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**Diuron effects on photosynthesis and vertical migration of microphytobenthos:
potential rapid bioassessment of herbicide toxicity in coastal sediments**

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Abstract

The effects of herbicide diuron on photosynthesis and vertical migration of intertidal microphytobenthos (MPB) assemblages were investigated using chlorophyll fluorometry. The results shown diuron $\leq 60 \mu\text{g}\cdot\text{L}^{-1}$ had no obvious effect on MPB vertical migration during 24 h indicated by consistent rhythm. Low concentration of $10 \mu\text{g}\cdot\text{L}^{-1}$ diuron had no significant influence on MPB photosynthesis throughout, however, high concentrations of 40, 50, and $60 \mu\text{g}\cdot\text{L}^{-1}$ had significant impacts exhibited by decreased parameters of maximum relative electron transport rate ($r\text{ETR}_{\text{max}}$), maximal PS II quantum yield (F_v/F_m) and non-photochemical quenching (NPQ). For middle concentrations of 20 and $30 \mu\text{g}\cdot\text{L}^{-1}$, above decreased 3 parameters recovered sooner or later after 2 h or 16.5 h. Comparatively, $r\text{ETR}_{\text{max}}$, F_v/F_m and NPQ are concentration dependent and more sensitive than other parameters in assessing diuron toxicity. This study revealed the potential of using MPB assemblages and chlorophyll fluorometry for rapid assessing diuron toxicity in coastal sediments.

Keywords: microphytobenthos, photosynthesis, vertical migration, diuron, herbicide toxicity, bioassessment

Heading: diuron effects on microphytobenthos

1. Introduction

Diuron (3-(3,4-dichlorophenyl)-1,1-dimethylurea, DCMU), as a urea-type herbicide has been effectively used to control a wide variety of grasses and weeds on cotton, coffee, sugar cane, and citrus farms (Coelho-Moreira et al., 2018). Apart from this, the organic biocide diuron has gained popularity as an antifouling agent in paints, especially to prevent fouling of algae on ship hulls and boats (Chesworth et al., 2004; Mukherjee et al., 2009). Incidentally, this kind of herbicide has been released to the freshwater and marine environments inevitably for more than 20 years. Due to its low solubility (42 mg mL⁻¹ at 25 °C), long persistence in soil (4–8 months) and chemical stability in water, diuron tends to accumulate in the environment (Tixier et al., 2001). It has been detected not only in ground and freshwater systems (Nitschke and Schüssler, 1998; Bruggen and Vandecasteele, 2003), but also in coastal water and sediments (Jones et al., 2003; Ali et al., 2014; Holmes, 2014).

As a universal PSII inhibitory herbicide, diuron can directly inhibits non-target photosynthetic organisms (Haynes et al., 2000; Jones, 2005). Besides, diuron's harms to human and animals have already been reported in numbers of literatures, causing diseases such as mutagenic, metabolic, abnormalities of the liver and spleen, and problems in the transport and release of natural hormones (US Government, 2005; Wong et al., 2013; Simões et al., 2017). All these make diuron a threat to the environment and human being, therefore, its ecological risk has been keenly concerned by countries over the world. It is declared that in 2011 Australia government has banned using it, and a series of assessment reports (US Government, 2005; Australian Government, 2011) and related researches have been carried out over the past two decades (OECD, 2006; Magnusson et al., 2008; Kumar et al., 2010; Negri et al., 2015).

In the published literatures, there are some standards dataset for accessing the toxicity of diuron on terrestrial plants (Australian Government, 2011; Holmes, 2014). In coastal ecosystems, some publications showed the toxicity of diuron for benthic photosynthetic organisms exposed through the water phase, which has been implicated in negative impacts on seagrass (Haynes et al., 2000), mangroves (Duke et al., 2005; Duke, 2008), coral (Jones et al., 2003; Negri et al., 2005, 2011), cyanobacteria (Stingaciu et al., 2019), and benthic microalgae (Magnusson et al., 2008, 2012). Standard toxicity tests have been conducted by exposing photosynthetic organisms to diuron in water (USEPA, 1996; OECD, 2006). In terms of risk characterization, the dataset had been consequently

1 applied in risk quotients of these organisms (Australian Government, 2011). Following these criteria,
2 within the catchments and waters of the Great Barrier Reef, diuron is frequently detected in aquatic
3 and marine water quality monitoring programs (Shaw et al., 2010; Kennedy et al., 2012; Lewis et al.,
4 2012; Smith et al., 2012). However, those datasets have not yet established for benthic flora or fauna
5 when exposed to diuron through the sediment due to short of investigations.

6 Microphytobenthos (MPB) is a group of photosynthetic organisms living in the upper several
7 millimetres of sediments where light reaches, and frequently dominated by diatoms in coastal flats
8 (Admiraal et al., 1984; MacIntyre et al., 1996; Underwood and Kromkamp, 1999). With short life
9 cycle and relevant stable position in the sediment, benthic diatoms are very suitable indicators for
10 monitoring the status of ecological environment. In freshwater system, there have been already
11 numerous publications deliberated it (Kelly and Whitton, 1995; Potapova and Charles, 2007;
12 Stevenson et al., 2010; Bere and Tundisi, 2012; Chen et al., 2016). It is widely used benthic diatoms
13 as Biological Diatom Index (BDI; Coste et al., 2009) in the Water Framework Directive (EU
14 Directive 2000/60/EC) for assessing freshwater system. Theoretically, MPB is very potential
15 bioindicator to monitor or detect toxicity of pollutants in coastal ecosystems. Desrosiers et al. (2013)
16 has proposed that using benthic diatoms for assessing water quality of coastal environments.

17 MPB shows high photosynthetic capacity and efficient protection against photoinhibition
18 through both physiological and behavioural mechanisms (Serôdio et al., 2006; Cartaxana et al., 2011;
19 Lavaud and Goss, 2014; Barnett et al., 2015; Laviale et al., 2015). Physiological photoprotection
20 mainly depends on distinct xanthophyll pigments, diadinoxanthin and diatoxanthin, which support
21 the dissipation of excessive light energy (the so-called non-photochemical quenching process) and
22 protect PSII reaction centres (Barnett et al., 2015; Cartaxana et al., 2016; Frankenbach et al., 2018;
23 Pniewski and Piasecka-Jędrzejak, 2020). Behavioural photoprotection is realized through the vertical
24 migration of MPB within the uppermost layers of sediment, by which motile diatoms position
25 themselves at suitable depth for optimal light conditions (Consalvey et al. 2004; Barnett et al., 2020).
26 Disturbed or inhibited motility can impair MPB photosynthetic capacity, especially under high
27 irradiance and/or prolonged illumination (Laviale et al., 2015; Cartaxana et al., 2016; Du et al., 2018;
28 Frankenbach et al., 2018).

29 This study aimed through applying gradients of diuron on natural MPB assemblages to detect

1 their effects on MPB photosynthetic activities and behaviour characteristics of vertical migration,
2 and monitor the duration of these effects during a whole day. The imaging chlorophyll fluorescence
3 techniques were used to ensure prompt and non-destructive measurement, and assist further
4 assessment of diuron toxicity in sediment.

6 **2. Materials and methods**

7 Sediment samples were collected from the intertidal mudflat of Aiguillon Bay (47°00' N, 1°05'
8 W) La Rochelle, France, on 18th February, 2015. The sampling site is composed of fine muddy
9 sediments (more than 90% of grain size < 63µm) where microphytobenthic biofilm is largely
10 dominated by epipellic diatoms, especially species of genera *Navicula* and *Gyrosigma* (Du et al.
11 2017). The surface layers of sediment (top ~0.5 cm) were collected using a spatula.

12 In the laboratory, the sediment was sieved through a 500 µm mesh to remove the meio- and
13 macrofauna, such as nematodes and sand hoppers, and was thoroughly mixed and spread in 4 cm
14 deep plastic trays. The stock solution of diuron (0.05 g L⁻¹) was made up in 95% ethanol. In
15 individual glass beakers, corresponding amounts of the solution were diluted with dH₂O and mixed
16 well with sediment samples to reach final concentrations of 0, 10, 20, 30, 40, 50 and 60 µg L⁻¹.
17 Beside the blank control of 0 µg L⁻¹ diuron, to check the influence of solvent, the same amount of
18 95% ethanol that equal to that of stock solution used for 60 µg L⁻¹ diuron was applied in sediment
19 samples to be as the ethanol control. Each 3 mL well mixed sediments were transferred into a well of
20 24-well plates using pipette. Each concentration had 3 replicates, and two sets of well-plates were
21 prepared. A minimum of 30 min was allowed for the well-plate system with diuron and MPB
22 assemblages to stabilize before measurements started.

23 Chlorophyll fluorescence was measured using an Imaging-PAM fluorometer (Max/L, Walz,
24 Germany) for 24h during 18th to 19th February. Before measuring fluorescence, areas of interest
25 (AOIs) were defined under Live Video Mode, with circular samples of the same size as internal
26 diameter of well in 24 well-plates. The same AOIs were used consistently over the course of the
27 experiment. Fluorescence was induced by royal blue (450 nm) 3W Luxeon LEDs, with a standard
28 intensity of 0.5 µmol photons m⁻² s⁻¹ and a modulation frequency between 1 and 8 Hz. The current
29 fluorescence yield, F_t , for monitoring surface MPB abundance were measured at 900 s intervals

using one set of well-plate samples for 24 h. The other set of well-plate samples was incubated under “ambient filtered true light” conditions (i.e., light passing by the window of the laboratory, 24-h natural photoperiod, no direct sunlight). The room conditions in laboratory were stable around 15°C temperature and 60% humidity. The rapid light curves (RLCs) were measured to assess the photosynthetic activity, by exposing to 7 incremental steps of actinic light (30 s per step) ranging from 0 to 610 $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ (PAR, photosynthetically active radiation). The measurement of RLCs were taken after 15 min dark-adaptation at 6 different times during the whole experiment. To check the short-time effect of diuron, RLC measures were carried at 1 h intervals in the daytime of the first day. To monitor the persisting of diuron effect or MPB recovery from it, in the next day, RLC measures were taken at the beginning and the end of the day respectively. For each irradiance level (E), the relative electron transport rates (rETR) was calculated from the product of E and the PSII effective quantum yield ($Y(\text{II}) = (F_m' - F)/F_m'$), $\text{rETR} = E \times (F_m' - F)/F_m'$ (Genty et al., 1989), where the F is fluorescence yield, F_m' is maximum fluorescence under the highest actinic light of RLC. The photosynthetic parameters, the maximum relative electron transport rate (rETR_{max}), maximum light utilization coefficient (α) and the minimum saturation irradiance (E_k) were derived from the RLCs, referring the model of Platt et al. (1980), $\text{ETR} = \text{ETR}_{\text{max}} (1 - \exp(-\alpha \times E/\text{ETR}_{\text{max}}))$, $E_k = \text{ETR}_{\text{max}}/\alpha$. The maximal PS II quantum yield (F_v/F_m) was calculated as $(F_m - F_0)/F_m$, where F_0 is the dark fluorescence yield and F_m is maximum fluorescence yield after dark adaptation. The non-photochemical quenching (NPQ) was calculated as $(F_m - F_m')/F_m'$ (Maxwell and Johnson, 2000).

Statistical analysis was carried out using SPSS 21.0 (SPSS Inc., Chicago, IL, USA). To test the effect of diuron on migration, separate one-way ANOVAs were run on F_t for each treatment, followed by the Tukey’s HSD- post-hoc tests to compare paired difference. To compare the effects of different diuron concentrations on photosynthesis, T-test were run for each treatment on the variables of rETR, rETR_{max} , α , E_k , F_v/F_m and NPQ at individual measuring time.

3. Results

3.1 Chlorophyll fluorescence yield (F_t) variations

Indicated by F_t , distinct variations of surface MPB abundance along with experiment time were illustrated in Figure 1. Under weak measuring light of F_t ($0.5 \mu\text{mol photons m}^{-2} \text{s}^{-1}$), the natural

1 MPB assemblage which had been artificially mixed homogenously with sediments still exhibited
 2 their endogenous migration up to the surface. Besides the initial rapid increase at the beginning, there
 3 was steeply ascending F_t from AM 3:30 to 5:00 during the early morning of the next day before
 4 sunrise, and followed by relatively smoothly increase till the midday around 12:00. Relatively
 5 decreased F_t happened around PM 20:30 and AM 2:00 (the next day) during night. In the next day,
 6 after midday peak, the F_t slowly declined until the end of experiments. Averagely, the F_t of the next
 7 day was twice higher than that of the first day. Overall, there was no significant difference between
 8 the different treatments for all measured F_t (Tukey HSD test, $P > 0.05$), especially for treatments of
 9 diuron concentration less than $40 \mu\text{g L}^{-1}$, there was no concentration dependent difference along with
 10 the experiment time. However, it illustrated two trends which are worthy of further study. One is the
 11 blank control ($0 \mu\text{g L}^{-1}$) presented the largest variation (SD, ± 0.717) throughout the whole
 12 experiment with relatively lower F_t during night and higher F_t during day than other treatments. The
 13 another is the high concentration of 50 or $60 \mu\text{g L}^{-1}$ generally induced the lowest F_t . Especially for $60 \mu\text{g L}^{-1}$
 14 $\mu\text{g L}^{-1}$ in the next day, the difference became significant with 20 or $40 \mu\text{g L}^{-1}$ since 10:49 (after 18.5h
 15 exposure, paired t-test, $P < 0.05$), but still not significant with the remained treatments including
 16 controls (paired t-test, $P > 0.05$). Overall, the results indicated that no significant diuron effects on
 17 vertical migration of MPB within $60 \mu\text{g L}^{-1}$ concentration during 24h experiment period.

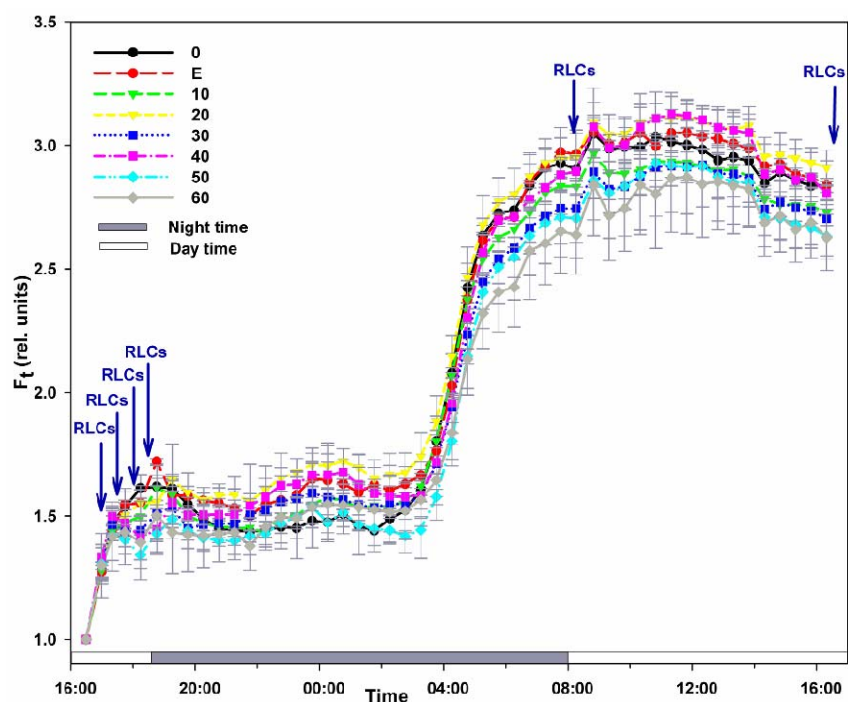


Fig. 1. F_t variations along with time under different treatments (shown as 30 min interval and normalized to initial values). Concentrations of 0, 10, 20, 30, 40, 50, 60 $\mu\text{g L}^{-1}$ diuron are represented by numbers, and ethanol control is labelled as E; dark grey bar, night time; blank bar, day time of natural photoperiod. RLCs (rapid light curves) arrows point the timing at which RLCs were performed.

3.2 Rapid light curves (RLCs) and photosynthetic parameters

RLCs generated by PAM fluorometers presented MPB photosynthetic activity of different treatments. It showed a similar relation pattern of different treatments at 6 measuring time (Fig. 2). The RLCs of blank and ethanol control always accompanied closely and presented increasing rETR along with incremental light intensity more rapidly than other treatments. On the contrary, the high diuron concentrations of 50 and 60 $\mu\text{g L}^{-1}$ constantly induced the most slowly increasing rETR along with light intensity. After 1.5 h exposure, the RLCs of 10 $\mu\text{g L}^{-1}$ diuron almost overlapped with those of blank or ethanol control. The RLCs of 40 $\mu\text{g L}^{-1}$ diuron always closed to those of 50 $\mu\text{g L}^{-1}$ diuron. The RLCs of 20 and 30 $\mu\text{g L}^{-1}$ diuron were always in the middle of all curves at each measuring time.

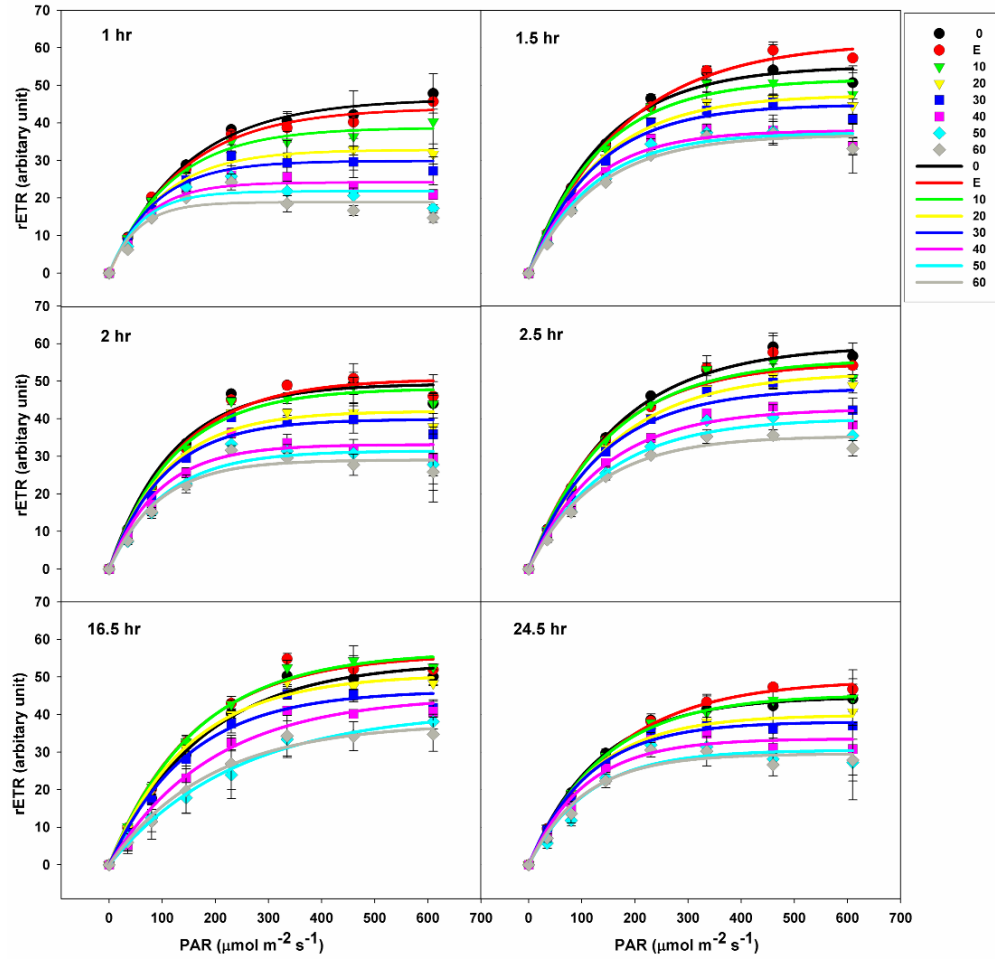


Fig. 2. Rapid light curves (RLCs) with corresponding fitting curves of the different treatments at 6 measuring time, which are marked by blue arrows in Fig.1 with starting timing 0 hr of the experiment beginning. Treatments symbols of 0, 10, 20, 30, 40, 50, 60 $\mu\text{g}\cdot\text{L}^{-1}$ diuron are represented by numbers and ethanol control by E. The monochrome lines represent corresponding fitting curves (Platt et al., 1980).

Five key photosynthetic parameters derived from fluorescence measurement were illustrated their variation under different treatments at 6 measuring time (Fig.3).

Revealed by $r\text{ETR}_{\text{max}}$, the diuron toxicity distinctly affected MPB photosynthetic ability under different treatments (Fig. 3a). It showed that the $r\text{ETR}_{\text{max}}$ under high concentrations of 40, 50 and 60 $\mu\text{g}\cdot\text{L}^{-1}$ were significantly lower than other treatments ($P < 0.01$ or 0.05), especially at the early period ($P < 0.01$). There was no difference of 10 $\mu\text{g}\cdot\text{L}^{-1}$ diuron with controls on $r\text{ETR}_{\text{max}}$ throughout the whole experiment ($P > 0.05$). Comparing to blank control, the inhibition of 20- 60 $\mu\text{g}\cdot\text{L}^{-1}$ diuron on $r\text{ETR}_{\text{max}}$ was more distinct under 1 h exposure, but for lower concentrations of 20 and 30 $\mu\text{g}\cdot\text{L}^{-1}$ diuron, the inhibition recovered from 2 h. However, for high concentrations of 40 to 60 $\mu\text{g}\cdot\text{L}^{-1}$

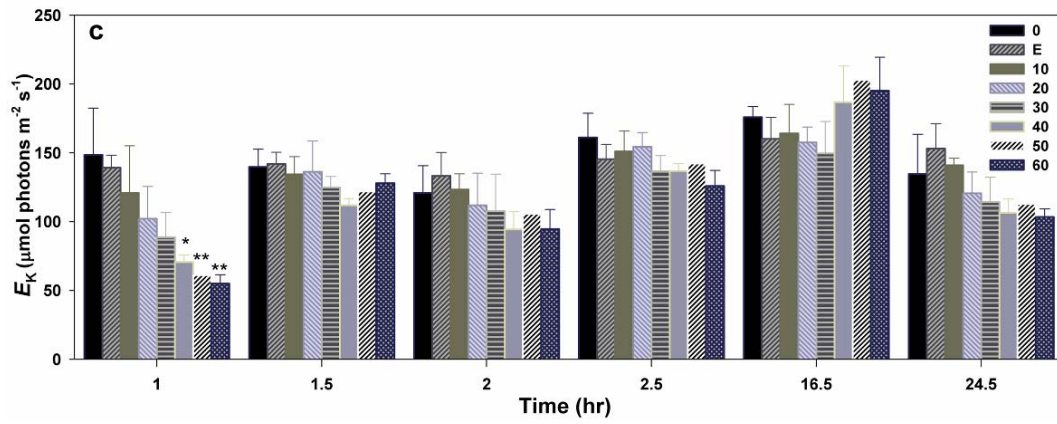
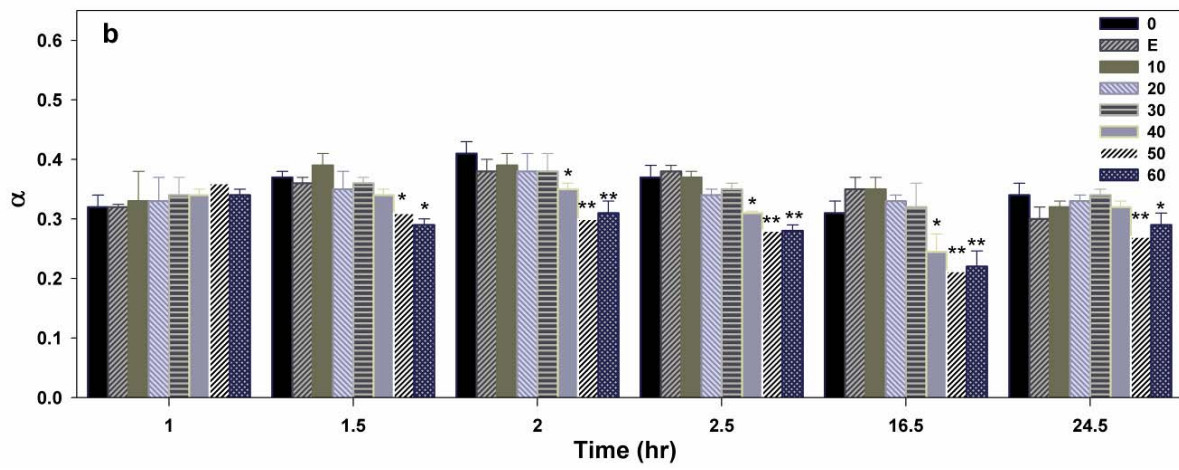
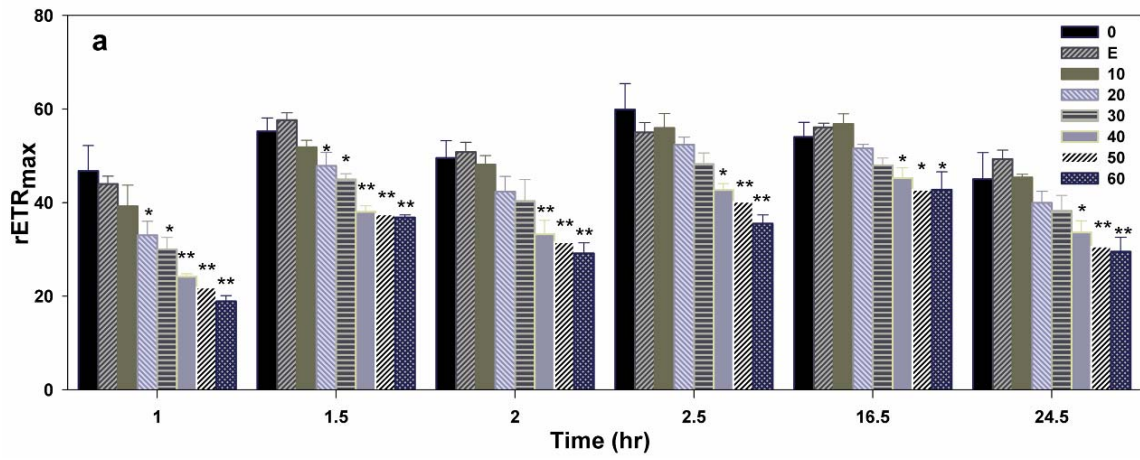
1 diuron, no recovery happened until the end of the experiment (24.5 h).

2 The diuron effect on the maximum light utilization coefficient (α) exhibited less difference than
3 on $rETR_{max}$ (Fig. 3b). There was no difference of α among all treatments under 1 h exposure.
4 Comparing to blank control, 10-30 $\mu\text{g L}^{-1}$ diuron had no significant effect on α throughout the
5 experiment ($P > 0.05$), 40 $\mu\text{g L}^{-1}$ diuron induced significantly different α from 1.5 h to 16.5 h
6 exposure ($P < 0.01$ or 0.05), while 50 and 60 $\mu\text{g L}^{-1}$ diuron resulted in significant difference from 1.5
7 h till to the end of the experiment (24.5 h) ($P < 0.01$ or 0.05).

8 For the minimum saturation irradiance (E_k), only 40-60 $\mu\text{g L}^{-1}$ diuron caused significantly
9 different E_k with controls under 1 h exposure ($P < 0.05$, Fig.3c). After 1 h, no significant difference
10 existed in E_k of each treatment from control.

11 The maximal PS II quantum yield (F_v/F_m) which indicated the photosynthetic capacity presented
12 the similar trend as the $rETR_{max}$ (Fig. 3d). Besides no difference on F_v/F_m between 10 $\mu\text{g L}^{-1}$ diuron
13 and controls ($P > 0.05$), the difference among treatments was still distinct throughout the experiment.
14 For 20 and 30 $\mu\text{g L}^{-1}$ diuron, the significant impacts on F_v/F_m recovered for a longer time as 16.5 h
15 and 24.5 h respectively, however, for high concentrations of 40 to 60 $\mu\text{g L}^{-1}$ diuron, the effects were
16 significant throughout the whole experiment ($P < 0.01$ or 0.05).

17 Comparing to above 4 parameters, the non-photochemical quenching (NPQ) exhibited more
18 distinct difference among treatments at each measuring time, except at the dawn of the second day
19 (16.5h, Fig.3e). The 10 $\mu\text{g L}^{-1}$ diuron still showed no significant effect on NPQ indicated by no
20 difference with the controls ($P > 0.05$). The 20 and 30 $\mu\text{g L}^{-1}$ diuron had identical significant effects
21 on lowering NPQ from 1 h to 2.5 h exposure ($P < 0.01$ or 0.05). At the dawn of the second day
22 (16.5h), it showed the effects of 20 and 30 $\mu\text{g L}^{-1}$ diuron were not significant due to the whole
23 dropping of NPQ values, however, for 30 $\mu\text{g L}^{-1}$ diuron the effect became significant again at 24.5 h
24 along with increased NPQ of all treatments ($P < 0.05$). The high diuron concentrations of 40, 50 and
25 60 $\mu\text{g L}^{-1}$ brought significant impacts on NPQ over whole experiment ($P < 0.01$ or 0.05) and
26 exhibited significant difference among themselves in the first day from 1.5 h to 2.5 h exposure ($P <$
27 0.01 or 0.05).



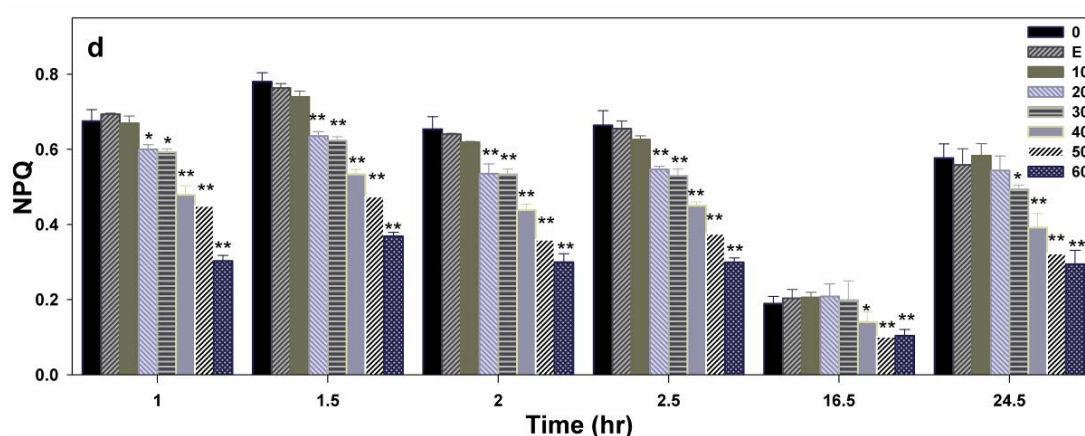
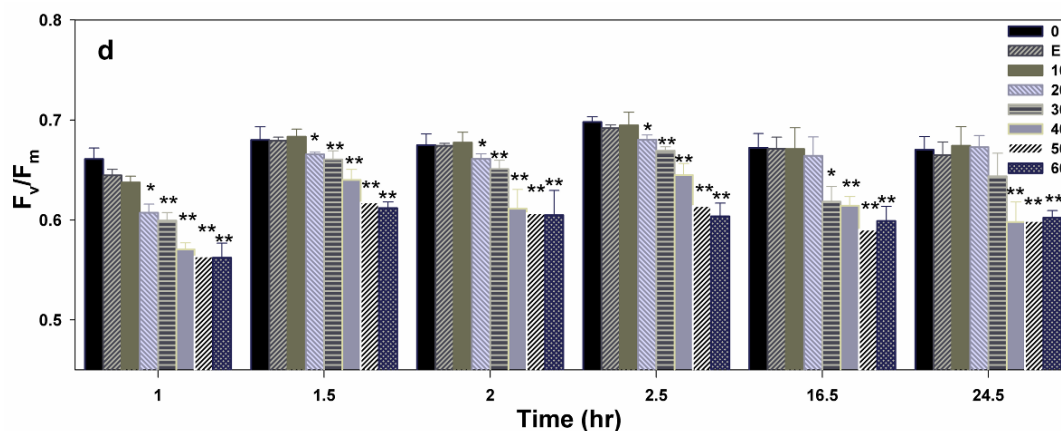


Fig 3. Variations of photosynthetic parameters under different treatments as a function of experiment time (hr). a, maximum relative electron transport rate ($rETR_{max}$); b, light utilization coefficient (α); c, minimum saturation irradiance (E_k); d, maximal PS II quantum yield (F_v/F_m); e, non-photochemical quenching (NPQ). Concentrations of 0, 10, 20, 30, 40, 50, 60 $\mu g \cdot L^{-1}$ diuron are represented by numbers, and ethanol control is labelled as E. Symbols * and ** denote the statistical difference of each treatment from the blank control (0) significantly by $P < 0.05$ and $P < 0.01$, respectively.

4. Discussion

Diuron effects on microphytobenthos (MPB) photosynthesis

Diuron inhibition on photosynthesis has been observed in all kinds of aquatic plants and algae, with differences in both toxicity degree and minimum toxicity concentration both of which depend on exposure time. For brown macroalga *Saccharina japonica*, after 2 weeks exposure to diuron, it

presented a concentration dependent decrease on photosynthetic parameters of F_v/F_m and ETR_{max} along with incremental concentration from 0.1 to 0.4 mg L⁻¹ (in water) and a complete inhibition of electron transport at 0.4 mg L⁻¹ (Kumar et al., 2010). For seagrass *Zostera marina*, the EC20 of diuron revealed that the lowest concentration was 0.5 µg L⁻¹ for significant effect on growth and photosynthetic efficiency in 10 days (Chesworth et al., 2004). Other two tropical seagrass *Halodule uninervis* and *Z. mueller* were significantly inhibited by 0.3 µg L⁻¹ diuron on their photosynthetic efficiency (F_v/F_m and Y(II)) over a 11-week exposure period (Negri et al., 2015). In the first study of diuron toxic effects on tropical MPB (diatom *Navicula* sp. and chlorophyte *Nephroselmis pyriformis*), growth and photosynthetic efficiency were reduced by 10% when exposed to diuron concentrations as low as 0.5~2µg L⁻¹ for 3 days (Magnusson et al., 2008).

These above studies were all carried out for a longer exposure time from 3 days to 11-week and tested the diuron toxicity in water. However, in the present study, natural MPB assemblages was used to test the diuron toxicity in sediments during shorter time within 24.5 h. It shown that the lowest diuron concentration of 10 µg L⁻¹ had no obvious effect on MPB photosynthesis, while, from 20 µg L⁻¹ to 60µg L⁻¹, it presented a concentration dependent decrease in the photosynthetic activity. Under this situation, the minimum toxicity concentration on MPB could be 40µg L⁻¹ for 1 h till to 24.5 h. Because the impact of 20 or 30 µg L⁻¹ was mostly evident within 2 h and could recovered sooner or later (2 h or 16.5 h) according to individual parameters, if using them as minimum toxicity concentration, the testing time and parameters should be considered together. Nevertheless, our study suggested a very short time as 1 h for assessing diuron toxicity. Cho et al. (2008) also proposed that short time as 2 h for suspended microalgae to predict EC50 values using a photosynthetic activity measurement system (substantially dissolved oxygen meter) for rapid toxicity assessment. It corroborated our study on supplying a potential rapid method instead of the conventional algal assay, in which 96 h is recommended for assessing toxicity based on algal growth rate (U.S. EPA, 1996 and OECD guidelines, 2002).

Previous studies almost tested diuron toxicity in water on aquatic photosynthetic organisms. Even for MPB Magnusson et al. (2008 and 2010) used suspended cells of tested species and developed an artificial epiphytic biofilm (2012) for assessing, it still tested water phase toxicity. Without directly applying diuron in sediments, it hardly reflects the natural response of benthic flora

1 in coastal sediments. Furthermore, diuron, and its toxic degradation products, could persist longer in
2 sediments than in water, potentially imposing longer lasting toxicity (Tixier et al., 2001). Therefore,
3 through mimicking the exposure of MPB in freshly collected mudflat sediments to diuron, our study
4 is specifically practical for assessment diuron toxicity in coastal sediment.

5 It was also noteworthy that, in this study, the recovery of MPB photosynthetic ability under 20
6 and 30 $\mu\text{g L}^{-1}$ diuron concentrations was different according to respective parameters, i.e. recovery
7 was observed after 2 h exposure for $rETR_{\text{max}}$, while it needed the overnight period for NPQ and
8 F_v/F_m . Such recovery was also observed in other aquatic photosynthetic organisms and very much
9 species-dependent (Negri et al., 2005; Magnusson et al., 2012; Burns et al., 2015; Esteves et al.,
10 2017). Among them, diatoms are often considered as more resistant to and more rapidly recoverable
11 from diuron toxicity (Juneau et al., 2007; Magnusson et al., 2012; Stachowski-Haberkorn, et al.,
12 2013), which possibly most thanks to the capability of diatom frustules (porous silica cell-wall) to
13 sort and filter nutrients from harmful agents (De Tomassi et al., 2017).

14
15 *Diuron toxicity assessment on MPB using chlorophyll fluorescence-derived photosynthetic*
16 *parameters*

17 To assess the environmental toxicity of a contaminant, a biological sensitive parameter with
18 clear concentration-dependent ranging values needs to be defined as a solid toxicity indicator. The
19 chlorophyll fluorescence-derived photosynthetic parameters firstly contribute to a rapid, sensitive
20 and non-invasive assessment based on photosynthetic activity (Juneau et al. 2007; Baker 2008;
21 Murchie et al. 2013). Furthermore, coupled with a well-plate system, the imaging fluorometry
22 (Imaging-PAM) used in this study realized a monitoring on multi-samples at the same time.

23 As referenced above, several previous studies also used chlorophyll fluorometry to assess the
24 toxicity of diuron on benthic-living photosynthetic organisms, such as F_v/F_m and ETR_{max} used for
25 brown macroalga (Kumar et al., 2010), F_v/F_m and $Y(\text{II})$ for tropical seagrass (Negri et al., 2015),
26 $Y(\text{II})$ for tropical benthic microalgae (Magnusson et al., 2008 and 2010). Juneau et al. (2007)
27 reviewed chlorophyll fluorescence-derived parameters and considered the F_v/F_m may not be the best
28 parameter for evaluating toxic effect of herbicides due to its relatively low sensitivity, and rather,
29 proposed better use $Y(\text{II})$ and NPQ since they better integrate the physiological status of the plant.

In the present study, five derived parameters, $rETR_{max}$, α , E_k , F_v/F_m , as well as NPQ, were assessed in determining diuron effect. Although all photosynthetic parameters were lowered during the 24.5 h diuron treatment, the effective concentrations as well as the timings differed among parameters. The α showed a delay reaction over 1 h, and was the most insensitive of the parameters examined, exhibiting relatively stable values especially under 0-30 $\mu\text{g L}^{-1}$ diuron range. The other derived parameter, E_k presented a more volatile and irregular trends. Comparatively, $rETR_{max}$, F_v/F_m and NPQ were more sensitive and reliable parameters with clear concentration-dependent ranging values. Although for most tested parameters, the minimum toxicity concentration could be 40 $\mu\text{g L}^{-1}$ diuron as its throughout significant influence, on the other hand, with respect to individual parameters such as $rETR_{max}$ and F_v/F_m , it could be the lowest as 20 $\mu\text{g L}^{-1}$ for only 1 h exposure time. Besides, compared to others, NPQ could better distinguish the difference among individual diuron concentrations. Juneau et al. (2007) proposed to use a combination of various fluorescence parameters to provide complementary information on the mode of herbicide action. Therefore, based on our study and previous researches, we suggested $rETR_{max}/F_v/F_m$ and NPQ as the combination indicators for diuron toxicity in sediments. Our study also validated the utility of imaging chlorophyll fluorometry as a rapid and reliable technique to measure sub-lethal toxicity threshold.

Indirect effect of diuron on MPB photosynthesis inhibition

Because photosynthesis is a central process that supports the whole cell metabolism, its inhibition could cause potential effects on other activities. Staats et al. (2000) observed that the secretion of exopolysaccharide (EPS) by motile benthic diatoms was decreased after diuron inhibition on photosynthesis. EPS keenly associates with diatoms motility, especially in building the trails for diatom motility on substrate surface (Edgar and Pickett-Heaps, 1984; Hoagland et al. 1993, Molino and Wetherbee, 2008). Even Coquillé et al. (2015) suggested to use diatom motility features as endpoints of herbicide metolachlor toxicity. However, in our study, there was no evidence of diuron-driven differences in vertical migration of natural MPB assemblages during 24.5 hours. Instead, in our conditions, MPB abundance at the surface of sediment indicated by F_t went on varying at the function of photoperiod (and probably of the tidal cycle), with striking increase of F_t before day starts (Fig.1) as previously reported before (Consalvey et al. 2004; Barnett et al. 2020). It

1 was consistent with the observation of Staats et al. (2000) that *in situ* patterns of diuron-treated-MPB
2 vertical migration in the dark did not differ from those exposed to light during 24 h field experiment
3 on mudflat surface. Both studies indicated that although light steers MPB vertical migration, the
4 decrease in photosynthetic activity itself does not seem to directly influence this directive motility of
5 MPB in a short time as 24 h. This phenomenon, from other side, also endows MPB a special ability
6 to actively avoid further exposing to diuron as benthic motile diatoms did in escaping from herbicide
7 metolachlor (Coquillé et al. 2015) and directing to dissolved silicate (Bondoc et al. 2016). Further
8 study needs to explore the possible combined results, because it is crucial in the framework of using
9 chlorophyll fluorometry to bioassess diuron and photosynthesis-impacting contaminants in coastal
10 sediments.

11 As natural assemblages in coastal flats, MPB community dynamics in species composition and
12 biomass are close related to a complex set of environmental factors (MacIntyre et al. 1996;
13 Underwood and Kromkamp 1999). The effect of diuron on MPB also depends on environmental
14 factors such as light and temperatures (Camuel et al., 2017; Chaumet et al., 2020). Therefore, it
15 should be considered these factors when assessing diuron toxicity *in situ* in different seasons and
16 regions. Furthermore, the influence of diuron on MPB photosynthesis inevitably firstly impacts MPB
17 functions as primary producer in coastal ecosystem. Secondly, according to the findings of
18 class-specific responses of marine benthic microalgae (Magnusson et al., 2008, 2010 and 2012) and
19 species-specific sensitivity of benthic diatoms (Larras et al., 2012; Wood et al., 2016) to herbicide,
20 chronic exposures to diuron also could in turn affect the sensitivity and community structure of MPB
21 (Magnusson et al., 2012).

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Declaration of competing interest

The authors declare that we have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

CRediT authorship contribution statement

Guo-ying Du: planning, conceptualization, methodology, performed the experiments and drafted the original manuscript, **Xue-feng Zhong** and **Shuai Che:** performed the data analysis, editing the manuscript; **Christine Dupuy:** project administration and supervision, funding acquisition, **Johann Lavaud:** planning, methodology, supervision, reviewing and editing the manuscript.

References

- Admiraal, W., Peletier, H., & Brouwer, T. (1984). The seasonal succession patterns of diatom species on an intertidal mudflat: an experimental analysis. *Oikos*, 30-40. <https://doi.org/10.2307/3544606>
- Ali, H.R., Arifin, M.M., Sheikh, M.A., Shazili, N.A.M., Bakari, S.S., & Bachok, Z. (2014). Contamination of diuron in coastal waters around Malaysian Peninsular. *Marine pollution bulletin*, 85(1), 287-291. <https://doi.org/10.1016/j.marpolbul.2014.05.049>
- Australian Government. (2011). Diuron environment assessment. Australian Government-Australian Pesticides and Veterinary Medicines Authority.
- Baker, N.R. (2008). Chlorophyll fluorescence: a probe of photosynthesis in vivo. *Annual Review of Plant Biology* 59:89-113. DOI: [10.1146/annurev.arplant.59.032607.092759](https://doi.org/10.1146/annurev.arplant.59.032607.092759)
- Barnett, A., Méléder, V., Dupuy, C., & Lavaud, J. (2020). The vertical migratory rhythm of intertidal microphytobenthos in sediment depends on the light photoperiod, intensity, and spectrum: Evidence for a positive effect of blue wavelengths. *Frontiers in Marine Science*, 7. <http://dx.doi.org/10.3389/fmars.2020.00212>
- Barnett, A., Méléder, V., Blommaert, L., Lepetit, B., Gaudin, P., Vyverman, W., Sabbe, K., Dupuy, C., & Lavaud, J. (2015). Growth form defines physiological photoprotective capacity in intertidal benthic diatoms. *The ISME journal*, 9(1), 32-45. <https://doi.org/10.1038/ismej.2014.105>
- Bere, T., & Tundisi, J.G. (2012). Diatom communities as indicator of ecological impairment in rivers: Conservation and water quality management. Lambert Academic Publishing. Saarbrücken, Germany. DOI:10.13140/2.1.3087.8722.
- Bondoc, K., Heuschele, J., Gillard, J., Vyverman, W., & Pohnert, G. (2016). Selective silicate-directed motility in diatoms. *Nature communications* 7, 10540. <https://doi.org/10.1038/ncomms10540>
- Bruggen, B.V.D. & Vandecasteele, C. (2003). Removal of pollutants from surface water and groundwater by nanofiltration: overview of possible applications in the drinking water industry. *Environmental Pollution*, 122, 435–445. [https://doi.org/10.1016/S0269-7491\(02\)00308-1](https://doi.org/10.1016/S0269-7491(02)00308-1)
- Burns, M., Hanson, M.L., Prosser, R. S., Crossan, A.N., & Kennedy, I.R. (2015). Growth recovery of *Lemna gibba* and *Lemna minor* following a 7-day exposure to the herbicide diuron. *Bulletin of environmental contamination and toxicology*, 95(2), 150-156. <https://doi.org/10.1007/s00128-015-1575-8>
- Camuel, A., Guieysse, B., Alcántara, C., & Béchet, Q. (2017). Fast algal eco-toxicity assessment: influence of light intensity and exposure time on *Chlorella vulgaris* inhibition by atrazine and DCMU. *Ecotoxicology and environmental safety*, 140, 141-147. <https://doi.org/10.1016/j.ecoenv.2017.02.013>
- Cartaxana, P., Cruz, S., Gameiro, C., & Köhl, M. (2016). Regulation of intertidal microphytobenthos photosynthesis over a diel emersion period is strongly affected by diatom migration patterns. *Frontiers in Microbiology*, 7, 872. <https://doi.org/10.3389/fmicb.2016.00872>
- Cartaxana, P., Ruivo, M., Hubas, C., Davidson, I., Serôdio, J., & Jesus, B. (2011). Physiological versus behavioral photoprotection in intertidal epipelagic and epipsammic benthic diatom communities. *Journal of Experimental Marine Biology and Ecology*, 405(1-2), 120-127. <https://doi.org/10.1016/j.jembe.2011.05.027>
- Chaumet, B., Mazzella, N., Neury-Ormanni, J., & Morin, S. (2020). Light and temperature influence on diuron bioaccumulation and toxicity in biofilms. *Ecotoxicology*, 29(2), 185-195. <https://doi.org/10.1007/s10646-020-02166-8>
- Chen, X., Zhou, W., Pickett, S.T., Li, W., Han, L., & Ren, Y. (2016). Diatoms are better indicators of urban stream conditions: A case study in Beijing, China. *Ecological Indicators*, 60, 265-274. <https://doi.org/10.1016/j.ecolind.2015.06.039>
- Chesworth, J.C., Donkin, M.E., & Brown, M.T. (2004). The interactive effects of the antifouling herbicides Irgarol 1051 and Diuron on the seagrass *Zostera marina* (L.). *Aquatic toxicology*, 66(3), 293-305.

- <https://doi.org/10.1016/j.aquatox.2003.10.002>
- Cho, C.W., Pham, T.P.T., Jeon, Y.C., Min, J., Jung, H.Y., Lee, D.S., & Yun, Y.S. (2008). Microalgal photosynthetic activity measurement system for rapid toxicity assessment. *Ecotoxicology*, 17(6), 455-463.
- <https://doi.org/10.1007/s10646-008-0197-x>
- Coelho-Moreira, J.D.S., Brugnari, T., Sá-Nakanishi, A.B., Castoldi, R., De Souza, C.G., Bracht, A., & Peralta, R.M. (2018). Evaluation of diuron tolerance and biotransformation by the white-rot fungus *Ganoderma lucidum*. *Fungal biology*, 122(6), 471-478. <https://doi.org/10.1016/j.funbio.2017.10.008>
- Consalvey, M., Paterson, D.M., & Underwood, G.J. (2004). The ups and downs of life in a benthic biofilm: migration of benthic diatoms. *Diatom Research*, 19(2), 181-202. <https://doi.org/10.1080/0269249X.2004.9705870>
- Coste, M., Boutry, S., Tison-Rosebery, J., & Delmas, F. (2009). Improvements of the Biological Diatom Index (BDI): Description and efficiency of the new version (BDI-2006). *Ecological Indicators*, 9 (4), 621-650.
- <https://doi.org/10.1016/j.ecolind.2008.06.003>.
- Desrosiers, C., Leflaive, J., Eulin, A., Ten-Hage, L. (2013). Review Bioindicators in marine waters: Benthic diatoms as a tool to assess water quality from eutrophic to oligotrophic coastal ecosystems. *Ecological Indicators*. 32, 25-34.
- DOI :10.1016/j.ecolind.2013.02.021
- De Tommasi, E., Gielis, J., & Rogato, A. (2017). Diatom frustule morphogenesis and function: a multidisciplinary survey. *Marine genomics*, 35, 1-18. <https://doi.org/10.1016/j.margen.2017.07.001>
- Du, G.Y., Yan, H.M., & Dupuy, C. (2017). Microphytobenthos as an indicator of environmental quality status in intertidal flats: Case study of coastal ecosystem in Pertuis Charentais, France. *Estuarine, Coastal and Shelf Science*, 196, 217-226. <https://doi.org/10.1016/j.ecss.2017.06.031>
- Du, G.Y., Yan, H.M., Liu, C.R., & Mao, Y.X. (2018). Behavioral and physiological photoresponses to light intensity by intertidal microphytobenthos. *Journal of Oceanology and Limnology*, 36(2), 293-304. <https://doi.org/10.1007/s00343-017-6099-0>
- Duke, N.C. (2008). Corrections and updates to the article by Duke et al. (2005) reporting on the unusual occurrence and cause of dieback of the common mangrove species, *Avicennia marina*, in NE Australia. *Marine pollution bulletin*, 56(9), 1668. <https://doi.org/10.1016/j.marpolbul.2008.08.001>
- Duke, N.C., Bell, A. M., Pederson, D.K., Roelfsema, C. M., & Nash, S. B. (2005). Herbicides implicated as the cause of severe mangrove dieback in the Mackay region, NE Australia: consequences for marine plant habitats of the GBR World Heritage Area. *Marine Pollution Bulletin*, 51(1-4), 308-324. <https://doi.org/10.1016/j.marpolbul.2004.10.040>
- Edgar, L.A., & Pickett-Heaps, J.D. (1984). Diatom locomotion. In *Progress in phycological research*, Vol.3 (Round, F. E. & Chapman, G., editors), 47-88. Biopress, Bristol.
- Esteves, S. M., Keck, F., Almeida, S. F., Figueira, E., Bouchez, A., & Rimet, F. (2017). Can we predict diatoms herbicide sensitivities with phylogeny? Influence of intraspecific and interspecific variability. *Ecotoxicology*, 26(8), 1065-1077. <https://doi.org/10.1007/s10646-017-1834-z>
- Frankenbach, S., Schmidt, W., Frommlet, J.C., & Serôdio, J. (2018). Photoinactivation, repair and the motility-physiology trade-off in microphytobenthos. *Marine Ecology Progress Series*, 601, 41-57. <https://doi.org/10.3354/meps12670>
- Genty B, Briantais JM, Baker NR. 1989. The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta*, 990: 87 – 92. [https://doi.org/10.1016/S0304-4165\(89\)80016-9](https://doi.org/10.1016/S0304-4165(89)80016-9)
- Haynes, D., Ralph, P., Prange, J., & Dennison, B. (2000). The impact of the herbicide diuron on photosynthesis in three species of tropical seagrass. *Marine pollution bulletin*, 41(7-12), 288-293. [https://doi.org/10.1016/S0025-326X\(00\)00127-2](https://doi.org/10.1016/S0025-326X(00)00127-2)

- 1 Hoagland, K.D., Rosowski, J.R., Gretz, M.R., & Roemer, S.C. (1993). Diatom extracellular polymeric substances:
2 function, fine structure, chemistry, and physiology. *Journal of phycology*, 29(5), 537-566.
3 <https://doi.org/10.1111/j.0022-3646.1993.00537.x>
- 4 Holmes, G. (2014). Australia's pesticide environmental risk assessment failure: The case of diuron and sugarcane. *Marine*
5 *pollution bulletin*, 88(1-2), 7-13. <https://doi.org/10.1016/j.marpolbul.2014.08.007>
- 6 Jones, R. (2005). The ecotoxicological effects of Photosystem II herbicides on corals. *Marine Pollution Bulletin*, 51(5-7),
7 495-506. <https://doi.org/10.1016/j.marpolbul.2005.06.027>
- 8 Jones, R. J., Muller, J., Haynes, D., & Schreiber, U. (2003). Effects of herbicides diuron and atrazine on corals of the
9 Great Barrier Reef, Australia. *Marine Ecology Progress Series*, 251, 153-167. <http://doi.org/10.3354/meps251153>
- 10 Juneau, P., Qiu, B., & Deblois, C. P. (2007). Use of chlorophyll fluorescence as a tool for determination of herbicide toxic
11 effect. *Toxicological and Environ Chemistry*, 89(4), 609-625. <https://doi.org/10.1080/02772240701561569>
- 12 Kelly, M.G., & Whitton, B.A. (1995). The trophic diatom index: a new index for monitoring eutrophication in
13 rivers. *Journal of Applied Phycology*, 7(4), 433-444. <https://doi.org/10.1007/BF00003802>
- 14 Kennedy, K., Schroeder, T., Shaw, M., Haynes, D., Lewis, S., Bentley, C., Paxman, C., Carter, S., Brando, V.E., Bartkow,
15 M., Hearn, L. & Mueller, J.F. (2012). Long term monitoring of photosystem II herbicides–Correlation with remotely
16 sensed freshwater extent to monitor changes in the quality of water entering the Great Barrier Reef,
17 Australia. *Marine Pollution Bulletin*, 65(4-9), 292-305. <https://doi.org/10.1016/j.marpolbul.2011.10.029>
- 18 Kumar, K. S., Choo, K.S., Yea, S.S., Seo, Y., & Han, T. (2010). Effects of the phenylurea herbicide diuron on the
19 physiology of *Saccharina japonica* Aresch. *Toxicology and Environmental Health Sciences*, 2(3), 188-199.
20 <http://doi.org/10.1007/BF03216505>
- 21 Larras, F., Bouchez, A., Rimet, F. & Montuelle, B. (2012) Using Bioassays and Species Sensitivity Distributions to
22 Assess Herbicide Toxicity towards Benthic Diatoms. *PLoS ONE* 7(8): e44458.
23 <https://doi.org/10.1371/journal.pone.0044458>
- 24 Lavaud, J., & Goss, R. (2014). The peculiar features of non-photochemical fluorescence quenching in diatoms and brown
25 algae. In *Non-photochemical quenching and energy dissipation in plants, algae and cyanobacteria*, Advances in
26 Photosynthesis and Respiration 40. 421-443. Springer, Dordrecht. https://doi.org/10.1007/978-94-017-9032-1_20
- 27 Laviale, M., Barnett, A., Ezequiel, J., Lepetit, B., Frankenbach, S., Méléder, V., Serôdio, J., & Lavaud, J. (2015).
28 Response of intertidal benthic microalgal biofilms to a coupled light–temperature stress: evidence for latitudinal
29 adaptation along the Atlantic coast of Southern Europe. *Environmental microbiology*, 17(10), 3662-3677.
30 <https://doi.org/10.1111/1462-2920.12728>
- 31 Lewis, S. E., Schaffelke, B., Shaw, M., Bainbridge, Z. T., Rohde, K. W., Kennedy, K., Davis, A. M., Masters, B. L.,
32 Devlin, M. J., Mueller, J. F., & Brodie, J. E. (2012). Assessing the additive risks of PSII herbicide exposure to the
33 Great Barrier Reef. *Marine Pollution Bulletin*, 65(4-9), 280-291. <https://doi.org/10.1016/j.marpolbul.2011.11.009>
- 34 MacIntyre, H.L., Geider, R.J., & Miller, D.C. (1996). Microphytobenthos: the ecological role of the “secret garden” of
35 unvegetated, shallow-water marine habitats. I. Distribution, abundance and primary production. *Estuaries*, 19(2),
36 186-201. <https://doi.org/10.2307/1352224>
- 37 Magnusson, M., Heimann, K., & Negri, A.P. (2008). Comparative effects of herbicides on photosynthesis and growth of
38 tropical estuarine microalgae. *Marine Pollution Bulletin*, 56(9), 1545-1552.
39 <https://doi.org/10.1016/j.marpolbul.2008.05.023>
- 40 Magnusson, M., Heimann, K., Quayle, P., & Negri, A.P. (2010). Additive toxicity of herbicide mixtures and comparative
41 sensitivity of tropical benthic microalgae. *Marine Pollution Bulletin*, 60(11), 1978-1987.
42 <https://doi.org/10.1016/j.marpolbul.2010.07.031>
- 43 Magnusson, M., Heimann, K., Ridd, M., & Negri, A.P. (2012). Chronic herbicide exposures affect the sensitivity and
44 community structure of tropical benthic microalgae. *Marine Pollution Bulletin*, 65(4-9), 363-372.

- <https://doi.org/10.1016/j.marpolbul.2011.09.029>
- Maxwell, K., Johnson, G.N. (2000). Chlorophyll fluorescence--a practical guide. *Journal of experimental botany*, 51(345):659-668. [https://doi.org/10.1016/S0304-4165\(89\)80016-9](https://doi.org/10.1016/S0304-4165(89)80016-9)
- Molino, P.J., & Wetherbee, R. (2008). The biology of biofouling diatoms and their role in the development of microbial slimes. *Biofouling*, 24(5), 365-379. <https://doi.org/10.1080/08927010802254583>
- Mukherjee, A., Rao, K.M., & Ramesh, U.S. (2009). Predicted concentrations of biocides from antifouling paints in Visakhapatnam Harbour. *Journal of environmental management*, 90, S51-S59. <https://doi.org/10.1016/j.jenvman.2008.07.018>
- Murchie, E.H., Lawson, T. (2013). Chlorophyll fluorescence analysis: a guide to good practice and understanding some new applications. *Journal of Experimental Botany*. 64:3893-3898. DOI: [10.1093/jxb/ert208](https://doi.org/10.1093/jxb/ert208)
- Negri, A.P., Flores, F., Mercurio, P., Mueller, J.F., & Collier, C. J. (2015). Lethal and sub-lethal chronic effects of the herbicide diuron on seagrass. *Aquatic Toxicology*, 165, 73-83. <https://doi.org/10.1016/j.aquatox.2015.05.007>
- Negri, A.P., Flores, F., Röthig, T., & Uthicke, S. (2011). Herbicides increase the vulnerability of corals to rising sea surface temperature. *Limnology and Oceanography*, 56(2), 471-485. <https://doi.org/10.4319/lo.2011.56.2.0471>
- Negri, A., Vollhardt, C., Humphrey, C., Heyward, A., Jones, R., Eaglesham, G., & Fabricius, K. (2005). Effects of the herbicide diuron on the early life history stages of coral. *Marine Pollution Bulletin*, 51(1-4), 370-383. <https://doi.org/10.1016/j.marpolbul.2004.10.053>
- Nitschke, L., & Schüssler, W. (1998). Surface water pollution by herbicides from effluents of waste water treatment plants. *Chemosphere*, 36(1), 35-41. [https://doi.org/10.1016/S0045-6535\(97\)00286-5](https://doi.org/10.1016/S0045-6535(97)00286-5)
- OECD. (2002). Guideline for testing of chemicals. Proposal for updating guideline 201, freshwater alga and cyanobacteria, growth inhibition test., Organization for the Economic Cooperation and Development.
- OECD. (2006). OECD Guideline 201. Guidelines for the testing of chemicals. Freshwater alga and cyanobacteria, growth inhibition test., Organization for the Economic Cooperation and Development. <https://doi.org/10.1787/9789264069923-en>
- Platt, T. G. C. L., Gallegos, C. L., & Harrison, W. G. (1980). Photoinhibition of photosynthesis in natural assemblages of marine phytoplankton. *Journal of Marine Research*, 38(4), 687-701. <https://images.peabody.yale.edu/publications/jmr/jmr38-04-06.pdf>
- Pniewski, F., & Piasecka-Jędrzejak, I. (2020). Photoacclimation to constant and changing light conditions in a benthic diatom. *Frontiers in Marine Science*. <https://doi.org/10.3389/fmars.2020.00381>
- Potapova, M., & Charles, D.F. (2007). Diatom metrics for monitoring eutrophication in rivers of the United States. *Ecological indicators*, 7(1), 48-70. <https://doi.org/10.1016/j.ecolind.2005.10.001>
- Serôdio, J., Coelho, H., Vieira, S., & Cruz, S. (2006). Microphytobenthos vertical migratory photoresponse as characterised by light-response curves of surface biomass. *Estuarine, Coastal and Shelf Science*, 68(3-4), 547-556. <https://doi.org/10.1016/j.ecss.2006.03.005>
- Shaw, M., Silburn, D.M., Thornton, C., Robinson, B., & McClymont, D., (2011). Modelling pesticide runoff from paddocks in the Great Barrier Reef using HowLeaky. In *MODSIM 2011 - 19th International Congress on Modelling and Simulation - Sustaining Our Future: Understanding and Living with Uncertainty*. 2057-2063. <https://www.mendeley.com/catalogue/44ec9b6d-cdfe-321e-86a1-320f5e314d7f/>
- Simões, M.D.S., Bracht, L., Parizotto, A.V., Comar, J.F., Peralta, R.M., & Bracht, A. (2017). The metabolic effects of diuron in the rat liver. *Environmental Toxicology and Pharmacology*, 54, 53-61. <https://doi.org/10.1016/j.etap.2017.06.024>
- Smith, R., Middlebrook, R., Turner, R., Huggins, R., Vardy, S., & Warne, M. (2012). Large-scale pesticide monitoring across Great Barrier Reef catchments--paddock to reef integrated monitoring, modelling and reporting program. *Marine pollution bulletin*, 65(4-9), 117-127. <https://doi.org/10.1016/j.marpolbul.2011.08.010>

- Staats, N., Stal, L. J., de Winder, B., & Mur, L. R. (2000). Oxygenic photosynthesis as driving process in exopolysaccharide production of benthic diatoms. *Marine Ecology Progress Series*, 193, 261-269. <http://doi.org/10.3354/meps193261>
- Stachowski-Haberkorn, S., Jérôme M., Rouxel, J., Khelifi, C., Rincé, M., Burgeot, T. (2013). Multigenerational exposure of the microalga *Tetraselmis suecica* to diuron leads to spontaneous long-term strain adaptation, *Aquatic Toxicology*, 140–141, 380-388. <https://doi.org/10.1016/j.aquatox.2013.06.016>.
- Stevenson, R.J., Pan, Y., & Van Dam, H. (2010). Assessing environmental conditions in rivers and streams with diatoms. In *The diatoms: applications for the environmental and earth sciences*, 1(4),57–85. Cambridge University Press, Cambridge.
- Stingaciu, L.R., O'Neill, H. M., Liberton, M., Pakrasi, H. B., & Urban, V. S. (2019). Influence of chemically disrupted photosynthesis on cyanobacterial thylakoid dynamics in *Synechocystis* sp. PCC 6803. *Scientific reports*, 9(1), 1-9. <https://doi.org/10.1038/s41598-019-42024-0>
- Tixier, C., Sancelme, M., Sancelme, M., Bonnemoy, F., Cuet, A., & Veschambre, H. (2001). Degradation products of a phenylurea herbicide, diuron: synthesis, ecotoxicity, and biotransformation. *Environmental Toxicology and Chemistry: An International Journal*, 20(7), 1381-1389. <https://doi.org/10.1002/etc.5620200701>
- US Government. 2005. Bureau of Land Management. Diuron Ecological Risk Assessment. Utah State University. Bureau of Land Management, United States, Reno, Nevada. pp 105.
- Underwood, G.J.C. & Kromkamp, J. (1999). Primary production by phytoplankton and microphytobenthos in estuaries. *Advances in Ecological Research*. 29, 93–153. [http://doi.org/10.1016/S0065-2504\(08\)60192-0](http://doi.org/10.1016/S0065-2504(08)60192-0)
- USEPA. (1996). Ecological effects test guidelines OPPTS 850.5400. Algal toxicity, tiers I and II, United States Environmental Protection Agency.
- Wood, R.J., Mitrovic, S.M., Lim, R.P., & Kefford, B.J. (2016). How benthic diatoms within natural communities respond to eight common herbicides with different modes of action. *Science of The Total Environment*, 557–558, 636-643. <https://doi.org/10.1016/j.scitotenv.2016.03.142>.
- Wong, A., de Vasconcelos Lanza, M.R., & Sotomayor, M.D.P.T. (2013). Sensor for diuron quantitation based on the P450 biomimetic catalyst nickel (II) 1, 4, 8, 11, 15, 18, 22, 25-octabutoxy-29H, 31H-phthalocyanine. *Journal of Electroanalytical Chemistry*, 690, 83-88. <https://doi.org/10.1016/j.jelechem.2012.11.007>