

# Tropical mangrove forests as a source of dissolved rare earth elements and yttrium to the ocean

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1	Tropical mangrove forests as a source of
2	dissolved rare earth elements and yttrium.
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Abstract: Rare earth elements (REEs) and Y are commonly used as geochemical proxies for water chemistry, history of the continental crust and provenance studies. At the continent-ocean interface, the estuarine geochemistry of REEs is commonly considered to be driven by a large-scale removal. Consequently, the river-borne flux of dissolved REEs to the marine budget is assumed as a minor fraction. Here, we report a significant release of dissolved REEs, together with a fractionation between light, heavy REEs and Y, in the tropical mangrove estuaries of New Caledonia. These observations were associated with internal recycling within the redox-dynamic mangrove system. The possible contribution of groundwater can be ruled out based on water stable isotopes. These findings imply that tropical mangrove estuaries act as a sizeable source of REEs and Y to the ocean rather than as a buffer zone. We extrapolated our data toward global dissolved fluxes of REEs and Y through the mangroves. This preliminary calculation suggests that the mangrove system supplies 2.6-5% of global river-borne dissolved Nd, a REE with the most comprehensive mass balance. Therefore, oceanic mass balance budget models may need to consider this atypical system as an input to balance the global distribution of REEs and Y.

**Keywords:** Rare earth elements, yttrium, water stable isotopes, mangrove, estuary.

#### 1. Introduction

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Over the past decade, rare earth elements (REEs or the lanthanide series), Y and REE isotopes have received significant attention in geochemistry as tracers of the geological history of the continental crust and provenance study (Bayon et al., 2015). In paleoenvironmental studies, REEs are also commonly used as proxies to reconstruct water chemistry and oxygen saturation as per the distinct behaviour of Ce during redox transitions (Auer et al., 2017; Wallace et al., 2017). The fractionation between the light REEs (LREEs), heavy REEs (HREEs) and the non-lanthanide Y can also highlight the mechanisms of particle transport, sediment deposition and even geological features (e.g., iron formation) over the Earth's history (Bayon et al., 2015; Planavsky et al., 2010). More recently, because of the implication of REEs in novel technologies, REE anomalies (increased in concentrations of a specific REE relative to its neighbouring REEs) could also be used as a proxy for emission sources (Dang and Zhang, 2021; Ma et al., 2019; Tepe et al., 2014). However, the environmental chemistry of REEs remains fragmented, mostly regarding their anomalous behaviour across the river-estuarine-coastal water mixing zone where the fractionation between LREEs and HREEs is often associated with the redistribution of REEs among the dissolved and particulate fractions and the formation of stable dissolved complexes (Nozaki et al., 2000; Sholkovitz and Szymczak, 2000; Sholkovitz, 1995). As a consequence, the ocean mass budget of REEs remains unbalanced with missing fluxes at the global scale (Arsouze et al., 2009; Garcia-Solsona and Jeandel, 2020; Pourret and Tuduri, 2017; Tachikawa et al., 2003). The interpretation of estuarine REE behaviour is fundamental to highlight their geochemistry at the continent-ocean interface. One conventional feature derived from decades of investigation on REEs' estuarine geochemistry is that large-scale removal of dissolved REEs (especially LREEs) occurs in the estuarine mixing zone (Elderfield and Greaves, 1982; Lawrence and Kamber, 2006; Nozaki et al., 2000; Pourret and Tuduri, 2017; Sholkovitz, 1995). This phenomenon minimizes the effective continental flux of REEs to the ocean and subsequently affects the marine budget calculations of REEs (Nozaki et al., 2000). Various factors would lead to such extensive removal of REEs. These include planktonic uptake, coprecipitation with ferromanganese oxyhydroxides and salt-induced coagulation of colloids (Hoyle et al., 1983; Nozaki et al., 2000; Sholkovitz and Szymczak, 2000). However, a limited number of publications pointed out a supply of dissolved

61 REEs in the mid-estuary zones (Lawrence and Kamber, 2006; Nozaki et al., 2000). This REE 62 release could be associated with desorption from terrigenous suspended particles or 63 remineralization in the estuarine system. 64 Moreover, most of the estuarine systems investigated so far for REEs geochemistry were major 65 well-oxygenated riverine systems. However, it has been demonstrated that REEs cycling in 66 marine waters can be impacted by redox variations (Bau et al., 1997). This process is particularly 67 important as it can increase the effective continental REE flux to the oceans through enhanced 68 supply to the water column upon dissolution of REEs-bearing oxyhydroxide phases. Therefore, 69 further insights on REE behaviour across the redox-sensitive estuarine mixing zone are needed to 70 strengthen our understanding of the REE geochemistry at the continent-ocean interface. 71 Tropical mangrove forests are of great interest to investigate these environmental conditions. 72 First, they cover a large surface area (150,000 km<sup>2</sup>) where complex hydrological processes 73 balance abundant precipitation, surface runoff, and evaporation (Spalding, 2010). Moreover, the 74 combination of elevated and annually constant temperature patterns with high bioavailability of 75 natural organic matter boosts bacterial respiration, promoting hypoxia (Dubuc et al., 2019). To 76 date, very limited studies on REEs focused on this critical ecosystem as it is generally assumed 77 that the mangrove systems act as a buffer zone retaining trace elements and thus contribute a 78 minor input for REEs to the oceans (Marchand et al., 2012). 79 Here, we investigated REE internal cycling within two estuarine mangrove systems in the 80 subtropical region of New Caledonia. We meticulously measured dissolved REEs and Y over a 81 24-hour cycle in mangrove stations during a dry season and the spatial variations across the 82 estuary in a rain season. To track whether additional water masses alter the mixing between the fresh and seawater endmembers, we also measured stable water isotopes ( $\delta^{18}O$  and  $\delta D$ ) to assess 83 84 the hydrologic regime. We compared the stable water isotopes in these mangrove systems with 85 other river waters collected in southeastern New Caledonia to verify the signature of the 86 freshwater endmembers. Altogether, the data highlights REE behaviour during the estuarine 87 mixing and document the mangrove forest's role on REE cycling at the continent-ocean interface. 88 This dataset will help answer whether the REE continental flux from the tropical mangrove-89 dominated estuaries should be included in the REE marine budget.

#### 2. Materials and Methods

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2.1.Study site description

In the Southwest Pacific Ocean, New Caledonia is surrounded by the Coral Sea (Figure 1A,B). The main island "Grande Terre" is 400 km long and 64 km wide with a semi-arid tropical climate along the west coast. The whole island is also bordered by a coral reef enclosing a lagoon of over 20,000 km<sup>2</sup>. The rich reef diversity and associated ecosystems were inscribed as a UNESCO World Heritage Site in 2008. On the other hand, most of the island is mountainous with vast ultramafic masses, schists and outcrops of basalt (Lillie and Brothers, 1970). Weathering of the ultramafic rocks' surface led to the formation of thick lateritic regoliths (tens of meters) hosting deposits of transition metallic elements of prime economic importance (Dublet et al., 2014; Noël et al., 2014). Between the lucrative mineral-rich land and the sea of tremendous ecological importance, the mangrove system acts as a buffer zone to filter heavy metals from reaching the reef (Marchand et al., 2012). The hydrological behaviour of the Grande Terre is clearly distinct between the southwestern and northeastern coasts because of the 'rain-shadow' effect across the central massif divide (Terry and Wotling, 2011) (Figure 1B). Accordingly, the southwestern coast (leeward) tends to be drier than the windward northeastern coast. Overall, the hydrologic regime is characterized by high annual variability of rainfall amount and extreme events. Moreover, high and steep watersheds are associated with flash floods and thus rapid water transfer to the lagoons (Desclaux et al., 2018). The Voh-Kone region (Northwest of the Grande Terre, **Figure 1B**) is an area of great economic and ecological interest. The Koniambo Massif hosts a large mine (Koniambo mine, Figure 1C), one of the largest nickel reserves in New Caledonia. The ore is conveyed to a coastal smelting site (Koniambo Nickel SAS, Figure 1C). Besides, the hydrologic system mainly comprises of two major rivers (Temala and Voh) discharging into the mangroves, then the bays of Chasseloup and Vavouto, respectively. Moreover, the hydrologic system comprises also of relatively smaller streams, such as the Coco River. These small tributaries have their flows significantly reduced during the dry season.

This study focused on the Temala and Coco Rivers (**Figure 1C**) in the Voh region. The Temala River originates from outside of the Koniambo Massif and discharges into Chasseloup Bay. It is, therefore, considered not impacted by mining activities. On the other hand, the Coco River's sources are in the Koniambo Massif hosting the Koniambo Mine. Along its course toward Vavouto Bay, the Coco River meanders parallelly to the ore conveyor. The Koniambo Nickel SAS smelting site is indeed only ca. 1.2 km away from the stream (**Figure 1 C**). Therefore, it is highly probable that this river is subject to anthropogenic impacts. Besides this contrasted exposition to anthropogenic sources, the natural geological settings of both rivers also differ. The Temala River catchement is made of soils developed on volcano-sedimentary rocks (cherts, basalts), whereas the Coco River catchment is composed of Ni-laterites developed on ultramafic rocks (peridotites) (Merrot et al., 2019).

We also investigated river waters (Dumbea, Coulee, Tontouta, Pirogue, **Table 1**) around Noumea in the Grande Terre's Southeast (**Figure 1B**) as a reference site for hydrological assessment using salinity and water isotope mixing.

# 2.2.Water sampling

We organized multiple sampling trips to investigate the hydrologic regime and REE cycling. The two first sampling campaigns were organized in June 2019 (dry season) and March 2020 (rain season) to collect samples in the Voh-Kone region (Temala and Coco Rivers). We also collected samples in four southeastern rivers (Dumbea, Coulee, Tontouta, Pirogue) in January, March, October and November of 2018. The March samples were collected during and after the Hola tropical cyclone while other samples are collected during the dry season.

During a dry season (June 2019), the first campaign aimed for a high-temporal resolution sampling in the mangrove forests (sites C2 and T6, hereafter referred to as "mangrove stations", **Figure 1C**). For that purpose, two automatic samplers (ISCO) were deployed during high tide (8-9h AM). The samplers were programmed to collect water samples one hour after installation to ensure the resettlement of resuspended sediments during deployment. The first water samples were collected at 9h AM at C2, and 10h AM at T6. Following samples were collected every hour. At the end of the 24-hour automatic sampling cycle, all samples were filtered (0.22  $\mu$ m, cellulose acetate, Sartorius) directly on site. The filtered waters for stable water isotopes were filled to the

top to avoid water-air exchange, while samples for elemental analysis were acidified to pH<2 with ultra-trace HNO<sub>3</sub> (Merck). The samples were then immediately shipped by air to Trent University for analysis.

In 2020, during the rain season, a series of samples were collected across the salinity gradient along the Temala (T1-T8) and Coco (C1-C5) rivers (hereafter referred to as the "estuarine stations", **Figure 1C**). An additional sample was collected in the lagoon to serve as a marine endmember (M, **Figure 1C**). The samples were treated with the same protocol as in 2019 before being shipped to Trent University.

All the filters, sampling and storing bottles were previously rinsed with 10% HCl and milli-Q water (18 M $\Omega$  cm) in the laboratory and with river or lagoon water in the field.

# 2.3. Water stable isotopes

Water stable isotopes (δ¹8O and δD, deuterium) were measured using a Liquid Water Isotope
 Analyzer (LWIA-24d, Los Gatos Research) at the Water Quality Center (Trent University,
 Ontario, Canada). The delta notation was calculated relative to the V-SMOW (Vienna-Standard
 Mean Ocean Water) standard.

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$$\delta^{18}O = \left(\frac{\binom{180}{160}_{sample}}{\binom{180}{160}_{V-SMOW}} - 1\right) \times 1000(Eq. 1)$$

$$\delta D = \left(\frac{\left(\frac{D}{^{1}H}\right)_{sample}}{\left(\frac{D}{^{1}H}\right)_{V-SMOW}} - 1\right) \times 1000(Eq. 2)$$

Analytical recovery was assessed using two isotopic natural water reference materials (CRMs, Elemental Microanalysis, UK), covering the range of  $\delta$  values found in natural waters (**Table S1**). Averages and standard deviations of each measurement (standards and samples) were calculated based on eight injections (i.e., n = 8, 750 nL each). We also used ultra-high purity water (18.2 M $\Omega$  cm) as an in-house standard to bracket every two samples to monitor analytical

drift and eliminate any memory effects. However, no correction was necessary. Analytical recovery of stable water isotopes is reported in **Table S1**.

#### 2.4.Elemental analysis

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To accurately determine the concentrations of REEs and Y at the ng/L levels in complex environmental matrices (seawater), we have performed a preconcentration step using column chromatography before analysis by Inductively Coupled Plasma Mass Spectrometry (ICP-MS). A full description of the REE column chromatography and ICP-MS analysis were reported elsewhere (Dang and Zhang, 2021; Ma et al., 2019). Briefly, REEs and Y in the filtered water samples were preconcentrated using 0.2 mL Nobias Chelate-PA1 resin (Hitachi High-Technologies). An aliquot of 50 mL of samples buffered at pH 6 was loaded on previously washed and conditioned resin. The seawater matrix was then washed off with 40 mL 0.05 M acetate buffer solution (pH = 6). REEs and Y were collected by eluting 20 mL of a 1 M HNO<sub>3</sub> solution through the column. This aliquot was collected into a 30 mL PTFE vessel and completely evaporated on a hotplate at 100°C. A final 2 mL of 0.5 M HNO<sub>3</sub> was added to retake the REEs and Y. All acids used to clean and wash the resins were double-distilled trace metal grade. The acetate buffer was also passed through the Nobias column for purification. We also meticulously checked procedural blanks and column recovery in every batch of 10 samples. Procedural blanks were between 0.1 to 29 pg of REEs and Y and significantly less than their mass in natural samples. Several aliquots (50 mL) of certified reference seawater CASS-6 (National Research Council of Canada, NRCC) were also repeated to test column recovery (Ma et al., 2019). REEs and Y were determined using a triple quadrupole ICP-MS (8800 Agilent Technologies) at the Trent Water Quality Center. All REEs and Y were analyzed in MS/MS mode with O<sub>2</sub> as the reaction cell gas. Detection limits were in the range of 0.1-0.4 ng/L. Although there were no certified reference materials for REEs, we compared the REEs and Y concentrations measured in an NRCC natural water certified reference material (SLRS-6) to the values reported in a European interlaboratory calibration (Yeghicheyan et al., 2019). Excellent recovery was observed; the detailed data are reported in **Table S2**.

## 2.5. Normalization of REE patterns and calculations of anomalies

A common practice in the scientific community to eliminate the Oddo-Harkins effect and compare samples is to normalize the measured REE concentrations to geological materials (e.g., upper continental crust, shales, chondrite) or offshore seawater (Piper and Bau, 2013). Here, we normalized the measured concentrations of dissolved REEs to Post-Archean Australian Shale (PAAS (McLennan, 2001)) (Goldstein and Jacobsen, 1988; Ma et al., 2019; Piper and Bau, 2013). The shale-normalized concentrations are hereafter referred to as REE<sub>PAAS</sub>.

2015). The shale-normanzed concentrations are hereafter referred to as REEPAAS.

In the current literature, excess or depletion of an REE relative to its neighbouring REEs are referred to as positive or negative anomaly of this element (REE/REE\*). Accordingly, the REE anomalies could be determined by several approaches (e.g., arithmetic mean, geometric extrapolations, modelling the shape of PAAS-normalized patterns, third-order polynomial fit). Using all these approaches, Hatje et al. have demonstrated minor variability in the final results (Hatje et al., 2016). Therefore, we calculated Ce, Eu, and Gd anomalies using the following equations (arithmetic mean).

$$Ce/Ce = \frac{2 \times Ce_{PAAS}}{La_{PAAS} + Pr_{PAAS}} (Eq. 3)$$

$$Eu/Eu = \frac{2 \times Eu_{PAAS}}{Nd_{PAAS} + Sm_{PAAS}} (Eq. 4)$$

$$Ce/Ce = \frac{Gd_{PAAS}}{0.33 \times Sm_{PAAS} + 0.67 \times Tb_{PAAS}} (Eq. 5)$$

### 3. Results and Discussions

#### 3.1.Physical-chemical properties

At both mangrove stations (C2 and T6, **Figure 1B**), pH values varied within a narrow range (7.8  $\pm$  0.1, n = 48, **Table 1** and **Figure S1B**). Besides, the daily variations in salinity followed a sinusoidal curve, typical of estuarine systems (**Figure 2A**). This variation pattern is more noticeable at Temala with low tide (S = 26) recorded at 4h and 16h, and high tide (S = 33) at 22h. However, the salinity pattern at Coco is significantly distinct, with two events marked by steep drops at 13h, 19h and a third gradual decrease from midnight until 7h AM (1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup> as marked in **Figure 2A**). The variation range of salinity in Coco is also narrower (29.6 to 34.2) than in Temala (26.0 to 33.0) (**Table 1**).

- Temperature variations also help distinguish the two systems. Water temperatures increased from
- 225 16.8°C at 11h and held constant at ca. 17.5°C at Temala (**Figure S1A**, average of 17.3  $\pm$  0.2°C, n
- = 24, **Table 1**). However, the temperature pattern at Coco showed broader variations (15.5°C to
- 227 16.5°C, average of  $16.1 \pm 0.3$ °C, n = 24, **Table 1**) with temperature drops at 11h and 21h (**Figure**
- 228 S1A). Minimums in water temperatures at Coco are lagged by two hours relative to the salinity
- 229 minimums (13h and 19h, **Figure 2A**).
- Across the entire estuaries (estuarine stations), salinities ranged from 0 to 34.9 for the river water
- and marine endmembers, respectively (**Figure 2B**). On the other hand, all the river samples
- collected from the Southeast of New Caledonia are freshwater with a salinity of zero (**Figure**
- 233 **2C**).
- 234 3.2.Stable water isotopes and the hydrologic regime
- 235 The  $\delta D$  and  $\delta^{18}O$  pairing. As the mangrove stations have narrow daily salinity variations (**Table**
- 236 1), so too is the range of stable water isotope compositions relative to the whole estuaries (**Figure**
- 237 **2 D-I**). At the mangrove stations, the  $\delta D \delta^{18}O$  relationship has the equation of  $\delta D = 4.3 \times \delta^{18}O +$
- 5.5 (n=40, **Figure S2**). The slope of  $4.3 \pm 0.6$  is typically lower than other tropical Pacific regions
- 239 (5.1 to 6.5, Conroy et al. (2014)), and the global seawater average of 7.4 (Rohling, 2007). This
- indicates a strong influence of the evaporation process in the mangrove during the dry season.
- Across the estuaries, the  $\delta D$ - $\delta^{18}O$  relationship of Coco and Temala waters aligns with
- southeastern rivers with the equation of  $\delta D = 4.8 \times \delta^{18}O 4.6$  (n=29, r<sup>2</sup>=0.9, **Figure S2**). The
- slope  $(4.8 \pm 0.3)$  is very similar to the mangrove stations  $(4.3 \pm 0.6)$ , only the intercepts differ (-
- 5.3 across the estuary and +5.5 in the mangrove during the dry season).
- According to the interpretation of the local evaporation line, as described by Wolfe et al. (2007),
- local surface waters often plot in linear clusters to the right of the Meteoric Water Line along a
- slope in the range 4 to 6. Our observations of consistent slopes  $(4.3 \pm 0.6 \text{ and } 4.8 \pm 0.3)$  fall into
- 248 this conventional interpretation (**Figure S2**) considering the Local Meteoric Water Line (LMWL)
- of  $\delta D = 7.1 \times \delta^{18}O + 12.6$  (retrieved from the online isotopes in precipitation calculator,
- www.waterisotopes.org).

251 To explain the difference in the intercepts (higher values in the dry season than the rain season), 252 we calculated the deuterium excess (d-excess) using the conventional approach: d-excess =  $\delta D$  - $8 \times \delta^{18}$ O. This variable is valuable as it quantifies the deviation of a given dataset from the Global 253 Meteoric Water Line (GMWL) by differential kinetic fractionation effects between D and <sup>18</sup>O. 254 255 Such effects are related to humidity, moisture recycling and post-deposition process (Dansgaard, 256 1964). Low d-excess tends to reflect slow evaporation due to high humidity, while high d-excess 257 values indicate fast evaporation due to low humidity (Lee et al., 2003). This interpretation is coherent with our observations as high d-excess values (15.8±1.3% and 15.9±2.9% at Coco and 258 259 Temala, respectively) were observed at the mangrove stations in dry season while low d-excess values were typical of rain season and across the estuaries  $(0.4\pm5.8 \%)$  (**Table 2**). 260 **Pairing \delta D and \delta^{18}O with salinity.** The linear relationships between  $\delta^{18}O$ ,  $\delta D$  and salinity have 261 been used to estimate the stable water isotope composition of a freshwater endmember, defined 262 263 as the  $\delta^{18}$ O and  $\delta$ D values when salinity equals zero. This extrapolation should be interpreted 264 with careful consideration of the contributions of regional precipitation, river water and runoff. However, it has been demonstrated that the extrapolated  $\delta^{18}O$  and  $\delta D$  of freshwater endmembers 265 in the Great Barrier Reef (NE Australia) reflected the isotope composition of local river water 266 267 with relatively insignificant contributions of regional precipitation and runoff (Munksgaard et al., 268 2012). Accordingly,  $\delta^{18}$ O and  $\delta$ D values in the samples collected in the mangroves of Coco and Temala 269 varied linearly with salinity (**Figure 3** and **Table 2**). In addition, all linear relationships 270 271 converged to a common signature of the marine endmember (blue diamond in Figure 3). We 272 extrapolated these linear relationships toward salinity of zero to determine the isotope 273 composition of each river water endmember (**Table 2**). It is important to note that the 274 interpretation of such relationships often requires a spatial and temporal assessment of the two 275 systems (Conroy et al., 2017). As the samples were collected simultaneously in the dry season, 276 the difference in water isotope composition between the freshwater endmembers at Coco and 277 Temala can thus be only attributed to distinct water sources in each river system. 278 In the mangrove stations, the two linear regression lines are significantly distinct between Coco and Temala, except for a few stations at Temala that fall on the regression line of Coco (encircled 279

280 data, Figure 3B). These data represent the last samples collected during the high tide over the 24-281 hour series (3<sup>rd</sup> discharge event of the Coco River, **Figure 2A**). All physical-chemical properties and stable water isotopes of the two rivers overlap during this period (grey zone in Figure 2 A, 282 283 **D, G**). This probably reflects a common blackish water mass formed during the high tide. 284 Therefore, we excluded these data from Temala's  $\delta D$ -salinity regression because of the lack of 285 more robust proxies to confirm this phenomenon and assuming that it may be minor. The 286 exclusion of these three data did not affect the fact that the water isotope composition is significantly more depleted in the Coco River than Temala during the dry season. This depletion 287 is more noticeable for  $\delta D$  (-72.9±8.9% vs. -36.0±2.8%, respectively) than  $\delta^{18}O$  (-9.9±1.7% vs. -288 289  $8.4\pm1.2\%$ , respectively) (**Table 2**). However, across the estuaries and during the rain season, the  $\delta D$ -S and  $\delta^{18}O$ -S relationships for 290 both river systems are significantly similar (p<0.05) and align along with consistent equations 291 292  $(\delta D=1.0\times S-33.7 \text{ and } \delta^{18}O=0.2\times S-5.2, \text{ Table 2} \text{ and Figure 3A})$ . Both the slopes and intercepts are close to the Temala mangrove during the dry season ( $\delta D=1.0\times S-36.0$  and  $\delta^{18}O=0.2\times S-8.4$ , 293 294 **Table 2**). This observation suggests that the hydrologic regime in the Temala River differs only slightly within seasons with relatively constant water isotope compositions of the river 295 endmember ( $\delta D_{River}^{Temala} \sim -34$  to -36% and  $\delta^{18} O_{River}^{Temala} \sim -5.2$  to -8.4%). These values are close 296 to the four southeastern rivers over an annual cycle (**Figure 3A**) ( $\delta D_{River}^{SE} = -31 \pm 6\%$  and 297  $\delta^{18}O_{River}^{SE} = -5.8 \pm 1.0\%$ , **Table 2**), confirming reasonably consistent  $\delta D\text{-S}$  and  $\delta^{18}O\text{-S}$ 298 relationships and hydrologic regimes of major river systems along the western coast of New 299 300 Caledonia. 301 It is important to note that the Coco River is a small tributary with a relatively dry riverbed with 302 significantly decreased flow during the dry season. Consequently, the Coco River's isotopic 303 compositions differ substantially during the dry and rain seasons (Table 2) due to less 304 precipitation and enhanced evapotranspiration during the dry season. 305 In summary, the difference in stable water isotope composition between the Coco and Temala 306 rivers is related to the natural effects of a depleted hydrologic reservoir with enhanced

evaporation. This effect is naturally more noticeable in small reservoirs during the dry season,

such as the Coco River than the major streams (Temala and the four southeastern rivers). Overall,

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we observed a relatively conservative hydrologic system with the mixing of only two water masses: the river freshwater and the marine endmembers. Water isotopic data allow excluding the contribution of a third water mass (e.g., groundwater discharge) in these mangrove-dominated estuarine systems.

#### 3.3. Rare earth element cycling in the mangrove system.

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The PAAS-normalized REE patterns are typical of coastal seawater with a gradual enrichment of heavy REEs relative to light REEs (Hoyle et al., 1983; Nozaki et al., 2000; Piper and Bau, 2013) (Figure 4). Moreover, we observed a depletion of Ce but an enrichment of Eu and Gd relative to their neighbouring REEs that are commonly referred to as anomalies. The anomalies of Ce and Eu were already extensively discussed in the literature. The negative Ce anomaly was shown to develop progressively with increasing salinity as dissolved Ce(III) undergoes continued oxidation to insoluble Ce(IV) across the estuarine zone (Nozaki et al., 2000). The positive Eu anomaly was also reported in the dissolved fractions of various major rivers (Amazon, St Lawrence, Piper and Bau, (2013)) and the clay fractions of river sediments worldwide (Bayon et al., 2015). Several previous studies have recorded positive Eu anomaly in continental materials (e.g., suspended load, atmospheric aerosol and dust, Censi et al., 2004; Goldstein and Jacobsen, 1988). Therefore, the positive Eu anomaly observed in the dissolved loads was interpreted as the result of dissolution from suspended particles and formation of stable dissolved complexes during transport (Nozaki et al., 2000). On the other hand, the Gd anomaly is commonly associated with anthropogenic emissions, such as medical sources (medical resonance imagery) or wastewater (Hatje et al., 2016; Tepe et al., 2014). We observed Gd anomaly (Gd/Gd\* up to 3.5) only during the dry season in the Coco river, in the first and second freshwater discharge events (Figure S3 and Figure 2A). We have previously suggested that the 3<sup>rd</sup> event may be associated with a common brackish water at both Coco and Temala rivers formed during the high tide from 4h to 9h AM (Figure 2A), which is Gd-anomaly-free (**Figure S3D**). No Gd anomaly ( $Gd/Gd^* = 1$ ) was indeed observed in the marine endmember (blue diamonds, **Figure 4 C,D**). Given the absence of major medical facilities and considering the low population along the Coco river, a possible medical or wastewater origin for this Gd anomaly can be ruled out. However, the Coco River is associated with the nickel mined lateritic Koniambo regolith where a 13km long ore conveyor was built parallel to the river

340 While there is no precedent study reporting Gd anomaly associated with lateritic ores extraction 341 and processing, the Gd anomaly observed in the Coco river might be associated with those 342 activities. 343 Estuarine mixing behaviors of REEs and Y. The REE concentrations in the marine endmembers 344 are typically low; the sum of all REE concentrations (ΣREEs) approximates 6 ng/L. However, 345 there was a significant difference in REE total concentrations between the two river endmembers, with Temala containing more REEs than Coco (ΣREEs of 24 and 2.6 ng/L, respectively). This 346 347 trend was also observed at the mangrove stations (**Figure S3A**). The higher REE concentrations 348 measured in the Temala River waters compared to the Coco River are directly associated with the 349 geology of their watersheds. The Temala watershed is indeed made of soils developed on 350 volcano-sedimentary rocks, whereas the Coco river meanders on lateritic soils developed on 351 ultramafic rocks (Figure 1C) (Merrot et al., 2019). As volcaniclastic bedrock tend to contain higher REE and Y concentrations than the granitic regolith (Chapela Lara et al., 2018), the 352 353 difference in the watershed's geology could be directly associated with higher REE 354 concentrations in Temala river waters than Coco. 355 Moreover, we did not observe any REEs and Y removal during estuarine mixing between the 356 rivers and the marine endmember, as commonly assumed for REE estuarine geochemistry (see 357 section 1). In fact, we measured very high REE concentrations in the mangrove forest (salinity 358 range of 25-33, see La concentrations in **Figure 5A**). The high REE (La) concentrations in the mangrove area seem to be conservatively mixed with the freshwater mass (y=0.6x+3.5,  $r^2 = 0.97$ ) 359 and the marine endmember (y=-5.2x+183,  $r^2 = 0.84$ ). This situation was not reported previously. 360 361 It is commonly assumed that approximately 60-90% of REEs are removed from the low salinity 362 region of an estuary by flocculation/coagulation of dissolved organic-bound Fe and colloidal 363 materials depleting REEs from the dissolved pool (Lawrence and Kamber, 2006; Pourret and 364 Tuduri, 2017; Sholkovitz and Szymczak, 2000; Sholkovitz, 1995). Minor releases of REEs 365 during an estuary mixing were only reported in the turbid-clear water transition zone (S=12-15) 366 in the Chao Phraya estuary (Thailand) (Nozaki et al., 2000) and the mid-salinity zone of the 367 Elimbah Creek (Australia) (Lawrence and Kamber, 2006). Such REE release was attributed to 368 several mechanisms, including desorption from suspended particles, coastal erosion, and

and the Koniambo Nickel SAS smelting plant is adjacent to the Coco mangrove (**Figure 1C**).

369 mineralization during early diagenesis. In addition, the reductive dissolution of ferromanganese 370 oxides has been shown to significantly release REEs into the porewater, although LREEs tend to 371 sorb back onto the newly formed ferromanganese oxides at the sediment-water interface (Och et 372 al., 2014). 373 Although the La-salinity biplots (**Figure 5A**) could be interpreted as mixing of three water 374 masses (river, mangrove forest, and marine), stable water isotopes (Figure 3) allow to rule out 375 the possibility of a third water mass in the estuaries of Coco and Temala. Thus, the high La 376 concentrations in the mangrove are instead explained by specific biogeochemical processes 377 leading to the substantial release of dissolved REEs. Given the subtropical weather condition and the high availability of fresh organic matter in the mangrove forest (Noël et al., 2014), it is highly 378 379 probable that bacterial activity contributes to this REEs solubilization in the sediments porewater 380 (Och et al., 2014). In the mangrove system, the tidal conditions with water pushing up through 381 the porous sediment (seawater circulation cell, **Figure 6A**) would then favour REE 382 remobilization to the water column. A previous study has demonstrated significant fluxes of 383 REEs from such a seawater circulation through diagenesis-active sediments (i.e., organic matter 384 degradation and reductive dissolution of ferromanganese oxides (Paffrath et al., 2020), **Figure 6**). Other processes such as passive diffusion fluxes or sediment resuspension events induced by 385 386 waves and tides (Dang et al., 2020) or bioturbation could also contribute to the REE release 387 towards the water column (**Figure 6 C**). At a larger scale of the Northwestern Mediterranean Sea, 388 upward diffusion of REEs (Nd) from sediment porewater counts for 30% of the Nd marine 389 budget (Garcia-Solsona and Jeandel, 2020). Moreover, given the organic mater-rich surface 390 waters in the mangrove system and the high affinity between REEs and dissolved organic carbon 391 (Davranche et al., 2004; Marsac et al., 2010; Pourret and Davranche, 2013), it is also highly 392 possible that the formation of stable organic complexes could contribute to maintaining REEs in 393 solution in this mid-salinity region (25-33). 394 Fractionation of HREEs/LREEs and elemental ratios. All the water samples collected showed 395 a progressive enrichment of HREEs relative to the PAAS shale (Figure 4). This is consistent 396 with the conventional REE behaviours in the aquatic environment where the fractionation 397 between the dissolved and particulate loads of HREEs and LREEs leads to this characteristic 398 REE curve (Piper and Bau, 2013). Therefore, elemental ratios between representative HREEs and 399 LREEs (e.g., Er/Nd (Nozaki et al., 2000), Nd/Lu (Sholkovitz, 1995), Ho/Er (Lawrence and 400 Kamber, 2006)) could help determine whether this fractionation occurs in the freshwater 401 endmember or the estuary. As the interpretations using different ratios are very similar, we 402 choose to discuss only the Er/Nd (weight ratios). 403 The Er/Nd ratios in both UCC (Rudnick and Gao, 2003) and PAAS (McLennan, 2001) 404 approximate 0.08, while those in the river endmembers are significantly above this value (0.2 and 405 0.3 at Temala and Coco, respectively, **Figure 5B**). This indicates that the fractionation between 406 HREEs and LREEs occurred in the freshwater system. It is important to note that such 407 fractionation is pH-dependent. At the pH range observed in Temala and Coco rivers (7.7-7.9, 408 **Table 1)**, REE adsorption on particles is enhanced, leading to significant fractionations by 409 preferential LREE removal (Sholkovitz, 1995). Furthermore, the measured Er/Nd in the estuarine 410 samples varied with salinity along the conservative mixing line of Temala (Figure 5B), 411 suggesting no further fractionation (Lawrence and Kamber, 2006). This is consistent with the 412 occurrence of dissolved organic-REE complexes in the mangrove waters, as previously 413 suggested. Previous studies have demonstrated that the presence of dissolved organic matter 414 significantly reduced REE sorption on ferromanganese oxides and the subsequent fractionation between LREEs and HREEs (Davranche et al., 2005, 2004). 415 416 On the other hand, a negative departure from the conservative mixing line (i.e., Er/Nd ratios 417 decreased) was noticed at the mangrove stations (black arrow in **Figure 5B**). This indicates a 418 preferential release of LREEs (Nd) relative to the HREEs (Er). This observation is in agreement 419 with the hypothesis that LREEs previously bound to the solid fraction are released toward 420 solution when the oxide phases undergo reductive dissolution (Bau et al., 1997). High REEs 421 concentrations in porewater promote upward diffusion from the sediments by the seawater 422 circulation cell or diffusive flux (**Figure 6**). 423 Furthermore, variations in the Y/Ho ratios are also of great interest. Although these two elements 424 have similar ionic radii, their electronic configurations differ. Yttrium has no f-electron while Ho 425 has 10 electrons filling up the f orbitals (Lawrence and Kamber, 2006). Therefore, the ability to 426 form strong surface and solution complexes using the f orbitals (Byrne and Lee, 1993) drives the 427 fractionation of this pair during weathering, riverine transport or in the marine environment

128	(Lawrence and Kamber, 2006; Nozaki et al., 2000). As a result, Ho tends to form more stable
129	surface complexes than Y (Bau et al., 1997; Byrne and Lee, 1993; Lawrence and Kamber, 2006).
130	Therefore, two conditions could lead to Y/Ho fractionation during estuarine mixing. First,
131	coagulation, flocculation and sedimentation of Ho-bearing colloidal particles preferentially
132	remove Ho from solution. Second, as Y tends to form weaker surface complexes than Ho, this
133	fractionation is enhanced by the salt effect (competition for surface sorption sites), which favors
134	Y release from colloidal particles toward solution. Both processes were well documented in the
135	literature to support the increase in the Y/Ho ratio during the estuarine mixing (Figure 6B)
136	(Lawrence and Kamber, 2006).
137	The Y/Ho ratio in the Temala river endmember (28.4) approximates UCC and PAAS (25.3
138	(Rudnick and Gao, 2003) and 27.2 (McLennan, 2001), respectively), indicating that there was no
139	significant fractionation between Y and the REEs in the Temala river (Figure 5C) (Lawrence and
140	Kamber, 2006). However, in the Coco river endmember, the Y/Ho ratio is significantly lower
141	(18.6). A situation where Y-Ho fractionation leads to lower Y/Ho ratios relative to the UCC is
142	scarcely reported in the literature. While considered a minor process in natural water systems, it
143	was previously reported by Bau et al. in the slightly acidic solutions of low complex-forming
144	capacity of a stream from an abandoned mine (Bau et al., 1995). Interestingly, the Coco River,
145	where this phenomenon is suspected to occur, is connected to the Koniambo mine.
146	In the estuary, we also observed the conventional Y/Ho fractionation ( <b>Figure 6B</b> ) as the Y/Ho
147	ratios were beyond the conservative mixing lines (Figure 5C). However, the positive departure is
148	even more noticeable in the mangrove forest, with Y/Ho reaching up to 80 (black arrow in
149	Figure 5C). This suggests that additional processes took place in the mangrove forest area,
150	further fractionating this pair. A previous study has suggested that the fractionation between Y
<b>1</b> 51	and Ho could be redox driven (Bau et al., 1997). Accordingly, a redox change is expected to
152	occur at the sediment-water interface (Figure 6C) where the upward diffusive flux from
153	mangrove sediments confronts the freshly formed ferromanganese oxyhydroxides. As discussed
154	previously, these reactive surfaces tend to further remove Ho relative to Y. Such preferential
155	removal of Ho increases the V/Ho ratios as observed in Figure 5C

# 3.4. Calculations of REE flux from the mangrove systems

Fluvial fluxes of dissolved constituents in the oceans are essential to balance their marine budget.

The current ocean box models for REEs consider major inputs as river-borne dissolved and

particulate load, atmospheric dust, groundwater discharge, porewater diffusion and dissolution of

reworked sediments (Arsouze et al., 2009; Garcia-Solsona and Jeandel, 2020; Tachikawa et al.,

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Goldstein and Jacobsen have suggested a simple empirical approach to calculate the dissolved

REEs flux  $(R_i^{eff.})$  from the continent (Goldstein and Jacobsen, 1988) as detailed in the following

464 equation.

$$R_i^{eff.} = C_i^R \times F_R \times \phi_i \times CF \quad (Eq. 6)$$

where  $C_i^R$  is the dissolved concentration of an element in the continental endmember (ng/L),  $F_R$  is

the annual river discharge,  $\Phi_i$  is the effective factor, and CF is a correction factor to convert the

final units into kg/year.

Here, we applied this calculation to our preliminary dataset to determine whether the fluxes of

REEs and Y from the tropical mangrove system could be of interest in the global oceanic budget.

Therefore, it is important to note a few assumptions in this simple calculation. First, because of

the conservative mixing of REEs and Y between the mangrove system and the seawater (Figure

**5A**), we can assume a full effective factor (i.e., absence of estuarine removal or  $\Phi_i = 1$ ). Second,

the local Water Agency (DAVAR, Service de l'eau) monitored the hydrologic regime of the local

river systems. According to the most recent report, the monthly discharges of the rivers in the

leeward side (southwestern, e.g., Boghen, Poya, Pouembout-Boutana Rivers) of Grand Terre in

2019 and 2020 average 18.6 m<sup>3</sup>/s in February and 0.6 m<sup>3</sup>/s in May (the month preceding the two

sampling campaigns) (DAVAR Service de l'eau, 2020). The variability of the river discharge that

could differ Coco and Temala, although not individually determined, could be consisted within

the variability of major river systems of the leeward sides. Third, the concentrations of REEs and

Y of the mangrove endmember  $(C_i^R)$  are those of the samples with salinity in the range of 25-34

(the mangrove forest, **Figure 5A**). Finally, the calculated fluxes  $(R_i^{eff.})$  of dissolved REEs and Y

from the mangrove system are reported with standard deviations reflecting the errors propagating

through the calculations and variability of all variables.

Accordingly,  $R_i^{eff.}$  values of Temala and Coco in the dry and rain seasons are reported in **Table S3** while the annual average fluxes are showed in **Table 3**. The fluxes of REEs and Y are significantly higher in the rain season and dry season (**Table S3**), as reported in other major estuarine systems (Pourret and Tuduri, 2017). Overall, the annual averages of REE and Y fluxes from the Coco and Temala range from 0.05 (Lu) to 16.1 (Y) kg per year (**Table 3**).

To extrapolate our data toward a general perspective of the global mangrove system, we first normalized the calculated fluxes to mangrove watershed surface areas (Sw, km<sup>2</sup>) and the fraction of vegetation coverage (fv). Equation 7 allows the calculation of area-normalized fluxes of REEs and Y from the two mangrove forests ( $R_i^{S-MF}$ , kg/yr/km<sup>2</sup>).

$$R_i^{S-MF} = \frac{R_i^{eff.}}{S_W \times f_V} (Eq.7)$$

The watershed of the Voh region has a vegetation coverage of approximately  $43\pm16\%$  (Taureau et al., 2019), thus  $f_V$  =0.43. The surfaces of the Temala and Coco watersheds (Sw) approximates  $163 \text{ km}^2$  and  $42 \text{ km}^2$ , respectively (FalconBridge NC, 2001). The calculated area-normalized fluxes of REEs and Y range from  $1.6\times10^{-3}$  (Lu) to 0.4 (Y) kg/yr/km<sup>2</sup>.

From the  $R_i^{S-MF}$  values, we attempted an extrapolation to the global surface of mangroves (150,000 km²) (Spalding, 2010) and thus calculated the global fluxes of dissolved REEs and Y from this unique ecosystem (**Table 3**). These global fluxes calculated for the mangrove systems could then be compared to the published global river-borne dissolved Nd loads (Arsouze et al., 2009; Garcia-Solsona and Jeandel, 2020; Tachikawa et al., 2003). Currently, the Nd oceanic budget is the most comprehensive as extensive studies reported both Nd concentrations and isotope composition to calibrate its ocean box models. In these models, the river inputs of dissolved Nd range from 260 to 500 tons per year (or Mg per year) (Rousseau et al., 2015; Tachikawa et al., 2003). Relative to these values, we estimate that the mangrove systems supply  $13.3\pm5.6$  tons per year of dissolved Nd, representing 2.6-5% of the total river input of dissolved Nd. Compared to other sources (e.g., dust dissolution of 400 tons of Nd per year, Nd release from suspended particles of  $5,700\pm2,600$  ton per year) (Arsouze et al., 2009; Rousseau et al., 2015; Tachikawa et al., 2003), the mangrove systems appear to be a minor source. However, it is important to note that even though this study documents a significant release of REEs and Y

within the mangrove system for the first time, we assess only two small mangrove systems representing <1% of the global mangrove surface. Further studies are therefore required to better characterize the contribution of the mangrove system to the global marine budget.

# 4. Implications for the role of mangrove on REE cyling and oceanic budgets.

The conventional estuarine REE behaviour often implies a sharp removal of REEs when a

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riverine water mass enters an estuary. However, our data revealed a significantly release of dissolved REEs and Y (7.3 times for Y, 6.4 times for La and 3.3 times for Lu) in the mangrove system. Although other water masses could supply REEs to the water column (e.g., groundwater discharge (Kim and Kim, 2011)), our stable water isotopes data confirm a sole binary mixing between the fresh and seawater endmembers in the mangrove estuaries of New Caledonia. This release of REEs and Y has thus to be associated only with internal recycling or biogeochemical processes within the mangrove forests. This dataset thus strengthens the hypothesis that tropical mangroves act as a source of dissolved REEs and Y to the marine system. This finding first redefines the role of tropical mangroves in the cycling of trace elements at the continent-ocean interface. While mangrove forests are often referred to as a buffer zone to filter trace metals from reaching the open sea (Marchand et al., 2012), this assumption might be valid only for the particulate loading. This specific ecosystem produces large amounts of organic matter that is bioavailable for bacterial degradation (Marchand et al., 2012) and thus boosts early diagenesis. These biogeochemical processes lead to the complex recycling of REE-bearing phases. The reductive dissolution of ferromanganese oxides occurs within the subsurface layers of the sediments and is thus responsible for releasing trace elements (Dang et al., 2015, 2014). However, the redox boundary at the seawater-sediment interface leads to forming a thin layer of newly formed Fe- and Mn-oxyhydroxides with large reactive surface areas that efficiently sorb trace elements and thus minimize their upward diffusive flux (Dang et al., 2015; Rigaud et al., 2013). Although we could not evaluate the relative amounts of REEs being diffused upward from the sediment porewaters relative to REEs retained at the interface, the diel variation in oxygen concentrations associated with hypoxia events in the mangrove (Dubuc et al., 2019) strongly suggests that the fraction of REEs being retained would be minor regarding the low stability of Fe- and Mn-oxyhydroxides in such redox-dynamic conditions (Bau et al., 1995).

Over the past 30 years, several works aimed to understand and revisit REE behaviour during
estuarine mixing and accumulation in sediments (Bayon et al., 2015; Lawrence and Kamber,
2006; Sholkovitz and Szymczak, 2000; Sholkovitz, 1995). This first dataset reporting REEs
release in a tropical mangrove may provide an important insight into the contribution of
mangrove forests to the oceanic budgets and mass balance of REEs and Y. However, these box
models remain unbalanced with a significant missing flux (e.g., 800 Mg per year) (Tachikawa et
al., 2003). Pourret and Tuduri (2017) recently suggested the continental shelves as a potential
resource of REEs to be included in the oceanic mass balance of REEs. Moreover, mangrove is
growing in 123 tropical and subtropical countries and covers a total surface of 150,000 km <sup>2</sup>
(Spalding, 2010). Our simple flux calculation suggests that this atypical system supplies a
significant proportion of REEs toward the oceans (2.6-5% of the global dissolved loading).
However, it remains uncertain about the representativity of the Coco and Temala systems of the
global mangroves. Therefore, further research on the REE cycling within mangrove systems is
needed to determine whether the mangrove estuaries should be considered a significant input in
ocean mass budget to balance the global REE distribution.
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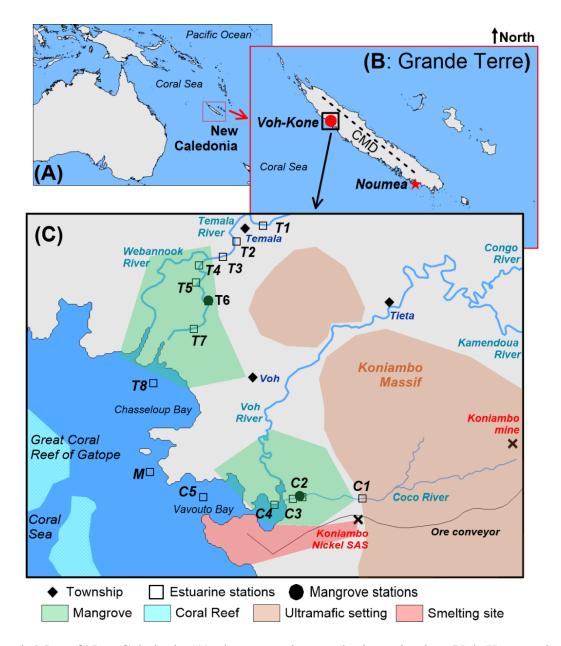
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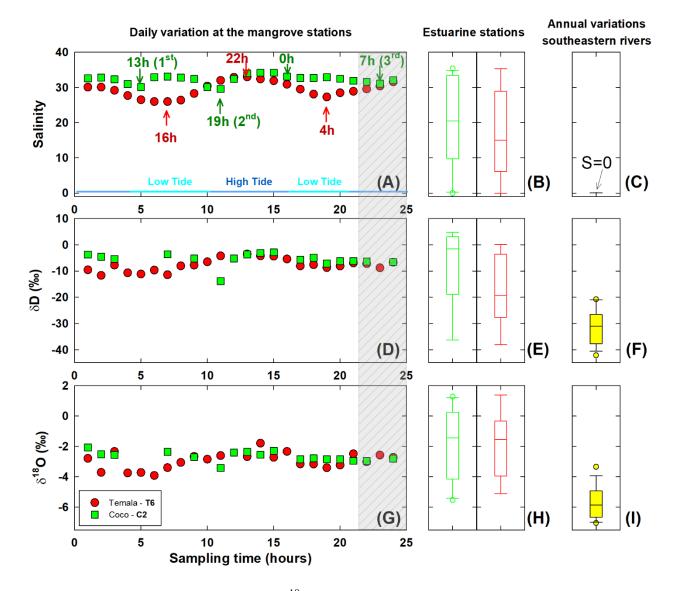
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**Figure 1:** Map of New Caledonia (A), the two regions under investigation: Voh-Kone region (red circle) and four rivers in the Southeast of Grande Terre (red star), around Noumea (B), and a zoom into the Voh-Kone region with sampling locations (C). The position of the CMD (Central Massive Divide) in B is approximative. The southwestern basins of the CMD are considered leeward while the northeastern are windward. See text for more details on the study sites.



**Figure 2**: Variations in salinity,  $\delta D$  and  $\delta^{18}O$  in waters collected from the Temala (red symbols) and Coco (green symbols) Rivers of the Voh-Kone regions and four rivers of the Southeast (yellow boxes). Left panels (A, D, G) show the daily variations recorded within 24 hours in the mangrove forest (stations T6 and C2). The arabic numbers (1<sup>st</sup>, 2<sup>nd</sup>, 3<sup>rd</sup>) indicates the three events where freshwaters were discharged from the Coco river toward the lagoon. The greyed zone shows the last samples collected of the 24-hour series during high tide where all physical-chemical properties and stable water isotopes of the two rivers overlaps. Those data are excluded from the  $\delta D$ -salinity regresssion of Temala River as justified in text. Middle box plots (B, E, H) summarize the spatial variations of Temala and Coco estuaries. The right box plots (C, F, I) show the annual variations of the four rivers in the Southeast of New Caledonia. For all box plots, the horinzonal lines in boxes show median values while box boundaries indicate the 25<sup>th</sup> and 75<sup>th</sup> percentiles. The whiskers demark the 10<sup>th</sup> and 90<sup>th</sup> percentiles and scatters are values of the 5<sup>th</sup> and 95<sup>th</sup> percentile range.

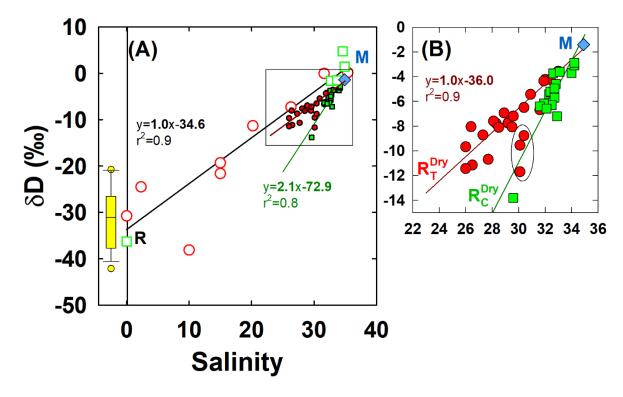
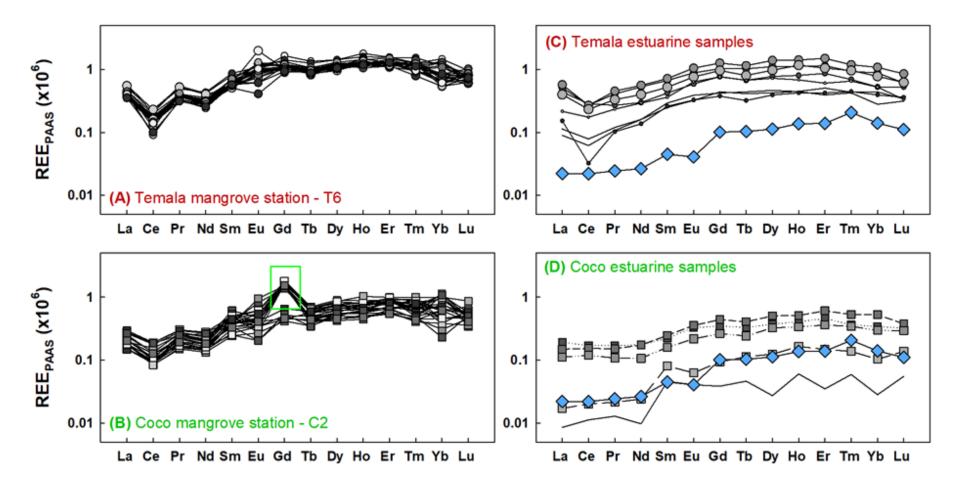
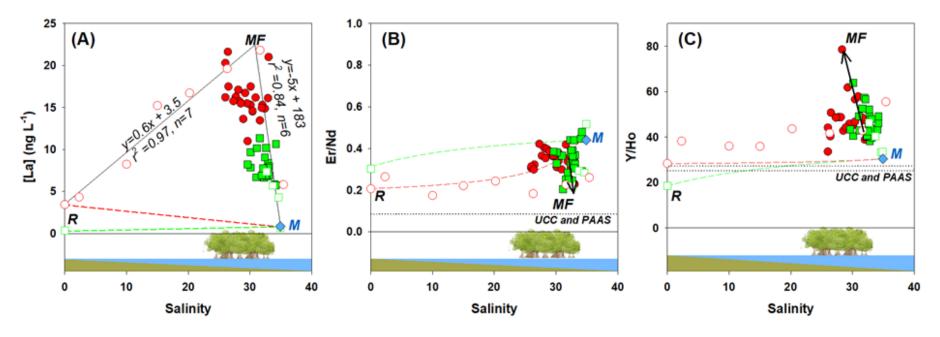


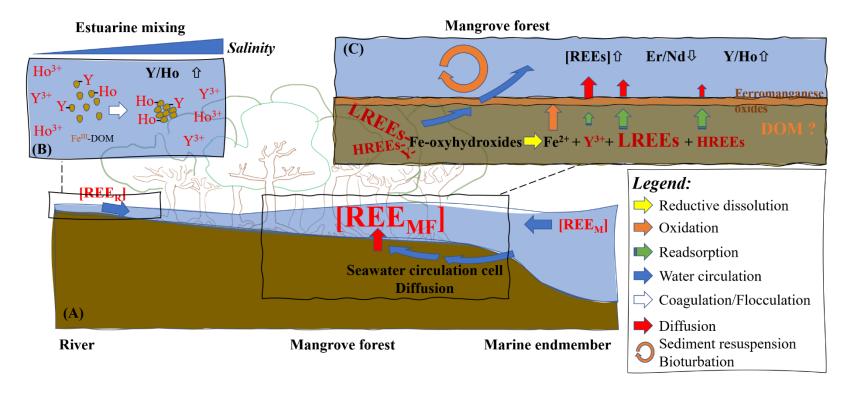
Figure 3: (A)  $\delta$ D-Salinity biplots of all waters collected from the Coco (green symbols), Temala (red symbols) and the box plot of four rivers in the Southeast (yellow, salinity = 0, see legend of Figure 2 for more details). Samples collected in the mangrove forests (full red and green symbols), across the estuaries (open red and green symbols) and the marine endmember (blue diamond) are also shown. A zoom in the Coco and Temala system during dry season is provided in panel (B). Their respective regression lines are shown in red (Temala) and green (green) while the black regression line represents all samples from Temala and Coco estuaries. The regression for Temala River excludes the data (encircled in B) that are observed on the Coco River regression lines. See text for justification.



**Figure 4:** Rare earth element patterns in the waters of the mangrove stations (T6 and C2) at Temala (A) and Coco (B), and across the estuarine mixing of Temala (C) and Coco (D). The blue diamonds show the REE patterns in the marine endmember. The gray intensity of the symbols in panels A and B is associated with the 24-h time-series (black symbols corresponding to the first sampling). The symbol size in panels C and D reflect the salinity (small symbols reflecting freshwater). The green rectangle in B demarks Gd anomalies observed in Coco River water.



**Figure 5:** Variations of La concentrations (A), Er/Nd (B) and Y/Ho (C) ratios as the function of salinity at Temala (red symbols) and Coco (green symbols). Samples in the mangrove forests are shown by full symbols while estuarine samples are presented as open symbols. The blue diamond symbols represent the marine endmember. The horizontal dotted lines show the values in the Upper Continental Crust (UCC) and PAAS shale. The dotted green and red lines are conservative mixing line for Coco and Temala estuaries, respectively.



**Figure 6:** Schematic represention of the REE cycle in tropical mangrove-dominated estuaries (A). Panels (B) and (C) provide a zoom into the mechanisms occurring across the estuarine mixing and within the mangrove forest, respectively. In (B), the flocculation of either dissolved organic Fe<sup>III</sup> complexes (Fe<sup>III</sup>-DOM) or colloidal particles is presented.

**Table 1:** Variations in the physical-chemical properties of water collected over a 24-hour cycle in the dry season in the Temala (T6) and Coco (C2) mangrove forest.

Parameters	Temala (n=24)			Coco (n=24)		
rarameters	Range	Average	SD	Range	Average	SD
Temperature	16.8-17.7	17.3	0.2	15.5-16.6	16.1	0.3
pН	7.7-7.9	7.8	0.1	7.6-8.0	7.7	0.1
Salinity	26.0-33.0	29.5	2.1	29.6-34.2	32.3	1.2

**Table 2:** Summary of the  $\delta D$ -S and  $\delta^{18}O$ -S relationships in the Coco and Temala rivers during rain and dry seasons. Values in brackets show the  $r^2$  values. The stable water isotope compositions of the Coco and Temala endmembers ( $\delta D_i$  and  $\delta^{18}O_i$ ) were calculated by the extension of the  $\delta D$ -S and  $\delta^{18}O$ -S relationships. For the Southeastern rivers and the marine endmembers, the  $\delta D_i$  and  $\delta^{18}O_i$  values were calculated as the average of all river samples with salinity = 0 and the values recorded at station M, respectively. For the See text for more details.

	Endmembers	δD-S relationship	δ <sup>18</sup> O-S relationship	δD <sub>i</sub> (‰)	δ <sup>18</sup> O <sub>i</sub> (‰)	d-excess (‰)
Mangrove stations – Dry	Coco (n=17)	y=2.1x-72.9 (0.8)	y=0.2x-9.9 (0.6)	-72.9±8.9	-9.9±1.7	15.8±1.3
season	Temala (n=19)	y=1.0x-36.0 (0.9)	y=0.2x-8.4 (0.6)	-36.0±2.8	-8.4±1.2	15.9±2.9
Estuarine stations – Rain	Temala and Coco (n=15)	y=1.0x-34.6 (0.9)	y = 0.2x - 5.0 (0.9)	-34.6±1.9	-5.0±0.4	0.4±5.8
season	Marine (n=1)			-1.6±0.2	-2.1±0.2	
Freshwater stations	Southeastern rivers (n=14)			-31.2±6.2	-5.8±1.0	

**Table 3:** The fluxes  $(R_i^{eff.}, \text{kg/yr})$  of REEs and Y from the two mangrove systems in New Caledonia (NC) and the extrapolation toward the global flux (ton, or Mg per year).

	Annual f	Global flux		
	Temala			
Y	16.1±4.6	6.1±1.3	42.1±21.2	
La	$6.2 \pm 1.2$	$2\pm0.6$	$14.8 \pm 7.6$	
Ce	$6.4 \pm 1.7$	$3.9 \pm 1.3$	$22.8 \pm 14$	
Pr	$1.1\pm0.3$	$0.4\pm0.1$	$3.0 \pm 1.5$	
Nd	$5.5 \pm 1.1$	$1.8 \pm 0.4$	$13.3 \pm 5.6$	
Sm	$1.2 \pm 0.2$	$0.4\pm0.1$	$2.9 \pm 1.4$	
Eu	$0.3\pm0.1$	$0.1\pm0$	$0.8\pm0.7$	
Gd	$1.6 \pm 0.4$	$0.6\pm0.3$	$4.2 \pm 3.3$	
Tb	$0.3\pm0$	$0.1\pm0.03$	$0.6\pm0.3$	
Dy	$1.8 \pm 0.4$	$0.6\pm0.2$	$4.5\pm2.7$	
Ho	$0.4\pm0.1$	$0.1\pm0$	$1.0\pm0.6$	
Er	$1.1\pm0.3$	$0.5\pm0.2$	$3.1 \pm 1.8$	
Tm	$0.1\pm0.03$	$0.1\pm0.02$	$0.4\pm0.3$	
Yb	$0.8\pm0.3$	$0.4\pm0.2$	$2.4\pm2.5$	
Lu	$0.1\pm0.03$	$0.05\pm0.01$	$0.3\pm0.2$	