

Indoor use of essential oil-based cleaning products: Emission rate and indoor air quality impact assessment based on a realistic application methodology

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- Indoor use of essential oil-based cleaning products: Emission rate
- 2 and indoor air quality impact assessment based on a realistic
- 3 application methodology

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ABSTRACT

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Cleaning is performed to increase hygiene, esthetics and material preservation. Despite its benefits, cleaning also poses risks, potentially contributing to nearly 20 % of indoor pollution. As indoor air quality has become a major human health concern, "natural-scented" cleaning products, formulated with essential oils, have become market leaders among household products. However, these consumer products have benefitted from skillful marketing strategies based on the ambiguity of the words "green" and "natural". The characterization of the emission processes studied through 1-m³ chamber experiments under a realistic scenario suggests variable maximum total terpene concentrations from 150 to 300 ppb. The estimated emission rate profiles confirm that the liquid-to-gas transfer of terpenes is driven by (i) the formulation of the product matrix inducing specific chemical affinities, (ii) the liquid content of individual terpenes, and (iii) the intrinsic volatility of terpenes. A unique formaldehyde emission kinetics profile was observed, suggesting the presence of a unique emission source: formaldehyde-releasers. Consequently, the use of essential-oil-based cleaning products might generate a long-term increase in the indoor formaldehyde concentration, and the maximum levels might be sustained for several hours after cleaning. Thus, essential-oil-based cleaners should be seriously considered as versatile and significant sources of fragrance molecules and formaldehyde.

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KEYWORDS: cleaning products; emission rates; indoor air quality; terpenes

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- 43 **ABBREVIATIONS: TerVOCs** Terpenic volatile organic compounds; **AER** Air exchange rate;
- 44 OVOCs Oxygenated volatile organic compounds; VP Vapor pressure; PROBAS Danish
- 45 Product Register Database; **BNPC** National Database for Products and Compositions

1 INTRODUCTION

Major efforts have been focused on addressing indoor air problems since daily activities mostly occur in confined environments. Indoors, people are exposed to a wide variety of chemical, biological and physical pollutants that (i) infiltrate from outdoor air, (ii) are released from building materials and furnishing, and (iii) are generated by occupants and their indoor activities (Missia et al., 2012; Weschler, 2001). The scientific community has focused on chemical pollutants, principally volatile organic compounds (VOCs), defined as organic molecules with boiling points ranging from 50 to 100 °C on the low end and from 240 to 260 °C on the high end (Brown et al., 1994; World Health Organisation, 2010). Among indoor sources of pollutants, the main origins of chemical contaminants are linked to occupant activities, such as interior renovation or decoration, smoking, cooking, or housekeeping (Kirchner et al., 2001).

Among the thousands of cleaning products available and marketed, people are tempted to use scented products because a pleasant odor is associated with a cleaner environment. Therefore, despite all the benefits of cleaning activities, such as increased hygiene, esthetics, and material preservation, the chemical and physical pollutant exposure risks posed by such activities remain unclear. Previous studies (Nazaroff and Weschler, 2004; Wolkoff et al., 1998) have shown that atmospheric VOC emissions per capita from general purpose cleaners and air fresheners contribute the most to indoor VOC pollution, with values ranging from 200 to 230 mg per day. The intake fraction is defined as the pollutant mass inhaled per unit of pollutant mass emitted (Lai et al., 2000). A typical intake fraction from indoor emissions is on the order of 10⁻³ and 10⁻¹, while that from outdoor emissions is on the order of 10⁻⁶ and 10⁻³. These data highlight the fact that the proportion of emissions inhaled is higher when emissions occur indoors rather than outdoors (Nazaroff and Weschler, 2004).

Unexpectedly, the largest indoor sources of terpene VOCs are not usually the recognized, regulated and controlled sources. Indeed, the composition of VOCs emitted from household products has been associated with health hazards to indoor occupants (Dimitroulopoulou et al., 2015; Nazaroff and Weschler, 2004; Steinemann, 2009, 2016; Trantallidi et al., 2015; Wolkoff et al., 1998). Nevertheless, limited information is available about the composition, quantification and emission dynamics associated with housekeeping activities in confined environments. Available scientific data are, in most cases, incomparable due to wide variations and heterogeneity in the experimental methodologies used for the evaluation of primary and secondary pollutant emissions from fragranced household products (Angulo et al., 2020). These variations are related to two key parameters: (i) the application protocol (quantity, mode of operation) and (ii) the test environment (air exchange rate (AER), temperature, chamber size, and relative humidity (RH)) (BEUC, 2005; Chesnais and Marchais, 2014; Colombo et al., 1991; Delmas et al., 2016; Höllbacher et al., 2017; Nicolas and Chiappini, 2013; Norgaard et al., 2014; Sarwar et al., 2004; Singer et al., 2006a, 2006b; Solal et al., 2014; Uhde and Schulz, 2015). Therefore, comparing the results from one investigation to those of another may be difficult and inaccurate.

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Fragranced household products can be formulated with (i) synthetic fragrances, (ii) pure essential oils and (iii) mixtures of synthetic fragrances and essential oils to reduce production costs. Essential oils are used in "naturally" formulated cleaning products either to provide them with a pleasant odor, to hide odors from other chemicals in the formulation, or to enhance their antibacterial performance (Rastogi, 2002; Steinemann, 2009). Essential oils are extracted from plants by water vapor distillation, dry distillation or mechanical extraction, which does not involve temperature changes (Delmas et al., 2016). They usually contain more than 100 different

chemicals that compose their characteristic smell, and most of these chemicals are terpenes: monoterpenes, sesquiterpenes, terpene alcohols and terpenoids (denoted TerVOCs) (Babu et al., 2002; Lassen et al., 2008; Zuzarte and Salgueiro, 2015). The characteristic smell of an essential oil often originates from its major chemical components; therefore, the typical "scent" of an essential oil directly depends on its chemical composition. In contrast, fragrance mixtures combine differently sourced compounds (synthetic, natural-like or extracted) to replicate a "natural" scent (Teixeira et al., 2013; Zuzarte and Salgueiro, 2015).

Although diverse sanitation benefits of essential oil-based cleaning products have been recognized, several risks associated with their use in indoor environments are still questioned. Is there a potential exposure risk to consumers posed by TerVOCs emitted from household products?

This study is developed within the framework of the ESSENTIEL research project, aiming to characterize the impact of essential oil-based products on indoor air quality (IAQ) through investigation of their realistic emissions and indoor fate. The objectives of this investigation are to evaluate the correlation between the fragrance chemicals contained in cleaning products and their volatile fraction through determination of the gas-transferred concentration by micro-chamber testing and to characterize and assess their emissions in a 1 m³ experimental chamber reflecting a realistic application scenario. The proposed experimental approach and the results provided in this study are required (i) to propose harmonized and realistic test protocols and (ii) to define risk scenarios and evaluate the human exposure to TerVOCs associated with the use of essential oil-based cleaning products.

2 MATERIAL AND METHODS

2.1 Cleaning product benchmarking

To frame the approach of the present study, it is necessary to define the type of cleaning products considered. Essential oil-based cleaning products are defined in this work as naturally scented and certified organic products used in a nonpermanent way in indoor environments for housekeeping purposes. For their selection, a benchmark analysis was performed among cleaners certified as *ecological products* by different European labels. A total of 108 essential oil-based products were identified from the European market. Among them, 6 representative cleaning products were selected in the framework of this work and classified by category, i.e., intended use: (i) 2 kitchen degreasers, (ii) 2 general (multiuse) cleaners, (iii) 1 surface cleaner, and (iv) 1 glass cleaner. This selection considered (i) diverse intended uses to assess the effect of different solvent formulations, (ii) a variety of application modes and (iii) a diversity of essential oils used as natural fragrances in the formulation of the cleaning products. The selected cleaning products were purchased from environmentally friendly and ecologically conscious retail stores in France.

Importantly, the solvent matrix composition could vary from one product to another in terms of the organic content (% ethanol) or the presence of texturing agents (gums). The contents of the fragrance chemicals and their volatile fractions were characterized for the six selected

cleaning products, whereas only three products were considered for the 1 m³ emission test. Table

1 details the characteristics of the six selected products.

Table 1. Specifications of the selected cleaning products.

					C	haracteriz	zation	
						perform	ed	
Product	Product category	Diffusion mode	Natural fragrance ¹	General product formulation ²	Liquid composition	Volatile fraction (micro- chamber)	Realistic emissions (1 m³ test chamber)	
KC - 1	Kitchen	Trigger spray	Trigger spray	Lemon oil	< 5 % surfactant (anionic and nonanionic), citrus oil, citric acid, water, methylisothiazolone, 5-chloro-2-methyl-4-isothiazolin-3-one, nitrilotriacetic acid trisodium salt monohydrate	х	х	
KC – 2	cleaner		Eucalyptus oil	< 5 % surfactant (nonanionic), essential oils (including eucalyptus oil²), water, alcohol², preservatives, acid correctors	X	x	X	
MC – 1	Multiuse	Liquid	Citrus oil	< 5 % surfactant (anionic and nonanionic), water, alcohol, citrus oil², citric acid	X	X	X	
MC – 2	cleaner		Lemon oil	< 5 % surfactant (anionic and nonanionic), water, alcohol, lemon oil ² , lactic acid, fatty acids, sodium citrate, glyceryl caprylate	X	X		
SC - 1	Surface cleaner	Trigger spray	Lavender oil	70 % ethanol ² , $<$ 1 % lavender oil ² , $<$ 1 % sage oil ² , water	X	X	X	
GC - 1	Glass cleaner	Trigger spray	Eucalyptus oil	> 30 % water, 5 – 15 % alcohol ² , < 5 % surfactant (anionic and nonanionic), citric acid, essential oils ² (including eucalyptus oil), amphoteric surfactants	х	X		

^{138 1:} Certified organically grown

^{2:} Based on information given by the manufacturer

2.2 Characterization of the liquid composition

Several extraction parameters were optimized to maximize the extraction of terpenes: (i) the organic extraction solvent, (ii) the mass ratio of the cleaning product to the solvent, (iii) the contact time between the solvent and the cleaning product, and (iv) the number of consecutive extractions. The extraction procedure and the analytical methods and parameters are detailed in a supplementary information sile (SI file).

2.3 Determination of the gas-transferred concentration by micro-chamber testing

2.3.1 Micro-chamber description

The liquid-to-gas phase transfer capacity of each terpene identified in the cleaning products is defined as the gas-transferred concentration and was determined by using a stainless steel cylindrical micro-chamber with a 44 mL volume (M-CTE-120 Markes International®). The device was equipped with restrictors allowing the selection of a low-flow range and a high-flow range of 10-70 mL/min and 50-500 mL/min, respectively. In this work, the low-flow range was used and set at 50 mL/min. The dry air generation is ensured by an air compressor combined with membrane for humidity removal. Before entering the micro chamber, the dry air is purified by an activated carbon filter in order to remove particles and trace pollutants. The top of the chamber acts as a door and is sealed with a silicon gasket for leak control. After optimization, the temperature and air flow were set at 40 °C and 50 mL/min, respectively. Dry air flow enters each micro-chamber through the lid and is preheated to the micro-chamber temperature of 40 °C. A total volume of 50 ± 1 μ L of each selected fragranced cleaner was introduced into the micro-chamber. Blank measurements were performed to determine the background levels of VOCs in the chamber outlet before running each set of experiments. Note that the mean concentration of

blank measurements in the micro-chamber did not exceed the quantification limits for the background concentrations of the individual target VOCs. The detailed characteristics of the micro-chamber are presented in Fig. 1a.

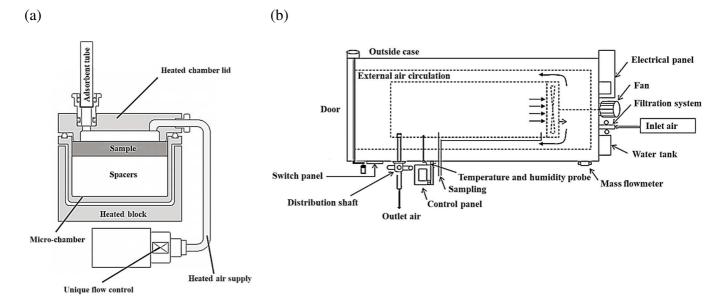


Fig. 1. (a) Schematic of the M-CTE-120 micro-chamber from Markes International®. (b) Schematic of the 1 m³ emission chamber VCE1000 Classic from Vötsch.

2.3.2 Analytical strategy

The TerVOCs emitted from the investigated products inside each micro-chamber were monitored using an off-line device for their identification and quantification. Subsequently, two consecutive Tenax TA® cartridges collected volatilized compounds at the device outlet for 15 minutes. The total sampled volume was 0.6 L. Afterward, these samples were desorbed at 280 °C under a helium flow of 50 mL/min using a PerkinElmer® thermal desorber (model: Turbo Matrix 350). Then, the desorbed compounds were thermally transferred via the helium flow and refocused on a Carbotrap B/C trap at 5 °C with an outlet split flow of 70 mL/min. The gas

chromatographic analyses were performed using a 7890A instrument from Agilent Technologies equipped with a Restek 10189 chromatographic column (60 m x 320 μ m, 1 μ m film thickness) connected to two different detectors: (i) an FID and (ii) an MS (5975B inert MSD) from Agilent Technologies. The typical detection limit of this instrument for hydrocarbon VOCs with the abovementioned sampling conditions is 0.5 μ g/m³. Calibration for the identified and quantified TerVOCs was achieved by individual liquid doping of the high-purity certified standards from Sigma-Aldrich® on Tenax TA® absorbent tubes.

2.4 Emission chamber test: simulation of a realistic scenario - correlation method for a

realistically sized room

2.4.1 Emission chamber parameters

The experiments reported in this paper were conducted in an experimental chamber with a 1 m³ volume (Vötsch VCE1000 Classic) that complied with the ISO 16000-9 (2006) standards. The chamber was operated at a controlled temperature, humidity and air exchange rate (AER) of 23 ± 1 °C, 50 ± 5 % and 0.3 h⁻¹ ± 0.05, respectively. The humidity and temperature were set according to ISO 16000-9. The established AER corresponds to the 25th percentile reported in French dwellings by the Indoor Air Quality Observatory (OQAI) (Kirchner et al., 2001; Lucas et al., 2009). Detailed characteristics of the experimental chamber are presented in Fig. 1b. Before loading the test chamber with the selected product, blank measurements were performed to determine background levels of TerVOCs and oxygenated volatile organic compounds (OVOCs). Note that the mean concentration determined from blank measurements in the

experimental chamber did not exceed the standardized background concentrations of individual VOCs.

2.4.2 Definition of a realistic scenario for the use of essential oil-based household products

The assessment of TerVOC emissions in the experimental test chamber has been primarily focused on surface cleaning scenarios. The product application procedure must be based on the typical consumer use pattern to mimic realistic application in the evaluation of cleaning product emissions. The key parameters required to define a specific and standardized product application procedure are (i) the quantity of product to apply and (ii) the surface to clean. These parameters were estimated from the loading factor and the final application yield of the cleaning product after wiping, called product yield is this study.

Loading factor

The loading factor is defined as the ratio between the emissive surface area, i.e., the cleaned area, and the chamber volume. This is the key parameter to transfer the real application scenario at scale 1:1 to the 1m3 test chamber. It has to be keot constant between both chamber volume as reported in Table 2. Conventionally, a reference room is used to extrapolate measured VOC concentrations in test chambers with small volumes when assessing human exposure. The reference room considered for the study is the IRINA experimental room characterized by a floor area of 12 m², a ceiling height of 2.5 m and a volume of 40 m³. (Harb et al., 2016) In regard to the product application for surface cleaning, a typical table area of 2 m² was considered in the reference room. It results in a loading factor of 0.05 m²/m³. For the correlation between the 1 m³ emission test chamber and the 40 m³ reference room, the loading factor must be identical (Table

2). Based on this consideration, the *surface cleaning scenario* was defined and the surface to clean was determined for the 1 m³ emission test chamber as reported in Table 2, along with the corresponding masses of cleaning products. Table 2.

Table 2. Application parameters for the surface cleaning scenario: reference values from the 40 m3 room for the product yield and loading factor and subsequent quantities of products to be applied in both chambers 40 m³ and 1m³.

Application par	cameters to mimic a 2 m² si	urface cleaning in a 40 m³ room to th	ne 1m³ test chamber
		(40 m ³)	$1 m^3$
		REFERENCE ROOM	TEST CHAMBER
SURFACE TO CLEAN		2 m ²	$0.05 m^2$
LOADING FACTOR		0.05	0.05
(m^2/m^3)		0.05	0.05
	SC - 1	8.7 (g.r	n ⁻²)
PRODUCT YIELD	MC -1	7.2 (g.r	n ⁻²)
	KC - 2	12.0 (g.	m ⁻²)
	SC - 1	18.0 g	3.2 g
MASS APPLIED	MC -1	15.0 g	4.4 g
	KC - 2	24.0 g	2.7 g

Product yield

The product yield is defined as the effective mass of product remaining on a 1 m² surface; it includes the total mass applied balanced by the losses induced by wiping. This parameter was estimated for each selected cleaning product. First, the average amount of cleaning product required to clean 1 m² of surface was determined from the cleaning of a 1 m² surface by 10 different operators. Subsequently, the average mass of each product dispersed by a single spray from each trigger spray mechanism was determined for a set of 15 successive sprays performed

by the same operator. Then the mass of product lost on wipes is determined by weighing. This procedure allows the estimation of the effective mass of product remaining on a 1 m² surface. The mass of product required for the *surface cleaning scenario* in the 1 m³ emission test chamber is determined from this product yield by proportionality.

For each experiment, the calculated mass of product was evenly drizzled or sprayed on a 0.05 m^2 stainless steel surface placed at the center of the 1 m³ experimental chamber. Once the simulation of the cleaning activity was completed, the experimental chamber remained closed during the emission monitoring. For each cleaning product, a set of three experiments was performed to evaluate the repeatability of the experimental protocol. Additionally, during the experiment, t = 0 corresponded to the time when the application of the cleaning product was completed inside the chamber.

2.4.3 Sampling and analytical parameters

Regarding the analytical methodology, the temporal dynamics of the TerVOC and OVOC concentrations inside the experimental chamber were monitored using off-line devices that allowed their quantification with high temporal resolution. Several preliminary tests were performed to optimize the experiment duration depending on the cleaning product. On the basis of these preliminary results, the duration of a typical emission experiment was 5 hours.

Off-line sampling was performed according to ISO 16000-3 and 16000-6 by using (i) Tenax TA® cartridges for terpene compounds and (ii) 2,4-dinitrophenylhydrazine (DNPH)-impregnated silica cartridges (Sep-Pak DNPH-Silica Plus Short Cartridges, Waters) for carbonyl compounds. Sampling was performed using a pump (KNF®) connected to a mass flow controller with specific flow rates and durations optimized for each cleaning product tested, as described in Table 3.

Table 3. Experimental protocol and optimized parameters for the off-line sampling systems.

Evaluation of emissions from cleaning products						
Sampling device Terpenes (TerVOCs): Tenax TA® adso Carbonyl Compounds (OVOCs): DNPH c						
Dur	ration of experiment* (h)	5 h				
Sampling	TerVOCs	0.150 L/min				
flow*	Carbonyl compounds	0.800 L/min				
Sampling dur	ration (min) and time interval (h)*	From 0 to 1 hour; continuous sampling for 15 minutes; total of 4 samples From 1 – 5 hour; continuous sampling for 60 minutes; total of 4 samples				

^{*}Experimental parameters that were optimized

The analytical protocol to quantify TerVOCs was the same as that reported in section 2.3 for the gas-transferred concentration determination. In contrast, derivatized carbonyl compounds (OVOCs) were extracted from DNPH cartridges with 3 mL of acetonitrile and analyzed with a Thermo Dionex Ultimate U3000 liquid chromatograph. Compounds were eluted on an Acclaim RSLC Carbonyl column (2.1 x 150 mm) and detected by RS variable-wavelength UV absorption. The acquisition was performed at 365 nm wavelength. Calibration for the carbonyl compounds was achieved using high-purity certified standard solutions (Aldehyde/Ketone-DNPH Stock Standard-13). Under these conditions, the detection limit for the carbonyl compounds was 0.01 ppb.

3 RESULTS AND DISCUSSION

3.1 Chemical composition: terpene (TerVOC) content of natural, certified organic cleaning

278 products

The terpene contents of the six selected cleaning products are listed in Table 4. Note that of the identified compounds, the predominant terpenes are written in bold. The most frequently identified terpenes were limonene, linalool, and α -terpineol, which were present in all the evaluated cleaning products. To date, five studies have assessed the presence of fragrance chemicals in cleaning product formulations on the European market. Among the 121 cleaning products evaluated, limonene and linalool have been reported with the highest occurrence rates: 84.2 % and 60.8 %, respectively (Nicolas and Chiappini, 2013; Pors and Fuhlendorff, 2003; Rastogi, 2002; Rastogi et al., 2001; Uhde and Schulz, 2015). Worldwide, the most commonly used essential oils in household product formulations are citrus oil, lavender oil, pine oil, eucalyptus oil, tea tree oil and rose oil. In contrast, several terpene derivatives, such as camphor and bergamol, detected in SC-1 and KC-2, have been more rarely reported (Ezendam et al., 2009; Huang et al., 2011; Jo et al., 2008; Kwon et al., 2007).

Table 4. Liquid composition (expressed in μg of terpene per g of product: $\mu g/g$) of the six selected cleaning products and their occurrence rate in the literature. NB: the mass concentration of the predominant TerVOCs in each cleaning product is written in bold.

	Occurrence rate (%)						
	in the literature						
Identified Terpenes	MC -1	MC -2	SC - 1	KC - 1	KC – 2	GC - 1	[121 cleaning products] (Nicolas and Chiappini, 2013; Rastogi, 2002;
Diffusion mode	Liquid	Liquid	Spray	Spray	Spray	Spray	Rastogi et al., 2001; Uhde and Schulz, 2015)
α-Pinene	24.2 ± 6.2	23.3 ± 4.8	37.6 ± 5.0	-	35.6 ± 3.8	-	36 %
Limonene	852.9 ± 32.9	472.0 ± 8.2	25.5 ± 0.7	6.3 ± 0.5	173.3 ± 36.1	2.2 ± 0.3	84 %
Linalool	10.2 ± 0.6	6.0 ± 0.9	390.0 ± 5.0	6.5 ± 0.5	4.3 ± 0.7	1.5 ± 0.3	61 %

Eucalyptol	424.1 ± 37.0	-	213.4 ± 4.7	-	2230.8 ± 71.4	18.0 ± 1.5	38 %
Cymene	158.7 ± 23.9	8.5 ± 0.2	-	-	76.2 ± 11.5	-	26 %
Bergamol	-	-	279.0 ± 2.0	-	-	-	NS
Camphor	-	-	256.8 ± 2.5	-	19.3 ± 2.5	-	NS
β-Pinene	69.9 ± 3.6	146.2 ± 5.0	21.8 ± 0.4	3.4 ± 0.4	3.1 ± 1.4	-	40 %
γ-Terpinene	15.9 ± 2.6	61.4 ± 5.7	2.6 ± 0.2	-	7.5 ± 0.5	-	20 %
α-Terpineol	29.7 ± 7.4	2.9 ± 0.8	16.4 ± 0.5	10.0 ± 0.3	18.2 ± 0.8	12.8 ± 2.0	NS
Menthol	-	-	-	1.0 ± 0.3	570.9 ± 25.2	3.3 ± 0.7	NS
Other TerVOCs	121.5 ± 15.8	79.1 ± 7.4	247.9 + 6.9	10.3 ± 0.6	77.2 ± 18.5	1.30 ± 0.3	-
TOTAL TerVOCs	1682.8± 131.3	776.0 ± 28.1	1453.7 ± 22.6	37.6 ± 2.5	3180.7 ± 200.5	39.1± 4.5	

3.1.1 Total TerVOC mass concentration

Regarding the total terpene content in household products, the mass concentrations varied widely, ranging from 37.6 ± 2.5 to 3180.7 ± 200.5 µg/g (of product). Nevertheless, these concentrations accounted for only 0.01 % to 0.17 % w/w of the total composition of the selected products, as shown in Table 4. Sarwar et al. (2004) reported the quantification of TerVOCs in four different types of cleaning products used as floor cleaners and general purpose cleaners. The main detected terpenes were α -pinene, β -pinene, 3-carene, limonene and α -terpinene, with individual mass concentrations ranging from 0.2 % w/w to 2.0 % w/w. The proportions of fragrance chemicals in industrial formulations of cleaning products do not exceed 5.0 % w/w (Ezendam et al., 2009; Rastogi, 2002; Steinemann, 2009; Steinemann et al., 2011).

3.1.2 Individual mass concentration of TerVOCs

Among the selected cleaning products, a total of 27 terpenes were detected: (i) 14 monoterpenes, (ii) 8 terpene alcohols, (iii) 4 terpene derivatives, and (iv) 1 sesquiterpene. Per product, the diversity of odorous chemicals varied from 6 to 20 different species. Furthermore, all tested products, except for KC – 1, showed a correlation between the predominant terpene detected and the essential oil indicated by the manufacturer. KC – 1 was reportedly formulated with lemon oil; however, the major terpene was α -terpineol instead of limonene. In terms of the product categories, limonene was the predominant terpene contained in the multiuse cleaners, both fragranced with citrus oils. In the kitchen cleaners tested, the main detected terpene differed: eucalyptol in KC – 2 and α -terpineol in KC – 1. Concerning the mass concentration of terpenes, the predominant species in the selected products were limonene, linalool, eucalyptol, and α -terpineol, detected at concentrations ranging from $10.0 \pm 0.3 \,\mu\text{g/g}$ to $2230.8 \pm 71.4 \,\mu\text{g/g}$.

The chemical composition of a cleaning product might vary widely depending on its usage purpose and diffusion mode. For instance, when a cleaning product emphasizes a disinfecting property, complementary chemicals are added to its formulation to suppress or inhibit microbial activity. In so-called "green" cleaning product formulations, specific essential oils might be added to increase the concentration of terpenes with antibacterial and antiseptic activity, such as eucalyptol and geraniol. Therefore, no correlation was evidenced between the diffusion mode of the selected cleaning products and their total terpene mass concentration. This finding is suggested to result from the intrinsic industrial formulations designed to achieve a particular performance.

Finally, the contrasting terpene contents bring up questions about the intrinsic emission potential of the cleaning products. The diversity in the constituent volatility affects the emission dynamics associated with the cleaning activity. *Does a high terpene content necessarily imply high emission levels? What is the effective amount of terpenes eventually transferred to the gas phase? What is the impact of the cleaning product matrix on the gas phase transfer? How long is the user impacted by the emissions?*

To provide key elements needed to answer these questions, a screening of their ability to be transferred from the liquid to the gas phase, defined in this paper as the gas-transferred concentration, was performed to identify the emission potential of each cleaning product.

3.2 Gas-transferred concentration vs. liquid composition

3.2.1 Total gas-transferred concentration

The total gas-transferred concentration of terpenes corresponds to the sum of the concentrations, subsequently expressed in $\mu g/m^3$, of terpenes collected at the outlet of the micro-chamber setup. This concentration can be used to assess the ability of a given cleaning product to release terpenes to the gas phase. Comparison of the six selected products revealed very different total gas-transferred concentrations, ranging from 125 ± 15 to $85\ 225 \pm 5\ 500\ \mu g/m^3$. The glass cleaner GC – 1 was the least emissive product, with a total gas-transferred concentration of $125 \pm 15\ \mu g/m^3$, while the gas-transferred concentrations of the surface cleaner SC – 1 and the kitchen cleaner KC – 2 were higher by 2 orders of magnitude. These two products transferred analogous total terpene concentrations to the gas phase, considering the standard deviation: $85\ 000 \pm 5\ 500\ \mu g/m^3$ and $81\ 000 \pm 12\ 000\ \mu g/m^3$, respectively.

Fig. 2 shows the correlation between the total terpene mass concentrations in the liquid phase of the selected products (μ g/g of product) and the total gas-transferred concentrations of TerVOCs (μ g/m³) for the same products. For the multiuse cleaners (MC – 1 and MC – 2), the volatile concentrations reached equivalent levels of 27 000 μ g/m³, but their total terpene mass concentrations in the liquid phase differed by a concentration ratio of 2:1. The pattern between the surface cleaner SC – 1 and the kitchen cleaner KC – 2 was similar, with gas-transferred concentrations that both reached approximately 80 000 μ g/m³, considering the standard deviation, but a terpene content of the surface cleaner SC – 1 that was half that of the kitchen cleaner KC – 2 (3200 ± 270 μ g/g). In contrast, the two household products SC – 1 and MC – 1 had equivalent terpene mass contents but gas-transferred concentrations that differed by a factor of 3. Consequently, no direct correlation was evidenced between the total terpene mass content and the total gas-transferred concentration among the six selected products. Therefore, the amount of terpenes emitted by a given product cannot be estimated from its liquid terpene content, and emission tests are required.

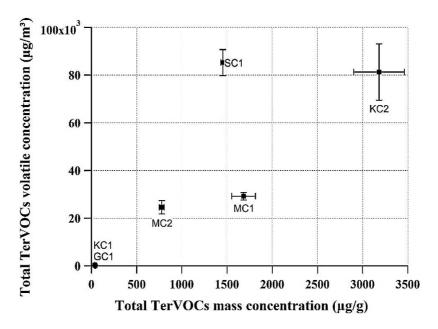


Fig. 2. Total TerVOC gas-transferred concentration from micro-chamber testing ($\mu g/m^3$) as a function of the total terpene mass concentration in the liquid form ($\mu g/g$) for the six selected cleaning products (T = 40 °C, dry air flow = 50 mL/min, micro-chamber volume = 44 mL).

3.2.2 Individual gas-transferred concentrations

Only 19 terpene molecules from the 27 TerVOCs contained in the cleaning products were identified in the gas phase during the micro-chamber evaluation, and the number of emitted terpenes varied from 1 to 14 per cleaning product. Table 5 lists the (i) volatile/liquid fraction concentration ratio (V/L) of each cleaning product and (ii) various physical and chemical properties of the individual terpene molecules. For discussion purposes, in this paper, the V/L concentration ratio is defined as the ratio of an individual gas-transferred concentration to its mass fraction in the liquid phase for each identified terpene. Higher is the V/L value, greater is the ability of the considered molecule to be transferred from the liquid to the gas phase.

Table 5. Comparison of the total TerVOC gas-transferred concentrations (T = 40 °C, dry air flow = 50 mL/min, micro-chamber volume = 44 mL) with the total TerVOC mass fractions in the liquid form for each cleaning product, individually. NB: The physical and chemical properties of identified terpenes (VP, boiling point and molar mass) from the literature are detailed.

	RATIO	– Gas-trar		ncentration s work)	/Mass frac	etion (V/L) -	Vapor pressure (VP) from the	Boiling point
TERPENES	MC - 1	MC -2	SC - 1	GC - 1	KC - 1	KC - 2	literature (Pa)*	(°C)
α-Pinene	1.15	1.39	-	-	-	1.27	529- 599,	156
Limonene	1.15	1.02	0.68	-	3.99	1.51	189- 213	176

Linalool	-	0.47	0.97	-	-	1.17	20-27	198
Eucalyptol	1.27	-	1.13	2.17	-	0.80	230- 260	176
Cymene	0.58	0.99	-	-	-	1.73	192-195	177
Bergamol	-	-	1.15	-	-	-	NS	220
Camphor	-	-	1.25	-	-	1.02	28	209
β-Pinene	0.49	1.02	0.53	-	2.83	1.12	391-400	166
γ-Terpinene	0.17	1.19	0.64	-	-	1.99	103-145	183
α-Terpineol	-	-	1.31	-	-	-	**	175
Menthol	-	-	-	-	-	1.49	11-18	212

NS: Not specified in the literature; *: (Harb et al., 2016; ISO 16000-9, 2006; López and Gómez, 2000; Lucas et al., 2009; Rastogi et al., 2001; Teixeira et al., 2013; Espinoza et al., 1999; Hoskovec et al., 2005; Štejfa et at., 2014); **Vapor pressure near 0 Pa

Among the physical and chemical properties of TerVOCs, the saturation vapor pressure (VP) of a pure molecule (expressed in Pa) is an inherent property required for the investigation of vapor-liquid equilibrium and a key parameter for emission assessment. It is defined as the pressure exerted by a vapor in equilibrium with its pure condensed phase at a given temperature in a closed system. VPs have been experimentally determined in the literature for most TerVOCs, and the corresponding values are given in Table 5. The reported VPs of terpenes are highly varied, ranging from 0.3 Pa for α -terpineol to 599 Pa for α -pinene.

As shown in Table 5, α -pinene had a V/L ratio higher than 1, ranging from 1.15 to 1.39 among the cleaning products in which this molecule was detected. This suggests a significant ability of α -pinene to be transferred to the gas phase independent of the solvent matrix composition and could be related to its high VP of 599 Pa, the highest among all identified terpenes. In contrast, for the other compounds, the relation between the VP and V/L ratio was not predictable. Unique behaviors were observed for different terpenes: (i) the V/L ratio of linalool, with low volatility (VP values below 30 Pa), differed greatly depending on the cleaning product,

ranging from 0.47 for MC – 2 to 1.17 for KC – 2, while (ii) V/L ratio of β -pinene, with chemical and physical properties analogous to those of α -pinene, widely varied from 0.49 for MC – 1 and 2.83 for KC – 1. Thus, for most terpene species, the transfer to the gas phase is not exclusively driven by the intrinsic properties of the pure molecule but more by the solvent properties. The proportion of organic solvent and the presence of texturing agents in the aqueous media of a cleaning product play key roles in flavor release (Jouquand et al., 2008). Interactions between flavor compounds and a matrix have been extensively studied in the field of food sciences (Guichard, 2002). The determination of the gas-liquid partition coefficient under equilibrated conditions specific to each VOC/matrix pair could be used to explain the various observed behaviors (Ettre et al., 1993).

Consequently, terpene emissions from essential oil-based household cleaning products cannot be directly predicted from the liquid composition of the product. Instead, the concentrations of TerVOC species released from such products could be governed by three main factors: (i) the inherent volatility of the TerVOCs, (ii) the chemical affinities between the TerVOCs and other compounds in the formulations and (iii) the mass concentrations of terpenes in the liquid phase. In the micro-chamber tests, the major compounds emitted and transferred to the gas phase were identified, but this experimental approach is not a substitute for emission testing using experimental chambers of conventional size. Moreover, micro-chamber tests are not conducted under realistic conditions of temperature, humidity and air velocity, but under conditions which potentially enhance the transfer from the product to the gas phase. Next, primary emissions were investigated from a representative cleaning product for each of the three product categories of the six selected products.

3.3 Emission of TerVOCs under realistic application conditions: 1 m³ emission test chamber

To assess the primary emissions associated with the application of essential oil-based products, one representative product from each category was selected. This selection was based on the previously discussed results regarding the correlation between the gas-transferred concentration and the liquid content of TerVOCs. The cleaning product in each category attesting the highest ratio between its gas-transferred and liquid concentrations (V/L) was selected: (i) the surface cleaner SC - 1, (ii) the multiuse cleaner MC - 1, and (iii) the kitchen cleaner KC - 2. Regarding the diversity of the species emitted, 19 out of the 23 TerVOCs contained in the three liquid cleaners were identified in the gas phase. Fig. 3b shows the contributions of the top three emitted terpenes to the total TerVOC concentration after the separate application of the selected cleaners.

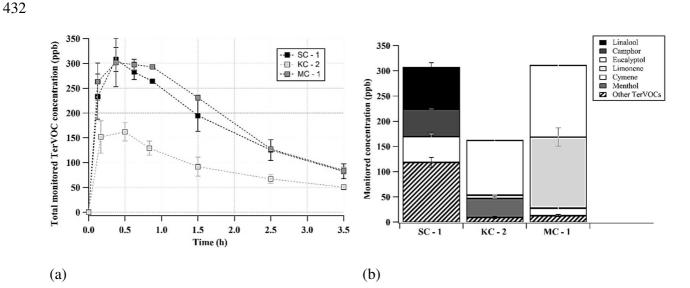


Fig. 3. (a) Temporal evolution of the total TerVOC concentration during cleaning with SC -1 (black symbols), MC -2 (dark gray symbols) and KC -2 (light gray symbols) (T = 23 °C, RH = 50 %, air renewal rate = 0.3 h⁻¹, chamber volume = 1 m³). (b) Peak concentration of total TerVOCs and the contributions of the 3 predominant terpenes after the application of SC -1, MC -1 and KC -2.

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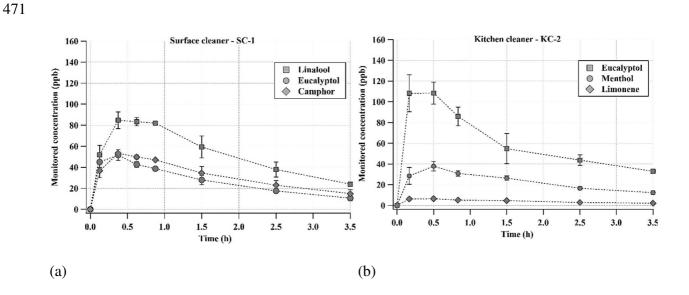
3.3.1 Emission Profile of total TerVOCs

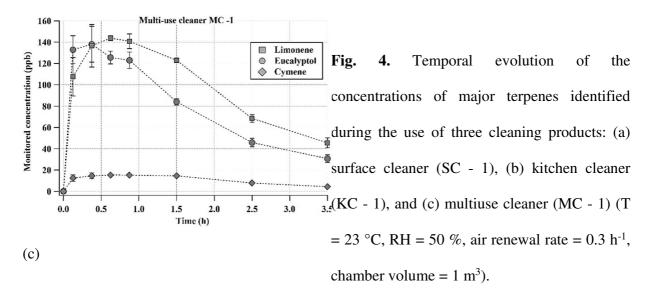
The temporal profiles of the total TerVOC concentrations in SC - 1, KC - 2 and MC - 1, reported in Fig. 3a, are characterized by the three typical phases of transient emission: (i) a sudden increase in the total TerVOC concentration, reflecting the rapid transfer of terpenes from the applied cleaning product layer to the gas phase until a peak concentration is reached (< 30 minutes); (ii) a long decrease, corresponding to the gradual depletion of the VOC source and removal of VOCs from the experimental chamber; and (iii) a final decrease exclusively associated with the evacuation of the gas phase terpenes by air exchange and possible deposition on the inner walls of the experimental chamber. Note that in any of the three phases, possible losses of terpenes from deposition on the chamber walls must be considered. As observed in Fig. 3a, SC – 1 and MC – 1 had similar, considering their respective standard deviations, peak total TerVOC concentrations of approximately 300 ppb within a half-hour after application. The maximum emitted concentration of TerVOCs KC-2 (160 ppb) was lower than those of the other two cleaning products but was reached at an analogous time. However, considering the total terpene composition in the liquid phase (section 2.1), KC – 2 was characterized by the highest mass concentration of TerVOCs (3200 \pm 200 μ g/g) among the three cleaners. These results confirm the absence of correlation between the liquid content of terpenes and the resulting concentrations of terpenes in the 1 m³ emission test chamber. Based on the investigated products, the temporal emission dynamics of essential oil-based cleaning products cannot be directly predicted from their liquid composition.

In comparison with other indoor terpene sources, primary emissions from a wood-based building product evaluated by Harb et al. (2018) produced a peak concentration of total

TerVOCs of 31 ppb under realistic conditions. Brown (1999) identified similar emitted concentrations from a 16-mm pine softwood particleboard. Thus, the contribution to the indoor total concentration of TerVOCs made by cleaning activities may transiently exceed that made by wood-based materials. However, emissions from cleaning products are characterized by temporal dynamics different from those of indoor material emissions; the former emissions mostly act as punctual or transient sources of pollutants. Thus, the cumulative exposure to TerVOCs from cleaning products is directly related to cleaning frequency and human factors. This behavior contrasts with wood-based materials, which mostly act as long-term and continuous emission sources.

3.3.2 Individual emission profiles of TerVOC





The individual emissions of 9 monoterpenes, 7 terpene alcohols, 1 oxygenated terpene, and 1 sesquiterpene were quantified. SC – 1 was characterized by the largest diversity of terpenes, since it emitted a total of 16 different terpene molecules. Fig. 4 shows the individual profiles of the predominantly emitted terpene species. The individual profiles were characterized by the same three phases observed in the emission profiles of the total TerVOCs (Fig. 3a). Nonetheless, some differences can be observed among the tested products.

The predominant terpenes released from *surface cleaner SC* – 1 were linalool, eucalyptol and cymene, which had peak concentrations of 85 ± 10 ppb, 50 ± 5 ppb and 55 ± 5 ppb, respectively. Their cumulative concentration accounted for 47.6 % of the peak total TerVOC concentration. Interestingly, regarding the emission kinetics, despite these compounds showing contrasting physical and chemical properties (Table 5), their peak concentrations were reached at equivalent emission times, i.e., nearly 25 minutes after the cleaning activity was completed (Fig. 4a). This observation suggests that determinants other than the physical and chemical properties of individual TerVOCs are involved in the emission of TerVOCs during cleaning. For instance, in the liquid composition of SC – 1, a significant concentration of ethanol of 70 % w/w was

detected. In general, in their formulations, surface cleaners contain water and various water-miscible solvents, i.e., alcohols and glycols, which are included to (i) enhance the "fast-dry effect" and (ii) dissolve grease on the surface being cleaned (Magnano et al., 2009; Missia et al., 2012; Steinemann et al., 2011; Wolkoff et al., 1998). The ethanol in surface cleaner SC – 1 was probably included for those purposes and may have accelerated the emission kinetics of TerVOCs due to the fast drying rate.

The major terpenes emitted from *kitchen cleaner KC* – 2 were eucalyptol, menthol, and limonene, as reported in Fig. 4b. Their respective peak concentrations were 110 ± 10 ppb, 38 ± 5 ppb, and 7 ± 1 ppb, contributing to 96 % w/w of the total TerVOC peak concentration. Analogous times (15 minutes) were observed for the peak concentrations of eucalyptol and limonene (Fig. 4b). Noticeably, the emission profile of eucalyptol was characterized by a sharp increase in its concentration and a subsequent clearly identifiable maximum concentration ca. 15 minutes. The emission kinetics of eucalyptol contrasts with those of menthol, which had a peak concentration at 30 minutes after the cleaning product was applied. The lower VP of menthol of approximately 18 Pa may explain this different behavior.

In the case of the *multiuse cleaner* MC – 1, limonene, eucalyptol and cymene were the main terpenes identified in the gas phase. As shown in Fig. 4c, limonene and eucalyptol had peak concentrations of approximately 145 ppb. However, despite their similar VPs, several differences can be noted in the emission profiles of these terpenes. Limonene and eucalyptol were characterized by a sharp increase up to their peak concentrations within 8 minutes, and 90 minutes after the cleaning activity was completed, their gas phase concentrations still accounted for more than 60 % of their peak concentrations. This behavior suggests that after the intense emission within the first minutes post cleaning, their emissions may continue over a long time

scale. This phenomenon can be evaluated through the determination of individual TerVOC emission rates as a function of time. In contrast, cymene presents a concentration profile characterized by a broad emission peak corresponding to a low peak concentration but possibly emission over a longer time scale.

Comparing the emission dynamics of eucalyptol (commonly released from all cleaners), rapid emission kinetics were observed for MC - 1, from which the maximum emitted concentration of eucalyptol was reached within the first 8 minutes after its application. The peak eucalyptol concentration (140 ppb) emitted from this product (MC - 1) was higher than that from the other tested products; however, the liquid form contained only 424 μ g/g eucalyptol. In contrast, slower eucalyptol emission dynamics were observed from KC - 2, with the peak concentration occurring nearly 23 minutes after the application of the kitchen cleaner. KC - 2 contained the highest concentration (2230 μ g/g) of eucalyptol in its liquid phase, but its maximum emitted eucalyptol concentration was only 110 ppb. These observations suggest that the liquid concentration of individual terpene molecules is not the only key factor driving the transfer from the liquid to the gas phase. To assess the emission of individual TerVOCs from each cleaner, the mass emission rate was calculated from the concentration profiles and assessed in the next section.

3.4 Estimation of TerVOC mass emission rates from monitored concentration profiles

In this study, the emission rate of a cleaning product is defined as the mass of volatile compound emitted per gram of applied product per unit time for a given duration *t* from the beginning of the emission. The estimated mass emission rate from a source is of major importance since (i) it provides key information to understand and describe the emission from an indoor source of

pollutants; (ii) it is a representative parameter that can be extrapolated to realistic cases, allowing the prediction of pollutant emissions in other scenarios; and (iii) it is required for the evaluation of human exposure and indoor quality control technologies.

Indoor pollutant concentrations are governed by various processes, including ventilation, pollutant emission, and pollutant transformation, such as uptake or chemical interaction. The same phenomena could be observed in an emission test chamber. Therefore, it is necessary to address the fate of pollutants considering the contribution of each process separately. In a typical indoor environment, the variation in the concentration of an individual VOC is driven by the mass conservation equation shown in Eq. 1.

$$\frac{d[X]}{dt} = -\frac{d[X]_{Out}}{dt} + \frac{d[X]_E}{dt} - \frac{d[X]_{Dep}}{dt} + \frac{d[X]_{Dep}}{dt} + \frac{d[X]_{Des}}{dt} + \frac{d[X]_{Des}}{dt} + \frac{d[X]_{Hom}}{dt} + \frac{d[X]_{Hom}}{dt} + \frac{d[X]_{Hom}}{dt} + \frac{d[X]_{Hot}}{dt} + \frac{d[X]_{Hot}$$

In controlled environments, such as the emission chamber used for our experiments, several processes included in $Eq.\ 1$ can be ruled out because of the design and operation of the chamber. First, the concentration of one of the main oxidants in the experimental chamber was continuously monitored during experiments; the ozone level remained lower than the detection limit of the ozone analyzer (0.4 ppb) during all experiments, suggesting that the contribution of ozone-induced homogeneous and heterogeneous reactions can be neglected. Harb et al. (2016) and Haghighat and De Bellis (1998) evidenced that stainless steel is the most common chamber material in reviewed publications due to its inert properties. The use of stainless steel prevents the reaction of deposited TerVOCs with the inner material surface of the chamber. This is supported by the fact that no terpenes other than those monitored in the liquid composition and no carbonyls other than those reported in the literature were detected in the gas phase in the

experimental chamber. Therefore, surface reactivity on stainless steel was disregarded. Additionally, desorption is endothermic. Moreover, in a temperature-controlled environment, the sorption of TerVOCs is suggested to be irreversible, and desorption is considered negligible.

Thus, in the emission experiments with the cleaning products, the TerVOC concentrations in the emission test chamber were predominantly controlled by (i) the air exchange, (ii) the emission rate from the TerVOC sources, and (iii) the loss due to sorption to the inner walls of the chamber (k_D). Therefore, $Eq.\ 2$ describes the resulting pollutant concentration [X] in the experimental chamber. Determining the TerVOC emission rates from their monitored concentration profiles requires knowledge of the contributions of their respective AERs and deposition rates.

$$\frac{d[X]}{dt} = -\frac{d[X]_{Out}}{dt} + \frac{d[X]_E}{dt} - \frac{d[X]_{Dep}}{dt}$$
Source
$$\frac{d[X]}{dt} = -\frac{d[X]_{Out}}{dt} + \frac{d[X]_E}{dt} - \frac{d[X]_{Dep}}{dt}$$
Deposition on surfaces
$$Eq. 2$$

3.4.1 Air exchange

Air exchange removes species X via the air renewal system of the emission test chamber and is characterized by k_{AER} , expressed in h^{-1} . The air exchange can be calculated considering the ventilation flow and the volume of the chamber or experimentally determined using a relevant tracer gas. Based on the inlet ventilation flow of our emission test chamber, k_{AER} was fixed at 0.3 h^{-1} in all sets of experiments.

$$\left(-\frac{d[X]_{Out}}{dt}\right) = -k_{AER} \times [X]$$
Eq. 3

3.4.2 Emission rate

The emission rate of species X is determined from its release from the mass of applied product per unit of time. For cleaning products, the mass emission rate is determined using Eq. 4, where m_p is the mass of cleaner applied over the surface depending on the cleaning purpose, i.e., floor or surface cleaning, and $V_{chamber}$ corresponds to the volume of the experimental chamber.

$$\frac{d[X]_E}{dt}_{\text{Pollutant source}} = \frac{\tau(t) \times V_{chamber}}{m_p}$$
Eq. 4

3.4.3 Deposition on the inner surfaces of the emission test chamber

This phenomenon corresponds to the sorption of gas-phase TerVOCs to the surface of the stainless steel chamber. The nonreactive sorption of TerVOCs on indoor materials can be characterized by the deposition rate k_D expressed in h^{-1} and is driven by the concentration of the gas-phase species X, as reported in Eq. 5. Note that k_D depends on the nature of the material and the VOC and was assessed for each set of experiments to provide a comprehensive evaluation of the fate of emitted TerVOCs and its impact on the resulting TerVOC concentrations in the gas phase.

$$\left(-\frac{d[X]_{Dep}}{dt}\right) = -k_D \times [X]$$
Eq. 5

As finite sources of TerVOCs, fragrance molecules in cleaning products may become depleted. Thereafter, the drivers of the TerVOC concentrations in the experimental chamber are only (i) the air renewal at a known decay rate (k_{AER}) and (ii) the deposition on walls at a decay rate (k_D). This situation is described in Eq.~6, the linearized form of Eq. 2. The air renewal rate was set to 0.3 h⁻¹ in our emission test chamber and was therefore easy to account for. The method

to determine k_D is illustrated with the temporal profile of linalool emitted from the surface cleaner SC – 1 in Fig. 5a. After 1 hour of emission, the peak in linalool emission has past, and the experimental data inside the box (Fig. 5a) correspond to a decaying linalool concentration due to air exchange and possible sorption on the emission chamber walls. Based on Eq. 6, this portion of the data series is plotted in Fig. 5b as ln ([linalool]) as a function of time (h). The linear relationship indicates pseudo-first-order decay. The slope of the straight line in Fig. 5b is the sum of k_{AER} and k_D . Since k_{AER} is a known experimental constant, the deposition rate of linalool under our experimental conditions can be determined as k_D (linalool) = 0.16 h^{-1} .

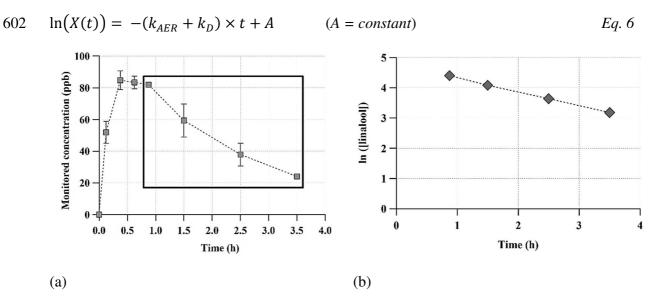


Fig. 5. (a) Temporal evolution of the linalool concentration (ppb) during cleaning using the surface cleaner SC – 1 (T = 23 °C, RH = 50 %, air renewal rate = 0.3 h^{-1} , chamber volume = 1 m^3); the data series in the box corresponds to linalool decay after depletion of its source. (b) Plot of ln ([*linalool*]) as a function of time (h); NB: in b), only the data series within the box in (a) is plotted.

The deposition rates (k_D) of the 6 predominant released TerVOCs were experimentally determined during the application of the surface cleaner SC-1, the kitchen cleaner KC-2, and

the multiuse cleaner MC – 2 (Table 6) and ranged from 0.12 to $0.52 \, h^{-1}$. These values clearly show that the deposition rates of TerVOCs on the chamber walls have to be determined to interpret the concentration profiles since they impact the resulting concentrations by the same order of magnitude as the AER $(0.3 \, h^{-1})$.

Table 6. Comparison of the deposition rate coefficient k_D (h⁻¹) of the predominant terpenes from the three selected cleaning products: (i) the surface cleaner SC – 1, (ii) the kitchen cleaner KC – 2, and (iii) the multiuse cleaner MC – 1.

Deposition rate $k_D(h^{-1})$								
TERPENE	Cleaning Product							
TEXTENE	SC - 1	KC – 2	MC – 1					
α-Pinene	0.52	0.46	0.26					
Limonene	0.12	0.11	0.24					
Linalool	0.16	ND	ND					
Eucalyptol	0.19	0.38	0.30					
Cymene	Ø	0.22	0.31					
Bergamol	0.12	Ø	Ø					
Camphor	0.12	ND	Ø					
Menthol	Ø	0.08	Ø					

 \emptyset = This terpene was not identified in a cleaning product in its liquid form.

ND: This terpene was not a predominantly emitted terpene from a cleaning product; therefore, it was not detected in the gas phase.

According to Table 6, the k_D values for the same species may differ from one applied product to another. For instance, the k_D of α -pinene was 0.52, 0.46, and 0.26 h^{-1} for SC – 1, KC – 2, and MC – 1, respectively. These differences are suggested to be associated with the cleaning product formulations. Among cleaning product constituents, water can be present in formulations at mass percentages ranging from 95 % and 50 %. Subsequently, during cleaning, the RH of the

confined environment increases. Springs et al. (2011) evidenced that the number of moles of terpenes taken up per unit of surface area tends to decrease with increasing RH, suggesting competitive interactions with water molecules for uptake surfaces. Consequently, the deposition rate (k_D) of fragrance molecules on indoor materials could be impacted by two main factors: (i) competition for uptake among terpenes and cleaning constituents and (ii) the increase in RH related to the water content of the cleaner. Consequently, the k_D value for a given terpene might differ depending on the cleaning product formulation.

3.4.4 Determination of the mass emission profiles from the concentration profiles

The mass conservation equation under our experimental conditions during cleaning is described by Eq. 7, where [X] is the concentration of species X, m_p is the mass of applied cleaning product, $V_{chamber}$ corresponds to the volume of the chamber (m^3) , and k_{AER} and k_D are the decay rates (h^{-1}) of species X related to air renewal and deposition.

$$\frac{d[X]}{dt} = \frac{\tau(t) \times V_{chamber}}{m_p} - \frac{(k_D + k_{AER}) \times [X]}{\text{Concentration of X removed by air exchange and deposition}}$$

Eq. 7

Based on Eq. 7, the emission rate of a given TerVOC can be retrieved from its concentration profile corrected by the air renewal rate and by the depletion rate. Fig. 6 shows the temporal evolution (a) of linalool emitted during cleaning using the surface cleaner SC - 1 and (b) of eucalyptol released during cleaning using the kitchen cleaner KC - 2. The concentrations of linalool and eucalyptol directly monitored in the 1 m³ emission test chamber are plotted as black squares in Fig. 6a and b. First, the raw profile was corrected by the air renewal rate, resulting in linalool and eucalyptol concentration profiles that would have been recorded if no air

exchange occurred in the emission test chamber (light gray squares in Fig. 6). Then, these profiles were corrected by the specific deposition rate of each terpene, producing final profiles that correspond to the concentrations if no linalool and eucalyptol had been removed from the gas phase in the chamber, neither by sorption on the chamber walls nor by air exchange (dark gray circles in Fig. 6). The resulting profiles reflect only the linalool and eucalyptol emissions from SC - 1 and KC - 2, respectively. Therefore, the times when these terpenes reach steady state indicates the end of their emission from their respective cleaners. Moreover, this final profile can be used to calculate the emission rate of linalool and eucalyptol according to Eq. 8. The same calculations were performed for all terpenes detected in the gas phase during the use of the three cleaners.

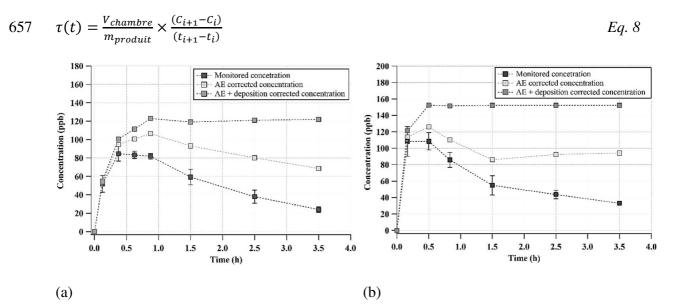
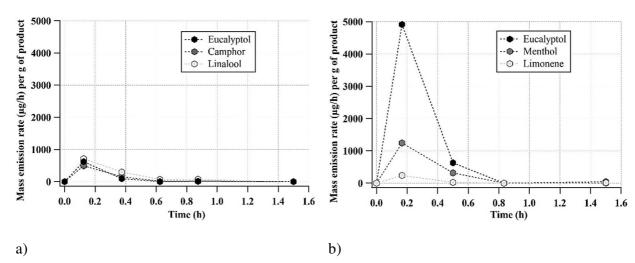


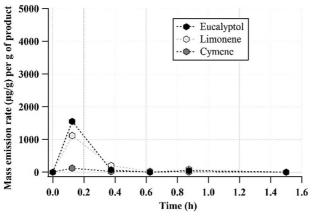
Fig. 6. (a) Temporal evolution of the (i) monitored concentration (ppb), (ii) air exchange-corrected concentration (ppb), and (iii) air exchange + deposition-corrected concentration (ppb) of linalool during cleaning using the surface cleaner SC – 1. (b) Temporal evolution of the (i) monitored concentration (ppb), (ii) air exchange-corrected concentration (ppb), and (iii) air exchange + deposition-corrected concentration (ppb) of eucalyptol during cleaning using the

kitchen cleaner KC – 2 (T = 23 °C, RH = 50 %, air renewal rate = 0.3 h^{-1} , chamber volume = 1 m^3).

3.4.5 Emission rate profiles of individual TerVOCs

Based on Eq. 8, the temporal evolution of the mass emission rate was calculated for each cleaner and for each TerVOC emitted. Fig. 7 presents the temporal evolution of the mass emission rates of the three predominant TerVOCs monitored during the use of (a) SC – 1, (b) KC – 2 and (c) MC – 1. The temporal profiles of the emission rates allow the determination of (i) the time required to reach the maximum emission rate, (ii) the value of the maximum emission rate, and (iii) the time at which the emission source is depleted.





c)

Fig. 7. Temporal evolution of the mass emission rates of the three predominant terpenes detected in the gas phase during the use of (a) the surface cleaner SC - 1, (b) the kitchen cleaner KC - 2 and (c) the multiuse cleaner MC - 1 applied in the 1 m³ emission test chamber (T = 23 °C, RH = 50 %, air renewal rate = 0.3 h⁻¹, chamber volume = 1 m³).

Based on Fig. 7, irrespective of the cleaning product, the time required for a given terpene to reach the maximum emission rate was similar for all monitored terpenes: within 8 minutes after cleaning was completed. However, the magnitude of the peak emission rate of the predominant terpene released clearly differed from one cleaning product to another: (i) $710 \pm 15 \, \mu g/h$ linalool from SC – 1, (ii) $1550 \pm 90 \, \mu g/h$ eucalyptol from MC – 1, and (iii) $5000 \pm 120 \, \mu g/h$ eucalyptol from KC – 2.

Noticeably, the emission of eucalyptol and menthol from KC-2 was exhausted within 50 minutes and after 30 minutes for limonene. Of the three selected categories, this product category had the highest emission rates. However, SC-1 and MC-1 showed shorter emission durations than KC-2. Their total TerVOC emissions were exhausted within only 24 minutes, as shown in Fig. 7a and b.

Regarding the emission of individual terpenes, eucalyptol was only terpene emitted from all three tested cleaners. However, its peak emission rate varied widely, reaching $620 \pm 20 \,\mu\text{g/h}$, $4\,900 \pm 110 \,\mu\text{g/h}$ and $1460 \pm 30 \,\mu\text{g/h}$ for the surface cleaner (SC – 1), the kitchen cleaner (KC –

2) and the multiuse cleaner (MC -1), respectively. Similar emission rate variability was observed for the limonene emitted from two products: $240 \pm 30 \,\mu\text{g/h}$ and $1100 \pm 80 \,\mu\text{g/h}$ for KC -2 and MC -1, respectively.

The assessment of the emission rates confirms that in addition to the liquid TerVOC content in the cleaner, the emission dynamics are also driven by (i) the solvent matrix in which the fragrance molecules are contained due to chemical affinities that might be generated among species, delaying the emission kinetics, and (ii) the volatility of individual terpene molecules.

3.5 Emission of OVOCs from natural cleaning products under realistic application conditions:

1 m³ emission test chamber

Carbonyl compounds were monitored during each set of emission experiments. Three targeted oxygenated VOCs emitted from the application of SC -1, KC -2 and MC -1 were identified and quantified: formaldehyde, acetaldehyde and acetone. Their individual maximum emitted concentrations varied from one cleaning product to another, ranging from (i) 6 ± 1 to 18 ± 3 ppb formaldehyde, (ii) 22 ± 3 to 210 ± 20 ppb acetaldehyde, and (iii) 18 ± 4 to 108 ± 15 ppb acetone. Among the identified OVOCs, formaldehyde was classified in 2016 by the European Commission as carcinogenic (Group 1) and genotoxic (International Agency for Research on Cancer, 1979). Additionally, the long-term exposure limit was established at 9 ppb by the World Health Organisation (2010). Thus, in the framework of this study, formaldehyde is of particular interest because of its toxicity.

3.5.1 Formaldehyde emission profiles from the use of natural cleaning products

Fig. 8a presents the temporal evolution of the concentration of formaldehyde emitted by cleaning with SC - 1, KC - 2 and MC - 1. Unlike the terpene emission profiles, the temporal evolution of the formaldehyde concentration in Fig. 8 is characterized by two phases: (i) a marked increase confirming the rapid transfer of formaldehyde from the product to the gas phase during the first 50 minutes of emission and (ii) either a slow increase for SC – 1 and MC – 1 or steady state for KC - 2. In contrast to the terpene concentration profiles, the emission profiles of formaldehyde do not follow a typical transient emission process. As observed in Fig. 8a, the maximum emitted concentration of formaldehyde was 9 and 16 ppb for the surface cleaner (SC - 1) and for the multiuse cleaner (MC - 1), respectively. These values were observed 3.5 hours after the cleaning was finished. For the kitchen cleaner KC – 2, the concentration of formaldehyde reached 6 ppb at 0.8 hours and then remained constant for at least 3.5 hours after the product application. These observations suggest that even after 3.5 h, the emission source of formaldehyde was not exhausted, reflecting delayed emission kinetics that may not solely be related to a primary release from the cleaning product itself. In agreement with our findings, (CSTB, 2006); Solal et al. (2014) assessed the emissions of formaldehyde from 35 consumer products in different categories applied under realistic conditions in the experimental house MARIA. Among the evaluated products, the uses of (i) 2 vacuum fresheners, (ii) 2 carpet cleaners, (iii) 3 toilet cleaners, (iv) 4 glass cleaners and (v) 2 furniture polishes were associated with increases in the formaldehyde concentration in the experimental house. The observed concentrations were described by the same two phases shown in Fig. 8a: (i) a sharp increase in the HCHO concentration confirming the rapid transfer of formaldehyde from the product to the gas phase and (ii) a slower increase or steady state. The maximum concentrations that arise from the

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emission were 6 to 16 ppb, depending on the category of product applied. These values were observed at 120 minutes after the cleaning was completed.



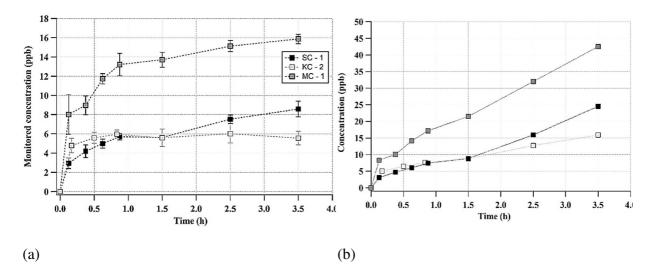


Fig. 8. (a) Temporal evolution of the concentration of formaldehyde (ppb) from the use of three cleaning products: (i) surface cleaner (SC-1), (ii) kitchen cleaner (KC-1) and (iii) multiuse cleaner (MC-1) (T = 23 °C, RH = 50 %, AER = 0.3 h⁻¹). (b) Temporal evolution of the *air exchange-corrected* concentration of formaldehyde (ppb) as a function of time during cleaning with the three cleaners: (i) SC – 1, (ii) KC – 2, and (iii) MC – 1 (T = 23 °C, RH = 50 %, AER = 0.3 h^{-1}).

To confirm these observations, the emission profile was corrected by the air exchange in the experimental chamber, i.e., k_{AER} of 0.3 h^{-1} . The corrected profile is presented in Fig. 8b, which depicts the temporal evolution of the concentration of formaldehyde that would have been monitored if no air exchange occurred in the emission test chamber. The concentration variation resulted from only the transfer of formaldehyde to the gas phase and the possible sorption on the inner walls of the experimental chamber. However, the mass emission rate of formaldehyde

cannot be calculated by the estimation method in section 3.4 since the emission source could not be assumed to be exhausted and the deposition rate (k_D) could not be determined. As shown in Fig. 8b, the corrected profile of formaldehyde emission from SC-1, MC-1 and KC-2 was characterized by two main phases: (i) a primary increase in the concentration, suggesting a release linked to a primary emission of formaldehyde potentially contained in the cleaning product, and (ii) a linear increase in the concentration of formaldehyde. The slope of the linear relationship of the corrected formaldehyde concentration varied depending on the cleaning product. Therefore, the temporal evolution of the corrected formaldehyde concentration, presented in Fig. 8b, did not correspond to a transient emission profile but suggested the presence of formaldehyde releasers in the cleaning product formulation, in which case, the concentration of formaldehyde might be associated with secondary emission sources.

The specific concentration dynamics associated with the emission of formaldehyde are suggested to be related to two mechanisms: (i) direct (or primary) emission of formaldehyde contained in the cleaning product and (ii) emission of formaldehyde from formaldehyde-releasing molecules present in the cleaning product formulations (Pastor-Nieto et al., 2017). Formaldehyde releasers are defined as chemical compounds that decompose to release formaldehyde when dissolved in aqueous solvents (De Groot et al., 2009; Flyvholm and Andersen, 1993; Kajimura et al., 2008). This process is characterized by an in situ degradation of non-formaldehyde-containing components, i.e., the self-oxidation of ethoxylated alcohols. Generally, these chemicals are used in consumer products as preservatives, antiseptics, and/or for enhancing their disinfectant performance. The concentration of formaldehyde or formaldehyde-releasing compounds added in product formulations might depend on their use purpose and manufacturer preferences. According to the National Database for Products and Compositions in

France (BNPC), formaldehyde releasers are included in the formulations of 78 % of household products as preservative agents, corresponding to a mass concentration ranging from 0.2 % to 0.3 % w/w (Affset, 2009).

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De Groot et al. (2009) assessed formaldehyde and formaldehyde releasers contained in consumer products in various categories by reviewing the Danish Product Register Database (PROBAS). His findings reveal that the most important product categories containing formaldehyde or formaldehyde releaser are (i) biocides or pesticides; (ii) paints, lacquers and varnishes; and (iii) cleaning and washing agents. In the cleaning and washing agents, 9 different formaldehyde-releasing compounds were identified. Among these chemicals, Bronopol® (2brono-2-nitropropane-1,3-diol) was the most frequently used formaldehyde releaser, being detected in 124 out of 275 cleaning products or washing agents. However, the authors indicated that a total of 19 out of the 42 detected formaldehyde-releasing compounds were not found in PROBAS. Similarly, the BNPC in France reported the presence of formaldehyde releasers in 236 out of 692 cleaning products on the European market (Affset, 2009). Additionally, the BNPC published and identified a list of 24 compounds confirmed as formaldehyde releasers, including Bronopol® and Methenamine as the most frequently used among household products. Kajimura et al. (2008) investigated the formation of HCHO from a common formaldehyde-releasing compound known as Bronopol® from a homemade cosmetic product prepared with 0.1 % w/v of this molecule. This study aimed to correlate the presence of Bronopol® and the respective formation of formaldehyde by headspace analysis. Their results verified an increase in formaldehyde concentration over time, reaching 20 ppm after 10 days, and this level was observed to be constant over 50 days. Note that this concentration cannot be compared to our results because of differences in the experimental protocol and approach.

Regarding the three cleaning products evaluated in this study, the presence of Bronopol® was listed by manufacturers for only the multiuse cleaner MC – 1. Its presence may explain the unique formaldehyde emission profile observed for that product. However, according to European legislation for the cleaning product industry, numerous formaldehyde releasers are not required to be included in the product ingredient list. With this lack of information, the correlation between the formaldehyde emission profile observed for the tested cleaning products and the presence of formaldehyde releasers in their composition is suggested but has not been confirmed.

Therefore, the use of natural cleaning products might generate a continuously increasing indoor formaldehyde concentration, in which the maximum emitted levels might remain in the confined environment for several hours after the cleaning is completed. Additionally, formaldehyde releasers included in consumer products represent a hidden source of formaldehyde, of which consumers might not even be aware, extending its impact on IAQ by several tens of ppb for several hours after household activity is completed.

3.6 Wrap-up of the terpene emissions from cleaning products: from their liquid composition to

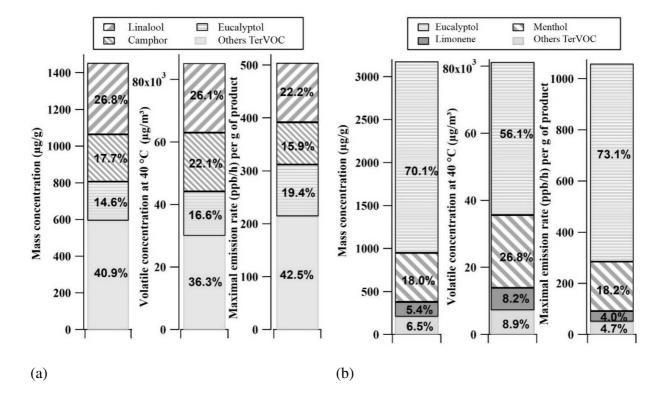
their mass emission rates

In this work, the TerVOC emissions from the use of 3 cleaners were investigated by comparing different scales, including their liquid composition, their gas-transferred concentration and their emission under realistic use patterns. This innovative methodology allowed the investigation of the emission of fragrance molecules at different scales to evaluate the impact on IAQ through an integrated approach, achieving an accurate assessment of the mass emission rates. These

estimated mass emission rates could be further implemented for the estimation of human exposure to these chemicals and to the definition of risk scenarios.

Fig. 9 shows the relative abundances expressed as percentages of the predominant TerVOCs and as their respective absolute scales for (i) the mass concentration detected in the liquid form (μ g/g of product), (ii) the volatile fraction (gas-transferred concentration) associated with the micro-chamber testing (μ g/m³), and (iii) the maximum emission rates per individual terpene (ppb/h per g of product) associated with the use of (a) the surface cleaner SC – 1, (b) the kitchen cleaner KC – 2, and (c) the multiuse cleaner MC – 1.





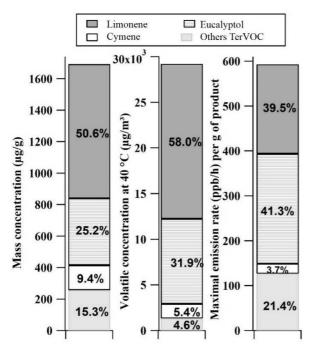


Fig. 9. Comparison of the relative abundances in the mass concentration (μ g/g) in the liquid phase, the volatile concentration (μ g/m³) (T = 40 °C, dry air flow = 50 mL/min, microchamber volume = 44 mL) and the maximum emission rate per gram of cleaning product (T = 23 °C, RH = 50 %, air renewal rate = 0.3 h⁻¹, chamber volume = 1 m³) for (a) the surface cleaner SC – 1, (b) the kitchen cleaner KC – 2, and (c) the multiuse cleaner MC – 1.

(c)

In terms of the absolute values, written in bold in Fig. 9, no correlation was found between them. As mentioned in section 3.2, the maximum emission rate of terpene molecules from the use of essential-oil-based cleaning products cannot be directly predicted from their content in the liquid form or from micro-chamber testing. Micro-chamber results can be used to describe key general trends of the predominant emitted terpenes but should not be considered for realistic emission assessment. Moreover, the relative distributions between terpenes for the liquid content and for the maximum emission rate are much closer rather than one for the gastransferred concentration. That can be explained by the fact that micro-chamber experiments were conducted at a temperature of 40 °C, enhancing the transfer from the liquid to the gas phase.

In regard to the behavior of specific compounds, such as eucalyptol and limonene, two species with similar VPs (260 and 219 Pa, respectively), substantial differences were observed. In all the investigated cleaning products, eucalyptol evidence higher proportions in the maximum mass emission rate if compared to its relative abundance in the liquid form. This observation confirms the ability of eucalyptol to be transferred from the liquid to the gas phase, irrespective of the solvent matrix of the cleaner. In contrast, for a given cleaning product, limonene had (i) a lower relative abundance at the maximum mass emission rate than in the liquid phase and (ii) a higher relative abundance in the volatile fraction than in the liquid form. These observations suggest that the emission of limonene might be more impacted by the solvent matrix in the cleaning product than by its volatility (VP = 219 Pa), and therefore, its mass transfer might be delayed due to chemical affinities to other compounds.

4. CONCLUSIONS AND PERSPECTIVES

This study has shown that the largest contributors of terpenes to indoor air are not only recognized, regulated and controlled sources, namely, building materials, but also "green" household products, which should be seriously considered as versatile and high-impact sources. These fragrance chemicals classified as allergens responsible for skin irritation, allergic rhinitis, and asthma are primarily associated with the use of cleaning products (Wolkoff and Nielsen, 2017). Essential-oil-based household products might release concentrations of several tens to hundreds of ppb TerVOCs, exceeding the exposure limits established by the European Union and the United States (Dimitroulopoulou et al., 2015; Kirchner et al., 2001; World Health Organisation, 2010). Regarding formaldehyde emissions from the investigated cleaning products, this study has confirmed that these products could represent a hidden source of formaldehyde in

confined environments. Indeed, consumers might increase the formaldehyde concentration indoors by several ppb without being aware of it due to the presence of non-regulated formaldehyde-releasing compounds contained in these types of products.

Several key factors were confirmed to drive the mass transfer from the liquid to the gas phase of terpene molecules contained in essential-oil-based cleaning products: (i) the mass concentration of individual terpenes in the liquid phase, since a correlation between the major contained and the predominantly emitted terpenes was observed for all the tested cleaning products; (ii) the solvent matrix included in the cleaning formulation, which may generate chemical affinities between terpenes and the cleaner constituents, impacting the emission dynamics; and (iii) the volatility of the individual terpene molecules.

Micro-chamber testing cannot replace standard emission test chambers but is a complementary tool that allows evaluation of the gas-transferred concentrations and rapid screening of the potentially released TerVOCs from essential-oil-based household products prior to a detailed product emissions characterization. Micro-chamber experiments need to be accompanied by an evaluation of the effective emissions under realistic conditions in a test chamber to accurately estimate the human exposure to cleaning product constituents under recommend application scenarios.

While this study aimed to implement realistic scenarios, the 1 m³ chamber imposes limitations regarding the application process. Due to space limitations, real consumer use patterns while performing a cleaning activity could not be completely reproduced. Without being wiped, trigger spray products were directly sprayed in the chamber and dispersed by droplets onto the surface, and liquid cleaning products were drizzled on the surface placed at the center of

880	the chamber. Therefore, further assessments of TerVOC emissions from essential-oil-based
881	cleaning products under real consumer use patterns in a full-scale room are highly recommended.
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899	The authors declare that they have no conflicts of interest.
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