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SUMOylation modulates the stability and function of PI3K-p110 β

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Running Title: Regulation of p110 β by SUMO modification

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Abstract

Class I PI3K are heterodimers composed of a p85 regulatory subunit and a p110 catalytic subunit involved in multiple cellular functions. Recently, the catalytic subunit p110 β has emerged as a class I PI3K isoform playing a major role in tumorigenesis. Understanding its regulation is crucial for the control of the PI3K pathway in p110 β -driven cancers. Here we sought to evaluate the putative regulation of p110 β by SUMO. Our data show that p110 β can be modified by SUMO1 and SUMO2 *in vitro*, in transfected cells and under completely endogenous conditions, supporting the physiological relevance of p110 β SUMOylation. We identify lysine residue 952, located at the activation loop of p110 β , as essential for SUMOylation. SUMOylation of p110 β stabilizes the protein increasing its activation of AKT which promotes cell growth and oncogenic transformation. Finally, we show that the regulatory subunit p85 β counteracts the conjugation of SUMO to p110 β . In summary, our data reveal that SUMO is a novel p110 β interacting partner with a positive effect on the activation of the PI3K pathway.

Keywords: p110 β , p85 β , stability, SUMO, transformation

Abbreviations

CHX, cycloheximide

PI3Ks, phosphatidylinositol-4,5,bisphosphate 3-kinases

SUMO, small ubiquitin-related modifier

SEN1, SUMO specific protease 1

Introduction

Class IA phosphatidylinositol-4,5,bisphosphate 3-kinases (PI3Ks) are heterodimeric proteins composed of one p110 catalytic subunit (p110 α , β , or δ) and one p85 regulatory subunit (p85 α , p55 α , p50 α , p85 β , and p55 γ) [1]. Mutational activation or altered expression of class IA PI3K results in enhanced PI3K signaling, which is associated with oncogenic cellular transformation and cancer [2-6]. The p85 subunit inhibits the p110 basal activity, allows its activation downstream RTKs, and is essential for its thermal stability [7-9]. Therefore, optimal signaling through the PI3K pathway depends on a critical molecular balance between the regulatory and the catalytic subunits [10]. The stabilization of p110 by p85 is not totally understood. Interaction between the iSH2 domain of p85 and the N-terminus of p110 stabilizes p110 in mammalian cells [11-14]. However, stabilization of p110 can be also achieved by protein synthesis at low temperature or the linkage of epitope tags to the N-terminus of p110 [8].

The regulatory activity of p85 on p110 is modulated by different p85 post-translational modifications such as the phosphorylation at tyrosine residues or its conjugation to the small ubiquitin-related modifier (SUMO) [15-17]. Conjugation of SUMO to the iSH2 domain on p85 reduces its tyrosine phosphorylation and reinforces its inhibitory effect on p110 [17]. Covalent interaction with SUMO is also a critical event in the regulation of other key PI3K signaling components such as AKT or PTEN [18-22]. So far, no direct post-translational modification of p110 by SUMO has been reported.

Here, we evaluated the potential regulation of the PI3K catalytic subunit p110 β by SUMO. *In vitro* and *in vivo* SUMOylation assays show that p110 β is an authentic SUMO substrate, that lysine residue K952,

located at the activation loop in p110 β , is essential for SUMO conjugation, and that SUMOylation increases the stability of p110 β . In addition, functional studies reveal that mutation of the SUMOylation site in p110 β downmodulated the p110 β -mediated PI3K activation downstream pathway, its cell proliferation-promoting activity, and the oncogenic capability. Finally, our data show that the regulatory subunit p85 β negatively modulates the interaction of p110 β with SUMO. These results indicate that SUMO conjugation to p110 β has a positive impact on the activation of the PI3K pathway, and its modulation could represent a novel strategy for PI3K regulation.

Materials and Methods

Cell culture, plasmids and transfections

HEK-293, PC3, Rat-2 and NIH-3T3 cells were purchased from ATCC. Primary chicken embryo fibroblasts (CEF) were obtained from specific-pathogen-free (SPF) 10-day-old eggs (Intervet, Salamanca, Spain) following the protocol previously described [23]. HEK-293, PC3, and CEF cells were cultured in complete medium (DMEM supplemented with 10% fetal bovine serum, 1% penicillin/streptomycin). Rat-2 and NIH-3T3 were cultured in DMEM supplemented with 5% fetal bovine serum and 1% penicillin/streptomycin. The cells were transiently transfected using PEI (Polysciences, Inc), according to the manufacturer's instructions. pcDNA-His6-SUMO1, pcDNA-His6-SUMO2, and pcDNA-SV5-Ubc9 plasmids were previously described [24, 25]. The vector encoding the HA-tagged form of p85 β (HA-p85 β) and p85 β SUMOylation mutant (HA-p85 β K535RK592R) were previously described [17]. The myc-tagged p110 β expression plasmid [26] was kindly provided by Jonathan M Backer. The HA-tagged p110 α expression plasmid (Addgene #1397) was kindly provided by Lewis Cantley. Myc-p110 β construct was mutated using Quickchange site directed mutagenesis (Stratagene) and the oligonucleotides p110 β -K647R-F-5'-GTGCAAGTGTTAAGATATGAGCCTTTTC-3', p110 β -K647R-R-5'-GAAAAGGCTCATATCTTAACACTTGCAC-3', p110 β -K952R-F-5'-CTAAGTTTGGCATTAGAAGGGAGCGAGTG-3', and p110 β -K952R-R-5'-CACTCGCTCCCTTCTAATGCCAAACTTAG-3'. All mutations were confirmed by sequencing. For generation of stably-expressing cell lines, NIH-3T3 or Rat-2 cells were co-transfected with pcDNA, myc-p110 β WT or myc-p110 β K952R and a vector with a puromycin resistance gene (ratio 5:1) and selected by incubating in 2.5 μ g/ml puromycin.

Focus formation assay

Stably transfected NIH-3T3 cells expressing p110 β or p110 β K952R were plated in six-well plates. Cells were grown for 20 days with changes of medium every three days. The cells were stained with crystal violet and the number of transformed foci per well was counted. In addition, NIH-3T3 cells stably transfected with empty vector or p110 β K952R were transduced with a lentiviral vector encoding an empty vector or p110 β WT, seeded to 6-well plates, and after 20 days incubation, focus formation was observed.

Antibodies

Anti-HA antibody (901513) was from Biolegend. Anti-SUMO2 (ab3742), anti-phospho-Ser-473 of AKT (9271S), anti-phospho-FoxO1 (Thr24)/FoxO3 (Thr32) (9464S), anti-FoxO3a (2497S), anti-lamin A/C (4C11), and anti-AKT (9272S) were from Cell Signaling. Anti-B23 antibody (B0556) was from Sigma.

Anti-myc antibodies were from Cell Signaling (9B11) and Santa Cruz Biotechnology (sc-40). Anti-GAPDH antibody (sc-32233) was from Santa Cruz Biotechnology. Antibodies against p110 β were purchased from Invitrogen (PA5-22152) and Cell Signaling (C33D4). Anti-Histidine antibody (MA1-21315) was from Invitrogen.

Nuclear-cytoplasm fractionation

The nuclear and cytoplasmic fractions were isolated using a nuclear-cytosol fractionation kit (BioVision), following manufacturer's instructions.

***In vitro* SUMOylation assays**

In vitro SUMO conjugation assays were performed on [³⁵S]methionine-labeled *in vitro*-transcribed/translated proteins as described previously [27], using recombinant E1 (Biomol), Ubc9, and SUMO1 or SUMO2. The *in vitro* transcription/translation of proteins was performed by using 1 μ g of plasmid DNA and a rabbit reticulocyte-coupled transcription/translation system according to the instructions provided by the manufacturer (Promega). Briefly, [³⁵S]methionine-labeled *in vitro*-transcribed/translated proteins were incubated with E1 in a 10 μ l reaction including an ATP regenerating system (50 mM Tris (pH7.5), 5 mM MgCl₂, 2 mM ATP, 10 mM creatine phosphate, 3.5 U of creatine kinase /ml, and 0.6 U of inorganic pyrophosphatase/ml), 600 ng of Ubc9 and 10 μ g of SUMO1 or SUMO2, at 37°C for 90 min. Reactions were stopped by adding SDS-PAGE loading buffer, boiled for 5 min at 100°C, analyzed by SDS-PAGE and detected by autoradiography.

***In vitro* deSUMOylation assay**

In vitro deSUMOylation assay with recombinant GST-SENP1 (Biomol) was performed on p110 β -SUMO1 or p110 β -SUMO2 as described previously [27]. p110 β -SUMO1 or p110 β -SUMO2 proteins obtained after *in vitro* SUMOylation assay were incubated with GST-SENP1. Reactions were incubated at 37°C for 1 h, terminated by adding SDS-PAGE loading buffer, boiled for 5 min at 100°C, analyzed by SDS-PAGE and detected by autoradiography.

Immunofluorescence staining

Immunofluorescence staining and confocal analysis were performed as described previously [27]. Briefly, cells cultured on coverslips were fixed with 2% paraformaldehyde, permeabilized with 0.25% Triton X-100 in PBS and blocked in 2% bovine serum albumin (BSA). Preparations were incubated with primary antibodies overnight in a moist chamber at 4°C. Coverslips were washed with PBS and incubated with Alexa conjugated secondary antibody for 1 h at room temperature. Nuclei were stained with DAPI (4',6'-diamidino-2-phenylindole), and coverslips were mounted with ProLong. Mouse anti-myc antibody was used at a dilution of 1:200. Secondary Alexa 488-conjugated antibody was obtained from Invitrogen. Analysis of the samples was carried out on a Leica TCS SP5 confocal laser microscope. Images were exported using Adobe Photoshop.

Purification of His-tagged conjugates

Cells were lysed in G-buffer (6M Guanidine hydrochloride, 100 mM sodium phosphate pH 8.0, 50 mM Tris-HCl pH8.0) and incubated with pre-equilibrated Ni²⁺-NTA-agarose beads for 2 h at room temperature. Beads were washed four times with U-buffer (8 M urea, 100 mM sodium phosphate, 10 mM Tris-HCl pH8.0), resuspended in SDS-PAGE loading buffer and analyzed by SDS-PAGE followed by Western-blot.

Immunoprecipitation assay

Immunoprecipitation of endogenous SUMO1 or SUMO2 modified proteins was performed using denaturing cell lysis as described previously [28]. Briefly, cells were lysed with lysis buffer (PBS, 1% (wt/vol) SDS, 5 mM EDTA, 5mM EGTA, 10 mM NEM) containing a protease inhibitor cocktail. Cell lysates were sonicated to reduce viscosity. DTT (50 mM final concentration) was added and cell lysates were boiled at 97°C for 10 min, diluted (1:10) in cold RIPA buffer and supplemented with NEM (10 mM final concentration). Lysates were then cleared by centrifugation at 16,000 x g for 15 min, and the supernatant was incubated overnight at 4°C with SUMO1 or SUMO2 antibody-coupled beads. Control lysate was mixed with normal mouse IgG and beads. The next day, the beads were then washed three times with RIPA buffer, immunoprecipitated proteins were resuspended in SDS-PAGE loading buffer, boiled for 5 min at 100°C and subjected to immunoblot analysis with the indicated antibodies.

Coimmunoprecipitation of p110 β with p85 β subunit of PI3K was evaluated in cells lysed in BC-100 buffer (20 mM Tris, pH 8, 0.1 M KCl, 10% glycerol, 1mM EDTA, 1 mM DTT, 0.1% Nonidet P40) containing 10 mM NEM and a protease inhibitor cocktail. Cell lysates were cleared by centrifugation at 20,000 x g for 10 min, and the supernatant was incubated overnight at 4°C with the primary antibody. The next day, lysates were incubated with protein A-sepharose beads for 2 h at 4°C. Control lysate was mixed with normal mouse IgG and beads. The beads were then washed four times with BC-100 buffer, resuspended in SDS-PAGE loading buffer, boiled for 5 min at 100°C and subjected to immunoblot analysis with the indicated antibodies.

Stability assay

HEK-293 cells were transfected with the indicated plasmids and 24 h after transfection cells were treated with cycloheximide (100 μ g/ml). At different times after treatment cells were analyzed by Western-blot with the indicated antibodies. Bands intensity was measured using ImageJ software. P110 β bands intensity was normalized to a housekeeping protein.

Statistical analysis

Statistical analysis was performed using Student's t test for comparison between two groups. We used the one-way Analysis of Variance (ANOVA) followed by Dunnett's test using the GraphPad Prism 8.0.2 software for comparisons of more than two groups. All data are presented as mean +/- standard deviation (SD). The significance level chosen for the statistical analysis was p<0.05.

Results

Modification of p110 β by SUMO

We carried out an *in vitro* SUMOylation assay in the presence of SUMO1 or SUMO2 using *in vitro* translated [³⁵S] methionine-labeled p110 β protein as a substrate. The p110 β protein was detected as a single band of the expected 110 kDa molecular weight. When the reaction was incubated with SUMO1 or SUMO2 we observed a strong additional band of around 130 kDa and fainter higher bands of around 150 and 170 kDa, which disappeared after incubating the reaction with the recombinant SUMO specific protease 1 (SENP1) (Fig. 1a). These results demonstrate that p110 β is SUMOylated by SUMO1 and SUMO2 *in vitro*. In addition, the presence of several bands corresponding to p110 β -SUMO1 in the *in vitro* assay indicates that SUMO1 can conjugate to more than one lysine residue, although the much stronger intensity of the 130 kDa band in comparison with the higher ones also suggests that the predominant form of SUMOylated

p110 β has only one SUMO1 molecule conjugated at a time. Then, in order to determine whether p110 β also conjugates to SUMO within the cell, HEK-293 cells were co-transfected with myc-tagged p110 β together with pcDNA or Ubc9 and His6-tagged SUMO1 or SUMO2 plasmids. At 48 h after transfection, His6-tagged proteins were purified in denaturing conditions using nickel beads. Western-blot analysis of the purified extracts with anti-p110 β antibody revealed bands of the expected size corresponding to p110 β -SUMO1 and p110 β -SUMO2 only in those cells co-transfected with His6-SUMO1 and His6-SUMO2, respectively, indicating that p110 β is modified by SUMO1 and SUMO2 in transfected cells (Fig. 1b). Finally, we decided to evaluate the SUMOylation of endogenous p110 β protein by using a method that allows high enrichment of SUMO targets under physiological conditions [28]. Protein extracts from PC3 cells were lysed under denaturing conditions, diluted in non-denaturing buffer and subjected to immunoprecipitation with validated anti-SUMO1 or anti-SUMO2 antibodies (SUMO1 21C7 or SUMO2 8A2 Hybridoma Bank). Finally, the immunoprecipitated extracts were evaluated by Western-blot analysis with anti-p110 β antibody. As shown in Fig. 1c, we detected SUMOylated p110 β bands in the SUMO-immunoprecipitated extracts but not in the anti-IgG immunoprecipitated extracts, indicating that endogenous p110 β is SUMOylated in the cell without overexpression of the SUMO machinery. Altogether these results demonstrate that p110 β protein is modified by SUMO.

Lysine residue K952 in p110 β is a SUMO acceptor site

To map the SUMO acceptor lysine of p110 β , we constructed single mutants in two candidate lysines (K647 and K952) identified by the GPS-SUMO software as present within a type I SUMO consensus motif, and analyzed their capability to conjugate to SUMO *in vitro*. We did not observe differences in the SUMO1 conjugation between the WT and the K647R mutant p110 β proteins (Fig. 2a, upper panel). However, SUMO1 modification was reduced after mutation of lysine K952 to arginine (Fig. 2a, upper panel), thus identifying this residue, located at the activation loop of p110 β , as the main SUMO acceptor site. Similarly, a reduction in the *in vitro* SUMO2 modification of the K952R mutant relative to the WT p110 β protein was observed (Figure 2a, lower panel). To verify the involvement of this residue in p110 β SUMOylation, we evaluated the SUMOylation of WT and K952R p110 β proteins in cells. We did not observe modification of p110 β K952R with SUMO1 (Fig. 2b) or SUMO2 (Fig. 2c), confirming that lysine residue K952 in p110 β is a SUMO acceptor site.

The lysine residue that corresponds to K952 of p110 β is conserved in the activation loop of p110 α (Fig. 2d). In addition, residues 941 to 950 in p110 α are disordered [29], a feature that has been reported to facilitate SUMO conjugation [30]. Therefore, we hypothesized that p110 α could be also SUMOylated. To test this hypothesis, we carried out an *in vitro* SUMOylation assay in the presence of SUMO1 or SUMO2 using *in vitro* translated [³⁵S] methionine-labeled p110 α protein as a substrate. We observed that p110 α is SUMOylated by SUMO1 and SUMO2 *in vitro* (Fig. 2e), suggesting that p110 β is not the only PI3K catalytic subunit modulated by SUMO.

SUMO modulates the stability of p110 β

SUMOylation can have varied effects on the target protein including the regulation of its subcellular localization, stability, transcriptional activity and interaction with other proteins [31]. We first studied whether mutation of the SUMOylation site in p110 β altered its subcellular localization. Confocal analysis

of NIH-3T3 cells transfected with myc-tagged p110 β WT or K952R mutant plasmids revealed that both proteins, the WT and the SUMOylation mutant, were detected mainly at the cell cytoplasm (Fig. 3a). Similar results were observed after Western-blot analysis of nuclear and cytoplasm fractions obtained from cells transfected with myc-tagged p110 β WT or K952R mutant using anti-myc antibody (Fig. 3b, upper panel). In addition, we did not observe a change in the subcellular distribution of endogenous p110 β in HEK-293 cells, detected using a cell fractionation assay, after transfection of SUMO1 or SUMO2 (Fig. 3b, lower panel), suggesting that SUMO does not affect the subcellular localization of p110 β .

To determine whether SUMO had an impact on p110 β stability we first treated cells with two different concentrations of the SUMOylation inhibitor ML-792 and evaluated by Western-blot the levels of the endogenous p110 β protein. We observed a significant decrease in p110 β levels after treatment with 500 nM of the inhibitor (Fig. 3c, left panel). Similar trends were observed on p110 α , with a statistically significant reduction in protein levels at the highest dose of ML-792 (Fig. 3c, right panel). Therefore, we speculated that SUMO may modulate the stability of the protein. Cycloheximide chase experiments were then carried out to determine the stability of p110 β WT and K952R mutant. p110 β K952R exhibited a shorter half-life than the WT protein (Fig. 3d). These results suggest that SUMO conjugation stabilizes p110 β protein.

The p85 regulatory subunit interacts with and stabilizes p110 [8]. Therefore, we decided to evaluate whether this interaction is modulated by SUMO conjugation. HEK-293 cells were co-transfected with p110 β WT or K952R together with HA-p85 β WT or the p85 β SUMOylation mutant, and 48h after transfection p110 β -p85 β interaction was analyzed by co-immunoprecipitation. We did not observe differences in the interaction between p110 β WT or p110 β K952R, and p85 β WT or the p85 β SUMOylation mutant (Fig. 3e), which suggests that an alteration in the p110 β -p85 β interaction is not responsible for the stabilization of p110 β by SUMO.

Mutation of the SUMOylation site in p110 β has a negative impact on the activation of the PI3K pathway mediated by the protein

To determine the consequences of p110 β SUMOylation on signaling, NIH-3T3 cells transfected with pcDNA, p110 β WT or K952R were serum deprived for 16 h, and activation of the AKT pathway was analyzed by Western-blot with a specific anti-phospho Ser473-AKT antibody or with anti-phospho-FoxO-specific antibody. Overexpression of p110 β led to phosphorylation of FoxO and to AKT activation, as previously reported [3, 32]. The levels of phosphorylated AKT or FoxO proteins in cells expressing p110 β K952R protein were higher than in pcDNA transfected control cells but lower than those detected in p110 β WT transfected cells (Figure 4a, upper panel). Similar results were observed after analysis of phosphorylated AKT or FoxO proteins in both CEF and Rat-2 cells transfected with p110 β WT or the SUMOylation mutant (Fig. 4a, middle and lower panel, respectively), indicating that mutation of the SUMOylation site in p110 β has a negative impact on the activation of the PI3K pathway mediated by the protein.

p110 β induces oncogenic transformation in cell culture [3]. Therefore, we decided to analyze the transforming activity of the p110 β SUMOylation mutant. We generated NIH-3T3 cells that stably overexpress WT or the K952R mutant p110 β protein and tested their ability to form foci of transformed

cells. As shown in Fig. 4b, we detected focus formation in cells transfected with p110 β WT, as previously reported [3, 32]. However, p110 β K952R was not able to induce foci (Fig. 4b). In addition, we evaluated the formation of transformed foci of NIH-3T3 cells stably transfected with an empty vector or with p110 β K952R as the result of transduction with a lentivirus expressing p110 β WT. The number of transformed foci was similar in both conditions (Fig. 4c). These results suggest that SUMOylation of p110 β is required for cellular transformation. Finally, we also generated Rat-2 cells that stably overexpress p110 β WT or p110 β K952R protein and proliferation of the two cell lines was compared by the MTT assay. We observed that the cells expressing p110 β WT grew at a higher rate than the K952R mutant cells (Fig. 4d). Altogether, these results suggest that SUMO conjugation to p110 β positively modulates its oncogenic capability, its cell proliferation-promoting activity, and the PI3K downstream pathway activation.

The PI3K regulatory subunit p85 β negatively modulates the SUMOylation of p110 β

Our data indicate that SUMO conjugates to a lysine residue located within the second basic box of the activation loop of p110 β . Interaction of p85 nSH2 domain with p110 has been proposed to confine the activation loop into a collapsed conformation [33, 34]. Therefore, we speculated that p85 may modulate the accessibility of SUMO to conjugate to p110 β . To evaluate this possibility, we first carried out an *in vitro* SUMOylation assay with SUMO1 using *in vitro* translated [³⁵S] methionine-labeled p110 β protein as a substrate, in the presence or absence of *in vitro* translated p85 β protein. In addition, since p85 β protein can also be SUMOylated [17], and to discard that any alteration in the SUMOylation of p110 β is due to a competition for limiting amounts of SUMO in the reaction, we also carried out the experiment in the presence of a p85 β SUMOylation mutant. We observed a down-modulation of p110 β SUMOylation by p85 β and this effect was independent of the p85 β SUMOylation (Fig. 5a). To confirm these results, we studied the SUMOylation of p110 β in cells co-transfected with pcDNA, p85 β WT or the p85 β SUMOylation mutant. Co-expression of p85 β WT or p85 β SUMOylation mutant led to a reduction in p110 β SUMOylation (Fig. 5b, left panel). In addition, we also observed a reduction in the SUMOylation of p110 β after transfection of increasing amounts of the p85 β SUMOylation mutant (Fig. 5b, right panel). These results suggest that p85 β modulates the covalent interaction of SUMO with p110 β .

Discussion

Here we demonstrate that the PI3K catalytic subunit p110 β can be modified by SUMO *in vitro*, in transfected cells and under completely endogenous conditions, indicating that SUMOylation of p110 β is physiologically relevant. We identified lysine residue K952, located at the activation loop in p110 β , as the SUMO-acceptor site. The lysine residue that corresponds to K952 of p110 β is conserved in a disordered region of the activation loop of p110 α [29], suggesting that p110 α may be also SUMOylated. This hypothesis is supported by the detection of p110 α protein conjugated to SUMO by *in vitro* SUMOylation analysis.

SUMOylation can regulate the stability of the target proteins [31]. Here we show that the endogenous p110 β and p110 α protein levels decrease after SUMOylation inhibition and that mutation of the SUMOylation site in p110 β reduces the stability of the transfected protein but did not affect its interaction

with p85 β . These results are in agreement with the observed reduction in the levels of PI3K in response to knockdown of SUMO in microglia [35] and led us to propose that SUMO increases the stability of p110 β . The PI3K catalytic subunit p110 is thermally unstable and can be conformational stabilized by different mechanisms including the interaction with the regulatory subunit p85, the synthesis at low temperature or the linkage of an epitope tag to the N-terminus of the protein [8]. Based on these data, we propose that SUMO conjugation may stabilize the overall conformation of the catalytic subunit. Interestingly, SUMOylation has been recently suggested as a mechanism that might promote protein thermal stability during mitosis [36]. In agreement with the role of SUMO in the stability of p110 β protein, mutation of the SUMOylation site in p110 β reduced its ability to activate the AKT signaling pathway, promote cell proliferation and induce transformation of NIH-3T3 cells, suggesting that SUMO positively modulates the activity of the p110 β protein and that the full oncogenic activity of p110 β may require SUMO conjugation. Supporting this hypothesis, it has been previously reported that the proliferation of the prostate cancer cell line PC3 depends on both, the SUMO conjugating enzyme Ubc9 [19] and p110 β [37].

Interaction of p85 with p110 inhibits its activity [8]. Here we show that p85 also inhibits the SUMOylation of p110 β . Recently, Zhang et al. proposed that the interaction of the nSH2 motif of p85 with the second basic box (KRER) of the activation domain, confines the activation loop into a collapsed conformation, inhibiting p110 [33]. The restriction in the flexibility of the activation loop exerted by p85 may then explain the inhibition of the SUMOylation of p110 β mediated by p85 β .

In summary, we have identified SUMOylation as a new mechanism of regulation of p110 β . Our data indicate that SUMO conjugation to p110 β has a positive impact on the activation of the PI3K pathway likely through increasing its stability. These results together with previous data demonstrating that SUMOylation of p85 β negatively regulates PI3K [17], indicate that SUMO can positively or negatively modulate the signaling pathway, depending on the targeted protein, and suggest that the interaction of SUMO with each PI3K component must be tightly regulated. Moreover, the results shown here suggest that p85 may be one of these regulators. This information could be useful for designing PI3K inhibitors.

Declarations

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Competing interests

The authors have no relevant financial or non-financial interest to disclosure.

Ethics approval

Not applicable

Consent to participate

Not applicable

Consent for publication

Not applicable

Availability of data and material

All data generated or analysed during this study are included in this published article. The materials used in this study are available from the corresponding authors, CR, upon reasonable request.

Code availability

Not applicable

Authors contributions

AEM, CFC-H, SV, RS, YHB, and MB-M conducted the experiments; ME and MSR generated reagents; AV, MSR, ME, MC and CR analyzed the results; CR, designed the experiments and wrote the paper.

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Figure legends

Fig. 1 p110 β protein is modified by SUMO1 and SUMO2 in vivo and in vitro. **a** Modification of *in vitro* translated p110 β by SUMO1 or SUMO2. ³⁵S-methionine labeled *in vitro* translated p110 β was subjected to an *in vitro* SUMOylation assay in the presence of SUMO1 or SUMO2, as indicated. SUMOylated p110 β protein was then subjected to a deSUMOylation assay in the presence of SENP1. **b** SUMOylation of p110 β in transfected cells. HEK-293 cells were co-transfected with myc-p110 β together with pcDNA or Ubc9 and His6-SUMO1 or His6-SUMO2, and 48 h after transfection whole protein extracts (input) and histidine-tagged purified proteins were analyzed by Western-blot using the indicated antibodies. **c** SUMOylation of endogenous p110 β under denaturing conditions. PC3 cells were lysed under denaturing conditions, immunoprecipitated using anti-SUMO1 or anti-SUMO2 antibodies and analyzed by Western-blot with anti-p110 β antibody

Fig. 2 Lysine residue K952 in p110 β is the main SUMO conjugation site. **a** Modification of *in vitro* translated p110 β WT, p110 β K647R or p110 β K952R by SUMO1 or SUMO2 *in vitro*. ³⁵S-methionine-labeled *in vitro* translated WT or mutant p110 β proteins were subjected to an *in vitro* SUMOylation assay in the presence of SUMO1 (upper panel) or SUMO2 (lower panel). **b** SUMO1 modification of p110 β WT or p110 β K952R in transfected cells. HEK-293 cells were co-transfected with myc-p110 β or myc-p110 β K952R together with pcDNA or Ubc9 and His6-SUMO1, and 48 h after transfection whole protein extracts and histidine-tagged purified proteins were analyzed by Western-blot with the indicated antibodies. **c** SUMO2 modification of p110 β WT or p110 β K952R in transfected cells. HEK-293 cells were co-transfected with myc-p110 β or myc-p110 β K952R together with pcDNA or Ubc9 and His6-SUMO2, and 48 h after transfection whole protein extracts and histidine-tagged purified proteins were analyzed by Western-blot with the indicated antibodies. **d** Amino acid sequence homology between the activation loop of p110 α and p110 β . Lysine residue involved in SUMO conjugation to p110 β and the corresponding lysine residue in p110 α are labeled in bold. **e** Modification of *in vitro* translated p110 α by SUMO1 or SUMO2. ³⁵S-methionine labeled *in vitro* translated p110 α was subjected to an *in vitro* SUMOylation assay in the presence of SUMO1 or SUMO2, as indicated

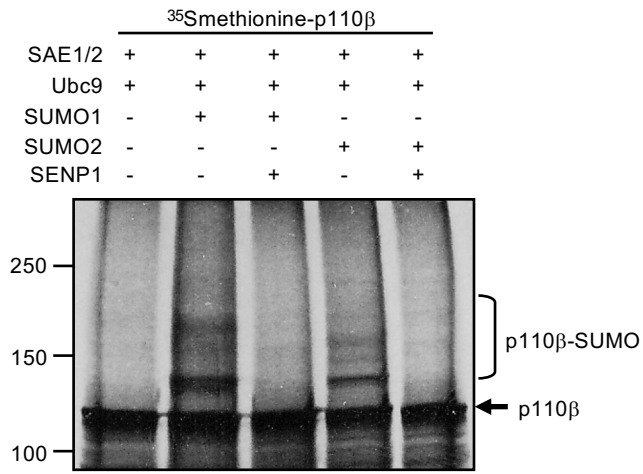
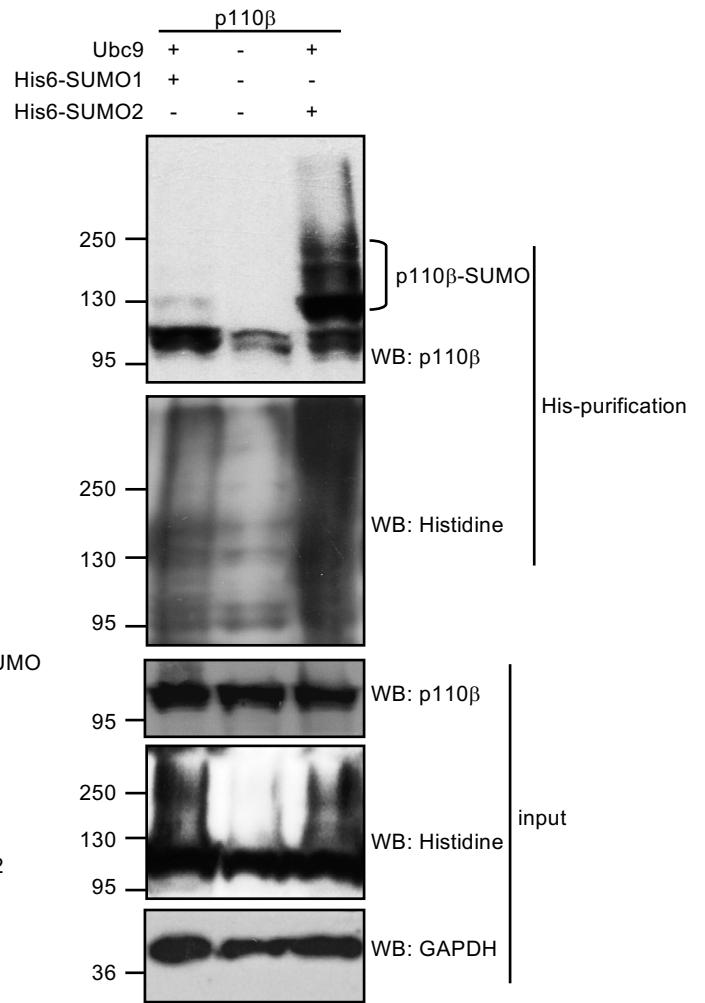
Fig. 3 SUMOylation of p110 β enhances its protein stability. **a** Subcellular localization of p110 β WT or p110 β K952R in NIH-3T3 cells. NIH-3T3 cells were transfected with p110 β WT or p110 β K952R. At 36 h after transfection, cells were stained with mouse anti-myc-tag antibody and Alexa-488 conjugated donkey anti-mouse secondary antibodies. Nuclei were visualized by DAPI staining (white). **b** Subcellular

localization of p110 β WT or p110 β K952R in HEK-293 cells (upper panel). HEK-293 cells were transfected with p110 β WT or p110 β K952R. At 48 h cells were subjected to subcellular fractionation and then analyzed by Western-blot with the indicated antibodies. Subcellular localization of endogenous p110 β protein in HEK-293 cells transfected with SUMO1 or SUMO2 (lower panel). Transfected cells were subjected to subcellular fractionation and then analyzed by Western-blot with the indicated antibodies. **c** PC3 cells were treated with the indicated concentrations of the SUMOylation inhibitor ML-792 and 12 h after treatment cells were harvested for Western-blot analysis with the indicated antibodies (upper panel). Quantification of endogenous p110 β or p110 α protein levels relative to GAPDH expression from at least three independent experiments is shown in the lower panel. Data are mean \pm SD of three experiments. Statistical analysis was assessed by one-way ANOVA followed by Dunnett's test. *, $P < 0.05$. **d** HEK-293 cells transfected with p110 β WT or p110 β K952R were incubated in the presence of cycloheximide (CHX). At the indicated times after CHX treatment protein extracts were analyzed by Western-blot analysis with the indicated antibodies (left panel). The stability of each protein (mean \pm SD) from three independent experiments is shown (right panel). *, $P < 0.05$, Student's paired t test. **e** Interaction between p110 β and p85 β is not altered by p110 β SUMOylation. HEK-293 cells were co-transfected with myc-p110 β WT or myc-p110 β K952R together with HA-p85 β WT or HA-p85 β K535RK592R, as indicated. At 48h after transfection, cells were lysed, immunoprecipitated using anti-HA antibodies and analyzed by Western-blot with anti-p110 β antibody

Fig. 4 Lysine residue K952 in p110 β is important for PI3K pathway activation and p110 β -induced transformation and proliferation. **a** Analysis of AKT or FoxO phosphorylation in NIH-3T3, CEF or Rat-2 cells expressing the indicated constructs and incubated for 16 h in serum free media. **b** Focus formation assay. NIH-3T3 cells stably expressing p110 β WT or p110 β K952R protein were plated and left to grow to confluence for 20 days when foci were stained and counted (upper panel). Data are mean \pm SD of triplicate samples for one experiment. Experiments were repeated twice. A representative image is shown (lower panel). **c** NIH-3T3 cells stably transfected with empty vector or p110 β K952R were transduced with lentivirus expressing empty vector or p110 β WT, plated and left to grow to confluence for 20 days when foci were stained and counted (upper panel). Data are mean \pm SD of triplicate samples for one experiment. A representative image is shown (lower panel). **d** Rat-2 cells stably expressing p110 β WT or p110 β K952R protein were plated in 96-well plates, incubated for the indicated times and assayed using the MTT assay (left panel). Data are mean \pm SD of triplicate samples. *, $P < 0.05$, **, $P < 0.005$, Student's paired t test

Fig. 5 Regulation of p110 β SUMOylation by p85 β . **a** p85 β negatively modulates the SUMOylation of p110 β *in vitro*. ³⁵S-methionine labeled *in vitro* translated p110 β protein was subjected to an *in vitro* SUMOylation assay with SUMO, in presence or absence of *in vitro* translated HA-tagged p85 β WT or p85 β SUMOylation mutant (p85 β K535RK592R) proteins. **b** HEK-293 cells were co-transfected with myc-p110 β and pcDNA or myc-p110 β , Ubc9, His6-SUMO2, and pcDNA, HA-p85 β WT or a HA-tagged p85 β SUMOylation mutant (HA-p85 β K535RK592R) (left panel). 48 h after transfection whole protein extracts and histidine-tagged purified proteins were analyzed using the indicated antibodies. HEK-293 cells were co-transfected with myc-p110 β together with Ubc9, His6-SUMO2, and increasing amounts of the HA-

tagged p85 β SUMOylation mutant (HA-p85 β K535RK592R) plasmid (right panel). 48 h after transfection whole protein extracts and histidine-tagged purified proteins were analyzed using the indicated antibodies

a**b****c**