



## **Assessing extracted organic matter quality from river sediments by elemental and molecular characterization: Application to the Tietê and Piracicaba Rivers (São**

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**Assessing extracted organic matter quality from river sediments by elemental and  
molecular characterization: Application to the Tietê and Piracicaba Rivers (São  
Paulo, Brazil)**

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27 **Abstract**

28 Urban river pollution causes serious problems to the environment, human health, and  
29 water scarcity. Developing tools to identify and assess the health of aquatic river  
30 systems is essential for monitoring the quality of rivers and implementing actions. This  
31 study assesses the elemental and molecular characteristics of organic matter (OM)  
32 extracted from river sediments and associates them with river quality. To assess the  
33 quality of sediment cores, the most reactive and available OM from two urban rivers  
34 (Tietê and Piracicaba Rivers, São Paulo, Brazil) was isolated with alkaline and water  
35 solutions. It was then characterized by elemental composition, ultraviolet-visible  
36 spectroscopy, and fluorescence excitation-emission matrix with parallel factor analysis.  
37 The average yield of alkaline extraction was  $40.71\% \pm 5.52\%$  of OM present in the bulk  
38 sediments. Extracted organic matter from sediments (EOMSed) from the Tietê River  
39 presented the highest average concentrations of non-purgeable organic carbon (NPOC)  
40 and total organic nitrogen (TON), and the lowest average NPOC/TON molar ratios and  
41 specific UV absorbance at 254 nm ( $SUVA_{254}$ ). Considering the high degree of  
42 eutrophication in the Tietê River, these results suggest a greater input of simple OM  
43 with nitrogenous structures. The humic-like component in EOMSed was the most  
44 abundant in both rivers. The aromaticity of EOMSed from the Piracicaba River was  
45 evidenced by the greater contribution of complex structures in the form of aromatic and  
46 polyaromatic moieties and higher  $SUVA_{254}$ . EOMSed from the Tietê River also featured  
47 enhanced biological activity due to the greater contribution of microbial-derived  
48 products and the presence of small molecules and nitrogenous structures. As this  
49 combination of elemental and spectroscopic techniques successfully identified the

characteristics of extracted OM, it can be used as a tool to assess the global river quality.

**Keywords:** River sediments, Extracted organic matter, Fluorescence, Parallel factor analysis.

## 1. Introduction

Sediments consist of particle accumulation with different physical and chemical properties that are composed of inorganic and organic material. The particles are transported by water, air, erosion, and weathering processes and distributed along rivers (Murdoch and MacKnight, 1991; Schorer and Eisele, 1997). The sediments are biogeochemically active due to the presence of many microbes and the continuous input of organic and inorganic substances (He et al., 2019; Kumwimba et al., 2017).

Organic matter (OM) in the sediments can originate from external sources (allochthonous) or from the aquatic ecosystem itself (autochthonous) (Brailsford et al., 2019; Kindler et al., 2011). Moreover, rivers are responsible for transferring carbon (C) between the terrestrial biosphere and oceans, where this element is transformed and stored (Cole et al., 2007). In sediments, bacteria consume or decompose organic particles, thus reducing the mass of OM (Bloesch, 2009). The organic material resistant to microbial action and any remaining products are deposited on the bottom of rivers, along with terrestrial sediments (Burdige, 2007; Henrichs, 1992). OM is then remineralized, altered, or preserved during early diagenesis, which records input history (Engel and Macko, 2013).

The mobilization and quality of sediments are partially affected by anthropogenic actions that cause extensive changes in the aquatic environment (Bloesch, 2009). The excessive load of nutrients leads to the eutrophication of rivers,

which is a process of increasing the OM load in an aquatic ecosystem (Nixon, 1995). In areas with a high population density, the main sources of nutrients are domestic sewage discharge, industrial effluent, and water drained from agricultural areas (Kubo and Kanda, 2020; Smith et al., 2014). Sediments with high OM content promote oxygen consumption through decomposition, which can harm aquatic animals and plants (Duarte, 1995).

Barra Bonita Reservoir in São Paulo State, Brazil, is formed by the damming of the Tietê and Piracicaba Rivers, and it is the first of six reservoirs of the Middle Tietê River Basin (Rodrigues et al., 2020). The Tietê and Piracicaba basins have different water resource management strategies, with different types of land use and occupation (Cruz et al., 2017). Both rivers receive a high degree of organic compounds, although the Tietê River is more eutrophic than the Piracicaba River, because it receives a large load of industrial and domestic waste (dos Santos et al., 2006; Morais et al., 2021; Rodrigues et al., 2020). The Middle Tietê River had a biochemical oxygen demand (BOD) of 33,636 t day<sup>-1</sup> in 2017 (CETESB, 2018). Before reaching the reservoir, the Tietê River passes through the metropolitan area of São Paulo (Upper Tietê River), which has a high population density and presented a BOD of 612,069 t day<sup>-1</sup>. This represents 57% of BOD of the Tietê River. By comparison, the Piracicaba Basin had a BOD of 94,818 t day<sup>-1</sup> (CETESB, 2018). According to the State of São Paulo Environmental Company (CETESB), the courses of both aquatic bodies are situated in industrial areas marked by a high demographic density and intense land use (CETESB, 2018). However, regarding the course of the Middle Tietê River, its non-compliance with water-quality standards stands at around 100% due to the untreated sewage discharge in this stretch of the Tietê River (CETESB, 2018).

99           In the current scenario of scarce freshwater resources in terms of both the quality  
100 and quantity available for use, it is urgent to restore water bodies. One way to do so is to  
101 focus on assessing OM quality in sediments. Furthermore, finding the tools to identify  
102 and assess the health of aquatic river systems is essential for the implementation of  
103 actions. Molar C to nitrogen (N) ratios can be useful to identify the origin of OM  
104 (Burone et al., 2003; Meyers, 1994), whereas determining the optical properties of OM  
105 present in sediments is also a rapid and efficient way to assess the quality of water  
106 resources and obtain information about anthropogenic activities (Du et al., 2021). OM is  
107 a small fraction of the sediment compared to the inorganic fraction. The extraction is  
108 carried out to isolate part of the OM and obtain information about its origin and quality.  
109 For this purpose, water or basic extractants have shown their potential for determining  
110 the quality and origin of OM in sediments (Brym et al., 2014; Funkey et al., 2019). The  
111 neutral aqueous medium allows for the extraction of polar substances of small  
112 molecular weight, which are free from the mineral fraction in the sediments. Water  
113 alkaline extraction favors the extraction of more polar or reactive substances, even  
114 allowing for the extraction of substances linked to minerals (Olk et al., 2019). After the  
115 extraction with neutral and basic water, there remains only an insoluble organic fraction  
116 that is strongly linked to minerals. This can be considered a stable or less reactive OM,  
117 which will hardly influence the chemical processes in the sediment (Guigue et al., 2014;  
118 Muller et al., 2014; Olk et al., 2019).

119           To assess the sediment quality, the most available and reactive OM was isolated  
120 with neutral and alkaline water solutions. It was then fully characterized by elemental  
121 composition, ultraviolet-visible spectroscopy (UV-Vis), and fluorescence excitation-  
122 emission matrix (EEM) processed with parallel factor analysis (PARAFAC). This study  
123 aimed to evaluate the elemental and molecular features of the extractable OM of

sediments taken from the Piracicaba and Tietê Rivers and associate this information with the global river quality.

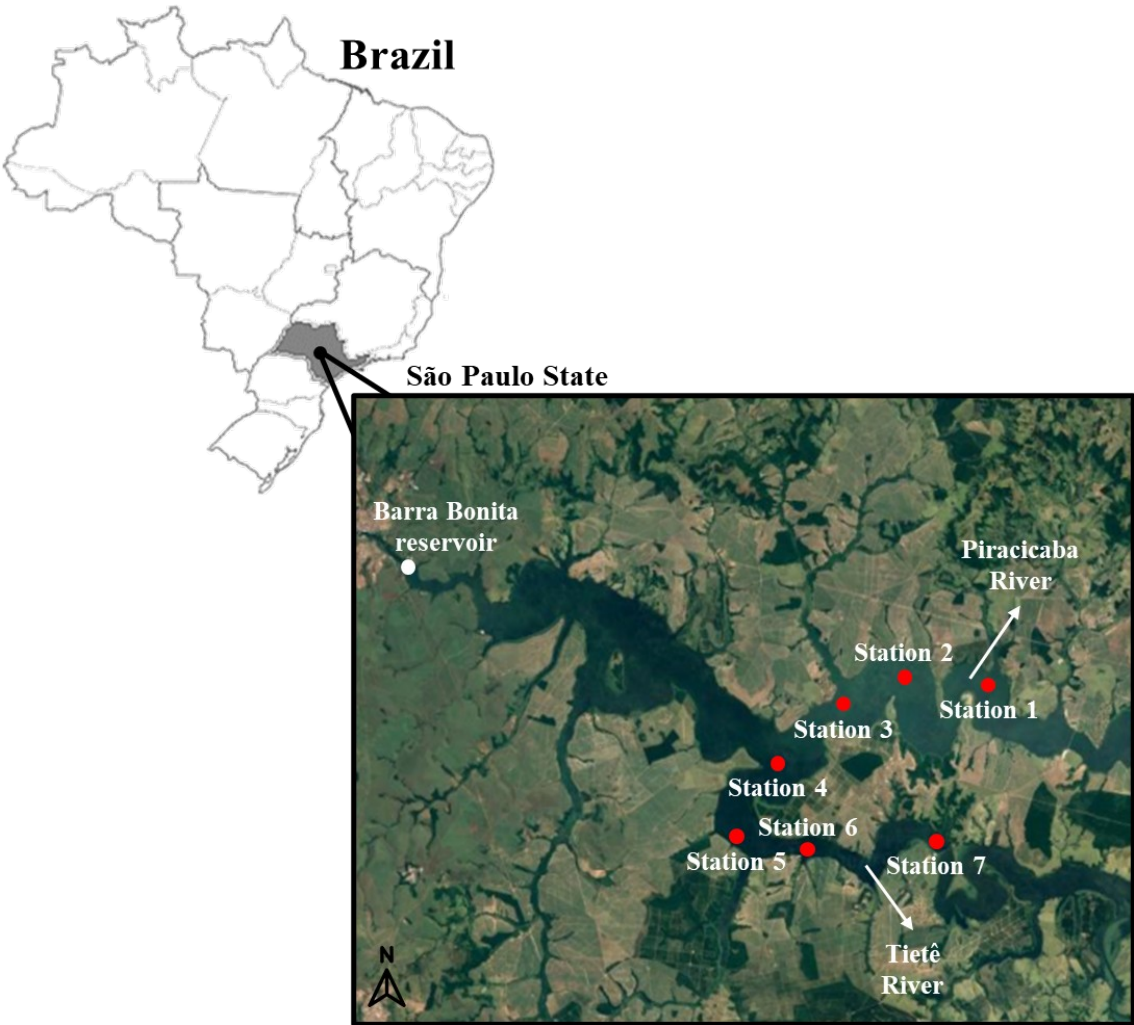
## **2. Materials and methods**

### *2.1. Study area description and sediment sampling*

The study area covers the Tietê and Piracicaba Rivers upstream from the Barra Bonita Reservoir. The Barra Bonita Reservoir, located between the cities of Barra Bonita and Igaraçu do Tietê, was created from the flooding and damming of these rivers (Bernardo et al., 2016). The Tietê and Piracicaba Rivers pass through large urban centers until their confluence. Both rivers receive a large load of organic and inorganic pollutants, which are delivered to the reservoir, although a greater load of pollutants is dumped into the Tietê River (Smith et al., 2014). The study area was chosen due to the different uses and occupations of the soil in this region, characterized by pollution issues and multiple uses of water. Furthermore, this region has suffered from many anthropic actions, mainly due to deforestation and flooding for the construction of six successive reservoirs, with the Barra Bonita Reservoir being the first (Rodrigues et al., 2020).

The sediment cores were collected at seven points: three on the Piracicaba River (stations 1, 2, and 3), one at the confluence region (station 4), and three on the Tietê River (stations 5, 6, and 7) (Figure 1) (Moraes et al., 2021). The collection points were georeferenced using a GPS Trimble Navigation unit as shown in the supplementary material (Table S1); the sampling took place on July 25, 2017 (austral winter). The sediment samples were collected using core sampling, which allowed us to maintain the original deposition of sedimentary layers. The horizontal slicing was performed at the same time as the sample collection, in different sizes (Table S2) under normal

atmosphere along the sediment cores without removing the interstitial water, thus totaling 69 sample layers. The sediment samples were immediately packed into polypropylene flasks and kept in a thermal box during transport to the laboratory to avoid the oxidation effect.



**Figure 1:** Location of the Barra Bonita Reservoir (white dot) and sediment sample collection stations (red dots).

## 2.2. Elemental analysis

The sediment samples were frozen, freeze-dried (L101, Liotop), crushed using an automatic mortar grinder (RM 200, Retsch), and sieved through a 100-mesh sieve. Total carbon (TC) content of the sediment samples was assessed using the dry



combustion method. For this purpose, 20 mg of the homogenized samples were accurately weighed directly in the tin capsules. The standard used in the analysis was aspartic acid, and the method was validated using the soil certified reference material (ThermoFisher Soil Reference Material NC PN338 40025). All measurements were performed in an elemental analyzer (Flash 2000, ThermoFisher Scientific).

### *2.3. Extractable organic matter from sediment samples*

#### *2.3.1. Extraction*

The alkaline extraction was performed with NaOH to isolate OM in the sediment samples taken from the Piracicaba and Tietê Rivers. The sediment extracts were denominated EOMSed (extracted organic matter from sediments). Approximately 1.0 g of each sample was placed in polypropylene flasks with 45.0 mL of 0.1 mol L<sup>-1</sup> NaOH (ThermoFisher Scientific) and shaken for 24 h in an overhead shaker (Rex 20, Heidolph) at 10 rpm. The samples were then centrifuged (Sigma 3-18K, Grosseron) at 10,000 rpm for 10 min and filtered over 0.45 µm syringe filters (Minisart NML, Sartorius). Furthermore, another extraction method was conducted in the same way as previously described with 45.0 mL ultrapure water (Ultrapure Water Cell, Starna Ltd) in which the extracts from sediments were denominated WEOMSed (water-extracted organic matter from sediments).

#### *2.3.2. Non-purgeable organic carbon and total organic nitrogen determination*

Non-purgeable organic carbon (NPOC) and total organic nitrogen (TON) from EOMSed were quantified after dilution with ultrapure water to fit the standard NPOC calibration curve (maximum 30 mg L<sup>-1</sup> of C). The extracts were acidified and measured using an elemental analyzer (TOC-V Shimadzu). The NPOC/TON molar ratios were

calculated. The extraction yield was obtained by dividing the quantity of organic carbon (OC) in the extracted solution in mg (NPOC multiplied by the extraction volume) by the carbon content in the corresponding sediment in mg (TC multiplied by extracted mass sample).

### 2.3.3. *UV-Vis spectroscopy*

Absorbance measurements were performed for all EOMSed and WEOMSed samples using a double beam UV-Vis spectrophotometer (UV 1800, Shimadzu) with a wavelength range from 200 to 700 nm in a 1 cm path length quartz cell and a scan speed of 300 nm min<sup>-1</sup>. A reference cell of NaOH and deionized water was used to measure EOMSed and WEOMSed, respectively. From the absorbance measurements, it was possible to define the dilution factor for fluorescence. Thus, all samples had the absorbance adjusted to 0.1 at 254 nm, thus allowing their comparisons by avoiding the inner filter effect for the fluorescence analysis (Kothawala et al., 2013). Specific UV absorbance at 254 nm (SUVA<sub>254</sub>) was calculated by dividing the UV absorbance measured at 254 nm by the NPOC concentration (mg of C L<sup>-1</sup>) and the optical path of the cuvette (0.01 m) (Weishaar et al., 2003).

### 2.3.4. *Fluorescence spectroscopy*

#### 2.3.4.1. *Sample preparation and analysis*

For the study of EEM, 1.0 mL of each sample diluted to absorbance at 254 nm equal 0.1 (1 cm path length) was placed in quartz cells along with 1.0 mL of 0.3 mol L<sup>-1</sup> HEPES (Acros Organics) for pH regulation at 7 and 1.5 mL of 0.1 mol L<sup>-1</sup> NaClO<sub>4</sub> (Sigma-Aldrich) with a purity of 99.99% to fix ionic strength. The samples were analyzed immediately after preparation.

All measured spectra were obtained with a fluorescence spectrofluorometer (F4500, Hitachi). The EEM spectra were acquired with an excitation range from 200 to 500 nm and an emission range from 250 to 700 nm at a scan speed of 2400 nm min<sup>-1</sup>. The steps and slits of emission and excitation were fixed at 5 nm, and the detector voltage was 700 V. Both emission and excitation detectors were corrected to prevent wavelength sensitivity. The Raman spectrum of ultrapure water (Ultrapure Water Cell, Starna Ltd) was measured on a daily basis with excitation at 350 nm, scan speed at 240 nm min<sup>-1</sup>, emission from 360 to 420 nm, and excitation and emission slit fixed at 5 nm and 700 V to normalize the EEM intensity with the Raman scattering peak (Lawaetz and Stedmon, 2009).

#### *2.3.4.2. Fluorescence data processing*

PARAFAC was conducted on the EEM dataset using PROGMEEF software (<https://woms18.univ-tln.fr/progmeeef/>) in Matlab language. All the EEMs were cleaned from the diffusion signals using Rayleigh scattering by cutting the diffusion band in 15 nm and using first- and second-order Raman scattering by applying the Zepp method (Mounier et al., 2011; Zepp et al., 2004). The number of evaluated components varied from 2 to 5 in a spectral range from 240 to 500 nm in excitation and from 250 to 700 nm in emission. The number of components was defined using the core consistency diagnostic (CORCONDIA). The optimal rank was the model giving the higher number of components with a CORCONDIA over 60% (Mounier et al., 2011). The excitation and emission of components obtained by PARAFAC were compared to the OpenFluor database (Murphy et al., 2014).

#### *2.4. Statistical analysis*

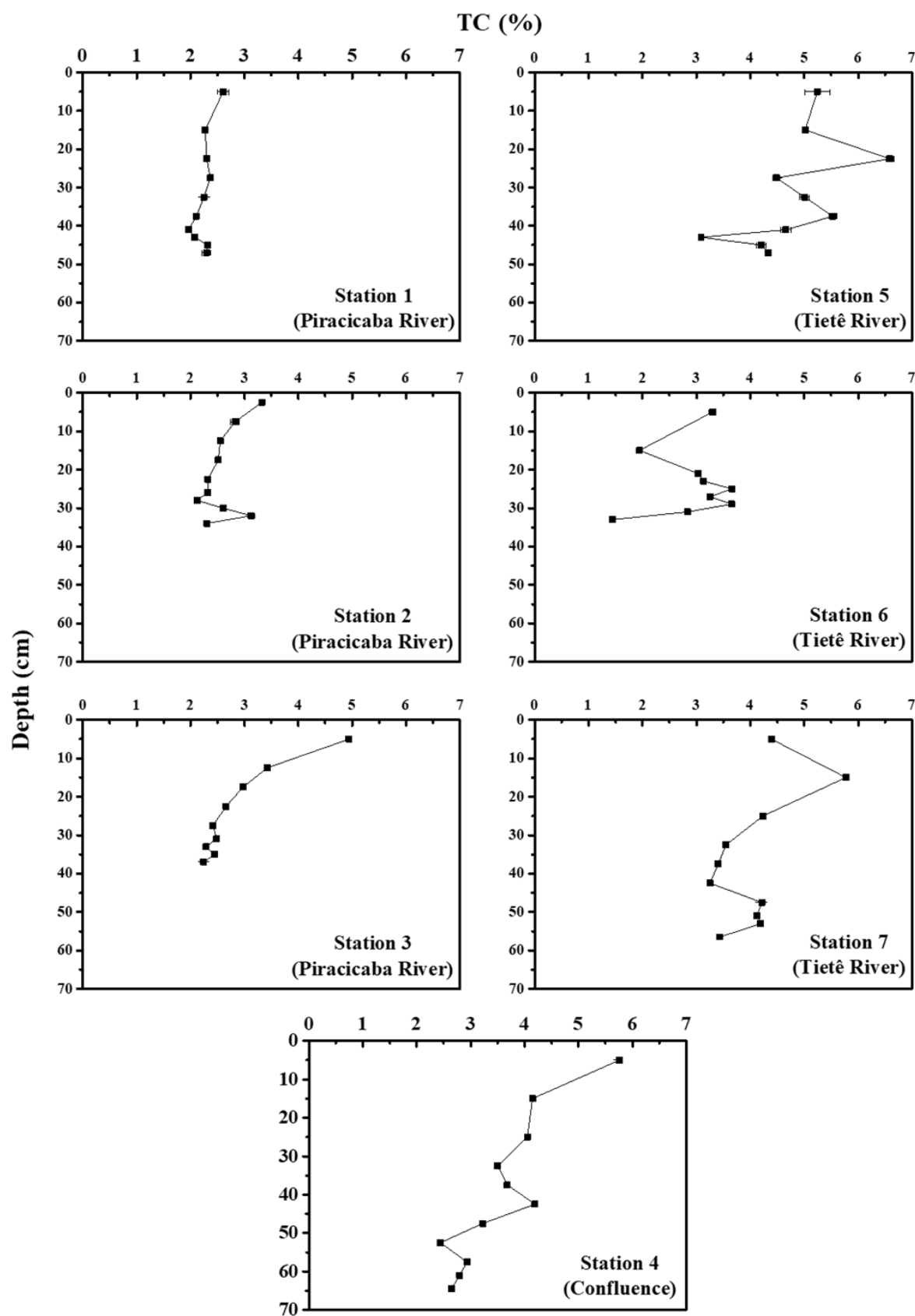
Unpaired t-tests were used to determine any statistical differences between the TC in bulk sediments from the Tietê and Piracicaba Rivers and NPOC, TON, NPOC/TON molar ratios, and SUVA values obtained for EOM<sub>Sed</sub> from the Tietê and Piracicaba Rivers. The correlation coefficient between the NPOC/TON molar ratios and the SUVA<sub>254</sub> values was calculated.

### 3. Results and discussion

#### 3.1. Total carbon characteristics in bulk sediments

The distributions of TC content in the bulk sediment cores in the Piracicaba and Tietê Rivers were different (Figure 2). In the collection stations of the Piracicaba River (stations 1, 2, and 3), the highest TC value was found in the upper layer of the sediment cores, and a decrease in TC content was observed at the lower layers of the vertical distribution. In station 1, TC varied from 1.96% to 2.60% with a coefficient of variation (CV) of 0.10, and the lowest value at a depth of 41 cm. In station 2, TC varied from 2.12% to 3.32% (CV = 0.15), and despite the decrease in TC when entering the sediment, it was possible to observe an increase in TC content between the depths of 30 cm and 32 cm. In station 3, TC varied from 2.24% to 4.94% (CV = 0.30), although it decreased exponentially from the surface to a depth of 27.5 cm, and from that depth, it remained almost constant. In the confluence region, TC varied from 2.44% to 5.76% (CV = 0.27), although the highest TC content is found in the surface layer, as in the collection stations of the Piracicaba River. Its behavior is more similar to the collection stations located on the Tietê River, which present a random TC distribution in the deeper layers of the sediment cores. TC variation in the Tietê River ranged from 3.09% to 6.59% (CV = 0.19), 1.44% to 3.66% (CV = 0.26), and 3.25% to 5.77% (CV = 0.19) at stations 5, 6, and 7, respectively.

261 TC content is a fundamental parameter to describe the abundance of OM and  
262 inorganic carbon (IC) in sediments (Veres, 2002). The random distribution of TC in the  
263 Tietê River may be associated with an irregular input of OM and IC. A unpaired t-test  
264 with a 95% confidence level showed that there were significant differences between the  
265 average TC of the rivers. The average TC content in the Tietê River ( $3.96 \pm 1.11$ ) is  
266 higher than in the Piracicaba River ( $2.57 \pm 0.58$ ). The highest TC content for the Tietê  
267 River suggests that this river is more eutrophic than the Piracicaba River, since the  
268 accumulation of nutrients, mainly P and N, in the aquatic bodies increases the synthesis  
269 of OM (Castillo, 2020). This hypothesis is corroborated by a study that we previously  
270 published on the quantification of total phosphorus (TP) in the same sediment samples  
271 studied here (Morais et al., 2021). As TP is a chemical variable used to assess the  
272 nutrient load and the extent of eutrophication in water bodies, the higher average TP  
273 content in the Tietê River ( $4316.16 \pm 1062 \text{ mg kg}^{-1} \text{ P}$ ) than in the Piracicaba River  
274 ( $1800.17 \pm 696.82 \text{ mg kg}^{-1} \text{ P}$ ) indicates that this river has a higher degree of  
275 eutrophication (Morais et al., 2021).



**Figure 2.** Total carbon distribution inside the sediment cores at the collection stations.

### 3.2. Characteristics of extracted organic matter from sediments

Table 1 shows the average concentration of NPOC and TON, alkaline extraction yield, NPOC/TON molar ratio, and SUVA<sub>254</sub> of EOMSed at the different stations. These values for each sample are shown in the supplementary material (Table S3). The extraction method with alkaline solution extracted an average of 40.71% ± 5.52% of OM present in the bulk sediments. The yield of OM extraction depends on the property of the extractant and is generally low. Among the different extractants used for OM extraction, NaOH seems to be the most effective (Kurek et al., 2020; Sire et al., 2009).

**Table 1.** Average concentration of non-purgeable organic carbon (NPOC) and total organic nitrogen (TON), alkaline extraction yield, NPOC/TON molar ratio, and specific UV absorbance at 254 nm (SUVA<sub>254</sub>) of the extracted organic matter from sediments at the collection stations.

Station	NPOC (mg C g <sup>-1</sup> )	TON (mg N g <sup>-1</sup> )	Extraction yield (%)	NPOC/TON ratio	SUVA <sub>254</sub> (L mg <sup>-1</sup> m <sup>-1</sup> )
1	9.02±0.84	1.14±0.11	40.63±5.17	6.80±0.20	6.36±0.53
2	10.65±1.71	1.25±0.24	41.03±3.58	7.37±1.00	5.99±0.69
3	12.59±1.98	1.75±0.38	45.23±6.56	6.22±0.49	6.11±1.08
4	13.54±4.82	2.40±0.76	37.17±5.61	4.78±0.30	4.54±0.73
5	19.60±5.74	3.01±0.74	40.11±5.17	5.53±0.50	3.92±0.61
6	11.90±3.54	2.00±0.52	40.50±3.96	5.05±0.37	3.70±0.51
7	16.24±2.31	2.51±0.45	41.05±6.36	5.59±0.32	4.44±0.83

292

293         A unpaired t-test at the 95% confidence level demonstrated that there were  
294 significant differences in the NPOC and TON averages of EOMSed in the Tietê and  
295 Piracicaba Rivers. The average concentration of NPOC and TON of EOMSed in the  
296 Tietê River ( $16.05 \pm 5.10 \text{ mg C g}^{-1}$  and  $2.52 \pm 0.70 \text{ mg N g}^{-1}$ , respectively) was higher  
297 than that in the Piracicaba River ( $10.69 \pm 2.11 \text{ mg C g}^{-1}$  and  $1.37 \pm 0.37 \text{ mg N g}^{-1}$ ,  
298 respectively). The highest NPOC concentrations of EOMSed in the Tietê River may be  
299 due to the fact that the river passes through the metropolitan area of São Paulo, where it  
300 receives a large load of domestic sewage and industrial effluents, resulting in a larger  
301 biological oxygen demand than the Piracicaba River (CETESB, 2018). Furthermore,  
302 taking into account the history of the Tietê River, its high degree of contamination  
303 (Rocha et al., 2011, 2010) and eutrophication (Morais et al., 2021), the higher TON  
304 concentration of EOMSed in the Tietê River when compared to the Piracicaba River  
305 may be due to the sewage discharge containing labile and fresh OM.

306         The average NPOC/TON molar ratios of EOMSed in the Tietê and Piracicaba  
307 Rivers were statistically different (95% confidence level, unpaired t-test) and were  
308 higher for the Piracicaba River ( $6.80 \pm 0.79$ ) compared to the Tietê River ( $5.39 \pm 0.46$ ).  
309 Considering the historical course of the Tietê River and its high degree of  
310 eutrophication (Morais et al., 2021), the lower ratios found in the river may suggest a  
311 larger input of simple OM with nitrogenous structures, which may indicate effluent  
312 input. In the confluence region, the NPOC/TON molar ratio of EOMSed was the lowest:  
313  $4.78 \pm 0.30$ . The lower value of the NPOC/TON molar ratio shows an increase in OM  
314 deposition with nitrogenous structures, suggesting that the confluence region is  
315 predominantly influenced by the Tietê River, which showed the highest nitrogen

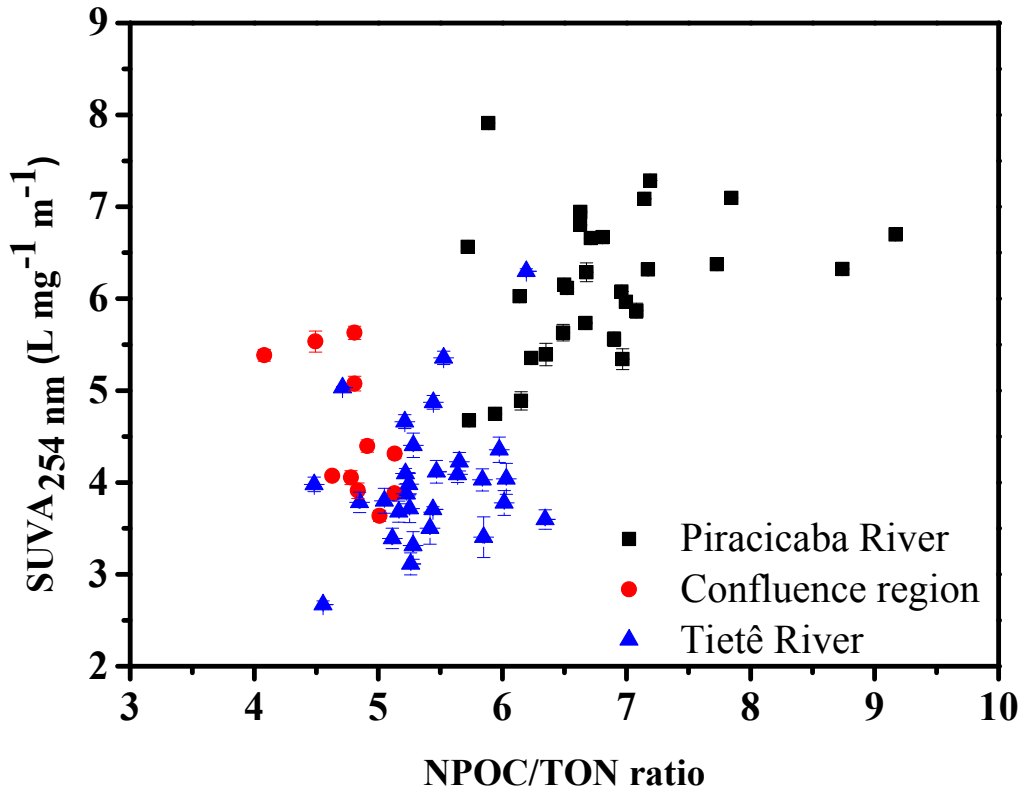


316 concentrations (range of 2.00–3.01 mg N g<sup>-1</sup>). Despite the difference in the NPOC/TON  
317 molar ratios of EOM<sub>Sed</sub> between the Tietê and Piracicaba Rivers, the studied EOM<sub>Sed</sub>  
318 showed a NPOC/TON molar ratio that varied from 4.1 to 9.2 (Figure 3). The  
319 NPOC/TON molar ratios between 4 and 10 categorize the sources of C and N of algae  
320 and aquatic plants (Kaushal and Binford, 1999; Meyers, 1994).

321 SUVA<sub>254</sub> is associated with the aromaticity of OM due to the carbon double  
322 bond (C=C) of aromatic moieties or polycondensated aromatic rings that absorb light at  
323 254 nm (Edzwald and Tobiasson, 2011). Therefore, high SUVA<sub>254</sub> values indicate an  
324 OM formed by more structures that absorb light at 254 nm per unit concentration of OC  
325 (Edzwald and Tobiasson, 2011). The average SUVA<sub>254</sub> values of EOM<sub>Sed</sub> in the  
326 Piracicaba and Tietê Rivers were statistically different (95% confidence level, unpaired  
327 t-test) and were higher for the Piracicaba River (6.16 ± 0.78) than the Tietê River (4.02  
328 ± 0.71). The confluence region showed SUVA<sub>254</sub> values of EOM<sub>Sed</sub> closer to the Tietê  
329 River (average SUVA<sub>254</sub> values was 4.54 ± 0.71 L mg<sup>-1</sup> m<sup>-1</sup>). SUVA<sub>254</sub> showed  
330 differences in the composition of EOM<sub>Sed</sub> between the two rivers, with the higher  
331 average SUVA<sub>254</sub> values for the Piracicaba River indicating that EOM<sub>Sed</sub> has a  
332 relatively high aromatic content compared to the Tietê River.

333 The NPOC/TON molar ratios of EOM<sub>Sed</sub> were correlated with the SUVA<sub>254</sub>  
334 values (Figure 3). Regression analysis found a positive relationship between the  
335 NPOC/TON molar ratio and SUVA<sub>254</sub> (r = 0.67; p < 0.001). By analogy to soil OM,  
336 these results could be associated with the functional complexity of EOM<sub>Sed</sub> (Dungait et  
337 al., 2012; Lehmann et al., 2020). During the degradation of OM, the more polar  
338 structures are more accessible to microorganisms and are thus consumed faster,  
339 resulting in an accumulation of OM that is thermodynamically unfavorable to  
340 microorganisms (hydrophobic and aromatic moieties). The relatively lower presence of

N suggests that these structures have already been totally or partly assimilated, resulting in the selective preservation and accumulation of more chemically stable structures, which led to greater aromaticity in the Piracicaba River. Conversely, the lower  $SUVA_{254}$  and NPOC/TON molar ratio in the Tietê River suggest the addition of fresh OM. This evidence is supported by the high BOD value of the Tietê River.



**Figure 3.** Relationship between specific UV absorbance at 254 nm ( $SUVA_{254}$ ) and non-purgeable organic carbon (NPOC) and total organic nitrogen (TON) molar ratios of extracted organic matter from sediments.

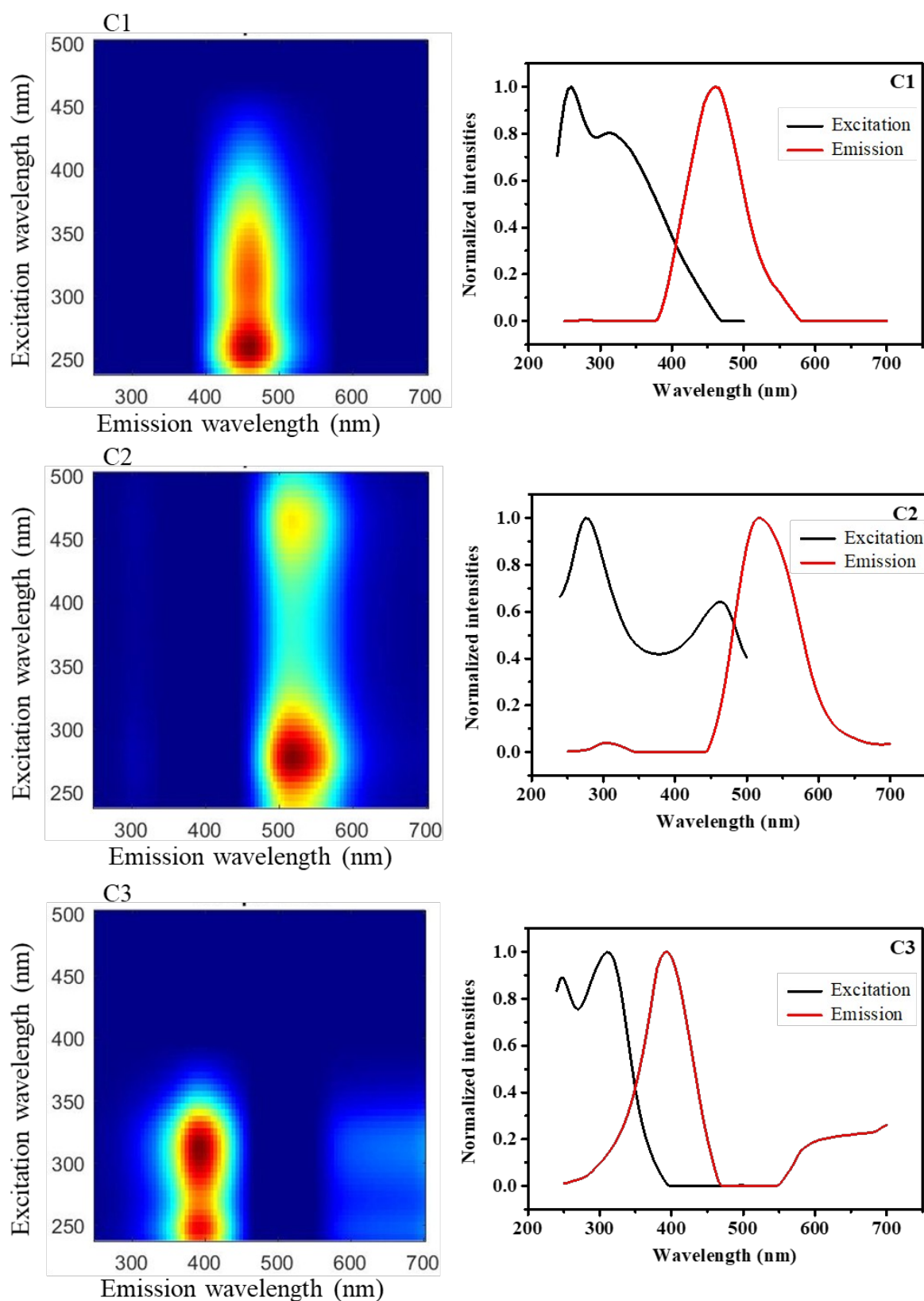
### 3.3. Fluorescence excitation-emission matrix spectra of organic matter extracted from sediments with alkaline and neutral water solutions

The PARAFAC model adjusted for EEM (N= 138) from EOMSed and WEOMSed diluted to the same optical density resulted in three fluorescent components (Figure 4) with a CORCONDIA of 89.98%. The spectral characteristics of the three

356 components were compared to the OpenFluor database (Murphy et al., 2014), and  
357 similarity was measured with the PARAFAC models from different aquatic  
358 environments using the EEM similarity score of 0.95. Components 1, 2, and 3  
359 respectively corresponded to PARAFAC models 35, 5, and 25 from the OpenFluor  
360 database.

361       Component 1 (C1) had two excitation maxima: one in 260 nm stimulated by UV  
362 excitation (peak A), and the other in 310 nm stimulated by visible excitation, with an  
363 emission maximum in the visible range at 465 nm for both excitation peaks (Coble,  
364 1996). Component 2 (C2) was comprised of two excitation maxima at 285 nm and 465  
365 nm, and an emission maximum at 520 nm (Osburn et al., 2011; Walker et al., 2013).  
366 The fluorescence emission is useful to access the structural arrangement of OM due to  
367 the fluorophore emission in short (blue-shift) or long wavelength (red-shift). C1 and C2  
368 had similar characteristics to soil fulvic and humic acids, respectively. C1 resembles  
369 fulvic acids due to the shorter wavelength emission, and the blue-shifted component is  
370 attributed to the lower molecular weight and a structural arrangement comprised of  
371 more alkyl moieties (Borisover et al., 2009; Kowalczyk et al., 2009; Osburn et al., 2016,  
372 2011; Shutova et al., 2014; Yamashita et al., 2010b, 2010a). C2 resembles humic acids  
373 from the soil due to the red-shifted component, which is attributed to a more complex  
374 structural arrangement mainly composed of aromatic moieties (Soares da Silva et al.,  
375 2020). Furthermore, the shift in the wavelength concerns the aggregation of humic  
376 structures in solution; a red-shifted humic structure can show a molecular arrangement  
377 with higher molecular aggregation, while a shorter wavelength suggests smaller  
378 molecules (Bento et al., 2020; Boguta and Sokołowska, 2020, 2016). Component 3 (C3)  
379 demonstrated two excitation maxima at 250 nm and 315 nm, and an emission maximum  
380 at 395 nm. C3 resembled microbial humic-like compounds from aquatic environments

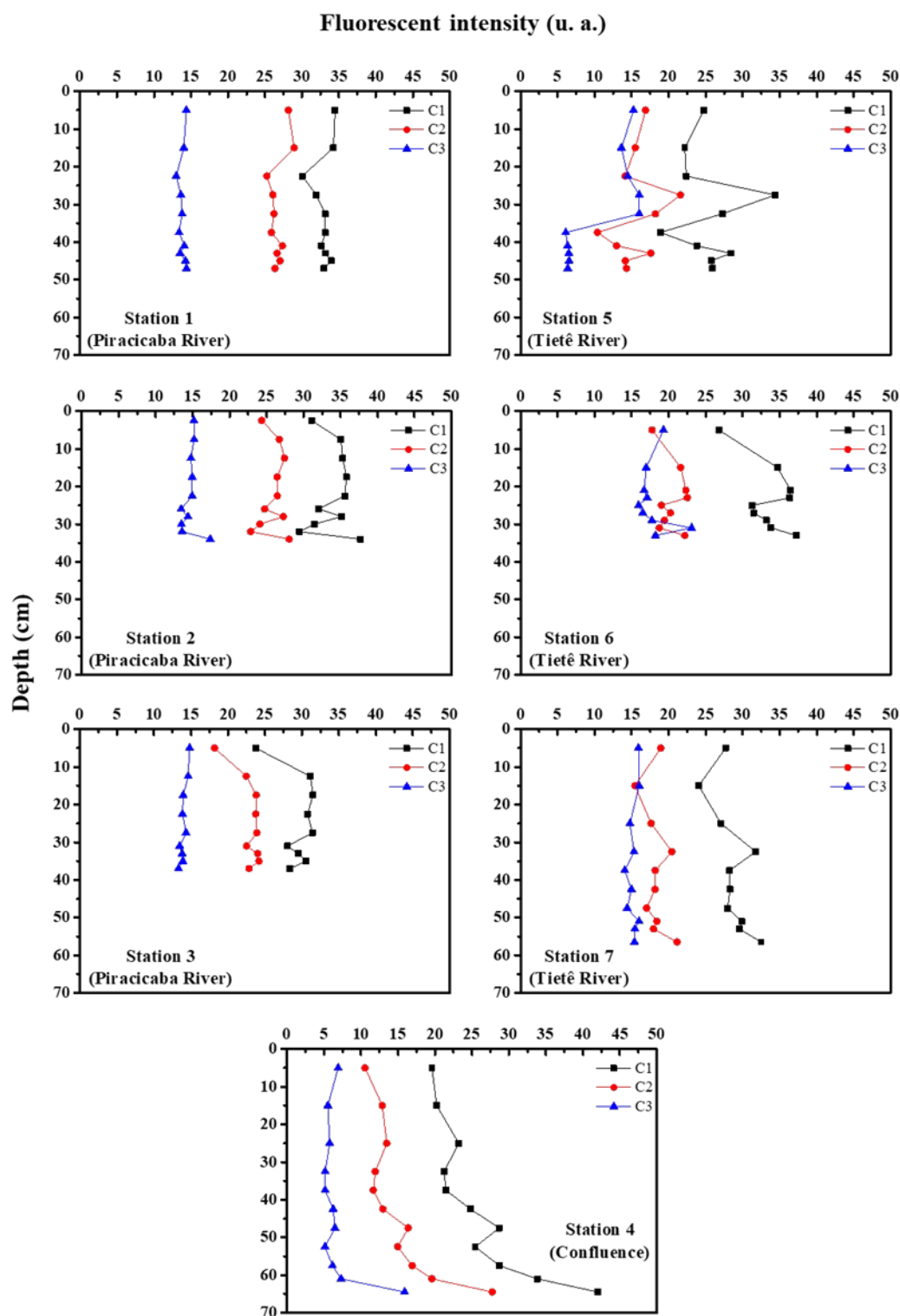
381 (Coble, 1996; Osburn et al., 2011). This component is probably a product of microbial  
 382 activity and thus of autochthonous origin (Yamashita et al., 2008).



383

**Figure 4.** Excitation-emission matrix contour plots of the three parallel factor analysis components (left) and their excitation and emission spectra (right).

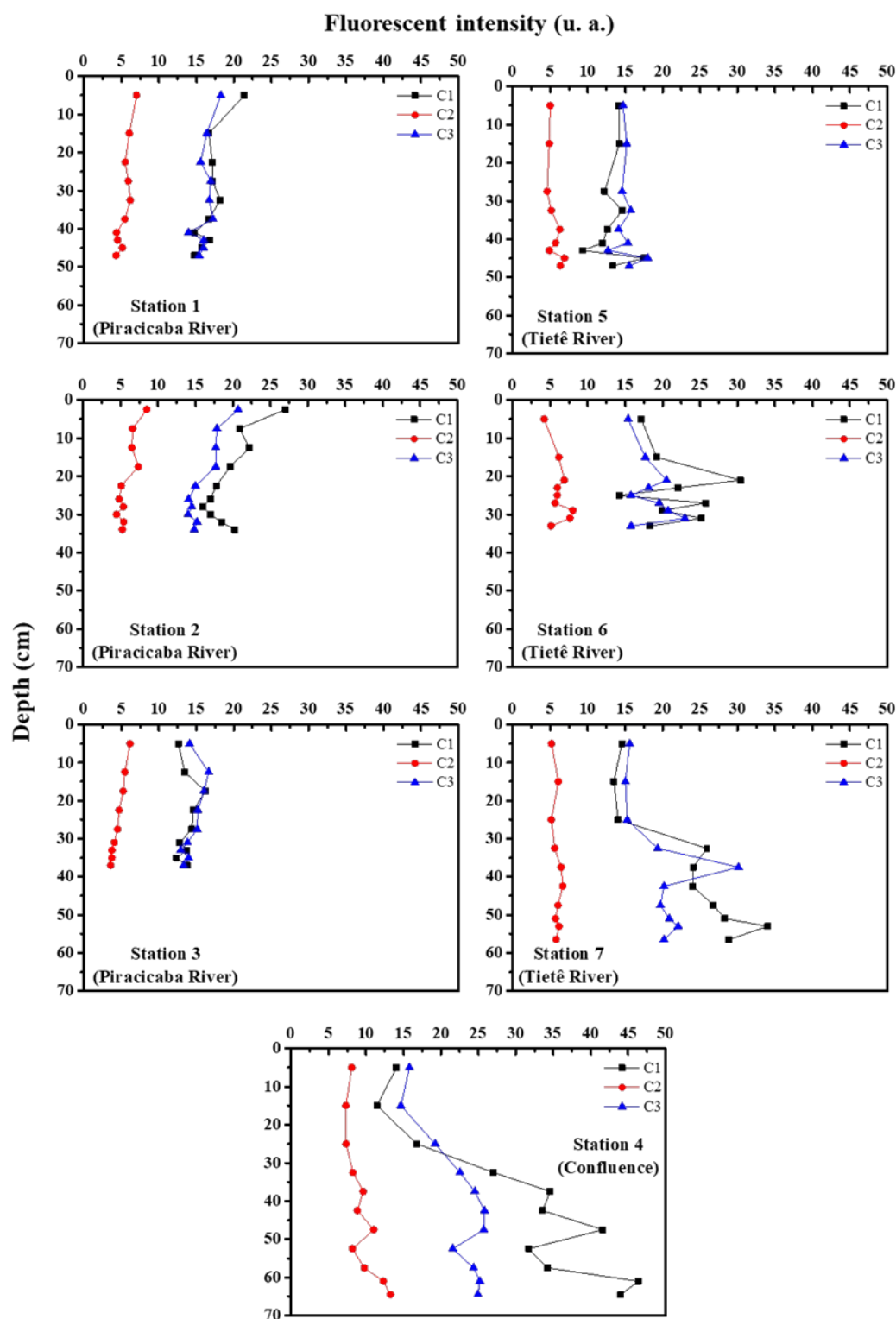
The PARAFAC model was made for a dataset with all samples, thus allowing us to compare the component contributions. Figures 5 and 6 show the fluorescent contributions of each component identified by PARAFAC for EOMSed and WEOMSed, respectively, along with the sediment cores in the different stations, corrected for the analytical dilution factor. For EOMSed (Figure 5), the average fluorescent contribution of C1 was higher in decreasing order for the Piracicaba River > Tietê River > confluence region. This shows that small molecules make a greater contribution to the EOMSed arrangement of the Piracicaba River. The average fluorescent contribution of C2 had the same decreasing order as that of C1 (Piracicaba River > Tietê River > confluence region), showing that complex structures in the form of aromatic and polyaromatic moieties make a greater contribution to the EOMSed arrangement of the Piracicaba River. The average fluorescent contribution of C3 was higher in decreasing order for the Tietê River > Piracicaba River > confluence region, showing that microbial-derived products make a greater contribution to the EOMSed arrangement of the Tietê River. Thus, EOMSed from the Piracicaba River contains more aromatics, while EOMSed from the Tietê River contains more microbial-derived products. These results are in agreement with the  $SUVA_{254}$  values and NPOC/TON molar ratios, which showed less aromatic EOMSed with more nitrogenous structures for the Tietê River compared to the Piracicaba River. The low NPOC/TON molar ratio may be the main reason for the higher microbial activity and the consequent presence of microbial derivatives (C3).



**Figure 5.** Fluorescent intensity of components (C1, C2, and C3) taken from the extracted organic matter from sediments along with the sediment core identified by parallel factor analysis.

In WEOM<sub>Sed</sub> (Figure 6), the average fluorescent contributions of C1, C2, and C3 were higher in decreasing order for the confluence region > Tietê River > Piracicaba River. The extraction of microbial-derived products (C3) was greater for WEOM<sub>Sed</sub> than for EOM<sub>Sed</sub>, and the C3 values were higher at the stations of the Tietê River than the Piracicaba River, indicating its greater microbial activity. The low abundance of C2 can be ascribed to the component structure, because humic-like substances with high aromaticity are more soluble in alkaline solution than in water, which may result in the higher relative content of C2 in alkaline solution; furthermore, the higher C2 concentration in alkaline extracts obfuscates the C3 concentration. The alkaline medium favored the greater extraction of C1 and C2 (humic-like compounds) due to the hydrolysis caused in the functional groups of humic molecules bonded to the surface of the mineral from sediment (Kleber and Lehmann, 2019; Olk et al., 2019).

The optical properties of EOM<sub>Sed</sub> proved to be more efficient for assessing the quality and composition of sediments than those of WEOM<sub>Sed</sub> due to the better discrimination between the fluorescence components in the alkaline extracts. Although water extraction has been used to assess OM sources and link contamination in polluted urban rivers (Chen et al., 2019; Zhang et al., 2020), in our study, alkaline extracts were better at describing the properties of OM.



**Figure 6.** Fluorescent intensity of components (C1, C2, and C3) taken from water-extracted organic matter from sediments along with sediment core identified by parallel factor analysis.



### 3.4. Features of extractable organic matter from sediments from the Piracicaba and Tietê Rivers

The neutral water solution enabled the better extraction of small molecules (blue-shifted), which are free from the mineral fraction in the sediments. While the alkaline solution favors the extraction of more complex substances (red-shifted). The results obtained with EEM-PARAFAC together with  $SUVA_{254}$  values and NPOC/TON molar ratio allowed for the total assessment of EOMSed quality.

The differences between the EOMSed compositions in sediments were highlighted by the  $SUVA_{254}$  values, NPOC/TON molar ratios, and EEM-PARAFAC. Although the most abundant component in EOMSed from the two rivers is humic-like with the lowest molecular weight (C1), the aromaticity of EOMSed from the Piracicaba River was evidenced by the greater contribution of C2 and  $SUVA_{254}$  values. Conversely, EOMSed from the Tietê River had greater microbial activity due to the greater contribution of C3, the presence of small molecules (lower C2 contribution), and the presence of nitrogenous structures. Based on the analyses performed, it was not possible to further identify the compositional alteration of EOMSed and WEOMSed, which may be related to their similar deposition.

The origin of EOMSed is inferred from the results obtained here. The NPOC/TON molar ratios calculated for EOMSed from the Piracicaba River indicate that EOMSed can form in the aquatic body (autochthonous). The contributions of two humic-like components in alkaline extracts indicate that EOMSed may also originate from soil OM (allochthon). According to the NPOC/TON molar ratios calculated for EOMSed from the Tietê River as well as the greater microbial activity in this aquatic ecosystem, it can be inferred that the origin of EOMSed is predominantly

allochthonous, mainly due to the discharge of domestic sewage, which is indicated by the higher content of TC present in the Tietê River sediments.

#### **4. Conclusion**

EOM<sub>Sed</sub> from the Piracicaba and Tietê Rivers was mainly characterized by an arrangement of low molecular weight compounds. However, EOM<sub>Sed</sub> from the Piracicaba River presented more aromatic moieties and fewer nitrogenous structures compared to the Tietê River. The presence of nitrogenous structures and fluorophores related to microbial activity can be an indicator of OM quality, which could be used as a global indicator of the river state. Thus, the combination of elemental and spectroscopic techniques allows for the successful deciphering of extracted OM characteristics. This approach can therefore be used as a tool to assess the quality of aquatic bodies.

#### **Declaration of competing interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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