

Origin, fate and ecotoxicity of manganese from legacy metallurgical wastes

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1	Origin, fate and ecotoxicity of manganese from legacy metallurgical wastes.
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26 Abstract

Over the course of history, mining and metallurgical activities have influenced the 27 28 socioeconomic development of human populations. However, these past and current activities can also lead to substantial environmental contamination by various metals. Here, we used an 29 30 interdisciplinary approach (incorporating archaeology, mineralogy, environmental chemistry and ecotoxicology) to investigate the origin, fate and potential ecotoxicity of anomalous 31 manganese (Mn) concentrations detected in the ancient mining district of Berthelange 32 (medieval period, eastern France). Mineralogical investigations of slag samples showed that 33 smelting temperature conditions in medieval bloomeries led to the production of slags mainly 34 composed of Fe- and Mn-rich olivine, *i.e.*, fayalites. Further mineralogical analyses of bulk 35 36 soil and clay fractions allowed us to identify the presence of serpentine. This evidence of 37 olivine weathering can account for the release of Mn from slags into the soil. In addition, chemical analyses of total and available (exchangeable and reducible) Mn concentrations in 38 39 soil samples clearly showed the contribution of slags deposited 1000 years ago to soil contamination. A complementary ecotoxicity bioassay performed on soils from a slag heap 40 using the land snail Cantareus aspersus confirmed that a significant fraction of the Mn 41 42 detected in soils remains available for partitioning with the soil solution and transfer to soil organisms. Although no growth inhibition of snails was observed after 28 days of exposure, 43 the animals accumulated quite elevated Mn concentrations in their tissues. Our study 44 emphasizes the environmental availability and bioavailability of Mn from ancient 45 metallurgical wastes to soil-dwelling invertebrates, *i.e.*, snails, even one millennium after their 46 deposition. Hence, as for more recent industrial sites, past mining ecosystems must be a cause 47 of concern for the scientific community and public authorities. 48

49 Keywords (x6)

50 slags; weathering; metals; chemical extraction; bioavailability; snails

51 **1. Introduction**

Throughout human history, and especially since the Bronze Age, socioeconomic 52 development has been strongly influenced by the exploitation of mineral resources, such as 53 copper (Cu), iron (Fe), lead (Pb) or silver (Ag) (Nriagu, 1996; Tylecote, 1987). However, 54 mineral-related activities also constitute the most ancient and significant source of waste 55 production (Macklin et al., 1997; Pyatt et al., 2005). These types of wastes, originating from 56 mining (ore dumps, ore-washing sediments) or metallurgical (smelter dusts, slags) operations, 57 are currently considered archaeological clues. Until quite recently, slags were mostly 58 considered chemically inert because of the strong association of metals with glass and mineral 59 phases, limiting their lability (Wilson et al., 1994). However, a growing body of evidence 60 shows that slags are affected by weathering and may be able to release metals into aquatic and 61 62 terrestrial compartments (Ettler, 2002 and 2016; Gee et al., 1997; Lottermoser, 2002; Parsons et al., 2001; Piatak et al., 2004; Seignez et al., 2006). 63

In addition to the modification of the geochemical cycles of metals, this contamination 64 may be responsible for the deterioration of water and soil quality and the impairment of 65 biodiversity (Ettler et al., 2009; Lottermoser and Cairns, 2005). For instance, Monna et al. 66 (2011) showed that wild brown trout living in past mining areas located in the Cevennes 67 (France) were contaminated by Pb originating from historical activities (15th and 20th 68 centuries). More recently, Mariet et al. (2016 and 2020) highlighted that metals originating 69 from mining operations (Pb and Ag) during the end of the Middle Ages in the Vosges 70 71 Mountains (France) remain bioavailable to soil invertebrates. However, the older the sites are, the scarcer the data are. In the Franche-Comté region (eastern France), Fe mining and 72 metallurgical activities began in the Early Middle Ages (5th century), with particular intensity 73 in the district of Berthelange (Forlin and Laurent, 2014; Laurent, 2016). Until the 15th century, 74 a direct ore-reducing process was conducted within clay-made blast furnaces. In these 75

bloomeries, temperatures exceeded 1000°C, allowing, with the addition of charcoal, the reduction of Fe oxides (Fe₂O₃). At the end of the smelting process, a heterogeneous block of Fe (sponge) was kept for further purification, and metal-rich residues, *i.e.*, slags, were dumped on site as heaps. Currently, more than 1000 years later, the environmental effects of potential metal releases have, to our knowledge, not been evaluated.

Therefore, this study aims to assess the effect of very ancient slag deposits on soil quality. Because a single chemical analysis of total metal concentrations in soils may be insufficient for ecological risk assessment purposes (Peijnenburg, 2020), we combined physical, chemical and biological tools in a holistic approach. We operationally investigated the metal distribution in and transfer between various matrices (slags, soil and invertebrates) to reveal the origin, fate and effects of legacy contamination.

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88 2. Materials and methods

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2.1. Study area and sampling design

This study was performed in the Bourgogne Franche-Comté region of eastern France, 90 91 where Fe mining and metallurgical exploitation were important activities during the Middle Ages (Jacob and Mangin, 1990). Over this period, Fe mining and metallurgical activities were 92 particularly intense in the district of Berthelange (140 km², 20 km west from Besancon). The 93 study area is composed of Jurassic limestones and marls (from Toarcian to Bajocian) partly 94 95 covered by clay and sandy alluvial Pliocene deposits (Dreyfuss and Kuntz, 1967). Here, more 96 than 120 direct Fe ore-reducing process sites were identified (Forlin and Laurent, 2014) because of the presence of slag repositories (from 10 to 20 m in diameter) (Figure 1A). In this 97 study, eleven slag repositories, located in forested regions and well preserved, were 98 99 investigated for the detection of potential anomalous metal concentrations in soils (Figure 1A,

Table 1). Among them, the Antorpe 9 heap (Figure 1B), containing slags produced during the 100 11th century (Forlin and Laurent, 2014), was selected for further investigations. More 101 precisely, slags and soils were sampled using a transect method following four directions (NE, 102 103 SE, SW and NW) from the centre of the slag heap (sampling point, SP, 4) according to Figure 1C. The reference site SP10 was located 100 m west of the heap. This remote station was 104 selected to represent the pedogeological background of the area studied, *i.e.*, the same 105 geological substrate and same vegetation cover, but without any smelting activities or slag 106 107 deposit. At each sampling point, three soil cores (0-100 cm) were taken (after removing surface vegetation and humus) using a soil auger. 108

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110 *2.2. Soil analyses*

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2.2.1. Physico-chemical characterisation

The soil samples were primarily sieved (2 mm mesh; slag fragments were sorted for 112 further analyses), dried at 40°C and homogenized. The soil pH was measured using a pH 113 metre (WTW, pH/ION 3310) after extraction with demineralized water (1:5, v:v: ISO 10390, 114 2005), and the organic matter (OM) content was determined via the loss of ignition (LOI) 115 method according to NF EN 15935 (AFNOR, 2013). Total metal concentrations were 116 determined after hot mineralization in aqua regia (1:2.5 HNO₃: HCl, v:v) using a block 117 digestion system (DigiPREP) and filtration (1 µm) by inductively coupled plasma atomic 118 119 emission spectroscopy (ICP-AES) (Thermo Scientific, iCAP 6000 Series). The reliability of 120 the analyses was assessed with certified reference material (calcareous loam soil: CRM n°141R from BCR). The recovery rate was on average 107±8% for all the metals analysed. 121

To identify potential decay products of slag minerals, mineralogical analyses of soil
samples (bulk and clay fraction) from different depths (0-20 cm, 20-40 cm, 40-60 cm, 60-80

cm and 80-100 cm) were obtained using X-ray diffraction (XRD) characterization and a D8 124 Advance Bruker diffractometer equipped with a LinxEye detector (CuKα radiation at 1.54184 125 Å, 40 kV/40 mA) hosted at the Utinam Institute (University Bourgogne Franche-Comté, 126 France). Powdered bulk rock samples were treated between 2θ values of 3° and 65° (divergent 127 slit) with a step scan of 0.019° (2 θ) and a scan rate of 1 s/step. The EVA software package 128 was used for data processing and phase identification. The extraction of clay fractions 129 required prior OM destruction by successive addition of 1 cm³ hydrogen peroxide (33% H₂O₂, 130 VWR Chemicals) to hydrated soil samples previously blended with water (1:5; w:v). To 131 increase the reaction rate (Holtzapffel, 1985), this mixture was heated at 60°C on a digital 132 hotplate (Stuart SD500). The samples were then subjected to repeated washing (n=4) using 133 ultrapure water (UP) and centrifugation (10 min, 8,000 rpm). Afterward, clay suspensions 134 were shaken in an end-over-end shaker for 10 min at 10 rpm. Finally, two clay fractions (<16 135 μ m and <2 μ m) were obtained by the sedimentation process according to Stoke's law, which 136 links the settling rate to the particle size, and mounted on glass slides for XRD analyses. 137 Before successive XRD analyses, the oriented preparations were sequentially air dried, 138 glycolated and heated for 4 h at 490°C for identification of the interlayer of clays, *i.e.*, the 139 (001) basal reflection positions after each treatment. The XRD patterns were obtained 140 between 2° and 32° (2 θ) with a step scan of 0.009° (2 θ) and a scan rate of 1.5 s/step. 141

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2.2.2. Environmental availability

The environmental availability was estimated using a chemical extraction method chosen to quantify the potential of metals for leaching or uptake by plants (Abedin et al., 2012). The extraction solution consisted of 1 mol.L⁻¹ ammonium acetate (NH₄OAc, purity >99%) added to a solution containing 2 g.L⁻¹ hydroxylamine hydrochloride salt (NH₂OH, HCl, purity >99%) and buffered at pH=7±0.05 using ammonium hydroxide (NH₄OH, purity >99%). Soil suspensions were obtained after shaking 37.5 mL of extraction solution and 1.5 g of dried soil in an end-over-end shaker for 2 h at 10 rpm. Suspensions were then centrifuged
at 3000 rpm for 10 min and filtered through a 0.45 µm cellulose acetate disk filter to obtain
the extracts (AFNOR, 2002). To ensure complete extraction, the soil residues were subjected
to four successive extractions of 2 h each. Finally, metal concentrations were determined in
the total extract solutions by ICP-AES.

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155 *2.3. Slag analyses*

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2.3.1. Total metal concentrations

Total metal concentrations in slags (bulk) were determined according to the 157 international standard ISO 14869-1 (2001). Briefly, 0.250 g milled slag samples (n=3) were 158 digested in PTFE dishes on a hot plate (150°C) after the addition of 5 mL of hydrofluoric acid 159 (47% HF) and 1.5 mL of perchloric acid (65% HClO₄). The mixture was allowed to evaporate 160 to near complete dryness, and the residue was dissolved in diluted nitric acid (HNO₃) before 161 analysis by ICP-AES. The reliability of the analyses was assessed with certified reference 162 material (calcareous loam soil: CRM n°141R from BCR). The recovery rate averaged 163 $104\pm6\%$ for all the metals analysed. 164

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2.3.2. Mineralogical characterisation

166 Slags were first ground, and the whole-rock powders were used for XRD analysis. 167 XRD patterns were obtained using the same method as for soil mineralogical characterisation 168 and allowed assessment of potential slag decay within soil through mineralogical analysis 169 (comparison of similar mineralogical phases within soil and slag samples).

Then, hand samples of slags were cut to prepare a 30 µm-thick polished thin section.
Optical investigations using scanning electron microscopy (SEM) in back-scattered electron
(BSE) mode were conducted at the Femto-ST Institute (University Bourgogne Franche-

Comté, France). Observations were complemented by energy-dispersive spectrometry (EDS) 173 mapping obtained at the University of Lausanne (Switzerland) using a Tescan Mira LMU 174 SEM equipped with a field emission gun (FEG). The composition of slag-forming minerals 175 and glass were obtained by electron probe microanalysis (EPMA) at the University of 176 Lausanne using a JEOL 8200 Superprobe equipped with five wavelength dispersion 177 spectrometers (WDS). Carbon-coated polished thin sections were analysed with a 15 kV beam 178 and a current of 15 nA. The resolution was ca. 1 µm. Oxides and silicates were used as 179 standards. In addition, X-ray intensity mapping was carried out to reveal the specific textures 180 caused by the chemical features of minerals. 181

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2.4. Ecotoxicity test: snail growth inhibition and bioaccumulation

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2.4.1. Animals

Land snails (*Cantareus aspersus*, Müller 1774, syn. *Helix aspersa*) were reared under controlled conditions according to Gomot-de Vaufleury (2000). For the bioassay, juvenile snails (n=105) were reared for 4 weeks and weighed 1.05±0.15 g prior to exposure.

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2.4.2. Exposure modalities

Following the standard ISO 15952 (2018), the juvenile snails were exposed for 28 189 days to three surface soils (0-20 cm) originating from increasing distance to the centre of the 190 smelting waste repository, *i.e.*, presenting a gradient in metal contamination (SP10, 8 and 4), 191 and to a control treatment (C) in which snails were sustained over dampened absorbent paper. 192 For each treatment, five transparent polystyrene containers of 3200 cm^3 ($24 \times 21 \times 8 \text{ cm}$; ref. 193 194 E1DBBAC001, Charles River IFFA-CREDO, 69 L'Arbresle) were used. Each container was filled with a 1.5 cm layer (200 g dry weight, DW) of soil or an absorbent paper layer, and five 195 snails were introduced. The snails were fed ad libitum with uncontaminated Helixal® snail 196

197 food placed in a Petri dish left on the bottom of the container. The day/night cycle 198 photoperiod was 18/6 h, the temperature was $20\pm1^{\circ}$ C, and the air relative humidity was 199 $90\pm5\%$. Three times a week, the containers were cleaned, the faeces were removed, and the 200 food was renewed.

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2.4.3. Measured endpoints

Each week, individual snail fresh mass (whole-body) and shell diameter were monitored as biological endpoints for the assessment of sublethal effects.

204 At the end of the exposure, one snail was randomly sampled in each replicate container for each exposure modality and subjected to a fasting period of 48 h to empty its gut 205 206 (the faeces were removed after 24 h to prevent their ingestion), then sacrificed by freezing at -207 80°C. After thawing, the whole soft body was removed from the shell and separated into two parts: the viscera (*i.e.*, the visceral complex containing the posterior gut, digestive gland, 208 kidney, mantle, and part of the reproductive tract) and the foot (containing the foot sensu 209 stricto, anterior gut, and remainder of the genital tract). The samples were then freeze-dried 210 and digested in HNO₃ (purity 99.9%) before analyses for metal content by ICP-AES. The 211 certified reference material was TORT-2 (lobster hepatopancreas from NRCC-CNRL, 212 Canada), and recovery rates ranged between 90% and 118% of the certified values for all the 213 metals. 214

215

216 *2.5. Statistics*

All data are presented as the mean \pm standard deviation (sd). First, correlation analyses were performed using the Spearman rank correlation test between the slag abundance observed at each sampling point and physico-chemical parameters, such as the pH, OM content and total and extractable metal concentrations. Significant differences among soil parameters, metal concentrations, snail growth and metal bioaccumulation were then analysed by one-way analysis of variance (ANOVA) followed by Tukey's HSD post hoc test for pairwise comparisons. For the case in which the normality of distribution and homogeneity of variances were not verified (using the Shapiro–Wilk and Bartlett tests, respectively), the Kruskal-Wallis test followed by the Kruskalmc post hoc test (pgirmess package) were used. For all statistical analyses, a *p*-value of 5% was used as a significance threshold.

All statistical analyses were performed with the free statistical software R (ver 3.4.0)(R Core Team, 2020).

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231 **3. Results**

3.1. Soil contamination in the Berthelange district

233 Metal concentrations in soils formed above the slag heaps and in reference soils, *i.e.*, soils that had never been subject to metallurgical activities, are presented in Table 1. In the 234 reference sites, metal concentrations are relatively low in comparison with values measured 235 236 on slag repositories. Indeed, among the metals that have been analysed, some exceed the maximum permissible concentrations in soils. For instance, low anomalous concentrations are 237 observed for arsenic (As) and zinc (Zn) in Petit-Mercey 25, Antorpe 9 or Ferrières 8. The 238 most marked contamination concerns major elements, such as Fe and Mn, with total 239 concentrations in soils of the latter element reaching more than $8500 \ \mu g.g^{-1}$ at the Antorpe 9 240 241 site. Therefore, further results and discussion will focus particularly on this site and this element. 242

243

244 *3.2. Characterisation of slags*

At the macroscopic scale, slags from the Antorpe 9 heap are generally homogeneous in shape, density and porosity. The morphology directly reflects the smelting technology in bloomeries. Slags are generally dark-coloured, ranging from blueish to brownish. Flow texture is frequently observed and is characteristic of slags formed in a molten state. Vesicles, ranging from micrometres to centimetres, are commonly met.

Metal concentrations in slags of Antorpe 9 (Table 1) reach an average of 250 approximately 15% Fe and 1.7%, 1.5% and 0.35% Mn, Ca and Mg, respectively. Other 251 quantified trace metal elements present lower concentrations ranging between 0.0003% (for 252 253 cadmium, Cd) and 0.008% (for Cu). At the mineralogical scale, XRD patterns allowed us to identify fayalite, an olivine group mineral, as the main component of slags (Figure 2A). 254 Petrographic investigations on polished thin sections showed that the phases present in 255 256 Antorpe 9 slags include olivine, glass, spinel, leucite, wustite and metallic Fe (Figure 3). Olivine group minerals commonly display a spinifex texture and elongated skeletal laths; the 257 258 WDS analyses revealed a fayalite (Fe₂SiO₄) composition. The glass phase is generally interstitial and contains subhedral spinel crystals (Figure 3B). Locally, wustite occurs as bleb 259 aggregates and is partially replaced by secondary Fe phases. Similarly, metallic Fe fragments 260 261 are corroded and transformed into oxides and hydroxides (Figure 3D). As shown in Table 2, Mn is heterogeneously distributed between minerals and glass. Fayalite, which may contain 262 up to 5% MnO, is the primary carrier of Mn, while glass and spinel display MnO contents of 263 less than 1.8% (Table 2; Figure 3C). Manganese is almost absent in metallic Fe and secondary 264 Fe phases (Figure 3F). 265

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3.3. Influence of the slag heap on soil characteristics

The edges of the heap were delimited using the relative abundance of slags in the surface layer (0-20 cm) of soils collected at each sampling point (Figure 4). Regardless of the transect direction considered, the slag abundance decreases with the distance from the centre
of the heap, dropping from approximately 70% at SP4 to less than 5% at the heap foot (SP3,
5, 7 and 8).

Significant positive correlations were identified between the slag abundance and some physicochemical parameters of soils (Figure 4), such as the OM content (Rho=0.94, p<0.001) and pH (Rho=0.54, p<0.001). Indeed, from the edge of the slag heap to its centre, the OM content and pH values increase from 8 to 19% and 4.8 to 5.4, respectively.

Variations in the clay composition of soils are also observed by means of XRD 277 analyses of soils (Figure 2B) and clay extract (Figures 2C to G). Mineralogical analyses of 278 clay extract from soil samples allowed us to identify different clay minerals that are present in 279 280 various proportions depending on the samples: illite, chlorite, kaolinite, serpentine (antigorite/chrysotile), and mixed-layer clay minerals. Kaolinite and serpentine exhibit very 281 similar diffraction patterns, but the absence of tetrahedral substitution in antigorite and 282 chrysotile induces rolling up of the layers (Holtzapffel, 1985; Moore and Reynolds, 1997). 283 This characteristic allowed us to distinguish samples without serpentine (Figure 2E) from 284 samples containing serpentine (Figures 2F and G). 285

The total Mn concentrations in the soils range from 1150 μ g.g⁻¹ at the reference site (SP10) to approximately 3500 μ g.g⁻¹ at the heap foot (SP3, 7 and 8) and up to more than 8000 μ g.g⁻¹ at the centre of the heap (SP4) (Figure 4). Manganese extractable concentrations display a similar pattern to total Mn with increasing values from SP10 (880 μ g.g⁻¹) to SP4 (2800 μ g.g⁻¹), representing extraction yields of 77% and 35%, respectively. Surprisingly, at SP9, the total and extractable concentrations were particularly low, *i.e.*, 400 and 170 μ g.g⁻¹, respectively.

3.4. Ecotoxicological bioassay

No snail mortality was recorded over the 28-day exposure periods. Concerning 295 sublethal effects, the growth of snails exposed to the four modalities did not show any 296 significant difference either in terms of body mass (Figure 5A) or shell diameter (data not 297 shown). Indeed, the final mass of snails reached 6.9 ± 1.2 g for the control modality and $7.3 \pm$ 298 1.4 g, 7.2 \pm 1.6 g and 7.2 \pm 1.1 g for snails exposed to soils located at 100 m, 5 m and the 299 centre of the heap, respectively. Similarly, the final shell diameter increased 28.12 ± 3.0 mm 300 for the control and 28.85 ± 2.38 mm, 28.92 ± 2.52 mm, and 29.29 ± 1.97 mm for snails 301 exposed to the three previously cited soils. 302

Growth measurements were completed with Mn bioaccumulation data in snail foot 303 304 and viscera after 28 days of exposure (Figure 5B). As snails grew over the 28-day duration of the exposure, we used body burdens rather than tissue concentrations because the latter can be 305 affected by rapid changes in the mass of the organism (Gimbert et al., 2006; Gimbert and de 306 Vaufleury, 2009). Snails significantly accumulate Mn in their viscera following a dose-307 response pattern according to the Mn concentrations in soils. Viscera internal burdens reached 308 $25.2 \pm 8.3 \ \mu g$ in the control snails (*i.e.*, snail food exposure only) and $93.4 \pm 1.1 \ \mu g$, $93.5 \pm 1.1 \ \mu g$, 93.5309 29.6 μ g and 154.0 \pm 34.4 μ g in snails exposed to soils from SP10, SP8 and SP4, respectively. 310 311 Accumulation in the foot is low, and body burdens do not vary regardless of the exposure modality (Figure 5B). 312

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314 **4. Discussion**

315 *4.1. Source of Mn contamination in soils*

316 Manganese concentrations measured in reference soils from the study area average 317 approximately $800 \ \mu g.g^{-1}$ and are in accordance with the Mn pedogeochemical background of

soils (Pinsino et al., 2012; Grygo-Szymanko et al., 2016). Varying from 480 to 1150 µg.g⁻¹, 318 319 these background concentrations nevertheless show a spatial heterogeneity that has to be related to the lithological nature of the soils (including within the same geological formation) 320 and to the local soil features (both abiotic and biotic). However, this natural variability is not 321 of the same order of magnitude as what is observed in soils formed on ancient slag 322 repositories. Indeed, Mn concentrations range from 1200 to more than 8000 µg.g⁻¹, testifying 323 to the influence of smelting wastes on soil contamination. These values exceed the maximum 324 permissible concentrations in soils; therefore, it is necessary to better understand the origin 325 and fate of this element in past mining ecosystems. 326

In slags, three main mineralogical phases are identified: metallic Fe residues, glass and 327 fayalites belonging to the olivine group. The presence of metallic Fe inclusions in slags attests 328 329 to the generally moderate extraction yields of Fe during the Early Middle Ages. Glass is ubiquitous in slags from Antorpe 9 and is primarily composed of Si, Ca and Al oxides, as 330 reported in the literature (Piatak et al., 2015). The presence of fayalite and its dendritic 331 shape are related to the temperature in the bloomeries (reaching more than 1000°C) 332 and the slow air-cooling conditions of molten slags at the end of the smelting 333 process (Piatak et al., 2004; Tossavainen et al., 2007). The manganese concentration is 334 1.7% in slag bulk samples and is primarily located in fayalites, where it reaches 4.5 335 wt.% MnO. In their review, Piatak et al. (2015) provided average concentrations of Mn 336 in steel and pre-1900 and modern Fe slags between 1 and 4 wt.% MnO. 337

The weathering of the slag primary phases after abiotic (air or water) or biotic 338 339 (microorganisms) interactions leads to the production of secondary products that can be used as indicators of the reactivity and alteration of slags (Piatak et al., 2015). In Antorpe 9 soils, 340 341 three clues of slag alteration have been identified. First, the alkalization of soil (+ 0.6 pH unit between the reference soil and the centre of the heap) may originate from the hydrolysis of 342 CaO and other Ca-Mg oxide and silicate phases of slags and the subsequent release of Ca²⁺ 343 and hydroxyl (OH⁻) ions (Riley and Mayes, 2015; Roadcap et al., 2006). This well-known 344 phenomenon led to the use of slags as a liming agent for soil quality restoration (Lopez et al., 345 1995; Pinto et al., 1995). Second, we identified serpentine minerals ((Mg, Fe, Ni, Mn, Al, Zn) 346 [Si₂O₅] (OH)₄) in soils from Antorpe 9 slag heap. Serpentine is a subgroup of minerals 347 belonging to the kaolinite-serpentine group and usually contains divalent cations in 348 octahedrally coordinated sites. This type of mineral is frequently formed after weathering of 349 350 olivine in rocks. This serpentinization process is also known for magnesiferous olivines in geological environments (basaltic rock alteration; Muntener, 2010), as well as for zinciferous 351 352 silicates in surface conditions (Choulet et al., 2016); to our knowledge, the process is rarely demonstrated for ferriferous and manganiferous olivines. Finally, along with alkalization and 353 serpentinization, Mn can be considered a release product of slag weathering leading to soil 354 contamination. 355

From an environmental point of view, this soil contamination raises the question of mobility and potential transfers to abiotic and biotic compartments of the current ecosystem. Chemical extraction using ammonium acetate and hydroxylamine hydrochloride allowed us to estimate the environmental availability of Mn in soils from Antorpe 9. This extractant can extract the exchangeable metal fraction (including water-soluble Mn held on the surface of negatively charged exchange complexes and Mn coprecipitated with the carbonate fraction) and the easily reducible fraction (including oxides) (McAlister and Smith, 1999). This result

explains the elevated Mn extraction yields in the reference soil (77%, *i.e.*, 880 µg.g⁻¹). In slag-363 364 impacted soils, Mn availability is higher and increases from the edge (approximately 1600 $\mu g.g^{-1}$) to the centre of the heap (2800 $\mu g.g^{-1}$). In contrast, the extraction yields decreased, 365 ranging from 66 to 35%. This result may be related to the fractionation of Mn, which relies on 366 (i) the soil characteristics affected by the slag deposit (*i.e.*, alkaline soil pH, high OM and clay 367 content) (Walna et al., 2010) and (ii) the sequestration of a substantial portion of the total Mn 368 369 concentrations in a soil fraction refractory to the extraction (*i.e.*, the mineral matrix) (Maskall et al., 1995). We have recently confirmed these findings with an in-depth study of the Mn 370 partitioning in soils from Roman and medieval slag heaps (Amnai et al., 2021). We have 371 372 indeed demonstrated that Mn was mainly found in the reducible (oxides) and residual (slag micro-fragments) fractions of soils. The anomalously low total and extractable concentrations 373 at SP9 may hence be explained by its localization in the vicinity of a small pond, which 374 375 modifies the hydromorphic conditions of soil (i.e., decreases the redox potential) and subsequently promotes the leaching of Mn (Walna et al., 2010). 376

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4.2. Manganese bioavailability, transfer and toxicity

The terrestrial snail Cantareus aspersus is well recognized as a biological indicator (de 379 Vaufleury, 2015), integrating three contamination routes: respiratory (from atmospheric 380 381 sources), digestive (from plant and soil sources) and dermal (diffusion of metals through the foot epithelium). In addition to the chemical assessment of metal environmental availability, 382 the analysis of metal bioaccumulation in snail tissues constitutes a way to estimate the 383 fraction of the total concentration in soil that is truly transferable in biota, *i.e.*, the metal 384 bioavailability (ISO 17402, 2008). The snail foot is primarily composed of muscles and does 385 not appear, as for other trace elements, to be an important sink for Mn (de Vaufleury et al., 386 2006). In viscera, the digestive gland (or hepatopancreas) is the main storage organ of Mn, 387

especially in intracellular granules occurring in large numbers in basophil cells (Mason and 388 389 Simkiss, 1982). The Mn burdens accumulated in snail viscera are in accordance with extractable concentrations and confirm that a substantial portion of the Mn released by slags 390 is effectively bioavailable to soil organisms and particularly snails. Despite the high total and 391 available Mn exposure concentrations in soils, we did not observe deleterious effects 392 regarding snail survival or growth, as already noticed with artificially contaminated soils (up 393 to 750 µg Mn.g⁻¹ soil; Bordean et al., 2014). These results are not obvious with respect to 394 ecotoxicity data available in the literature. Indeed, Kuperman et al. (2004) determined 395 laboratory LC₅₀ values (concentrations in the soil causing 50% mortality) of 389 μ g.g⁻¹, 1970 396 $\mu g.g^{-1}$ and 2575 $\mu g.g^{-1}$ for Enchytraeus crypticus, Eisenia fetida and Folsomia candida, 397 respectively. The absence of significant detrimental effects on snails may be related to (i) a 398 misestimation of toxicity metrics estimated using spiked artificial soils (Lock and Janssen, 399 400 2001); (ii) the presence in field soils of growth-promoting factors for snails, such as humic substances and oligo-elements (Gomot et al., 1989); and (iii) the ability of C. aspersus snails 401 402 to accumulate and detoxify Mn in their tissues using detoxification processes, such as metal-403 rich granules (Mason and Simkiss, 1982; Simkiss and Taylor, 1994). However, other sensitive biomarkers at the cellular and subcellular levels (e.g., cytotoxicity; Bradley and Runham, 404 1996) deserve to be investigated. At the ecosystem scale, snails, as possible prey for 405 numerous invertebrates and vertebrates, may contribute to the diffusion of metals in trophic 406

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409 **5.** Conclusion

chains (Moreno-Jiménez et al., 2011).

410 Although slag heaps may be regarded as punctual sources of metal release, their very411 large number, considerable cumulative volume and spatial coverage make them a significant

source of risk (Amnai et al., 2021). The combination of mineralogical, chemical and 412 ecotoxicological approaches allows us to show here that, in addition to the modification of 413 various soil physico-chemical characteristics, such as the pH and OM content, slag 414 weathering over centuries led to the release of metallic contaminants in the surrounding 415 environment. Among them, Mn ranks, based on a combination of its frequency, toxicity, and 416 potential for human exposure, 140th (out of 275) in the substance priority list of the Agency 417 for Toxic Substances and Disease Registry (ATSDR, 2019). We have emphasised not only the 418 environmental availability of this metallic palaeo-pollution but also its bioavailability to soil-419 dwelling invertebrates, such as snails. Hence, such legacy contaminated sites need to be more 420 421 clearly identified and included in environmental risk assessment procedures.

422

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611 **Table captions**

612

Table 1. Metal concentrations (μ g.g⁻¹) in soils from slag heaps and reference sites of the Berthelange district and in slags (bulk) from the Antorpe 9 repository. ^a: data from Crommentuijn et al. (2000). ^b: ecological soil screening level (Eco-SSL) from the US EPA (2007).

617

- **Table 2.** Chemical composition (mean ± standard deviation, SD) of slag-forming minerals
- and glass obtained by EPMA (% in weight).

621 Figure captions

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Fig. 1. A: map of the Berthelange ancient metallurgical district. Green areas are forests, red
dots correspond to archaeologically referenced slag deposits (from Forlin and Laurent, 2014)
and white stars correspond to the investigated slag heaps. B: photograph of the Antorpe 9 slag
heap. C: sampling map of the Antorpe 9 heap.

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Fig. 2. Diffractograms. A: slag; B: bulk soil (SP8, 40-60 cm); C and D: clay fraction in soil
(SP8, 40-60 cm and SP4 80-100 cm, respectively); E: photograph of glass slide with clay
fraction without serpentine (SP10 0-20 cm); F and G: photograph of glass slide with clay
fraction containing serpentine (SP4, 80-100 cm and SP8 40-60 cm, respectively).

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Fig. 3. Left (A to C): representative view of mineral-rich slag material from Antorpe 9. Right
(D to E): detailed view of a spherical metallic Fe particle trapped within slag from Antorpe 9.
A and D: BSE images; B and E: interpretative map of the mineral phases based on WDS and
compositional maps, respectively; C and F: Mn X-ray composition map based on WDS and
EDS, respectively. Brightness correlates with Mn content.

638

Fig. 4. Total (black bars) and extractable (white bars) manganese concentrations ($\mu g.g^{-1}$), the organic matter content expressed in % (dashed line) and the pH value and observed slag content (%) at each sampling point. The NW-SE transect is represented in the left panel (A), and NE-SW is represented in the right panel (B). Different letters correspond to significant differences (*p*-value<0.05). Code for slag abundance: AA (very abundant, *i.e.*, more than 50%), A (abundant, *i.e.*, between 25 and 50%), F (common, *i.e.*, between 20 and 25%), R
(rare, *i.e.*, between 10 and 20%), and T (trace, *i.e.*, less than 10%).

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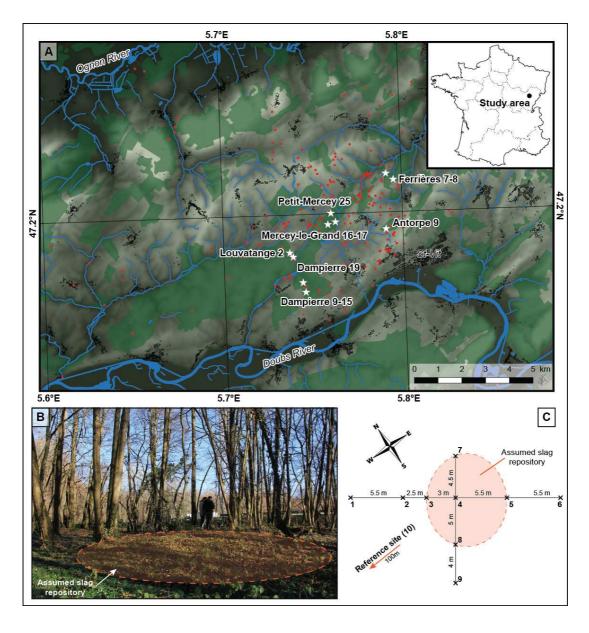
Fig. 5. A: Time course of snail growth (fresh weight) during the 4-week exposure to control conditions (C) and to surface soils from site Antorpe 9. No significant difference (*p*value<0.05) between treatments was identified. B: Mn bioaccumulation in *C. aspersus* foot (white) and viscera (black) (expressed in body burdens, μ g.snail⁻¹) after exposure (for 28 days) to control conditions (C) and to surface soils from site Antorpe 9. t0 corresponds to the metal body burdens in snails at the beginning of the experiment. For each snail part, different letters correspond to significant differences (*p*-value<0.05).

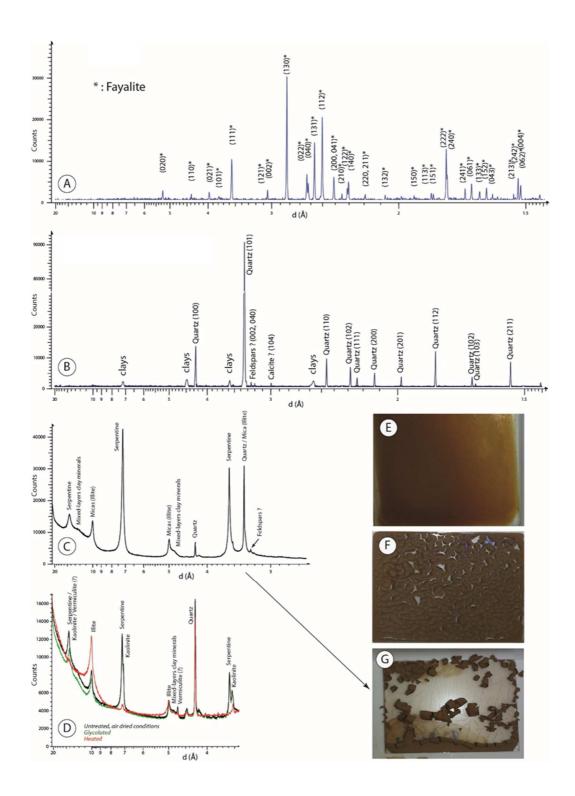
Table 1.

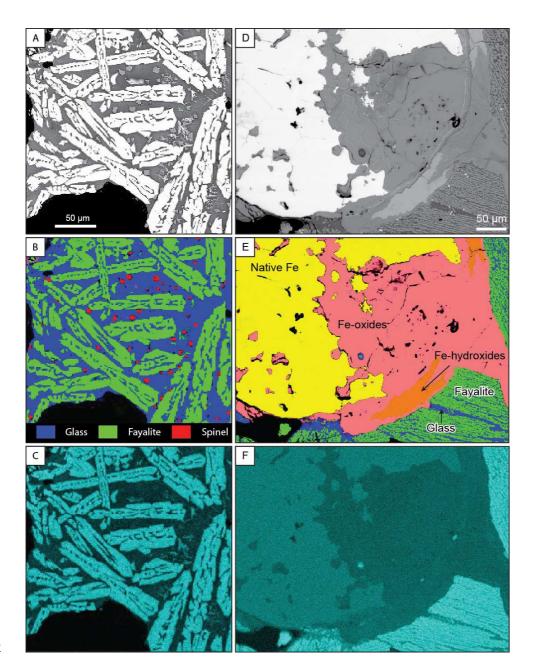
	As	Ca	Cd	Co	Cu	Fe	Mg	Mn	Ni	Pb	Zn
Ref 1	13.3	268.6	<dl< th=""><th>8.3</th><th>11.7</th><th>16,811.6</th><th>1402.7</th><th>483.5</th><th>12.4</th><th>41.4</th><th>33.2</th></dl<>	8.3	11.7	16,811.6	1402.7	483.5	12.4	41.4	33.2
Ref 2	26.2	403.7	<dl< th=""><th>16.8</th><th>20.3</th><th>32,814.5</th><th>1390.4</th><th>789.2</th><th>21.3</th><th>48.9</th><th>62.9</th></dl<>	16.8	20.3	32,814.5	1390.4	789.2	21.3	48.9	62.9
Ref 3	19.2	335.4	<dl< th=""><th>16.4</th><th>15.4</th><th>25,574.8</th><th>1849.5</th><th>1146.5</th><th>18.5</th><th>28.2</th><th>47.6</th></dl<>	16.4	15.4	25,574.8	1849.5	1146.5	18.5	28.2	47.6
Mercey-le-Grand 16	21.9	995.7	<dl< th=""><th>14.4</th><th>19.7</th><th>36,936.9</th><th>1663.0</th><th>1172.4</th><th>20.6</th><th>31.0</th><th>69.1</th></dl<>	14.4	19.7	36,936.9	1663.0	1172.4	20.6	31.0	69.1
Ferrières 7	23.3	775.9	<dl< th=""><th>22.4</th><th>19.0</th><th>35,746.0</th><th>1850.6</th><th>1208.2</th><th>25.9</th><th>26.6</th><th>195.7</th></dl<>	22.4	19.0	35,746.0	1850.6	1208.2	25.9	26.6	195.7
Dampierre 19	23.1	5461.2	0.6	15.0	21.3	27,745.6	1622.5	1422.5	24.4	29.8	61.2
Ferrières 8	35.4	797.4	0.3	22.5	21.0	40,902.3	1704.1	2088.4	32.3	31.6	135.7
Mercey-le-Grand 17	25.0	573.8	<dl< th=""><th>18.3</th><th>25.0</th><th>57,745.1</th><th>1391.5</th><th>2104.3</th><th>23.4</th><th>30.7</th><th>91.6</th></dl<>	18.3	25.0	57,745.1	1391.5	2104.3	23.4	30.7	91.6
Petit-Mercey 25	57.5	3807.3	<dl< th=""><th>24.0</th><th>29.5</th><th>60,975.6</th><th>1600.6</th><th>2236.7</th><th>31.0</th><th>40.3</th><th>104.1</th></dl<>	24.0	29.5	60,975.6	1600.6	2236.7	31.0	40.3	104.1
Louvatange 2	20.0	923.2	0.8	18.1	25.3	38,756.5	1294.2	2602.0	24.6	28.6	78.0
Dampierre 15	9.5	1075.6	<dl< th=""><th>14.3</th><th>16.9</th><th>32,110.0</th><th>1051.2</th><th>3165.8</th><th>16.9</th><th>31.2</th><th>70.2</th></dl<>	14.3	16.9	32,110.0	1051.2	3165.8	16.9	31.2	70.2
Antorpe 22	30.1	3369.2	0.2	20.4	38.8	82,160.3	2027.5	3207.0	36.4	38.3	121.0
Dampierre 9	16.8	2152.3	<dl< th=""><th>16.4</th><th>26.3</th><th>52,950.5</th><th>1383.0</th><th>4709.2</th><th>20.3</th><th>55.2</th><th>87.2</th></dl<>	16.4	26.3	52,950.5	1383.0	4709.2	20.3	55.2	87.2
Antorpe 9	51.8	6618.9	2.2	37.9	65.4	137,095.6	1553.7	8575.6	64.8	41.7	244.8
Maximum Permissible Concentration ^a	34.0		1.6	20.0	40.0			450 ^b	30.0	50.0	50.0
Antorpe 9 Slag #1	<dl< th=""><th>12,665.0</th><th>3.3</th><th>6.9</th><th>86.3</th><th>149,243.0</th><th>3296.6</th><th>17,590.5</th><th>27.3</th><th>11.5</th><th>27.2</th></dl<>	12,665.0	3.3	6.9	86.3	149,243.0	3296.6	17,590.5	27.3	11.5	27.2
Antorpe 9 Slag #2	<dl< th=""><th>14,899.2</th><th>3.3</th><th>10.8</th><th>87.4</th><th>154,248.9</th><th>3759.9</th><th>22,070.9</th><th>31.2</th><th>13.3</th><th>36.0</th></dl<>	14,899.2	3.3	10.8	87.4	154,248.9	3759.9	22,070.9	31.2	13.3	36.0
Antorpe 9 Slag #3	<dl< th=""><th>18,485.5</th><th>2.9</th><th>6.2</th><th>76.4</th><th>137,505.3</th><th>3436.2</th><th>11,135.5</th><th>25.0</th><th>9.0</th><th>28.8</th></dl<>	18,485.5	2.9	6.2	76.4	137,505.3	3436.2	11,135.5	25.0	9.0	28.8

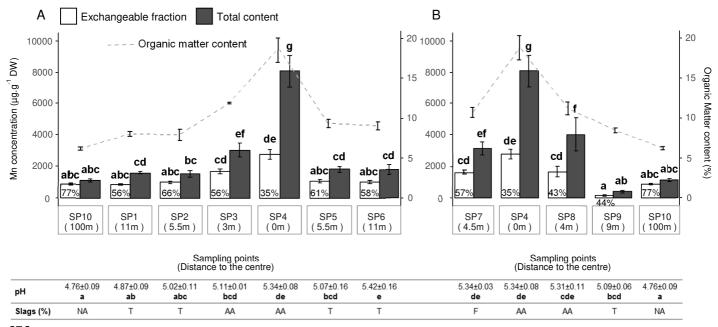
	SiO2	K20	Na2O	P205	MnO	Al2O3	CaO	MgO	TiO2	FeO	Cr2O3	NiO	Total
Glass (n=4)													
mean	40.16	3.12	0.25	1.55	1.69	20.23	9.07	0.02	1.22	22.35	0.01	0.02	99.68
SD	2.68	0.73	0.05	0.20	0.08	2.85	1.46	0.02	0.10	2.30	0.01	0.03	0.21
Fayalite (n=7)													
mean	29.24	0.07	0.01	0.52	4.51	0.59	0.51	1.00	0.13	63.72	0.02	0.01	100.32
SD	0.43	0.11	0.02	0.38	0.88	0.76	0.11	0.60	0.07	1.25	0.02	0.01	0.60
Leucite (n=3)													
mean	55.10	18.65	0.12	0.15	0.14	25.52	0.11	0.02	0.06	2.15	0.01	0.00	102.02
SD	1.85	1.10	0.03	0.18	0.14	0.37	0.12	0.01	0.03	2.31	0.01	0.00	0.50
Spinel (n=4)													
mean	3.01	0.13	0.02	0.02	1.56	49.23	0.08	0.25	2.41	44.60	1.02	0.00	102.32
SD	1.94	0.15	0.03	0.02	0.07	2.57	0.02	0.11	0.85	3.23	0.59	0.01	1.07
Wustite (n=2)													
mean	3.47	0.04	0.02	0.73	0.68	7.65	0.55	0.08	0.59	87.11	0.02	0.01	100.95
SD	2.19	0.01	0.02	0.58	0.06	7.93	0.39	0.02	0.18	5.13	0.01	0.01	1.83
Metallic Fe (n=2)													
mean	0.87	0.03	0.00	1.46	0.13	1.49	0.08	0.03	0.05	125.06	0.08	0.05	129.30
SD	0.39	0.03	0.00	1.91	0.15	2.10	0.10	0.02	0.06	1.89	0.12	0.07	1.54
Fe oxides (n=4)													
mean	2.21	0.01	0.04	0.09	0.05	0.01	0.02	0.02	0.01	87.65	0.01	0.02	90.12
SD	1.11	0.01	0.05	0.18	0.02	0.01	0.01	0.02	0.01	1.66	0.01	0.01	0.79
Fe hydroxides (n=4)													
mean	6.53	0.07	0.09	0.64	0.29	0.55	0.11	0.03	0.00	73.32	0.01	0.03	81.65
SD	4.08	0.04	0.10	0.77	0.23	1.06	0.04	0.03	0.01	2.86	0.01	0.03	1.45

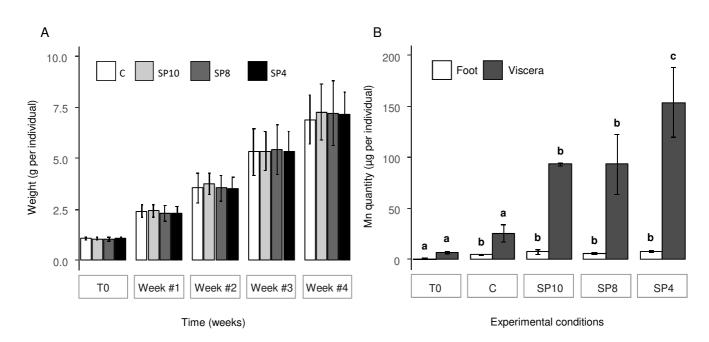
661 Figure 1.











Graphical Abstract

