 °C before functionalization.

General procedure for functionalization

Esterification using succinic or glutaric anhydride. Succinic anhydride (3.2 g, 32 mmol) or glutaric anhydride (3.6 g, 32 mmol) was solubilised in ethyl acetate (1 mL mmol⁻¹) and the solution was heated to reflux. The biomaterial (5 g) was added to the reaction mixture and the suspension was stirred for 1 hour



under reflux. Distilled water (50 mL) was then added to the suspension, which was stirred at room temperature for 15 minutes. The solid was filtered and washed several times with distilled water (15 minutes of stirring per washing) until pH was neutral. If necessary, an aqueous solution of NaOH (2 M) was added dropwise until pH was neutral. The functionalised biomaterial was filtered and washed with distilled water before being dried at 85 °C.

Autocatalyzed esterification using citric acid. 5 g of biomaterial was added to a solution of citric acid (4 g, 21 mmol) in anhydrous ethanol (20 mL). The mixture was refluxed for one hour. The solvent was removed under reduced pressure and the solid was stored at 120 °C for 12 hours. 50 mL of distilled water was then added to the solid. The suspension was stirred at room temperature for 15 minutes. The solid was filtered and washed several times with distilled water (15 minutes of stirring per washing) until pH was neutral. The solid was stored at 85 °C for 12 hours. Then to the solid was added 100 mL of distilled water. An aqueous solution of NaOH (2 M) was added dropwise until pH was neutral. Solid was filtered and washed with distilled water before being dried at 85 °C.

General procedure for biosorption

1 g of functionalised biomaterial was added to a solution of nickel sulfate (1 L of 12, 40 and 1024 mg L⁻¹). The suspension was stirred for 2 hours at room temperature. The solid was filtered and dried at 85 °C for 24 hours. The concentrations of nickel in solution were determined by MP-AES before and after biosorption.

Preparation of the catalyst Eco-Ni(HCOO)₂

The Ni-enriched biomaterial (Ec-CA-Ni) was heated at 550 °C under air for 4 hours. The resulting powder (243 mg) was heated in formic acid (7 mL) under reflux for 4 hours. The resulting blue-green suspension was cooled to room temperature, then filtrated and washed with 15 mL of formic acid. The resulting solid was rinsed with distilled water in order to dissolve the ecocatalyst. The resulting bright green solution was collected by filtration, then concentrated under reduced pressure, affording Eco-Ni(II) formate as a light green solid (160 mg).

General procedure for Suzuki–Miyaura coupling reaction

To a solution of aryl iodide (0.5 mmol, 1 eq.), boronic acid (0.6 mmol, 1.2 eq.) and potassium carbonate (1 mmol, 2 eq.) in a mixture of degassed *n*-butanol and glycerol (v/v = 1 : 1, 1 mL) was added the catalyst Eco-Ni(HCOO)₂ (5% mol). The suspension mixture was then stirred at 100 °C for 18 hours. The reaction was quenched by adding distilled water (5 mL) and ethyl acetate (10 mL). After separation of the layers, the aqueous layer was extracted 3 times with 10 mL of ethyl acetate. The combined organic layers were washed with a saturated solution of Eco-CaO (thermal treatment of *Crepidula fornicata*)²⁷ and then dried on anhydrous magnesium sulfate and filtered. The solvent was removed under reduced pressure conversions and yields were established by GC MS/FID, using *p*-cymene as intern standard, and then confirmed by ¹H NMR and ¹³C NMR.

Author contributions

C. G. conceived and designed the study. F. P. carried out the MP AES assessments and L. C. and P. A. carried out the experimental works of biosorption and organic synthesis. S. D. and C. B. performed XRD analysis. C. G. wrote the paper.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors would like to thank the French National Center for Scientific Research (CNRS) and the University of Montpellier for financial support. Sci-GuidEdit is acknowledged for writing assistance and proof reading the article.

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