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Reversible H₂ Oxidation and Evolution by Hydrogenase Embedded in a Redox Polymer Film

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Abstract

Efficient electrocatalytic energy conversion requires the devices to function reversibly, *i.e.* deliver a significant current at minimal overpotential. Redox-active films can effectively embed and stabilise molecular electrocatalysts, but mediated electron transfer through the film typically makes the catalytic response irreversible. Here, we describe a redox-active film for bidirectional (oxidation or reduction) and reversible hydrogen conversion, consisting of [FeFe] hydrogenase embedded in a low-potential, 2,2'-viologen modified hydrogel. When this catalytic film served as the anode material in a H₂/O₂ biofuel cell, an open circuit voltage of 1.16 V was obtained - a benchmark value near the thermodynamic limit. The same film also acted as a highly energy efficient cathode material for H₂ evolution. We explained the catalytic properties using a kinetic model, which shows that reversibility can be achieved despite intermolecular electron transfer being slower than catalysis. This understanding of reversibility simplifies the design principles of highly efficient and stable bioelectrocatalytic films, advancing their implementation in energy conversion.

Introduction

The distinction between bidirectional and reversible molecular catalysts of redox reactions has recently been emphasised.^{1–3} Bidirectional catalysts can catalyse a reaction at a significant rate in either direction. Reversible catalysts are a subgroup of bidirectional catalysts that catalyse the reaction at a significant rate in response to even a small departure from equilibrium, *i.e.*, in the case of a redox reaction, at a small overpotential.^{1–3} Reversible catalysts are required in energy conversion to minimise energy dissipation and thus to increase the energy efficiency of the process. Several reversible synthetic molecular catalysts^{2,4–8} and many redox enzymes^{1,9–14}

catalyse reactions of practical interest, such as the transformations of H_2 and CO_2 , at very high rates and reversibly. Their global implementation in sustainable energy schemes also requires that these catalysts are scalable and robust.¹⁵ Some bioinspired and biological catalysts are made of earth-abundant elements and could thus fulfill the scalability requirement,^{16–18} but their robustness is often insufficient for direct implementation in energy converting devices, especially because of their sensitivity to oxidative stress and molecular oxygen.^{19–23} A recently proposed solution to that problem, based on the incorporation of fragile catalysts into redox-active films serving as a protection matrix against oxidative deactivation,^{24–31} enabled us to use the highly O_2 -sensitive [FeFe] hydrogenase^{32,33} even under operating fuel cell conditions.^{34,35} However, the integration of biocatalysts such as hydrogenases or CO_2 -reducing enzymes into redox films has systematically led to a loss of their intrinsic reversibility:^{34,26–31} catalysis occurs only at a significant overpotential, which results in a dissipation as heat of a fraction of the input energy. Similarly, a large driving force is also typically required to obtain fast mediated electron transfer when unidirectional enzymes are immobilised in redox-active films.^{36,37} To the best of our knowledge, reversible catalysis in a redox-active bioelectrocatalytic film has neither been observed, nor explored theoretically.

Here, we show that reversibility of the catalysis by the [FeFe] hydrogenase from *Desulfovibrio desulfuricans* (DdHydAB) is preserved, over a large pH range, upon integration of the enzyme into a 2,2'-viologen based redox-hydrogel whose potential matches that of the $2H^+/H_2$ couple at pH 7 (Fig. 1). We show that this reversibility is an advantage when the electrode is used either as an anode in a H_2/O_2 fuel cell or as a cathode in a water electrolyser: combining the reversible hydrogenase electrode with an O_2 reducing biocathode gives a fuel cell whose open circuit voltage value is unambiguously attributed to the H_2/O_2 catalysed reactions, almost reaching the thermodynamic limit of 1.23 V. The reversible hydrogenase electrode also evolves H_2 with 100% Faraday efficiency. We propose a kinetic model that explains how both the kinetics and

thermodynamics of intermolecular ET between the mediator and the catalyst determine the properties of the system, including the effective catalytic bias, which is simply tuned by changing the pH.

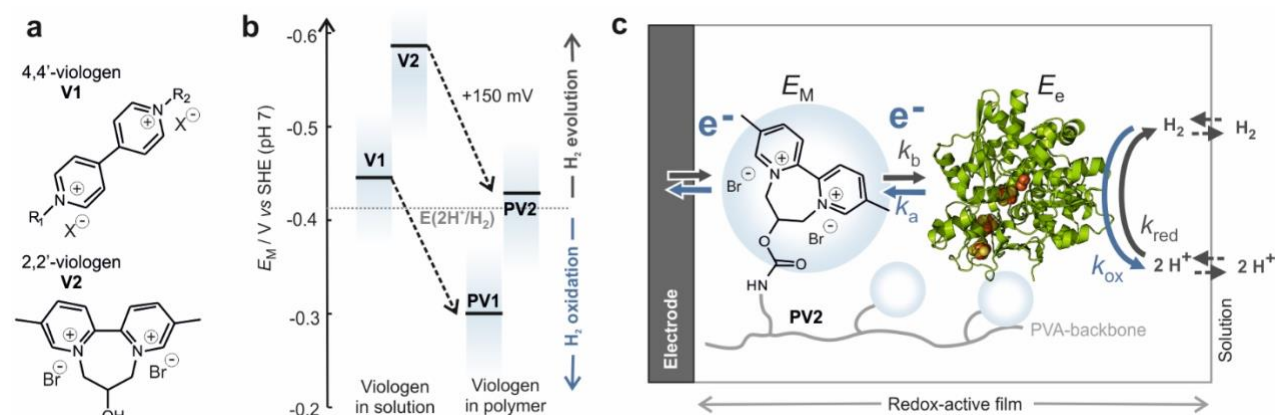


Fig. 1 Redox-active polymers for reversible hydrogenase catalysis. **a.** Structure of 4,4'-viologen (**V1**) and 2,2'-viologen (**V2**) moieties used in redox polymers for immobilisation of hydrogenase. **b.** Redox potential of freely diffusing viologens and of their polymer-bound analogues with respect to the $2H^+/H_2$ Nernst potential ($E(2H^+/H_2)$). The shaded areas illustrate the range of redox potentials observed for these viologens and viologen modified polymer analogues (depending on the nature of substituents, electrolyte composition, and polymer backbone); the black lines show the redox potentials of the 4,4'-methylviologen **V1** with $R_1 = R_2 =$ Methyl (reported in ref [38]), the 2,2'-methylviologen **V2**, the 4,4'-methylviologen-based polymer **PV1** (reported in ref [26] and the 2,2'-methylviologen-based polymer **PV2** (structure in Supplementary Figure 1). The backbone of **PV2** is polyvinyl-alcohol (PVA). **c.** Scheme of the mediated electron transfer (MET) and catalysis in the **PV2** based redox-hydrogel embedding [FeFe] hydrogenase from *Desulfovibrio Desulfuricans* (*DdHydAB*). The kinetic model developed in the Supporting Information relates the catalytic current to the values of the bimolecular electron

transfer rate constants k_a and k_b , the 1st order catalytic rate constants k_{ox} and k_{red} , and the redox potentials of the 2,2'-methylviologen mediator (E_M) and the enzyme (E_e).

Results

Design of the Low Potential 2,2'-Viologen Modified Polymer

Freely diffusing mediators such as 4,4'-viologen species have been used to mediate reversible hydrogenase catalysis,^{27,39–41} but when similar 4,4'-viologen moieties are covalently tethered to polymer or dendrimer backbones, electron transfer (ET) and catalysis with hydrogenases becomes unidirectional.^{25–27,31,34} We hypothesised that this is due to the functionalisation of the 4,4'-viologen moiety and its local environment within the polymeric matrices, making its redox potential more positive by as much as 150 mV (Fig. 1). Here, we used a low potential viologen based on alkylated 2,2'-bipyridine to compensate for the potential shift induced by tethering to the matrix backbone (Fig. 1b). The redox potentials of 2,2'-viologens **V2** (Fig. 1a) are typically more negative than that of 4,4'-viologens⁴² (Fig. 1a and 1b), because the strain imposed by the ring structure destabilises the radical cation and the methyl substituents on the viologen core are electron donating.⁴³ A hydroxyl group on the propylene bridge serves as the anchoring group (Supplementary Method 1 and Supplementary Figure 1 for the synthetic procedure and Supplementary Figures 2 - 5 for full characterisation). Cyclic voltammograms of **V2** show a pair of peaks (Supplementary Fig. 6) attributed to the first reduction of the viologen dication ($V2^{++}$) to the radical cation states ($V2^{+•}$). The redox potential of the $V2^{+•}/V2^{++}$ couple is $-585 \text{ mV} \pm 1 \text{ mV}$ ($n = 3$) vs SHE (172 mV more negative than the $2H^+/H_2$ Nernst potential at pH 7), and is mostly independent of pH and of the presence of H_2 .

The 2,2'-viologen moiety was covalently grafted to a polyvinyl-alcohol backbone via a di-isocyanate linker (Supplementary Method 1 for the synthetic procedure and Supplementary Figures 7 – 17 for full characterisation), which yielded polymer **PV2** with a high load of viologen (w/w 48% \pm 10%, Supplementary Figure 18; Supplementary Figures 19 – 21 for full characterization). A similar 2,2'-viologen moiety bearing an additional thioacetate crosslinking functionality was also synthesised and bound to a poly(amidoamine) dendrimeric scaffold using the same isocyanate coupling strategy to yield dendrimer **DV2** (Supplementary Method 2 and Supplementary Figure 22 for the synthetic procedure and Supplementary Figures 23 – 32 for full characterisation) with a high viologen functionalisation yield (99%, Supplementary Note 1). The polymer-based hydrogels are advantageous from an application point of view because of their scalability and suitability for generation of high catalytic currents. However, viologen modified polymers tend to aggregate, which makes the formation of very thin films inaccessible. In contrast, viologen modified dendrimers with crosslinking functionalities can be used to produce homogeneous thin films,^{44,45} whose thickness and current response are proportional to the amount of catalyst they embed.²⁴

The viologen modified polymer **PV2** was deposited onto the surface of a glassy carbon electrode by drop-casting. Cyclic voltammograms of the modified electrode revealed a redox potential for the $PV2^{2+}/PV2^{•+}$ couple in the polymer film of $E_M(PV2) = -429 \pm 8$ mV vs SHE ($n = 4$), which is 156 mV more positive than the potential of the freely diffusing viologen analogue **V2** (Supplementary Figure 33). Hence, the attachment to the polymer backbone brings the redox potential of the viologen within the film of **PV2** close to the Nernst potential of the $2H^+/H_2$ couple at pH 7. Moreover, it is independent of pH, the presence of H_2 and the presence of hydrogenase within the film. The electrochemical behaviour of a film made of dendrimer **DV2**, modified with the same viologen units, is very similar (Supplementary Figures 34 – 37).

Reversible Catalysis with PV2 / Hydrogenase Electrodes

Electrodes modified with [FeFe] hydrogenase from *Desulfovibrio desulfuricans* (DdHdAB) catalyse both H₂ oxidation and production under conditions of direct electron transfer (DET) (Fig. 2a) or mediated electron transfer (MET) within the film of **PV2** (Fig. 2b,c). The catalytic currents are high (in the mA cm⁻² range for both electrode types and in both directions of the H⁺/H₂ reaction) considering that planar electrodes are used. The hydrogenase undergoing DET shows a change in catalytic bias (the ratio i_{ox}/i_{red}) in favour of H₂ production and a decrease in catalytic potential (the midpoint potential of the wave E_{cat}) as the pH decreases (Fig. 2a), which is in agreement with previous reports on DET of hydrogenases from various organisms.^{46–49} With the hydrogenase embedded within the viologen modified polymer film undergoing MET, the bias i_{ox}/i_{red} also changes in favour of H₂ production as the pH decreases (Fig. 2b,c), but the catalytic waves remain centred on the pH-independent redox potential of the viologen (Fig. 2b). The same behaviour is observed for the hydrogenase embedded in the viologen modified dendrimer film (Supplementary Figure 38). This demonstrates that the redox-active film tunes the apparent catalytic properties of the embedded catalyst.

Control experiments based on buffer exchange and multiple cycle experiments exclude contributions from unbound viologen (Supplementary Figures 39 and 40), and thus confirm that ET is mediated by the viologen moieties that are tethered to the backbone of the immobilisation matrix. Moreover, the H₂ oxidation current is independent of H₂ at high H₂ concentration (Supplementary Figure 41), excluding limitation from H₂ mass transport. This shows that catalysis with the hydrogenase/**PV2** modified electrodes operates under thin film conditions.^{24,50–52} Moreover, the catalytic current for H₂ oxidation or production increases in proportion to the amount of dendrimer/enzyme mixture deposited onto the electrode when the amount is small, and then tends to a plateau (Supplementary Figure 42). This demonstrates that the amount of the

immobilised catalyst defines the catalytic current in the thinnest films (catalysis limited regime) and that charge transfer via electron hopping within the film defines the current for thicker films (electron transfer limited regime). Under catalysis limited conditions, the concentrations of H₂ and reduced/oxidised forms of the mediators are homogenous in the film.^{50,51,53}

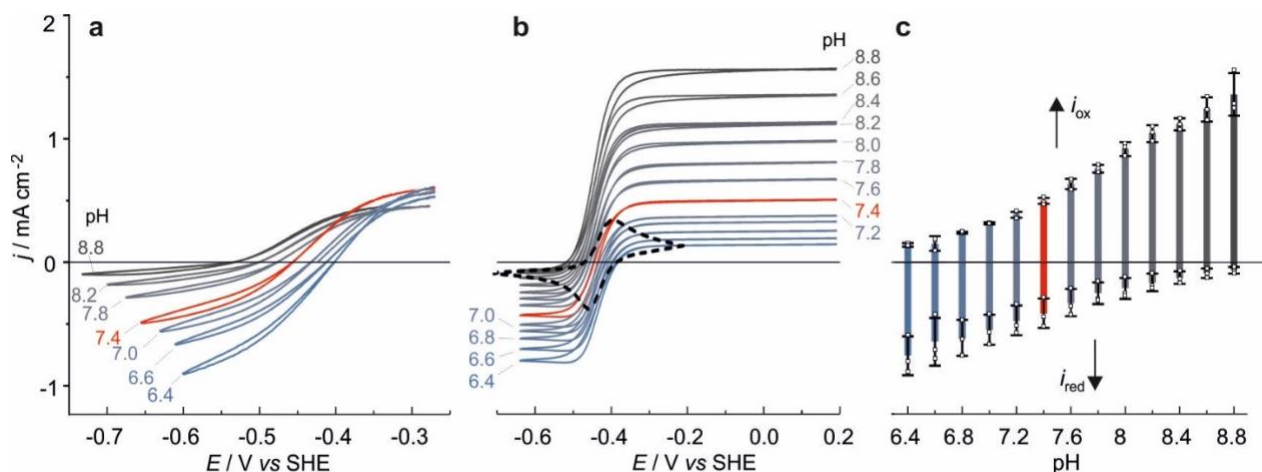


Fig. 2. Direct and mediated electrochemical responses of *Desulfovibrio desulfuricans* (DdHydAB) hydrogenase. **a.** Cyclic voltammograms (CVs) of reversible hydrogen oxidation and production under conditions of direct electron transfer (DET) with a rotating pyrolytic graphite edge disc electrode ($d = 3$ mm, scan rate = 10 mV s^{-1} , $T = 25 \text{ }^{\circ}\text{C}$, $p(\text{H}_2) = 1 \text{ atm}$, pH as indicated). CVs obtained for smaller pH increments are shown in Supplementary Figure 44; **b.** CVs obtained with the enzyme embedded in a redox hydrogel of **PV2** supported on a rotating glassy carbon disc electrode ($d = 3$ mm, scan rate = 10 mV s^{-1} , $T = 25 \text{ }^{\circ}\text{C}$, $P(\text{H}_2) = 1 \text{ atm}$, pH as indicated). The non-catalytic CV of the polymer film without enzyme is shown as a dashed line. The measurement at pH 7.4 used for comparison to simulation in Fig. 3 is shown in red; **c.** bar diagram of the limiting, mediated H₂ oxidation and production current (i_{ox} and i_{red} , as defined in Fig. 3b) as a function of pH. Data are presented as mean values \pm SD of three independent experiments ($n = 3$). The individual data points are shown as open squares. The catalytic currents are stable for at least 100 cycles over the course of 3 hours (Supplementary Figure 39a, b) over the entire pH range.

Kinetic Model of Mediated, Bidirectional Electrocatalysis

The comparison of Fig. 2a and 2b shows that interfacing the hydrogenase either directly (Fig. 2a) or via the viologen mediator embedded into the films (Fig. 2b) significantly changes the current response and, in particular, the catalytic bias (compare the triangles and squares in Fig. 3c). The DET responses can be modelled by a mechanism consisting of two electron transfer steps and one chemical reaction step (EEC-mechanism) taking into account slow interfacial ET.^{46,54–56} The MET response is much simpler, as it can be modelled by a 1-electron sigmoid (red line in Figure 3b), whose midpoint potential (the catalytic potential) is independent of pH. To explain this behaviour, we propose a simple theoretical model of the bidirectional current response of a 1-electron redox enzyme embedded in a thin film of polymer with a redox moiety that acts as a mediator, assuming the rates of catalysis and the concentration of reduced and oxidised viologen are homogeneous in the film. We show in Supplementary Note 2 that the current response reads:

$$i = \frac{i_{ox} \exp F(E - E_{cat})/RT - i_{red}}{1 + \exp F(E - E_{cat})/RT} \quad (1)$$

in which i_{ox} and i_{red} are the limiting oxidative and reductive currents, respectively, as defined in Fig. 3b, and the midpoint potential of the wave, E_{cat} , is given by

$$E_{cat} = E_M + \frac{RT}{F} \ln \frac{\alpha/\delta + 1}{\alpha\delta + 1} \quad (2)$$

in which: $\alpha = A^\Sigma \sqrt{k_a k_b} / (k_{ox} + k_{red})$ and $\delta = \sqrt{k_a/k_b} = \exp [F(E_M - E_e)/2RT]$. The rate constants are depicted in Fig. 1: k_a and k_b are the bimolecular rate constants for the oxidation and reduction of the enzyme by the mediator, k_{ox} and k_{red} are the 1st order rates of H₂ production and evolution by the enzyme, A^Σ is the total concentration of viologen in the film, E_M is the potential of the mediator, and E_e is the potential of the enzyme. Equation 2 shows that E_{cat} changes from the

potential of the mediator to the potential of the enzyme as the ratio of the rate of intermolecular ET over the rate of catalysis increases (Fig. 3a). Equation 1 perfectly reproduces the experimental catalytic currents of Fig. 2b (as exemplified in Fig. 3b), and we determined a value for E_{cat} of -435 ± 3 mV vs SHE at pH 7, identical to the potential of the mediator ($E_{\text{M}}(\text{PV2}) = -429 \pm 8$ mV vs SHE). We found that E_{cat} equals $E_{\text{M}}(\text{PV2})$ at all pH values below 8 and deviates only slightly under more alkaline conditions (Fig. 3c). Similarly, the catalytic potential is also equal to the potential of the mediator in the case of the hydrogenase embedded in the **DV2** films (Supplementary Figure 43a). Using Equation 2 we conclude that intermolecular ET is slower than catalysis ($\alpha \ll 1$, which corresponds to the shaded area in Fig. 3a).

Using the constraint that the current is zero at the equilibrium potential (i.e. when $E = E_{\text{eq}}$), equation 1 imposes a strict relationship between the catalytic bias (the ratio $i_{\text{ox}}/i_{\text{red}}$), the catalytic potential (E_{cat}) and the Nernst potential of the $2\text{H}^+/\text{H}_2$ couple (E_{eq}):

$$\frac{i_{\text{ox}}}{i_{\text{red}}} = \exp \frac{F(E_{\text{cat}} - E_{\text{eq}})}{RT} \quad (3)$$

From the above observation that $E_{\text{cat}} = E_{\text{M}}$, we conclude that the catalytic bias should be independent of the potential of the enzyme:

$$\frac{i_{\text{ox}}}{i_{\text{red}}} = \exp \frac{F(E_{\text{M}} - E_{\text{eq}})}{RT} \quad (4)$$

Equation 4 predicts that the catalytic bias changes with pH in our experimental case, since $E_{\text{M}}(\text{PV2})$ is pH-independent and E_{eq} shifts -60 mV per pH unit. The prediction is shown as a red line in Fig. 3d, together with the experimental values as squares. The very good agreement between the two shows that intermolecular ET is slower than catalysis over the whole pH range, yet this does not impair reversibility between pH 6.6 and 8.6 (Fig. 2b).

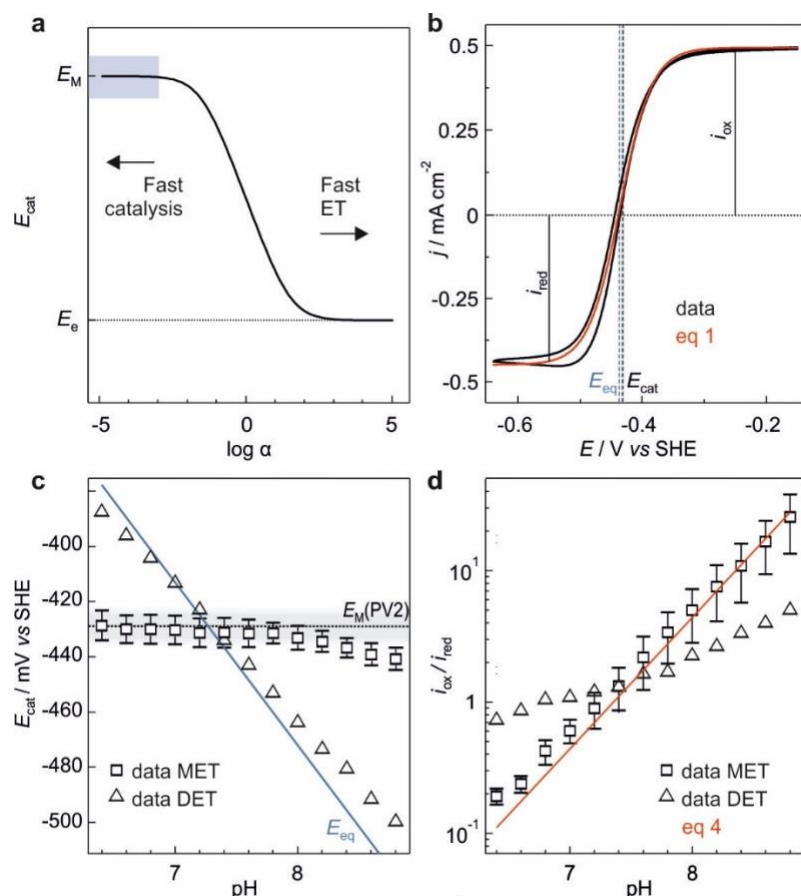


Fig. 3. Modeling of the mediated catalytic CVs. a. Effect of changing the rate of intermolecular ET between the mediator and the catalyst on the value of the catalytic potential, as predicted by Equation 2. The parameter $\alpha = A^{\Sigma} \sqrt{k_a k_b} / (k_{\text{ox}} + k_{\text{red}})$ measures the competition between intermolecular ET and catalysis. The catalytic potential E_{cat} continuously changes from the mediator potential E_M to the enzyme potential E_e as α increases. **b.** Equation 1 (red) is fitted to the experimental MET CV (black, data from Fig. 2b at pH 7.4) with i_{ox} and i_{red} being the plateau catalytic currents of hydrogen oxidation and hydrogen evolution, respectively. Fit parameters: $i_{\text{ox}} = 0.494 \pm 0.001 \text{ mA cm}^{-2}$, $i_{\text{red}} = 0.442 \pm 0.002 \text{ mA cm}^{-2}$, $E_{\text{cat}} = -0.435 \pm 0.001 \text{ V}$. **c.** Experimental catalytic potential (the half wave potential E_{cat}) as a function of pH. The dashed line is the potential of $E_M(\text{PV2})$ (mean value from four individual polymer batches ($n = 4$) shown in Supplementary Figure 33) with its standard deviation indicated by the shaded area. The solid blue line is the

calculated E_{eq} . **d.** Experimental catalytic bias (the ratio of $i_{\text{ox}}/i_{\text{red}}$ defined in panel b) as a function of pH. The prediction of Equation 4 (red line) is calculated from the pH-dependent $2\text{H}^+/\text{H}_2$ equilibrium potential (E_{eq}). The values of the catalytic potential and of the catalytic bias are obtained from the mediated electron transfer (MET) CVs from Fig. 2b and direct electron transfer (DET) CVs (triangles) from Fig. 2a and Supplementary Figure 44. For the MET-process E_{cat} and $i_{\text{ox}}/i_{\text{red}}$ are presented as mean values \pm SD derived from three independent experiments ($n = 3$). The corresponding data for **DV2** are shown in Supplementary Figure 43.

Use of Reversible Electrodes in Fuel Cells and Electrolysers

The use of an electrode that reversibly catalyses H_2 oxidation and H_2 production should maximise the energy conversion efficiency in H_2/O_2 fuel cells (high voltage) and in electrolyzers for H_2 production. The hydrogenase/**PV2** bioanode showed significant stability for more than 15 hours in the H_2 oxidising and H_2 producing modes (Supplementary Figure 45), which made it possible to test its fuel cell and electrolyser performance (Fig. 4). The fuel cells were built from a hydrogenase/**PV2** film either on a flat glassy carbon electrode (Fig. 4a) or on a large surface area gas breathing electrode (Fig. 4b). The H_2 oxidising bioanodes were combined with carbon felt electrodes modified with the enzyme bilirubin oxidase from *Myrothecium verrucaria* (electrode characterisation is detailed in Supplementary Figure 46).⁵⁷ This biocathode allows O_2 reduction to occur at small overpotential.^{57–59} The previously reported mechanism by which the redox polymer protects the enzyme from O_2 , relies on the reduction of O_2 in the outer layers of the thin redox-active films before it reaches the hydrogenase in the inner layers.²⁴ This mechanism remained operational in the films characterised here, as demonstrated by the **PV2**-catalysed O_2 reduction (Supplementary Figure 47) and the stable catalytic current for H_2 oxidation under a mixed gas-feed of H_2 and O_2 (Supplementary Figure 48). Moreover, although the anode based on the flat glassy carbon electrode experiences very oxidative potentials (of up to 0.79 V vs SHE)

due to the anode limiting conditions imposed by the use of an oversized cathode (Supplementary Figure 49), the embedded hydrogenase remains functional, demonstrating efficient protection of the enzyme from chloride-dependent high potential deactivation⁶⁰ by **PV2** (Supplementary Figure 50). This dual protection from aerobic and anaerobic oxidative stress made it possible to implement the reversible hydrogenase-**PV2** anode in either a one or a two-compartment H_2/O_2 fuel cell setup.

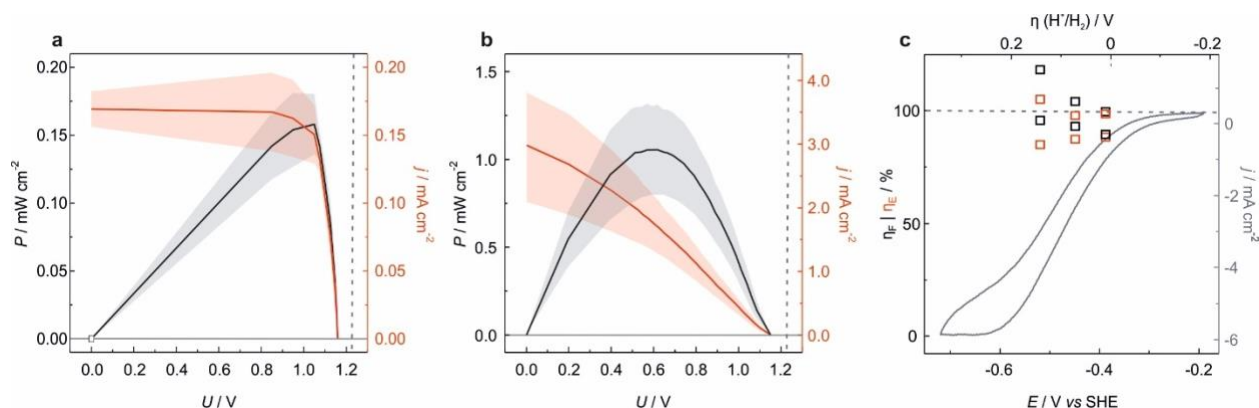


Figure 4. Reversible hydrogenase electrode for H_2/O_2 biofuel cell and electrolyser. **a.** Power curve and current / voltage (I/U)-curve of two-compartment biofuel cells operating with an anode consisting of **PV2**/*Dd*HydAB on a flat glassy carbon electrode (diameter = 3 mm). The setup is schematically shown in Supplementary Figure 51b. **b.** Power curve and I/U -curve of two-compartment biofuel cells with an anode consisting of **PV2**/*Dd*HydAB on a high surface area gas breathing electrode. The setup is schematised in Supplementary Figure 52. For both fuel cell setups, the cathode consisted of bilirubin oxidase immobilised on a high surface Toray paper electrode. The electrolyte of both compartments was potassium phosphate buffer (50 mM, pH 7.4). Averages and standard deviations (shaded areas) were measured from three repetitions of the biofuel cell experiments shown in Supplementary Figure 51c,d (flat electrodes) and Supplementary Figure 53b (gas breathing electrodes). The thermodynamic limit of a H_2/O_2 -fuel cell (1.23 V) is denoted as grey, dashed line. **c.** Faraday efficiency (η_F , black squares, defined as

the ratio of charge used for H₂ production over charge passing the electrode), energy efficiency of the cathodic half-cell (η_E , red squares, calculated by considering the cathodic overpotential and assuming a reversible oxygen evolving half cell^{61,62}) and cyclic voltammogram (grey solid line) of the electrolyser half-cell based on a **PV2/DdHydAB** film immobilised on a MTI carbon foam electrode operating in Tris-citrate buffer (200 mM; pH 6.4) with KCl (100 mM). Overpotentials (η) are with reference to the 2H⁺/H₂ redox couple. The setup is schematically shown in Supplementary Figure 54a. Hydrogen was quantified by gas chromatography (Supplementary Figure 54d). Data were collected from two individual electrodes per overpotential value (six electrodes in total).

The power curves were almost identical for the one-compartment and for the two-compartment setup based on the flat glassy carbon electrode (Fig. 4a and Supplementary Figure 51) and exhibited a maximum power output of $147 \pm 14 \mu\text{W cm}^{-2}$, a maximum current output of $163 \pm 26 \mu\text{A cm}^{-2}$ and an open circuit voltage (OCV) of $1.163 \pm 0.005 \text{ V}$ ($n = 3$). Power density and current density were increased by one order of magnitude to values of $1.06 \pm 0.26 \text{ mW cm}^{-2}$ and $3.0 \pm 0.9 \text{ mA cm}^{-2}$, while a high OCV value ($1.149 \pm 0.012 \text{ V}$, $n = 3$) was conserved (Fig. 4b) by using high surface area carbon electrodes modified with **PV2/DdHydAB** operated in a gas breathing mode in a two-compartment setup (Supplementary Figure 52). The use of the large surface area electrodes for both anode and cathode leads to high catalytic currents for both half-cells (Supplementary Figure 53a), which explains the difference in shape of the power curve (Fig. 4b) in comparison to the one of the fuel cell based on the flat glassy carbon electrodes (Fig. 4a) operated under anode limiting conditions (Supplementary Figure 49).

The high OCV values obtained for the fuel cells based on **PV2/DdHydAB** anodes match those previously reported for biofuel cells using hydrogenase under conditions of DET,^{63–67} with the added advantage of efficient protection from O₂ (Supplementary Figure 48) and high potential

(Fig. 2b and Supplementary Figure 50) for the hydrogenase anode incorporated in **PV2**. Earlier examples of H_2/O_2 biofuel cells based on bioanodes with the **PV1** type of polymer designed for hydrogenase protection had an OCV lower or equal to 1.08 V.^{26,34} A value of 1.13 V has also been reported,⁶⁸ but it included a pseudo-capacitance contribution which increases the OCV value but does not contribute significantly to power generation.⁶⁹

In addition to its use as a bioanode in the H_2/O_2 biofuel cell, we could use the reversible **PV2/DdHydAB** electrode as a biocathode in an electrolyser for H_2 production (Fig. 4c and Supplementary Figure 54). The catalytic currents were high ($6.2 \pm 2.1 \text{ mA cm}^{-2}$ at -0.62 V vs SHE, $n = 4$) and stable, so that the hydrogen produced at the cathode could be detected and quantified by gas chromatography even at overpotentials as low as 12 mV (Supplementary Figure 54d). High selectivity with Faraday efficiencies close to 100% and low overpotential requirements led to energy conversion efficiencies of the H_2 producing half cell (calculated based on a reversible O_2 producing half-cell^{61,62}) in excess of 95% even for current densities as high as 4 mA cm^{-2} (Fig. 4c).

Discussion

In this paper, we have demonstrated the possibility of reversible catalysis under conditions of mediated ET in a thin film. Reversibility is evidenced by the catalytic response cutting sharply across the potential axis at the equilibrium potential, allowing the fuel cell voltage to be at least 82 mV higher than in previous reports. The observed open circuit voltage value (1.16 V) was unambiguously attributed to the H_2/O_2 catalysed reactions, and close to the thermodynamic limit (1.23 V), with the residual overpotential resulting from the cathodic reactions.⁷⁰ Decreasing the overpotential for catalysis conserves energy that would otherwise be released as heat: a potential source of damage if the catalytic film is confined in a fuel cell stack. The same conclusions hold for the reverse reaction for use in electrolyzers: the reversible hydrogenase electrode generates

high current densities for H₂ production at minimal overpotential, thus ensuring maximised efficiency for the conversion of electricity into chemical energy.

In order to identify the requirements for reversibility, we have described a model of bidirectional mediated electrocatalysis, focusing on the case of a thin film. Our assumptions are different from those of the only existing previous model of mediated bidirectional catalysis.^{41,71} First, Kano, Ikeda and coworkers considered planar semi-infinite diffusion (which is not relevant to the case discussed here, a film of finite thickness). Second, they implicitly assume that the catalytic transformation is infinitely fast compared to intermolecular ET, whereas we explicitly consider the competition between the two half-reactions: reaction of the enzyme to produce and consume H₂, and intermolecular ET between the enzyme and the viologen. We reached the conclusion that reversible catalysis can operate under two distinct regimes (Fig. 3a): The fast catalysis regime in which the redox potential of the electron mediator defines both the position of the catalytic wave ($E_{\text{cat}} = E_{\text{M}}$) and the catalytic bias as a function of pH (eq. 4) and, the fast electron transfer regime in which the redox potential of the enzyme's active site defines the position of the wave ($E_{\text{cat}} = E_{\text{e}}$). These conclusions on the theory of reversibility in MET are the same as in the case of DET with an unidirectional enzyme that embeds a one electron redox relay between the surface and the active site.⁷²

The fact that the potential of the mediator is pH-independent while the potential of the enzyme is pH-dependent gives a decisive criterion for experimentally determining the relative kinetics of ET. The position of the catalytic wave in the MET case is mostly independent of pH and centered on the mediator potential demonstrating operation in the fast catalysis regime (Fig. 3c). This is further corroborated by the catalytic bias that changes as a function of pH and the properties of the mediator, but is independent of the properties of the enzyme (Fig. 3d). An important conclusion

here is that fast intermolecular ET between the mediator and the enzyme is not a requirement for observing a reversible response.

Under these conditions of fast catalysis, the requirement to experimentally achieve catalytic reversibility and high catalytic currents is a close match between the redox potentials of the mediator and the enzyme, because the catalytic bias depends exponentially on the difference between the two (Equation 4 and Fig. 3d). This is highlighted by the *DdHydAB* hydrogenase integrated in the redox-active polymer **PV2** film with matching potential at neutral pH conditions, which shows a transition from unidirectionality at pH 6.4 to reversibility around neutral pH and again to unidirectionality at pH 8.8 (Fig. 2c). Reversibility in a broader pH range can occur in the fast ET regime as observed for the hydrogenase in the DET configuration (Fig 2a).⁴⁶ The film properties that define reorganisation energy and interactions between the redox partners (Marcus model) could serve as a primary optimisation target to enhance ET, and thus transition the MET systems into the fast ET regime.

In conclusion, reversibility in mediated redox-active films can theoretically take place either in the fast electron transfer regime or in the fast catalysis regime, and we show that the latter is accessible experimentally. From a practical perspective, reversibility in electrocatalytic films is important for energy conversion applications because it combines the advantage of high current densities at low overpotential with the possibility for robust catalysis owing to the intrinsic protecting feature of the redox matrix.²⁴ Overall, this understanding of the different regimes for reversibility and the demonstration of its practical usefulness for conversion of small molecules may guide the future design of electrocatalytic films that have, until now, shown only unidirectionality despite the reversible nature of the catalysts.

Methods

Materials and instrumentation. The electrochemical measurements were carried out with a three-electrode setup consisting of a GCE-working electrode (for MET) or a PGE-working electrode (for DET), a Pt-counter electrode and an Ag/AgCl 3M KCl (for MET) or a saturated calomel (for DET) reference electrode. All MET CVs of *DdHydAB* embedded in **PV2** were recorded in Tris/citrate buffer (50 mM) with KI (100 mM) as a supporting electrolyte. The CVs of *DdHydAB* embedded in **DV2** were recorded in different electrolytes optimised for the requirements of the individual measurements as indicated. All DET CVs were recorded in a buffer mix containing MES, HEPES, TAPS, CHES, sodium acetate (15 mM each, pH as indicated) and NaCl (100 mM) as a supporting electrolyte.

Preparation of *DdHydAB*. *DdHydAB* was produced heterologously in *E. coli*, purified by affinity chromatography, artificially matured⁷³ and converted to an air-stable form as described previously.⁷⁴ The air-stable enzyme reactivates spontaneously in viologen modified films in presence of H₂.³⁵

Preparation of **PV2/*DdHydAB*-modified anodes.** The *DdHydAB* [FeFe] hydrogenase was immobilised via physical entrapment in the polymer network. For the preparation of the non-gas breathing anodes, an aqueous solution of **PV2** (10 μ L, 3 mg mL⁻¹) was mixed with a solution of hydrogenase (200 μ M in 50 mM Tris/HCl buffer, pH 7.2). The mixture was immobilised on a glassy carbon electrode ($A_g = 0.07$ cm²) by drop-casting and dried for at least 2 h at RT under glove box conditions with a moist atmosphere. To achieve sufficient stability, the anode was pretreated with KI before the biofuel cell experiments (Supplementary Figure 55). For the gas breathing anode a round piece of Carbon Toray Paper (Quintech, Freudenberg Gasdiffusionsanlage, H23-C9, 0.25 mm thick, $\delta = 135$ g cm⁻², $d = 2.2$ cm) was washed with ethanol (p.a.) followed by distilled water. The wet electrode was functionalised with a thin layer of **PV2** (35 μ g). The layer was allowed to dry at 8 °C overnight. On top of the layer a mixture of **PV2** (63 μ L, 1.5 mg mL⁻¹) and *DdHydAB* (6

μL , 200 μM in 50 mM Tris/HCl buffer, pH 8) was immobilised by drop-casting and was left to dry at 8 °C overnight under ambient air leading to catalytically active films with a geometric area of 0.196 to 0.259 cm^2 .

Preparation of PV2/DdHydAB-modified electrolyser cathode. A porous carbon foam electrode (MTI, EQ-bcgdl-1400S-LD, porous C, 6 mm x 13 cm x 0.454 mm, air permeability $\approx 10 \text{ mL cm}^{-2} \text{ s}^{-1}$, porosity $\approx 31 \mu\text{m}$ coated on one side with a Nafion/Teflon-based microporous film of 50 μm , carbon content 5 mg cm^{-2}) was washed with ethanol (p.a.) followed by distilled water. The wet material was functionalised with a thin layer of **PV2** (35 μg) and was allowed to dry at 8 °C overnight. A mixture of **PV2** (63 μL , 1.5 mg mL^{-1}) and DdHydAB (6 μL , 200 μM in 50 mM Tris/HCl buffer, pH 8) was immobilised by drop-casting on top of the underlying polymer layer. The films were left to dry at 8 °C overnight leading to a functionalised geometric surface area of 0.188 cm^2 .

Preparation of the bilirubin oxidase modified cathode. The cathode preparation was adapted from ref [57]. For the efficient orientation of bilirubin oxidase from *Myrothecium verrucaria* (MvBOx), a sheet of Toray carbon paper (6 cm^2) was electrochemically functionalised with negatively charged naphthoate moieties. The carbon paper was immersed in a solution of 5-carboxynaphthalene-2-diazonium tetrafluoroborate (0.87 mM) in MeCN containing tetrabutylammonium tetrafluoroborate (100 mM) as a supporting electrolyte (for the synthesis of the diazonium salt see ref [57]). The naphthoate was covalently attached by applying 10 reductive potential pulses (0.5 s, -0.1 V vs Ag/AgCl 3M KCl). The residual diazonium salt was washed off with acetonitrile (10 x 1 ml), followed by ethanol (10 x 1 ml). The organic solvents were removed by drying the carbon paper for at least two hours under air. The carbon paper was then wetted with potassium phosphate buffer (PB, 5 mL, 50 mM, pH 7.2). A solution of MvBOx (120 μL ; 2 mg mL^{-1}) in potassium phosphate buffer (50 mM, pH 7.2) was drop-cast on the wet sheet. The enzyme

was allowed to immobilise on the naphthoate moieties for 48 h at 4 °C. Unbound enzyme was removed by rinsing the electrode with potassium phosphate buffer (10 x 1 mL, 50 mM, pH 7.2).

Data availability statement

All source data that support the findings of this study (Data of the Figures 2 - 4, Supplementary Figures 2 - 16, 18 - 21, 23 - 51 and 53 - 55) are available in Zenodo with the identifier 10.5281/zenodo.4421107. Figure 1, Supplementary Figures 1, 22 and 52 are general schemes without underlying data. The underlying data of the crystal structure (Supplementary Figure 5) is deposited in the Cambridge structural database with the identifier CCDC 2046659.

References

1. Armstrong, F. A. & Hirst, J. Reversibility and efficiency in electrocatalytic energy conversion and lessons from enzymes. *Proc. Natl. Acad. Sci. U. S. A.* **108**, 14049–14054 (2011).
2. Dutta, A., Appel, A. M. & Shaw, W. J. Designing electrochemically reversible H₂ oxidation and production catalysts. *Nat. Rev. Chem.* **2**, 244–252 (2018).
3. Fourmond, V., Wiedner, E. S., Shaw, W. J. & Léger, C. Understanding and design of bidirectional and reversible catalysts of multielectron, multistep reactions. *J. Am. Chem. Soc.* **141**, 11269–11285 (2019).
4. Priyadarshani, N. *et al.* Achieving reversible H₂/H⁺ interconversion at room temperature with enzyme-inspired molecular complexes: a mechanistic study. *ACS Catal.* **6**, 6037–6049 (2016).
5. Dutta, A., DuBois, D. L., Roberts, J. A. S. & Shaw, W. J. Amino acid modified Ni catalyst exhibits reversible H₂ oxidation/production over a broad pH range at elevated temperatures. *Proc. Natl. Acad. Sci. U. S. A.* **111**, 16286–16291 (2014).
6. Cunningham, D. W., Barlow, J. M., Velasquez, R. S. & Yang, J. Reversible and selective

- CO₂ to HCO₂⁻ electrocatalysis near the thermodynamic potential. *Angew. Chem. Int. Ed.* **59**, 4443–4447 (2019).
7. Cunningham, D. W. & Yang, J. Y. Kinetic and mechanistic analysis of a synthetic reversible CO₂/HCO₂⁻ electrocatalyst. *Chem. Commun.* **56**, 12965–12968 (2020).
 8. Le Goff, A. *et al.* From hydrogenases to noble metal-free catalytic nanomaterials for H₂ production and uptake. *Science* **326**, 1384–1387 (2009).
 9. Del Barrio, M. *et al.* Electrochemical investigations of hydrogenases and other enzymes that produce and use solar fuels. *Acc. Chem. Res.* **51**, 769–777 (2018).
 10. Siritanaratkul, B. *et al.* Transfer of photosynthetic NADP⁺/NADPH recycling activity to a porous metal oxide for highly specific, electrochemically-driven organic synthesis. *Chem. Sci.* **8**, 4579–4586 (2017).
 11. Léger, C. *et al.* Enzyme electrokinetics: energetics of succinate oxidation by fumarate reductase and succinate dehydrogenase. *Biochemistry* **40**, 11234–11245 (2001).
 12. Wang, V. C. C., Can, M., Pierce, E., Ragsdale, S. W. & Armstrong, F. A. A unified electrocatalytic description of the action of inhibitors of nickel carbon monoxide dehydrogenase. *J. Am. Chem. Soc.* **135**, 2198–2206 (2013).
 13. Bassegoda, A., Madden, C., Wakerley, D. W., Reisner, E. & Hirst, J. Reversible interconversion of CO₂ and formate by a molybdenum-containing formate dehydrogenase. *J. Am. Chem. Soc.* **136**, 15473–15476 (2014).
 14. Zu, Y., Shannon, R. J. & Hirst, J. Reversible, electrochemical interconversion of NADH and NAD by the catalytic (I_λ) subcomplex of mitochondrial NADH:ubiquinone Oxidoreductase (Complex I). *J. Am. Chem. Soc.* **125**, 6020–6021 (2003).
 15. McKone, J. R., Lewis, N. S. & Gray, H. B. Will solar-driven water-splitting devices see the light of day? *Chem. Mater.* **26**, 407–414 (2014).
 16. Helm, M. L., Stewart, M. P., Bullock, R. M., DuBois, M. R. & DuBois, D. L. A synthetic nickel

- electrocatalyst with a turnover frequency above 100,000 s⁻¹ for H₂ production. *Science* **333**, 863–866 (2011).
17. Hoffert, W. A., Roberts, J. A. S., Morris Bullock, R. & Helm, M. L. Production of H₂ at fast rates using a nickel electrocatalyst in water-acetonitrile solutions. *Chem. Commun.* **49**, 7767–7769 (2013).
 18. Dutta, A., Roberts, J. A. S. & Shaw, W. J. Arginine-containing ligands enhance H₂ oxidation catalyst performance. *Angew. Chem. Int. Ed.* **53**, 6487–6491 (2014).
 19. Cracknell, J. A., Vincent, K. A. & Armstrong, F. A. Enzymes as working or inspirational electrocatalysts for fuel cells and electrolysis. *Chem. Rev.* **108**, 2439–2461 (2008).
 20. Wakerley, D. W. & Reisner, E. Oxygen-tolerant proton reduction catalysis: much O₂ about nothing? *Energy Environ. Sci.* **8**, 2283–2295 (2015).
 21. Cooney, M. J., Svoboda, V., Lau, C., Martin, G. & Minteer, S. D. Enzyme catalysed biofuel cells. *Energy Environ. Sci.* **1**, 320–337 (2008).
 22. Barton, S. C., Gallaway, J. & Atanassov, P. Enzymatic biofuel cells for implantable and microscale devices. *Chem. Rev.* **104**, 4867–4886 (2004).
 23. Leech, D., Kavanagh, P. & Schuhmann, W. Enzymatic fuel cells: recent progress. *Electrochim. Acta* **84**, 223–234 (2012).
 24. Li, H. *et al.* Complete protection of O₂-sensitive catalysts in thin films. *J. Am. Chem. Soc.* **141**, 16734–16742 (2019).
 25. Li, H. *et al.* Suppressing hydrogen peroxide generation to achieve oxygen-insensitivity of a [NiFe] hydrogenase in redox active films. *Nat. Commun.* **11**, 920 (2020).
 26. Plumeré, N. *et al.* A redox hydrogel protects hydrogenase from high-potential deactivation and oxygen damage. *Nat. Chem.* **6**, 822–827 (2014).
 27. Shiraiwa, S. *et al.* Reactivation of standard [NiFe]-hydrogenase and bioelectrochemical catalysis of proton reduction and hydrogen oxidation in a mediated-electron-transfer

- system. *Bioelectrochemistry* **123**, 156–161 (2018).
28. Yuan, M. *et al.* Creating a low-potential redox polymer for efficient electroenzymatic CO₂ reduction. *Angew. Chem. Int. Ed.* **57**, 6582–6586 (2018).
 29. Szczesny, J. *et al.* Electroenzymatic CO₂ fixation using redox polymer/enzyme-modified gas diffusion electrodes. *ACS Energy Lett.* 321–327 (2019).
 30. Sakai, K., Kitazumi, Y., Shirai, O., Takagi, K. & Kano, K. High-power formate/dioxygen biofuel cell based on mediated electron transfer type bioelectrocatalysis. *ACS Catal.* **7**, 5668–5673 (2017).
 31. Ruth, J. C., Milton, R. D., Gu, W. & Spormann, A. M. Enhanced electrosynthetic hydrogen evolution by hydrogenases embedded in a redox-active hydrogel. *Chemistry* **26**, 7323–7329 (2020).
 32. Esselborn, J., Kertess, L., Apfel, U.-P., Hofmann, E. & Happe, T. Loss of specific active-site iron atoms in oxygen-exposed [FeFe]-hydrogenase determined by detailed X-ray structure analyses. *J. Am. Chem. Soc.* **141**, 17721–17728 (2019).
 33. Kubas, A. *et al.* Mechanism of O₂ diffusion and reduction in FeFe hydrogenases. *Nat. Chem.* **9**, 88–95 (2017).
 34. Oughli, A. A. *et al.* A redox hydrogel protects the O₂-sensitive [FeFe]-hydrogenase from *Chlamydomonas reinhardtii* from oxidative damage. *Angew. Chem. Int. Ed.* **54**, 12329–12333 (2015).
 35. Oughli, A. A., Hardt, S., Rüdiger, O., Birrell, J. A. & Plumeré, N. Reactivation of sulfide-protected [FeFe] hydrogenase in a redox-active hydrogel. *Chem. Commun.* **56**, 9958–9961 (2020).
 36. Gallaway, J. W. & Calabrese Barton, S. A. Kinetics of redox polymer-mediated enzyme electrodes. *J. Am. Chem. Soc.* **130**, 8527–8536 (2008).
 37. Cai, R. & Minteer, S. D. Nitrogenase bioelectrocatalysis: from understanding electron-

- transfer mechanisms to energy applications. *ACS Energy Lett.* **3**, 2736–2742 (2018).
38. Bird, C. L. & Kuhn, A. T. Electrochemistry of the viologens. *Chem. Soc. Rev.* **10**, 49 (1981).
 39. Tsujimura, S., Fujita, M., Tatsumi, H., Kano, K. & Ikeda, T. Bioelectrocatalysis-based dihydrogen/dioxygen fuel cell operating at physiological pH. *Phys. Chem. Chem. Phys.* **3**, 1331–1335 (2001).
 40. Lojou, E., Giudici-Orticoni, M. T. & Bianco, P. Direct electrochemistry and enzymatic activity of bacterial polyhemic cytochrome c_3 incorporated in clay films. *J. Electroanal. Chem.* **579**, 199–213 (2005).
 41. Tatsumi, H., Takagi, K., Fujita, M., Kano, K. & Ikeda, T. Electrochemical study of reversible hydrogenase reaction of *Desulfovibrio vulgaris* cells with methyl viologen as an electron carrier. *Anal. Chem.* **71**, 1753–1759 (1999).
 42. Fultz, M. L. & Durst, R. A. Mediator compounds for the electrochemical study of biological redox systems: a compilation. *Anal. Chim. Acta* **140**, 1–18 (1982).
 43. Alber, K. S., Hahn, T. K., Jones, M. L., Fountain, K. R. & Van Galen, D. A. The electrocatalyzed reduction of fumaronitrile in aqueous solution by 4,4'-dimethyl-1,1'-trimethylene-2,2'-dipyridinium ion. *J. Electroanal. Chem.* **383**, 119–126 (1995).
 44. Li, H. *et al.* Preventing the coffee-ring effect and aggregate sedimentation by in situ gelation of monodisperse materials. *Chem. Sci.* **9**, 7596–7605 (2018).
 45. Buesen, D., Li, H. & Plumeré, N. The electron as a probe to measure the thickness distributions of electroactive films. *Chem. Sci.* **11**, 937–946 (2020).
 46. Caserta, G. *et al.* Engineering an [FeFe]-hydrogenase: do accessory clusters influence O_2 resistance and catalytic bias? *J. Am. Chem. Soc.* **140**, 5516–5526 (2018).
 47. Léger, C., Jones, A. K., Roseboom, W., Albracht, S. P. J. & Armstrong, F. A. Enzyme electrokinetics: hydrogen evolution and oxidation by *Allochromatium vinosum* [NiFe]-hydrogenase. *Biochemistry* **41**, 15736–15746 (2002).

48. Ceccaldi, P., Schuchmann, K., Müller, V. & Elliott, S. J. The hydrogen dependent CO₂ reductase: the first completely CO tolerant FeFe-hydrogenase. *Energy Environ. Sci.* **10**, 503–508 (2017).
49. Butt, J. N., Filipiak, M. & Hagen, W. R. Direct electrochemistry of *Megasphaera elsdenii* iron hydrogenase. Definition of the enzyme's catalytic operating potential and quantitation of the catalytic behaviour over a continuous potential range. *Eur. J. Biochem.* **245**, 116–122 (1997).
50. Andrieux, C. P., Dumas-Bouchiat, J. M. & Savéant, J. M. Catalysis of electrochemical reactions at redox polymer electrodes: Kinetic model for stationary voltammetric techniques. *J. Electroanal. Chem. Interfacial Electrochem.* **131**, 1–35 (1982).
51. Bartlett, P. N. & Pratt, K. F. E. Theoretical treatment of diffusion and kinetics in amperometric immobilized enzyme electrodes Part I: redox mediator entrapped within the film. *J. Electroanal. Chem.* **397**, 61–78 (1995).
52. Fourmond, V. *et al.* Mechanism of protection of catalysts supported in redox hydrogel films. *J. Am. Chem. Soc.* **137**, 5494–5505 (2015).
53. Andrieux, C. P., Dumas-Bouchiat, J. M. & Saveant, J. M. Catalysis of electrochemical reactions at redox polymer electrodes: effect of the film thickness. *J. Electroanal. Chem. Interfacial Electrochem.* **114**, 159–163 (1980).
54. Fourmond, V. *et al.* Steady-state catalytic wave-shapes for 2-electron reversible electrocatalysts and enzymes. *J. Am. Chem. Soc.* **135**, 3926–3938 (2013).
55. Fourmond, V. & Léger, C. Modelling the voltammetry of adsorbed enzymes and molecular catalysts. *Curr. Opin. in Electrochem.* **1**, 110–120 (2017).
56. Lampret, O. *et al.* Interplay between CN- ligands and the secondary coordination sphere of the H-cluster in [FeFe]-hydrogenases. *J. Am. Chem. Soc.* **139**, 18222–18230 (2017).
57. Lalaoui, N., Holzinger, M., Le Goff, A. & Cosnier, S. Diazonium functionalisation of carbon

- nanotubes for specific orientation of multicopper oxidases: controlling electron entry points and oxygen diffusion to the enzyme. *Chemistry* **22**, 10494–10500 (2016).
58. Mazurenko, I., Wang, X., de Poulpiquet, A. & Lojou, E. H₂/O₂ enzymatic fuel cells: from proof-of-concept to powerful devices. *Sustain. Energy Fuels* **1**, 1475–1501 (2017).
 59. Dos Santos, L., Climent, V., Blanford, C. F. & Armstrong, F. A. Mechanistic studies of the 'blue' Cu enzyme, bilirubin oxidase, as a highly efficient electrocatalyst for the oxygen reduction reaction. *Phys. Chem. Chem. Phys.* **12**, 13962–13974 (2010).
 60. Del Barrio, M. *et al.* Interaction of the H-cluster of FeFe hydrogenase with halides. *J. Am. Chem. Soc.* **140**, 5485–5492 (2018).
 61. Singh, M. R., Clark, E. L. & Bell, A. T. Effects of electrolyte, catalyst, and membrane composition and operating conditions on the performance of solar-driven electrochemical reduction of carbon dioxide. *Phys. Chem. Chem. Phys.* **17**, 18924–18936 (2015).
 62. Jouny, M., Luc, W. & Jiao, F. General techno-economic analysis of CO₂ electrolysis systems. *Ind. Eng. Chem. Res.* **57**, 2165–2177 (2018).
 63. Ciaccafava, A. *et al.* Electrochemistry, AFM, and PM-IRRA spectroscopy of immobilized hydrogenase: role of a hydrophobic helix in enzyme orientation for efficient H₂ oxidation. *Angew. Chem. Int. Ed.* **51**, 953–956 (2012).
 64. Xu, L. & Armstrong, F. A. Optimizing the power of enzyme-based membrane-less hydrogen fuel cells for hydrogen-rich H₂-air mixtures. *Energy Environ. Sci.* **6**, 2166–2171 (2013).
 65. Xia, H.-Q. *et al.* Dual gas-diffusion membrane- and mediatorless dihydrogen/air-breathing biofuel cell operating at room temperature. *J. Power Sources* **335**, 105–112 (2016).
 66. So, K. *et al.* Direct electron transfer-type dual gas diffusion H₂/O₂ biofuel cells. *J. Mater. Chem. A Mater. Energy Sustain.* **4**, 8742–8749 (2016).
 67. Gentil, S. *et al.* Oriented immobilization of [NiFeSe] hydrogenases on covalently and noncovalently functionalized carbon nanotubes for H₂/Air enzymatic fuel cells. *ACS Catal.*

- 8**, 3957–3964 (2018).
68. Szczesny, J. *et al.* A gas breathing hydrogen/air biofuel cell comprising a redox polymer/hydrogenase-based bioanode. *Nat. Commun.* **9**, 4715 (2018).
69. Conzuelo, F., Marković, N., Ruff, A. & Schuhmann, W. The open circuit voltage in biofuel cells: nernstian shift in pseudocapacitive electrodes. *Angew. Chem. Int. Ed.* **57**, 13681–13685 (2018).
70. Shleev, S. *et al.* Oxygen electroreduction versus bioelectroreduction: direct electron transfer approach. *Electroanalysis* **28** 2270–2287 (2016).
71. Sakai, K. *et al.* Interconversion between formate and hydrogen carbonate by tungsten-containing formate dehydrogenase-catalyzed mediated bioelectrocatalysis. *Sensing and Bio-Sensing Research* **5**, 90–96 (2015).
72. Léger, C., Lederer, F., Guigliarelli, B. & Bertrand, P. Electron flow in multicenter enzymes: theory, applications, and consequences on the natural design of redox chains. *J. Am. Chem. Soc.* **128**, 180–187 (2006).
73. Birrell, J. A. *et al.* Artificial maturation of the highly active heterodimeric [FeFe] hydrogenase from *Desulfovibrio desulfuricans* ATCC 7757. *Isr. J. Chem.* **56**, 852–863 (2016).
74. Rodríguez-Maciá, P. *et al.* Sulfide protects [FeFe] hydrogenases from O₂. *J. Am. Chem. Soc.* **140**, 9346–9350 (2018).

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Author contributions

N.P., C.L., V.F., J.A.B. and O.R. conceived the research. S.H and S.S. developed and carried out the synthesis of the polymers, and performed all electrochemical experiments involving the polymer. D.T.F. developed and carried out the synthesis of the dendrimer, and performed all electrochemical experiments involving the dendrimer. S.H. contributed with the fuel cell and electrolyser experiments. O.R. contributed with direct electron transfer experiments. V.F.

performed electrochemical modeling and data analysis. J.A.B. contributed the hydrogenase. All authors contributed to writing the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper.