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1	Empirical formulation for multiple groups of primary biological ice
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30 Abstract:

To resolve the various types of biological ice nuclei (IN) with atmospheric models, an 31 extension of the empirical parameterization (EP) (Phillips et al. 2008; 2013) is proposed to 32 predict the active ice nuclei from multiple groups of primary biological aerosol particles 33 34 (PBAPs). Our approach is to utilize coincident observations of PBAP sizes, concentrations, biological composition, and ice-nucleating ability. The parameterization organizes the PBAPs 35 into five basic groups: fungal spores, bacteria, pollen, viral particles, plant/animal detritus, 36 37 algae, and their respective fragments. This new biological component of the EP was 38 constructed by comparing predicted and observed concentrations of PBAP IN at the Amazon Tall Tower Observatory (ATTO) site located in the central Amazon. In addition to the 39 observations from the ATTO site, PBAP IN activity from the literature is used in constraining 40 the fitting parameters of EP. The parameterization has empirically derived dependencies on the 41 surface area of each group (except algae), and the effects of variability in their mean sizes and 42 number concentrations are represented via their influences on the surface area. The 43 44 concentration of active algal IN is estimated from literature-based measurements.

Predictions of this new biological component of the EP are consistent with previous laboratory and field observations not used in its construction. The EP scheme was implemented in a 0D parcel model. It confirms that biological IN account for most of the total IN activation at temperatures warmer than -20° C and at colder temperatures dust and soot become increasingly more important to ice nucleation.

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55 **1 Introduction**

Often referred to as bioaerosols, primary biological aerosol particles (PBAPs) are a 56 subset of biogenic particles emitted from the Earth's surface. They consist of biological 57 material such as viruses, algae, bacteria, fungal spores, pollen, plant and animal detritus, and 58 59 their fragments and secretions. In the ambient atmosphere, bioaerosols may be present in the form of clusters, single particles, and agglomerates and exist in a variety of shapes such as rods, 60 spirals, spherical, or spheroidal forms. A review article by Després et al. (2012) describes the 61 62 typical concentrations of these PBAPs in the ambient air, their sizes, and sampling techniques. Bioaerosols can act as ice nuclei (IN) to initiate ice at temperatures often warmer than -12° C, 63 well above the homogeneous freezing point (Prenni et al. 2009; DeMott and Prenni, 2010; 64 Morris et al. 2012; Fröhlich-Nowoisky et al. 2016). To maintain consistency with previous EP 65 papers (e.g. for mathematical symbols), we will use the term, 'IN' rather than 'ice nucleating 66 particles' (INPs, Vali et al. 2015) for this paper. They can also act as giant cloud condensation 67 nuclei (CCN) (Barahona et al. 2010; Delort et al. 2010) and may play an important role in the 68 69 hydrological cycle. Bioaerosols may influence the formation of precipitation, especially in 70 pristine air over densely vegetated areas (Phillips et al. 2009; Pöschl et al. 2010; Huffman et 71 al. 2013; Prenni et al. 2013).

72 In warm-based convective clouds, rain production can occur through collisioncoalescence that facilitates the heterogeneous raindrop freezing to form graupel ('warm rain 73 74 process'). Heterogeneous nucleation of ice crystals and their subsequent growth to form snow 75 and graupel particles ('ice crystal process') plays an essential role in rain formation in convective clouds with higher clouds bases (e.g. Rogers and Yau, 1989; Pruppacher and Klett, 76 1997). The role of bioaerosols in altering cloud microphysical properties, precipitation, and 77 78 ecosystem interactions has been recognized for many years and is a topic of increasing interest (e.g. Pöschl et al. 2010; Huffman et al. 2013; Amato et al. 2015). The PBAPs can play an 79

important role in altering the cloud microphysical processes and precipitation formation
through warm rain processes. However, large uncertainty still exists regarding their potential
to facilitate precipitation, particularly their impact on global or on regional scales (Hoose et al.
2010a; Morris et al. 2011).

84 As reviewed in Després et al. (2012), several previously published studies have reported 85 ice nucleation activities of various biological species, including fungi, pollen, algae, and bacteria. Among various PBAP species, higher ice nucleation activity is observed for bacterial 86 and fungal particles in clouds. The quantitative measurements of biological IN in the 87 atmosphere are rare. Biological IN in the atmosphere exhibit strong temporal and spatial 88 variations (Christner et al. 2008; Pouzet et al. 2017). For a better understanding of their impact 89 on clouds, more quantitative measurements of biological IN in the Earth's atmosphere are 90 required. 91

With the limited observations of biological IN, numerical models can serve as an 92 93 important tool for investigating their effect on cloud microphysical processes and hence on precipitation. Using a 1.5-dimensional cloud model, Levin et al. (1987) studied the efficiency 94 of cloud seeding with bacteria. Ariya et al. (2009) showed that biological IN could trigger cloud 95 glaciation through ice multiplication processes. Diehl and Mitra (2015) implemented PBAPs 96 97 (pollen and bacteria) in a parcel model using the ice-active mass site density approach (Kanji 98 et al. 2017 and references therein) and found that they were not involved in significant ice formation. Hummel et al. (2018) implemented PBAPs in the regional atmospheric model. They 99 showed that although PBAPs were very efficient IN at high subzero temperature, due to their 100 101 low atmospheric concentration, it is difficult to quantify their impact on cloud properties. However, they also showed that PBAP can influence the cloud ice phase in the clouds with 102 cloud temperatures below -15 °C in the absence of any other IN. Hiron and Flossmann (2015) 103 studied the potential impact of the presence of bacteria as an extremely IN active PBAP on 104

cloud and rain development. They noticed a significant increase in the rainfall flux in the 105 simulation with only bacteria as IN. However, in the simulations where all other IN modes 106 107 were also forming ice, their influence on rainfall became negligible. Phillips et al. (2009) showed that an increase in PBAP resulted in more numerous cloud droplets through their CCN 108 activity and reduced the precipitation production via warm rain process, boosting cloud lifetime 109 and extent. In the case of higher bioaerosol, the extra heterogeneous ice from biological IN 110 111 particles can only partially counteract that effect, indicating the CCN activity of PBAPs is more important than their IN activity for the simulated warm-based convection. Several studies have 112 113 used regional as well as global models to understand the geographical variation and climatic importance of bioaerosols (e.g. Burrows et al. 2009a, b; Hoose et al. 2010a; Sesartic et al. 114 2012). The empirical data included in these models are highly uncertain. Most of the 115 parameterizations of heterogeneous ice nucleation of biological particles are based on 116 laboratory measurements or derived from theory. 117

118 Consequently, the cloud-resolving numerical models need to be constrained and confirmed by observational data of biological IN to have a better estimation of their effect on 119 clouds. Using a 'bottom-up approach' (i.e. measurements of ice nucleation activity of 120 121 biological species in the laboratory to represent atmospheric ice initiation) in the numerical models can introduce significant bias in the active IN concentrations because the particular 122 123 biological species studied in the laboratory may not necessarily be a good representative of the corresponding wider PBAP population in the atmosphere. Instead, there is a need for a better 124 parameterization of heterogeneous ice nucleation of biological IN that accounts for diversity 125 in their physical and chemical characteristics in the atmosphere. We have followed a 'top-down' 126 approach in the current study to resolve the ice nucleation activity of multiple groups of PBAPs 127 collected directly from the atmosphere. The empirical parameterization (EP) for heterogeneous 128 ice nucleation presented here is based on the active surface site density approach (e.g. Phillips 129

et al. 2008; Niemand et al. 2012; Kanji et al. 2017 and references therein). The assumption of
active surface site density has some limitation on consistently describing freezing curves over
a wide range of surface area as described in Beydoun et al. (2016). They suggested an
alternative approach invoking a continuum assumption on the ice nucleating activity (contact
angle) of an aerosol particle's surface which does not require any assumptions about the size
or number of active sites.

The first empirical ice nucleation parameterization for biological particles was 136 proposed by Phillips et al. (2008) where biological particles were assigned as a part of insoluble 137 organic particles. An estimation of the number of active IN consisting of insoluble aerosol 138 species assumed that it was approximately proportional to the total surface area of its aerosol 139 particles. In the subsequent study, Phillips et al. (2013) revised the original EP scheme by 140 replacing insoluble aerosol species by the new group specifically for PBAP IN. However, their 141 scheme does not address the separate activity of various groups of PBAPs with different ice-142 143 nucleating abilities.

144 In the present study, a framework for parameterizing the heterogeneous ice nucleation 145 of biological particles is formulated and accounts for contributions from various groups of 146 PBAPs including

147 1) Fungal spores and associated fragments;

148 2) Bacteria and their fragments;

- 149 3) Pollen and their fragments;
- 150 4) Viral, plant/animal detritus;

151 5) Algae.

Matthias-Maser and Jaenicke (1995) first described these groups of PBAPs through
observations. The EP's treatment applies to deposition and condensation-/immersion-freezing
modes.

The proposed scheme is empirical, mostly based on coincident observations of PBAP size distributions and active biological IN sampled from the atmosphere over the Amazon Rainforest with abundant PBAPs. Prenni et al. (2009) have revealed that over the Amazon Rainforest the contribution of local biological particles to ice nucleation is increased at warmer atmospheric temperatures, whereas the contribution of dust particles is increased at colder temperatures, making this region ideal for biological IN measurements.

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2 Material and methods

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165 **2.1 Measurement site**

The samples for the present study were collected over the central Amazon basin at the 166 Amazon Tall Tower Observatory (ATTO; Andreae et al. 2015). The ATTO site is in the middle 167 of the Amazon Rainforest in northern Brazil (see Fig. 1), about 150 km northeast of Manaus 168 (120 m above sea level) with coordinates S 2° 08' 45.13" W 59° 00' 20.12". The measurement 169 facilities at the ATTO site consists of a 325 m tall tower and two 80 m towers, one of which is 170 a triangular mast currently used for a wide set of aerosol measurements (Andreae et al. 2015). 171 The immediate surroundings as well as the ATTO footprint region are covered by an old-172 growth undisturbed forest with minimal anthropogenic impact. 173

Meteorological conditions vary throughout the year with a wet (February–May), a 174 dry (August-November), and a transition from wet to dry season (December-January) 175 (Pöhlker et al. 2019). This classification of seasons is consistent with seasonality of 176 atmospheric pollution, rainfall, and air mass origin. Over the Amazon rainforest the PBAPs 177 are ubiquitous throughout the year (Martin et al., 2010). During the wet season, the atmospheric 178 boundary layer over the Amazon is relatively clean with low aerosol number concentrations 179 180 (Artaxo et al. 2002; Martin et al. 2010; Zhou et al. 2002) and the hydrological cycle is strongly influenced by biogenic aerosols. However, during the Amazonian dry season, the atmospheric 181 182 aerosol concentrations are considerably higher due to biomass burning (Andreae et al., 1988; Artaxo et al., 2002; Pöhlker et al. 2018; Holanda et al. 2019) and mixed-pollution scenarios 183 (Pöhlker et al. 2018). A detailed description of the aerosol size distribution and the CCN 184 characteristics at the ATTO site are given by Pöhlker et al. (2016, 2018). Additional details 185 on the diurnal temperature and wind variations, precipitation amount, and aerosol 186 characteristics at the ATTO site can be found in Andrea et al. (2015). 187

Sampling was performed during the dry season in the Amazon between 5 and 18 188 September 2018. The sample collection was interrupted on a few days including September 9 189 190 and 15 due to damage to power supply, so sample collection was only possible for the twelve days mentioned in Figure 5a. As shown in Fig. 1, the sampling took place at the triangular mast 191 192 site. The sampled air was carried through a stainless-steel tubing with a diameter of 0.75 inches to the aerosol sampling unit installed in a laboratory container located at the base of a triangular 193 mast. A Total Suspended Particles (TSP) inlet was installed on a triangular mast at 60 m above 194 ground level to ensure that the air was sampled above the canopy, which has an average height 195 of ~23 meters. To avoid condensation in the filter holder, the sampled air from the TSP inlet 196 was dried to a relative humidity < 40% using silica gel driers. 197

The inlet used for sample collection in the current study has been optimized to achieve 198 a maximum sampling efficiency (Moran-Zuloaga et al. 2018), however, a certain amount of 199 particle loss is unavoidable. The particles smaller than 100 nm are more prone to diffusive 200 losses and not relevant for the current study. Daniel Moran-Zuloaga et al. (2018) documented 201 potential losses in the accumulation and coarse mode particles of aerosol particles in this inlet 202 system which are more relevant to the current study. They calculated the transmission 203 204 efficiencies of the PBAPs for the inlet used in the current by using standard density of 1 g/cm^3 as best guess for typical PBAP densities. The aerosol transmission efficiency starts at about 205 206 100% (full transmission) for particles with diameter around 0.7 µm and then declines with increasing particle size. Based on their calculation typical transmission efficiencies for PBAPs 207 with diameter 1, 2, 4, 8, and 10 µm were 98, 97, 91, 72, and 55% respectively. Correction to 208 209 PBAP size distributions was applied accordingly.

210 A detailed analysis about the back trajectories arriving at the ATTO site is presented in Moran-Zuloaga et al. (2018) and Pöhlker et al. (2019). They showed that the ATTO-relevant 211 back trajectories followed a seasonal swing between a Northeast path during the wet season 212 and a Southeast path during the dry season as a response to the annual north-south Intertropical 213 214 Convergence Zone migration. Figure 1 shows the seven-day analysis from the Hybrid Single-Particle Lagrangian Integrated Trajectory Model (Draxler and Rolph, 2003) on three sampling 215 216 days (11-13 September 2018) at 60 m above ground. Back trajectories revealed that the anthropogenic emissions from Manaus did not significantly impact the sampled air mass, 217 however, the atmospheric conditions at the ATTO site during our sampling period were not 218 pristine. 219

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221 **2.2** Aerosol measurements and sample collection

222 2.2.1 Aerosol size distribution measurements

The aerosol size distribution and number concentration at the ATTO site was measured 223 224 by a Scanning Mobility Particle Sizer (SMPS; TSI Inc) and an Aerodynamic Particle Sizer (APS; TSI Inc). The SMPS measures the electrical mobility diameter (10-430 nm), while the 225 APS gives aerodynamic diameters (0.5-20 µm) of the particles. There were frequent 226 227 interruptions in the aerosol measurements during our observational period due to severe lightning strikes and associated damage to the power supply and installation. Therefore, 228 measurements of the aerosol size distribution from SMPS and APS was available only for a 229 few days. 230

231 2.2.2 Aerosol collection by filter method

232 Table 1 describes the sampling type, duration, flow rate, purpose, and corresponding instrument in which the samples were analyzed. The aerosol samples were collected from 233 September 5-18, 2018, for a twelve-day period at the ATTO site. The collection of aerosol 234 samples for the drop freezing tests and microscopic analysis was carried out via the TSP inlet 235 through a four-stage stacked filter unit. Four Polytetrafluoroethylene (PTFE) membrane filter 236 237 papers (25 mm diameter) (Sartorius Stedim Biotech) with the pore sizes of 5 (outermost), 1.2, 0.45, and 0.2 (innermost) µm were used for the sampling. The filters were placed in an airtight 238 aluminum filter holder in order of decreasing pore size. Samples were collected with a vacuum 239 pump (VWR vacuum gas PM20405-86). The flow rate was controlled using a flow meter 240 (Kytola instruments) equipped with a needle valve. Collection efficiencies of PTFE membrane 241 filters of pore size 5 and 1.2 µm are much higher compared to Millipore and glass filters of 242 243 similar pore sizes (Soo et al., 2016) and thus inevitably most of the aerosol particles are collected on the first two filters. 244

Cascade filter samples were collected with an average volumetric sample flow rate of 4 Lpm (Liters per minute) for approximately 4 hours each day. In addition, a set of cascade filters with a sample flow rate of 1.5 Lpm was collected for about 16 hours. A set of aerosol samples with a sample volume of about 200 L was collected using only a 0.2 µm filter to estimate IN concentrations activated by the immersion-freezing mode in Frankfurt Ice Nuclei Deposition Experiment (FRIDGE-IMM) chamber.

Samples were sealed carefully in a sterile container and stored at 4°C at the ATTO site. 251 The aerosol samples when stored for prolonged time under such conditions, can show changes 252 in their ice nucleation activity. Stopelli et al. (2014) found a degradation in the active IN 253 concentration by a factor of two at temperature between -8 to -12° C in snow water samples 254 stored at 4°C for 30 days. After the campaign was completed, the samples were transported to 255 the University of Sao Paulo and later to Lund (Sweden). The samples were stored in a deep 256 freezer at -25° C at Lund until they were analyzed by scanning electron microscopy (SEM) and 257 258 drop freezing tests.

259 2.2.3 Aerosol collection by Programmable Electrostatic Aerosol collector (PEAC7)

In addition to the filter method, aerosol sampling was performed using a Programmable Electrostatic Aerosol collector (PEAC7) (Schrod et al. 2016). The PEAC samples were analyzed to estimate IN activated by deposition and condensation-freezing mode in the FRIDGE-STD chamber. A total of twelve samples were collected using PEAC during the field campaign held from September 5-18, 2018 (see Figure 5).

In addition, PEAC samples were used for imaging PBAPs under SEM. On each sampling day, aerosol particles were collected at a volumetric flow rate of 2 Lpm using the PEAC7 for 50 minutes. The PEAC7 unit used in the current analysis is based on the single wafer electrostatic aerosol collector described in Klein et al. (2010). In PEAC7, aerosol

particles are introduced through a central inlet into the field of electrons emitted by 12 gold
electrodes. The aerosols are negatively charged and carried to the positively charged silicon
(Si) wafers (Wacker-Siltronic, thickness: 0.7 mm, diameter: 45 mm) where they are deposited
on the surface (Klein et al. 2010).

We have applied the correction to the IN concentrations and size distribution of bioaerosol particles by assuming the PEAC7 collection efficiency of 60% observed by Schrod et al. (2016). Klein et al. (2010) and Schrod et al. (2016) provide additional details on the design and working principle of PEAC7.

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278 2.3 Scanning electron microscopy analysis

279 Primary bioaerosol particles were analyzed by using a variable pressure Tescan Mira3 High-Resolution Schottky FE-SEM located in the Geology department at Lund University. It 280 281 should be noted that the SEM analysis was limited to three days: 9 (night-time; hereafter 0911N), 12 (day-time; hereafter 0912D) and 13 (day-time; hereafter 0913D) September, as an 282 intensive collection of SEM images characterizing the size distributions for all the samples was 283 not practical. During each of the three days, the size distribution and IN number were estimated 284 for total biological aerosols (5 + 1.2 μ m filters) and fractions obtained on 5 and 1.2 μ m filters. 285 Thus, nine pairs of measurements of active IN and size distributions were available from three 286 intensive sampling days mentioned above and will be referred to as the nine 'cases' in the 287 remaining part of the manuscript. 288

To reduce charging under the electron beam, the samples were coated with a thin layer (10 nm) of Palladium, sputter-deposited prior to the SEM analysis. The coated samples were examined at a beam size of 15 keV with a working distance of 10 mm. The working distance

was lowered (between 5 and 10 mm) for Si wafer samples to obtain clearer images of particles with diameter (*D*) < 1.5 μ m.

294 To quantify the PBAPs, the main method of identification was purely based on the morphological characteristics, following the method of Matthias-Maser and Jaenicke (1991, 295 1994). The organic nature of particles from energy-dispersive X-ray spectrometer (EDS) 296 analysis was confirmed if (C+O) > 75% and 1% < P, K, Cl < 10% (Coz et al. 2010). The SEM 297 system used in the current study was equipped with an energy-dispersive X-ray spectrometer 298 (EDS, X-Max^N 80, Oxford instruments, 124 eV, 80 mm²). The EDS could detect the 299 composition of individual particles semi-quantitatively for elements with an atomic number 300 greater than 6 including light elements such as C, N, O. Errors in measurements of the 301 composition of light elements by EDS vary for individual samples and are below 2 percent 302 overall. The EDS was performed at a beam intensity of 15 keV with a working distance of 10 303 304 mm. The Si and F peaks in the EDS spectra were not considered for particles from the Si wafer 305 and PTFE filter respectively, because of the interference from the substrate. The duration of each EDS collection was limited to 10-12 seconds to avoid particle damage. Because other 306 elements like S, Ca, or Si can also be tracers of PBAPs, this analysis should be treated as a 307 conservative estimate. 308

Before imaging the individual biological particles, wide reference areas each 100×100 µm in size were chosen randomly and corresponding images of those areas were saved as reference pictures along with their positions on the filter paper. Particles located in these predefined reference spots were counted, and high-resolution images were acquired for those particles. About five particles were observed roughly at each predefined reference spots on the filter samples. The maximum dimension of particles was recorded and saved. In this study the maximum dimension is treated as the diameter of the particle.

Based on the number of PBAPs detected in the scanned filter area, the bioaerosol 316 number was upscaled to the whole filter area. The size distribution of the PBAPs was classified 317 into 40 predefined size bins covering a size range from 0.2-40 µm. The measured number was 318 converted to an airborne concentration (per m³ of air) based on the sampled air volume. The 319 recorded data was then used to classify the particles into the basic PBAP groups: fungal spores, 320 bacteria, pollen, viral particles, plant/animal detritus, algae, and their respective fragments. The 321 322 primary classification of PBAPs into different groups was based on similarity in morphological characteristics with those reported in earlier studies. Despite all possible efforts to categorize 323 324 the identified PBAPs, not all of them could be attributed to any PBAP group. A small fraction of the PBAPs (~ 5%) which did not find any similarity in morphology with the PBAP groups 325 considered here were classified as 'unidentified' PBAPs. These unidentified particles were not 326 included in the size distributions of PBAPs presented here. 327

Since the emphasis of the current study was to characterize the size distribution of PBAPs, non-biological particles were excluded from the study. In addition to filter samples, Si wafers were analyzed under SEM using similar methodology. Additional details about SEM analysis including scanned area and number of bioaerosol images collected are given in Table 2. More than 0.1% of the total filter area was investigated under SEM and > 200 images were collected for each sampling day considered for SEM analysis. For Si wafers, more than 0.06% of the total area was scanned under SEM and ~100 images of PBAPs were collected.

Identification and counting of particles with $D < 1.5 \,\mu\text{m}$ (e.g. bacteria) on filter samples was challenging as they were often hidden inside the dense membrane of filter material. Therefore, the size distribution of biological particles with $D < 1.5 \,\mu\text{m}$ were determined mostly by SEM imaging of Si wafers. This was justified as the size distributions from the Si wafer and corresponding filter were in good agreement within the overlapping region between 1.5 and 5 μm . The collection efficiency of the 5 μm filter for various particle diameters was estimated

based on total aerosol number concentrations from the SMPS, APS and the size distribution of 341 total particles collected on the 1.2 µm filter. The SMPS and APS aerosol size distributions used 342 here were from 0914, a typical sampling day at the ATTO site. Based on the collection 343 efficiency of the 5 µm filter, the size distributions of biological particles on 5 and 1.2 µm filters 344 were estimated separately. The estimated collection efficiency for particles with a diameter 345 between 0.3-6 µm was variable between 0.81 and 0.99 whereas it was unity for other size 346 347 ranges. The collection efficiency of 1.2 µm filters was not measured and assumed to be unity (for particles with $D > 0.1 \,\mu$ m) based on measurements by Soo et al. (2016). 348

After acquiring the images of bioaerosols from SEM analysis, we classified them intocommon PBAP groups based on size and morphology.

351

352 2.4 Flow cytometry analysis

353 In addition to SEM, we conducted flow cytometry (FCM) analysis to estimate the total bioaerosol number concentration. FCM is widely used to detect and measure the physical and 354 chemical characteristics of cells or particles. This is a fluorescent technique, in which cells are 355 356 often labeled with fluorescent markers so that light is absorbed and then emitted in a wavelength band. For each sample extract, cell counts were performed on 450 µL triplicates 357 added with 50 µL 5% glutaraldehyde (stored at 4°C). For analysis, samples extracts were 358 stained with SYBRGreen I (Molecular Probes Inc.). This stains nucleic acids, and thus reveals 359 all DNA-containing particulate matter (e.g. viruses, bacteria, archaea, fungi, pollen, plant and 360 animal cells). The parameters used with FCM did not detect chlorophyll, proteins, and 361 carotenoids. The wavelengths for excitation and emission were 488 and 530 nm, respectively. 362 The FCM counts were performed for 3 minutes. The flow cytometry technique used in the 363 current analysis can detect particles as small as 0.3 µm (Lippé, 2018). 364

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2.5 Measurement of Ice nucleating particles

367 IN in the air were measured using the FRIDGE chamber of Goethe University, Frankfurt. FRIDGE operates in two different modes: a) as a low pressure diffusion chamber 368 (here called FRIDGE-STD) for the detection of deposition/condensation freezing IN collected 369 onto Si wafers (see 2.2.3), and b) as a cold stage freezing array (here called FRIDGE-IMM) 370 for the measurement of immersion freezing IN in extracts of filter samples (see 2.2.2). 371 372 Additional details have been described in previous studies (Klein et al., 2010; Schrod et al., 2016; DeMott et al., 2018, 2018a). 373

2.5.1 Measurement of IN activated by deposition and condensation freeing mode using 374 375 FRIDGE-STD

The FRIDGE-STD method applies a static low-pressure diffusion chamber for 376 377 activation, growth and counting of IN activated at temperatures between 0 and -30°C. For analysis, a Si wafer substrate that was previously laden with electrostatically precipitated 378 aerosol particles (using PEAC-7) was placed on the cold stage in the chamber. The chamber 379 380 was then closed, evacuated, and cooled to the desired temperature. The surface temperature was measured using a PT-1000 sensor. In the next step, water vapour was introduced into the 381 chamber by opening the connection to the vapour source. The supersaturation with respect to 382 ice over the Si wafer was then estimated as the ratio of vapour pressure in the chamber over 383 saturation vapour pressure at the surface temperature. The number of activated ice crystals 384 grown was counted using automatic picture analysis software. All the twelve samples collected 385 386 on Si wafer by PEAC-7 were processed for IN activation in the FRIDGE-STD mode (see Figures 5a-c). 387

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2.5.2 Measurement of IN activated by immersion freezing mode using FRIDGE-IMM:

The number concentrations of IN in the air activated by immersion freezing mode were 389 measured on the cold stage of FRIDGE-IMM with the drop freezing method of Vali (1971), as 390 391 described by Ardon-Dryer and Levin (2014) and DeMott et al. (2018). The aerosol samples collected on 0.2 µm filters were used for the drop freezing experiment. Each filter was placed 392 in 5 ml of deionized water and stirred vigorously for a few minutes to create particle 393 suspensions. Around 60 droplets, each with a volume of 2.5 µL were pipetted onto on a clean, 394 395 salinized Si wafer, with the chamber almost closed, but at ambient atmospheric pressure. The temperature of the cold stage was lowered by 1°C min⁻¹. A steady flow of synthetic air of 1 L 396 min⁻¹ through the chamber to avoid the condensation and rime effects. The number of drops 397 that froze as function of temperature was recorded by the charge-coupled device (CCD) 398 camera. Two runs of drop freezing experiments were conducted for each sample. In FRIDGE-399 400 IMM, eleven filter samples were treated to measure the IN concentrations activated by immersion freezing mode (see Figure 5d). 401

402 2.5.3 Quantification of biological IN

The concentration of biological IN in the air, active between $-5^{\circ}C$ and $-15^{\circ}C$, was 403 determined by conducting the drop freezing experiments in the LED-based Ice Nucleation 404 Detection Apparatus (LINDA) LINDA chamber as described below. The detailed technical 405 information and working principle of LINDA are described by Stopelli et al. (2014) and 406 407 summarized here. The LINDA device has an array (7×8) of red LEDs (Light Emitting Diodes; 645 nm wavelength). This array is mounted on a printed circuit board casted into a 408 polycarbonate housing and submersed in a cold bath. A liquid sample is distributed in a total 409 410 of 52 sample tubes (0.5 mL Eppendorf). These tubes are held in another polycarbonate plate and placed onto the LED array so that each tube is located above one LED. The temperature is 411 412 recorded by four sample tubes with cast-in Pt-1000 temperature sensors placed in the corner positions of the tube holder. 413

Before starting the freezing experiment, 75% of each filter sample was placed in 30 mL 414 of deionized water to create suspensions, of which 20 ml was used for the freezing experiment 415 in LINDA. The remaining 25% of the filter was saved and stored in a petri dish for SEM 416 analysis. The filter samples were cut using sterilized scalpel. To make the LINDA sample 417 extract clearer to facilitate the imaging, 2 ml of NaCl (in liquid form, 0.8% w/v) was added to 418 the solution. Adding NaCl to the LINDA sample extracts should not affect its freezing 419 420 characteristics since no freezing event was noticed for the solution containing deionized water and NaCl. The remaining 10 ml solution was saved and stored in a refrigerator (at 4°C) for 421 422 subsequent flow cytometry.

The sample extract is distributed in a total of 52 sample tubes each of which contain 423 400 μ L of the sample extract. The samples were analyzed at the cooling rate of 0.4 °C min⁻¹. 424 Images of the tubes are recorded every six seconds using a Monochrome camera, during which 425 temperature was changed by 0.04°C. For each filter sample extract, two sets of freezing 426 experiments were carried out. In the first set of experiments, sample extracts were untreated 427 (no heating) for total IN concentration measurements. In the second set, the samples were 428 heated to 95°C for 10 minutes to denature the IN activity of biological IN following Christner 429 430 et al. (2008) and Garcia et al. (2012). The concentrations of active biological IN were calculated as the difference between the concentrations of active IN measured with and without heating. 431 432 However, it should be noted that heat treatment may not inactivate every ice nuclei site of biological material and hence the values presented should be considered as the lowest possible 433 estimates of biological IN. 434

Previous findings based on heat treatment (e.g. Garcia et al. 2012; Joly et al. 2014) showed that the relative contribution of biological IN decreases with temperature, with 97% to 100% of total IN active at -8° C and 77% at -12° C as other particles including dust, become active at colder temperatures. Hill et al. (2014) showed that heat treatment (at 95°C) of bacterial 439 samples obtained by leaf washing reduced the ice nucleation activity by >75% at -10° C. 440 According to Conen et al. (2011), heating to 100° C reduced IN active at -12° C by 70–98%. 441 O'Sullivan et al. (2015) showed that heat treatment (90°C) of soil samples containing organic 442 decomposing matter reduced ice nucleation activity at -12° C to $\sim 1/10$.

In addition, the concentration of active biological IN at temperatures colder than -15 °C was estimated for the sample extract of 0911 (from section 2.5.1) based on heating tests (95°C for 10 minutes) in FRIDGE-IMM. The methodology of drop freezing experiment is already described in section 2.5.1.

447

448 2.6 Classification of PBAPs based on SEM images

After collecting the bioaerosol images, we classified them into common PBAP groups based on size and morphology. Table 3 presents the general size and morphological characteristics of different PBAP groups, based on the literature available. These characteristics form the basis for classifying PBAPs SEM images into basic groups. Typical images of the bioaerosols collected from each group during this study are shown in Fig. 2.

Fungal spores and associated fragments (group 1) were one of the most common classes 454 of bioaerosols observed during the campaign. Fungi can grow in almost every ecosystem and 455 456 are efficient in aerosolization (Adhikari et al., 2009; Löbs et al. 2020). The morphological characteristics of fungal spores have been well documented in global air and are summarized 457 in Table 3 (Després et al. 2012 and references therein; Shi et al. 2003; Tamer Vestlund et al. 458 459 2014; Valsan et al. 2015; Valsan et al. 2016; Priyamvada et al. 2017; Wu et al. 2019, Li et al. 2020). They appear in various forms including globular, sub-globular, elliptical, and elongated. 460 Many of the spores possess the attachment scar and appear either with smooth or ornamented 461 surfaces. The size range of particles from SEM images considered for classification of PBAPs 462

in this group is generally from 1 to 20 μm. The SEM images of fungal spores collected over
Amazon rainforest (e.g. Huffman et al. 2012, Cybelli et al. 2018, China et al. 2016) were also
referred while conducting the classification.

The images of bacteria (group 2) on Si wafer were clearer and were used for the size 466 distribution. The basic morphological features of bacterial particles are spheres (coccus) and 467 468 round-ended cylinders (bacillus). In addition, there may be few others with helically twisted shapes. Some bacterial cells grow separately while some rods or cocci grow in chains, grapelike 469 clusters or form a cubic packet characteristically. The surface structures include flagella, pili, 470 and capsules. A few representative images of bacillus and cocci shaped bacteria from the 471 ATTO samples are shown in Fig. 2. We attempted to grow the bacteria from filter samples in 472 agar solution, but no bacterial growth was observed. It should be noted that the filter samples 473 were not immediately analyzed for bacterial growth and were stored in a freezer at -25° C after 474 sampling, which could have affected the number of viable bacteria. 475

476 The pollen grains (group 3) were broadly distinguished from fungal spores based on size and morphology. Typically, its size ranges from 10 to 100 µm. PBAPs are classified into 477 pollen based on the general morphological characteristics of pollen grains given in table 3. 478 Cybelli et al. (2018) provided an extensive image database of pollen grains collected at the 479 ATTO site and was frequently used for classification. From our SEM imaging, the size range 480 481 of the pollen grains varied between 8 and 20 μ m. The pollen grains with a diameter of 10 to 30 um were frequently observed at the ATTO site and the large pollen grains were absent high 482 above the canopy except during some thunderstorm events (Cybelli et al. 2018). Accordingly, 483 484 our SEM analysis did not show pollen grain presence greater than 20 µm. Observed pollen grains exhibited different shapes including ellipsoid, fusiform, globose/spheroidal. 485

For a given scanned area under SEM, 2-4 brochosome clusters have been detected on 486 the filter for each case and are assigned to group 4. (viral, plant/animal detritus). Each cluster 487 consists of 3 to 40 brochosomes with an individual diameter of about 0.45 µm. 488 Brochosomes are intricately structured microscopic granules secreted by leafhoppers that are 489 typically found on their body surface and more rarely, eggs (Wittmaack, 2005). The group 4 490 was dominated by the presence of leaf litter followed by epicuticular wax of plants, insect 491 492 scales, and other vegetation detritus. The viral particles were hardly accessible in our SEM analysis. 493

Somewhat surprisingly, during a rather large number of SEM investigations, no particle 494 could be identified as algae (group 5; Sect. 1) and therefore algae are not shown in the size 495 distribution presented here. A near-by water source close to the ATTO site had no measurable 496 impact on algal aerosol number concentration. However, algae can grow on leaves and soil. 497 They could also be part of the long-range transport of marine particles from the Atlantic Ocean 498 into the Amazon Rainforest (Martin et al. 2010; Moran-Zuloaga et al. 2018). Backward 499 trajectories (at 2 km) using the HYSPLIT model (see Fig. 1) suggest that a marine air mass 500 from the Atlantic Ocean would have travelled over 5-6 days to reach the ATTO site. In 501 502 summary, the absence of algae in our SEM analysis can be attributed to their low concentration in the air at the ATTO site and possible limitation in identifying them using SEM. 503

The identification and classification of fragments of various PBAP is challenging due their complex and irregular shape. Consideration was given to the external features of the background PBAPs on the filter paper that were easy to detach from main PBAP particle. On this basis, the fragments in the form of external spines, hyphae, etc. are then assigned to the parent particle. In addition, the characteristics of the cell wall and other visible features of the fragments have been closely examined to identify its parent PBAP with matching features. The

510	fragments smaller than 1 µm that were not in the form of distinguished external features of
511	main particle are difficult to classify and referred to here as unidentified particles.
512	
513	

515

514

516 **3.1** The size distributions of PBAPs based on SEM analysis:

3 Results and discussion

Based on the SEM analysis of the filter and Si wafer samples, the size distributions for 517 total bioaerosol as well as for various PBAP groups were constructed for 0911N, 0912D, and 518 519 0913D and are shown in Fig. 3. In general, no difference in the size distribution of total PBAPs between day and nighttime samples is observed. The 'coarse mode' (diameter 1-10 µm) 520 521 bioaerosol particles were dominated by fungal spores as the corresponding size distribution shows a peak between $3-4 \mu m$. This is in agreement with the previous measurements by 522 Huffman et al. (2012) and Whitehead et al. (2016), in which they attributed the peak in the 523 524 coarse fluorescent biological size distribution to fungal spores. Several types of fungal spores such as Basidiomycota, Cladosporium, Inocybe calospora, Rhizopus were frequently observed. 525 526 Most pollen grains observed during SEM were with $D > 8 \mu m$. 'Fine mode' ($D < 1 \mu m$) bioaerosol particles were dominated by the plant or animal detritus as well as bacteria. Overall, 527 the total size distribution of PBAPs from the current study is in good agreement with previous 528 529 observations by Huffman et al. (2012) and Whitehead et al. (2016) from a similar geographical location (see Figure 3d). 530

531 The mean number concentrations of four PBAP groups including fungi, bacteria,
532 pollen, and viral, plant/animal detritus from SEM analysis are given in Table 3. Typically the

number concentrations of fungal spores and pollens at the ATTO are $\sim 10^4$ m⁻³ and $\sim 10^3$ m⁻³ respectively, following typical concentrations in vegetated areas (Després et al. 2012 and references therein). The estimated mean number concentrations of bacterial particles is $\sim 10^5$ m⁻³, which is greater by one order of magnitude than their typical concentration of $\sim 10^4$ m⁻³ in the air (Després et al. 2012 and references there-in).

To get an idea regarding the size distribution of total aerosol particles during the campaign, a combined aerosol size distribution from the SMPS and APS on 0914 is shown in Fig. 3d. The SMPS and APS data were not available at the ATTO site for other sampling days considered here, as these instruments were not working on those days. Using aerosol size distributions (by SMSP+APS) on 0914 as a reference, ~11% of coarse mode aerosol particles are of biological origin.

544 The total PBAP number concentration (with $D > 0.3 \mu m$) from SEM was compared with their independent estimations with FCM as shown in Fig. 4. Overall, a good agreement 545 546 is observed between SEM and FCM estimation of total PBAP number concentration for samples from all three days discussed here. The number concentration of PBAPs from FCM 547 was slightly lower than those from SEM, since abundance of PBAPs by FCM represents an 548 approximately lower limit for the actual abundance of PBAPs. The concentrations of 549 fluorescent biological aerosol particles (FBAP) from the FCM analysis were 3.3×10^5 , 550 1.06×10^6 , and 1.19×10^6 m⁻³ for 0911N, 0912D, and 0913D respectively. These values 551 are consistent with previously reported FBAP concentrations of about 2×10^5 – 552 1.2×10^6 m⁻³ by Whitehead et al. (2016) in the central Amazon forest. Huffman et al. (2012) 553 reported coarse mode FBAP concentration of about 7.3×10^4 m⁻³ over Amazon rainforest. 554

555

556 **3.2 Observations of total IN**

The observations of total IN including deposition and condensation-freezing IN and 557 immersion freezing IN are shown in Fig. 5. Figure 5 (a, b, and c) shows the concentration of 558 IN activated by deposition and condensation-freezing modes determined as a function of 559 temperature and relative humidity over ice (RHice) obtained by the FRIDGE-STD 560 measurements. No active IN at temperatures warmer than -20°C were detected with this 561 instrument, suggesting that their concentrations may have been lower than the detection 562 threshold of 0.01 L⁻¹. For a given temperature and RH_{ice}, a large day-to-day variability in IN 563 concentration was observed. The results indicate that higher IN concentration was associated 564 with higher RH_{ice} at a given temperature. The mean IN concentration at -25° C ranged from 565 0.03 to 0.43 L⁻¹ for RH_{ice} between 120% and 130%. During the field campaign, the highest 566 concentration of deposition and condensation freezing IN measured by FRIDGE-STD was 4.5 567 L^{-1} at $-30^{\circ}C$. 568

Figure 5d shows the cumulative number concentration of immersion freezing IN from 569 separate experiments in LINDA and FRIDGE-IMM as a function of temperature. Overall, the 570 concentration of immersion freezing IN was in the range of 0.0027-40 L⁻¹ at temperature 571 between -6 to -30° C. Absence of any active IN in FRIDGE-IMM at temperature > -15° C 572 could be associated with IN concentrations below the detection threshold of 0.15 L⁻¹. The 573 sample volume for LINDA samples was higher than the FRIDGE-IMM samples (see Table 1), 574 as a result LINDA had a lower detection threshold of 0.0014 L⁻¹. The concentrations of IN 575 from FRIDGE-STD at water saturation are lower than the concentrations of immersion freezing 576 IN from FRIDGE-IMM as FRIDGE-STD does not encapsulate the full immersion freezing. 577

578

579 **3.3 Biological IN concentration from drop freezing experiments**

The cumulative concentrations of biological and non-biological IN determined by drop 580 freezing arrays in LINDA are represented in Fig. 6 for 0911N, 0912D, 0913D. The 581 concentration of active biological IN between $-5^{\circ}C$ and $-15^{\circ}C$ was determined by comparing 582 measurements with and without heating. Freezing was observed with wide variance in onset 583 temperature. Heating the sample extracts decreased the highest temperature of freezing, by 1°C 584 in 0911N and 0913D to more than 4°C in 0912D. The active biological IN concentrations in 585 air ranged from 0.0015 to 0.047 L⁻¹ between -6.5° C and -15° C and are in the range of previous 586 biological IN measurements from rain samples (e.g. Bowers et al. 2009; Xia et al. 2013; Pouzet 587 588 et al. 2017). In general, the relative contribution of biological IN to the total IN decreased with decreasing temperature. The relative contribution of biological particles in the sample extracts 589 of 0911N and 0912D ranged from ~100% of the total IN active at -8° C to as low as ~ 86% at 590 -15°C. In comparison, the contribution of biological particles to the total IN concentration in 591 the sample extract of 0913D was much lower (~ 62% at -13° C). 592

593

594

4. Empirical parametrization of heterogeneous ice nucleation for multiple
 groups of biological particles

597

598 **4.1 General equations for prediction of biological IN number concentration**

599 *4.1.1 Overview of existing empirical parametrization of heterogeneous ice nucleation*

The EP of deposition and condensation-/immersion-freezing modes was proposed originally by Phillips et al. (2008, 2013). They assumed that for any scenario of aerosol loading, the number mixing ratio of active IN has contributions from the three groups including dust/metallic, black carbon, and organic aerosols. Thus, the number mixing ratio of active INcan be written as

$$n_{IN} = \sum_{X} n_{IN,X} \qquad (1)$$

606 where X = DM, BC, BIO and solO for dust/metallic, black carbon, primary biological and 607 soluble organic aerosols, respectively.

The number concentration of active IN activated by deposition and condensation-/immersion-freezing modes from group *X* is proportional with aerosol surface area as:

610
$$n_{IN,X} = \int_{\log [0.1 \, \mu m]}^{\infty} \{1 - \exp [-\mu_X]\} \times \frac{dn_X}{d \log D_X} d \log D_X,$$
(2)

611
$$\mu_X = H_X(S_i, T) \,\xi(T) \left(\frac{\alpha_X n_{IN,1,*}}{\Omega_{X,1,*}}\right) \times \frac{d\Omega_X}{dn_X} \quad \text{for } T < 0 \,^\circ \text{C and} \, 1 < S_i < S_i^w \quad (3)$$

The Ω_X is the total surface area of all aerosols with dry diameters larger than 0.1 µm, μ_X is the average of the number of activated ice embryos per insoluble aerosol particle of size D_X . The number mixing ratio of aerosols in group *X* is given by n_X . A simplified version of Eqs (2) and (3) are used for X=solO. The term $H_X(S_i, T)$ is the empirically determined fraction $(0 \le H_X \le 1)$ representing scarcity of nucleation under substantially subsaturated conditions. The α_X is a fractional contribution from aerosol group *X* to the IN concentration.

618 The number of ice crystals generated in a time step Δt is given by

619
$$\Delta n_i = \sum_X \left(n_{IN,X} - n_{X,a}, 0 \right) \equiv \sum_X \Delta n_{X,a}$$
(4)

where $n_{X,a}$ is the number mixing ratio of IN from group X that has already been activated. Additional details about the scheme and associated terms can be found in Phillips et al. (2008; 2013) 4.1.2 Modification in the existing scheme for parametrization of biological IN based on PBAP
IN examined here and literature

625 The original EP treated biological ice nucleation with only one species of IN. That626 assumption is changed as follows.

Here we now replace the previous X = BIO by five PBAP groups (1)-(5) (Sec. 1) 627 including fungal spores and associated fragments (FNG), pollen and their fragments (PLN), 628 bacteria and their fragments (BCT), viral, plant/animal detritus (DTS), and algae (ALG). 629 630 According to the EP for atmospheric ice nucleation by Phillips et al. (2008), the concentration of active IN from the given group within the given size interval $d\log D_X$ depends on surface 631 areas of given aerosol species. The empirical formulation mentioned below was not applied to 632 algal particles since their size distribution from SEM measurements was not available. We used 633 literature values of their frozen fraction to measure their ice nuclei activity, and will be 634 discussed in the later part of this section. 635

636 For
$$X$$
= FNG, PLN, BCT, and DTS

637
$$n_{IN_BIO,X} = \int_{\log [0.1 \ \mu m]}^{\infty} \{1 - \exp [-\mu_X]\} \times \frac{dn_X}{d \log D_X} d \log D_X, \tag{5}$$

638
$$\mu_X = H_X(S_i, T) \,\xi(T) \,\times \,\mathrm{MIN}\{[\exp(-\gamma_X T) - 1], 40\} \,\times \,\frac{1}{\omega_{X, 1*}} \frac{d\Omega_X}{dn_X} \quad \text{for } T < 0 \,\,^\circ \mathrm{C} \quad (6)$$

Here, $n_{IN_BIO,X}$ is the number mixing ratio of IN active at temperature *T* in the *X*th bioaerosol species; Ω_X is the total surface area mixing ratio of particles with diameters D_X greater than 0.1 μ m in group *X* and is a prognostic variable of the atmospheric model; $d\Omega_X/dn_X \approx \pi D_X^2$; $dn_X/d\log D_X$ is the normalized size distribution of given bioaerosol species *X*. The absence of smaller particles with diameter 0.1-0.2 μ m that were not measured during SEM analysis is associated with technical limitation of setup. This is expected to have an insignificant effect

on our empirical formulation as the contribution of these smaller particles to the overall surface 645 area mixing ratio is negligible. In the empirical formulation provided here, the surface area of 646 647 a particle is more important than its mass. For 0911N, the contribution of particles in the size bin 0.1-0.2 µm to the total surface area mixing ratio is about 0.15% when the number 648 concentration of particles in this size bin was assumed to be similar to that of in the neighboring 649 650 size bin with larger diameter. Shape factors should also be considered when estimating the 651 surface area of PBAPs. However, due to the complexity of the observed shapes of PBAPs we assumed spherical shape for all PBAPs and refrained from considering the shape factor in the 652 653 estimation of surface area.

The factor μ_X is the average number of potentially activated ice embryos per aerosol 654 particle with diameter D_X . For lower freezing fractions $\mu_X \ll 1$, the Eq (6) can be written as 655 $n_{IN_BIO,X} = H_X(S_i, T) \xi(T) [\exp(-\gamma_X T) - 1] \times \Omega_X / \omega_{X,1*}$ showing surface area dependence 656 of predicted IN number concentrations. The $\omega_{X,1*}$ is an unknown parameter and depends on 657 bioaerosol type with the dimensions of area (m²). In Eq (6), $H_X(S_i, T)$ is the empirically 658 determined fraction ($0 \le H_X \le 1$) representing suppression of nucleation under substantially 659 660 water-subsaturated conditions. The H_X is expressed as a function of S_i and T by Phillips et al. (2008) with $H_X=1$ at water saturation. The slope of the fitted curve predicting activated IN is 661 given by γ_X . The factor ξ considers the fact that drops are not observed to freeze at temperatures 662 warmer than a threshold in the laboratory observations. Phillips et al. (2008) proposed $\xi =$ 663 δ_1^0 (T, T₁, T₂), where ξ is 1 for $T < T_1$ and 0 for $T > T_2$ with cubic interpolation between and 664 assumed $T_1(-5^{\circ}C)$ and $T_2(-2^{\circ}C)$ for all aerosol species. 665

For temperatures colder than -20° C, lower slope values (45% of original) of the fit were prescribed based on observations of biological IN of 0911N at temperature between 668 -20° C and -27° C (see Fig. 9). The modified version of proposed parameterization (eq. 6 and 669 7) for any $T < 0^{\circ}$ C can be written as

670
$$n_{IN_BIO,X} = \int_{\log [0.1 \, \mu m]}^{\infty} \{1 - \exp [-\mu_X]\} \times \frac{dn_X}{d \log D_X} d \log D_X, \tag{7}$$

671
$$\mu_{X} = \begin{cases} H_{X}(S_{i},T) \ \xi(T) \ \times \ \text{MIN}\{[\exp(-\gamma_{X}T) - 1], 40\} \times \frac{1}{\omega_{X,1*}} \frac{d\Omega_{X}}{dn_{X}}, & \text{for } T \ > \ -20^{\circ}\text{C} \\ H_{X}(S_{i},T) \ \xi(T) \ \times \ \text{MIN}\{[\exp(-\gamma_{X}T) - 1], 40\} \times \exp[0.45 \times (T + 20)] \times \frac{1}{\omega_{X,1*}} \frac{d\Omega_{X}}{dn_{X}} \\ & \text{, otherwise} \end{cases}$$

Eqs (7) and (8) are applied for biological species, X = FNG, PLN, BCT, and DTS (groups 14, Sec. 1)

(8)

Regarding algal particles (group 5; Sec. 1), a separate representation is proposed as 675 follows. Since the concentrations of algal particles at the ATTO site were much lower than 676 our detection threshold (~ 0.5 m^{-3}) we could not use a similar empirical treatment for them as 677 was used for other PBAP groups. Generally, in atmospheric models there may be situations 678 679 where algae are more prolific and can influence the ice initiation. In such a scenario, their contribution to the total biological IN can not be excluded. We therefore used the estimation 680 of the frozen fraction for the algal particles (Diatom cell, T. pseudonana) available in the 681 682 literature (Wilson et al. 2015) to estimate the algal IN in the atmosphere. The relationship between the frozen fraction (f_{algae}) of algae and temperature was established and (see 683 extended data Fig. 6 in Wilson et al. 2015) is given by 684

685
$$f_{algae}(T) = A_1 + \frac{(A_2 - A_1)}{1 + 10^{(B+T) \times p}}$$
(9)

686 here $A_1 = -0.03$, $A_2 = 0.993$, B = 27.73, and p = 0.399.

687 with
$$f_{algae}(T) = 0$$
 at $T > -24 \, {}^{o}C$ and $f_{algae}(T) = 1$ at $T < -35 \, {}^{o}C$

688

If the concentration of algal particles (n_{algae}) in the air is known, then for X= ALG

$$n_{IN_BIO,X} = f_{algae} \times n_{algae}$$
(10)

Our scheme treats the algal particles over the ocean but does not apply to the prediction of biogenic organic macromolecules which can act as efficient IN. Although they are biological in origin, these organic macromolecules are not classified as PBAPs as they are dissolved organic matter and therefore not primary (Knopf et al. 2018; Pummer et al. 2012; Pummer et al. 2015; McCluskey et al. 2018). These ice nucleating macromolecules are accessible on the surfaces of leaves, wood and bark and can be washed down by rainwater (Felgitsch et al. 2018).

696

697 **4.2 Empirical determination of values of parameters**

698 Based on the IN (Fig. 7 and C1) and size distribution measurements (see Fig. 3 and B1) for nine cases mentioned earlier, the values of active biological IN from groups (1)-(4) were 699 700 estimated using proposed EP (Eqs. 5-8). Experiment-based estimate of all unknown parameters for each PBAP category is constrained by several technical aspects of PBAP collection and 701 processing for ice nucleation. A key problem we faced in the observation of our samples was 702 size distribution of various PBAP groups overlap greatly. Consequently, it was not possible to 703 704 isolate each PBAP group on a single filter. In addition, two of the four PBAP groups, including 705 pollen and leaf litter, naturally have lower IN activity compared to fungi and bacteria (Després et et al. 2012 and references therein). We have tried to overcome these difficulties by collecting 706 our samples on different days under contrasting conditions in conjunction with inverse 707 708 modeling. But there was a limit to the number of parameters that could be constrained by the 709 inverse modelling. Some of these parameters were therefore constrained based on laboratory measurements available in the literature. 710

The values of slope parameter γ at $T > -15^{\circ}$ C are prescribed based on observations 711 made by Vali (1994, equation 8 therein) and are assumed to be similar among PBAP groups 712 713 (1)-(4) (Sec. 1). This assumption has the basis as the slope of the observed biological IN concentration is consistent with the observations of Pouzet et al. (2017) (See Figure 7). The 714 concentration of total active biological IN from the EP is then compared to the observed 715 716 biological IN concentrations for each of the nine cases (see Fig 7 and C1). The relative abundance of the four groups of PBAPs and IN concentrations varies between the nine 717 contrasting cases. For agreement between observed and parameterized values, the least-square 718 719 fitting was used to estimate the universal law applicable to all nine cases to resolve the optimized values of the unknown parameters for PBAP groups (1)-(4). 720

The optimum values of the parameters inferred for the empirical scheme are shown in 721 Table 4. It should be noted that in the variational analysis the values of unknown parameter 722 $\omega_{X,1*}$ of comparatively less active PBAP groups including pollen and viral, plant/animal 723 detritus were constrained so that their estimated IN activity match the laboratory observation 724 reported in the literature. The estimations of $\omega_{X,1*}$ for group (4) are based on previous 725 observations of leaf litter (major part of group 4) from the tropics by Schnell and Tan-Schnell 726 727 (1982). For pollen, the $\omega_{X,1*}$ was constrained based on laboratory observations on active sites per gram of pollen mass (Betula pendula) by O'Sullivan et al. (2015). Additional details about 728 these measurements used in the construction of the EP are given in Table 5. 729

In the current study, the values of T_1 and T_2 for determining the factor ξ are assumed to be variable for each bioaerosol type and are estimated based on the literature of previous observations or laboratory measurements. Therefore, the values of T_1 and T_2 for each PBAP group are denoted by $T_{X,1}$ and $T_{X,2}$. Several studies have documented the activation of pollen at temperatures colder than -10° C (Després et al. 2012; Kanji et al. 2017, and references therein). Freezing tests on leaf litter showed its activation at temperatures ~ -7° C (Schnell and TanSchnell, 1982; Després et al. 2012). Based on these observations, T_I was assumed to be -10 to -7°C for pollen and plant/animal detritus respectively. Comparatively, fungi and bacteria are activated at temperatures warmer than -5°C (Richard et al. 1996; Després et al. 2012; Kunert et al. 2019) and therefore -7°C and -2°C, respectively, were assumed for T_I of both.

Fig. 7 and C1 shows the predicted number concentrations of crystals nucleated by 740 741 various biological species based on size distributions from the total and 5, 1.2 µm filters, respectively. The predicted values of total biological IN by the EP are of similar magnitude in 742 comparison with observations from the total $(5 + 2 \mu m)$ and 5 μm filters. The total number 743 concentration was dominated by fungi and bacteria. The contributions from pollen and detritus 744 were lower by two orders of magnitude than those from fungi and bacteria at a given 745 temperature. Pollen grains can swell and explode from high humidity exposure to emit 746 numerous nanoscale particles which are 10 to thousand times smaller than intact particles. 747 These smaller fragments may become aerosolized and act as efficient IN (O'Sullivan et al. 748 2015; Knopf et al. 2018). Given this, the contributions of pollen grains to total biological IN 749 may be more important than suggested by our measurements. The predicted total IN 750 concentration is lower by 1-2 order magnitude when compared with the values obtained for the 751 1.2 µm filters (See Fig. C1). It can be attributed to the uncertainties in estimating the collection 752 efficiency of 5 µm filters and hence to the size distributions of bioaerosol particles on 1.2 µm 753 filters. 754

755

756

5 Comparison of the empirical scheme with independent observations

The new EP is also compared with previous laboratory data and field observations, notused in its construction, as shown in Figure 7 and C1. Additional details on the origin of these

760 data sets and corresponding calculations are given in Table 5. The total concentration of PBAP IN predicted by EP is in good agreement with previous observations for rainwater samples 761 from Opme (France) by Pouzet et al. et al. (2017). For fungi, estimations of nucleation sites per 762 gram of fungal mass (Fusarium avenaceum) (O'Sullivan et al. 2015) and active IN per fungus 763 (Basidiomycota spores) (Haga et al. 2014) were used to calculate their contribution as IN for 764 765 the observed mass and number concentration of fungi over the Amazon. According to previous 766 studies by Murray et al. (2012) and references there-in., the number concentration of birch pollen as potential IN is 2×10^{-3} m⁻³ at -15° C. Based on ice nucleation activity of bacteria 767 isolated from cloud water by Joly et al. 2014, number concentrations of potential INA bacteria 768 were estimated ~ 0.32 and 5.8 m⁻³ of cloud air at temperatures -6 and $-8^{\circ}C$ respectively. 769 Murray et al. (2012) estimated the concentration of pollen IN at -15° C is 2.4×10^{-3} m⁻³. The 770 active IN concentration from leaf litter per m³ of air based on measurements by Schnell and 771 Vali (1976) for leaf litter from tropical regions is also shown for comparison with 772 parameterized values. When the uncertainty limits are considered, the EP's predicted PBAP IN 773 concentrations are in agreements with available literature data. 774

775 To represent ice nucleation abilities of various PBAP groups various metrics are used in the previous studies. Most used metric includes 1) ice-active surface site densities (n_s) , 2) 776 ice-active mass site densities (n_m) , an analogue to n_s , but mass equivalent, 3) activated fraction 777 778 (Kanji et al. 2017 and references therein). To represent proposed EP in terms of these common metric, we estimated the mean values of n_s , n_m for various PBAP groups based on PABP IN 779 activity, their surface area mixing ratio, and mass in nine cases considered in the study. Figure 780 8 shows the dependence of these variable from the EP on temperature. The values of these two 781 parameters for limited species of PBAP groups from the available literature are also shown for 782 783 reference. However, there is a considerable spread in the literature data.

At a given temperature, our estimated values of n_s and n_m for PLN and DTS by EP 784 were much lower (by three orders of magnitude) when compared with our prediction of FNG 785 786 and BCT as observed in the literature data (Kanji et al. 2017 and references therein). The values of n_m for group 4 (mostly leaf litter) are in good agreement with literature data. The literature 787 values of n_s and n_m for snowmax and *Pseudomonas syringae* bacteria are much higher than 788 our estimated values (see Figure 8). For a given PBAP group, the published observations of n_s 789 and n_m show large variations among different biological species (Kanji et al. 2017 and 790 references therein) which limits their direct comparison with our estimated values of the EP, 791 which apply to entire groups of PBAPs. At a temperature of -10° C, for example, n_m of three 792 species of fungal spores showed variation over four orders of magnitude. 793

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6 Application of empirical parameterization in a parcel model for an idealized case

798 The EP is applied in an idealized 0D parcel model to shed light on some ramifications for natural ice initiation arising from the formulation. It should be noted that there are no 799 microphysical processes included inside the parcel model except for the heterogeneous ice 800 nucleation. The adiabatic parcel starts at 20°C, rises to -50°C and humidity is prescribed 801 corresponding to exact water saturation. Input size distributions of PBAPs (group 1-4) to the 802 803 empirical scheme are prescribed based on observations from the case 0911N. An EP from Phillips et al. (2013) was applied for IN species including dust, soot and PBAPs. The 804 climatological values (from Model of Ozone and Related Chemical Tracers [MOZART]) of 805 mass mixing ratios for dust (2.9×10^{-10} kg/kg) and soot (3×10^{-11} kg/kg) over the ATTO 806 site were prescribed for estimating their contribution as IN. The total PBAP size distributions 807

were prescribed based on observations on 0911N. For algae, quantitative measurements of their
abundance in the ambient air are rare. The algal particles were not included in the parcel model
simulation shown here.

Figure 9 shows the predicted number concentration of active biological IN from 811 multiple species as well as the total IN. Compared with Phillips et al. (2013), the total biological 812 813 IN predicted by the proposed empirical scheme differs by maximum one orders of magnitude at $T > -15^{\circ}$ C and by half order of magnitude at $T < -15^{\circ}$ C. It should be noted that the EP by 814 Phillips et al. (2013) does not resolve individual groups of PBAPs and was based on the ice 815 nucleation of a bacterial strain observed in the laboratory that may not represent the large 816 variations in ice nucleation characteristics observed among different PBAP groups. Overall, 817 total IN concentrations predicted by the model are in good agreement with the concentrations 818 observed at the ATTO site. The relative contributions to the total IN concentration of the 819 different PBAP groups as well as dust and soot particles from this simulation are shown in 820 821 Figure 10. Biological particles dominate the predicted total IN concentrations at a temperature warmer than -20° C and at colder levels dust and soot particles became increasingly important 822 in determining the total IN concentrations in the cloud. 823

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825

826 **7 Conclusions**

A modification to the flexible framework for treating multiple species of atmospheric IN in terms of empirical observations from the real atmosphere, namely the 'empirical parameterization' is proposed in the current study. The previous single species for biological IN has been replaced by five major PBAP groups including FNG, PLN, BCT, DTS, and ALG. The new treatment has been fitted to the coincident observations of ice nucleation activity of
PBAPs and their size distributions based on observation from a field campaign at the ATTO
site located in the central Amazon. In addition to the observations of PBAP IN from the ATTO
site, the literature data are used for the construction of the EP. The new empirical treatment
will allow atmospheric models to explicitly resolve the IN activity of these PBAP groups.

PBAPs were collected on PTFE membrane filters and Si wafers for use in drop freezing
experiments, ice nuclei activation in cloud chamber, and SEM/EDX analysis. An EP was
constructed to predict the active IN from multiple groups of PBAPs based on coincident
observations of PBAP size, concentration, and ice-nucleating ability.

840 The measured IN concentrations activated by deposition and condensation-freezing modes varied over three orders of magnitude throughout the campaign and strongly depended 841 on the processing temperature and ice supersaturation in the chamber. At -25° C, the measured 842 IN concentrations activated by deposition and condensation-freezing mode ranged from 0.002 843 L^{-1} to 1 L^{-1} for ice supersaturations of 21% and 28% respectively. The filter samples were 844 845 analyzed in LINDA to estimate biological IN active between -5 and -15° C. Filter and Si wafer samples were analyzed under SEM to measure the size distribution of common PBAP species. 846 For each case, more than 300 images of bioaerosol particles were used for this purpose. 847

848 The observed bioaerosols included brochosomes, fungal spores, bacteria, canopy debris (e.g. leaf and insect fragments, plant waxes), and less commonly pollen and insect scales. 849 Overall, size distributions were observed to be bimodal and were like previous measurements 850 from similar geographical locations (e.g. Huffman et al. 2012; Whitehead et al. 2016). The 851 observed size distributions of PBAP groups have provided constraints primarily on their 852 853 surface areas in determining their ice nucleation activity. The size distribution of total bioaerosol particles was consistently dominated by a submicron peak associated with the viral, 854 855 plant and animal detritus and bacteria. The coarse mode of PBAP observed in the current study 856 was dominated by fungal spores consistent with previous observations (e.g. Huffman et al.

857 2012). The total number concentration of bioaerosols (with $D > 0.3 \mu m$) inferred by our SEM 858 analysis is in good agreement with FCM analysis.

859 The total number concentration of biological IN predicted by the proposed parameterization scheme was dominated by fungi and bacteria followed by pollen and detritus. 860 For PBAP groups considered in the construction of the scheme, the values predicted from the 861 scheme and independent observational data not used in the construction of the scheme were 862 863 consistent with concentrations of the same order of magnitude. Implementation of the scheme in the 0D parcel model showed that the biological IN account for most of the total ice nuclei 864 activation at temperatures $> -20^{\circ}$ C. At colder temperatures dust and soot become increasingly 865 more important to ice nucleation. Previous observations in the Amazon basin by Prenni et al. 866 (2009) also emphasize the importance of biological IN in determining total IN concentrations 867 at warmer atmospheric temperatures. They showed that the dust became increasingly more 868 important for ice nucleation at temperatures colder than -27° C. In their transmission electron 869 870 microscopy, no soot IN was observed, suggesting that biomass-burning particles were not a likely source of the carbonaceous IN. However, it should be noted that their observations were 871 limited to the wet season. The biomass burning conditions are characteristics of the Amazon 872 873 dry season and are maximum around September. Thus, the soot particles resulting from the biomass burning can play an important role in determining the ice nucleation at colder 874 temperature during the dry season, as suggested by our parcel model simulation. It should be 875 noted that the relative contributions of dust and soot to the ice nucleation shown in our parcel 876 mode simulations are limited to a case study and it will be highly variable depending on their 877 878 abundance in the atmosphere.

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Data Availability Statement

The data that support the findings of this study are available from the corresponding 882 author, Sachin Patade, on reasonable request. A copy of the EP scheme including the present 883 addition is also available on request. 884

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929	APPENDIX A
930	List of symbols
931	"Table A1 here"
932	
933	APPENDIX B
934	PBAP Size distribution from 5 and 1.2 µm filter
935	"Figure B1 here"
936	
937	APPENDIX C
938	Model prediction of biological particles for 5 and 1.2 μ m filters
939	"Figure C1 here"
940	
941	APPENDIX D
942	Implementation of empirical parameterization in an atmospheric model
943	The present EP scheme slots into the EP framework for heterogeneous ice nucleation
944	already described in Phillips et al (2008, 2013). Their framework represents all modes of
945	heterogeneous ice nucleation. The EP described in this study is for ice nucleation through
946	deposition, condensation- and immersion-freezing mode.
0/17	To implement the scheme in a cloud model, each group X of IN is partitioned into

To implement the scheme in a cloud model, each group *X* of IN is partitioned into prognostic components which are interstitial and immersed in cloud droplets to reflect the incloud scavenging of IN by precipitation. Extra scalars were introduced to track the depletion in the number mixing ratio of interstitial IN by becoming immersed in cloud liquid without necessarily freezing $(n_{X,a,liq})$ and the actual number mixing ratio of immersed IN $(n_{X,imm})$. The cut off size for immersed and interstitial IN size distributions is determined by $(n_{X,a,liq})$ and mixing ratio of aerosol in group $X(Q_X)$. To include the heterogeneous freezing of raindrops a temporary grid of size bins was created to discretize their size distribution. If the mass mixing ratio of each bin is dQ_r , the number of IN particles in the rain that activate in time Δt is given by

957
$$d(\Delta n_{IN,rain}) \approx \Delta t \text{MIN}[(w - v_t) \frac{\partial T}{\partial z}, 0] \frac{d}{dT} [\exp(-\gamma_X T) - 1] \sum_X \frac{d\Omega_{X,rain}}{\omega_{X,1,*}}$$
(11)

958 For a given raindrop size bin $d\Omega_{X,rain} = \Omega_{X,rain} \frac{dQ_r}{Q_r}$ is the surface area mixing ratio of IN 959 within the raindrops. The fall velocity of raindrops is given by v_t , while *w* is the vertical air 960 velocity.

We have adopted the approach followed by Phillips et al. (2008) for inside-out contact freezing. It assumes that a given IN has a freezing temperature for the contact nucleation which is $\Delta T_{CIN} \approx 4.5^{\circ}$ C higher than for the immersion and condensation freezing mode. The number mixing ratio of potentially active, interstitial IN activated by contact freezing ($n_{X,cn}$) is given by

966
$$n_{X,cn} \cong \xi(T) \left\{ \exp\left[-\gamma_X (T - \Delta T_{CIN})\right] - 1 \right\} \sum_X \frac{d\Omega_{X,int}}{\omega_{X,1,*}}$$
(12)

967 Here,
$$\Omega_{X,int}$$
 is the component of Ω_X for interstitial IN.

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1295	

1298 Tables:

Sr.	Type of aerosol	Time	Flow rate	Purpose	Instrument
No.	sampling	(UTC)	(sample volume)		used in the
					analysis
1	0.2 μm filter only	1400-1450	4 Lpm (200 L)	Immersion freezing IN	FRIDGE-IMM
2	PEAC7	1500-1550	2 Lpm (100 L)	Deposition and	FRIDGE-STD,
				condensation-freezing	SEM
				IN, SEM	
3	Cascading Filtering,	1600-2010	4 Lpm (1000 L)	SEM, Biological IN	LINDA, SEM,
	Day				flow cytometry
4	Cascading Filtering	2030-1230	1.5 Lpm (1440 L)	SEM, Biological IN	LINDA, SEM,
	Overnight	(next day)			flow cytometry

Tabe 1: Daily sampling plan along with sampling type, time duration, flow rate, volumesampled, sampling purpose, and the instrument used for sample analysis.

Sample	Sample Vol. in L	Scanned Area in % Filters (Si wefers)	Total images Filtors (Si wafers)
	ritters (Si waters)	ritters (Si waters)	Fillers (SI waters)
0911N	1500 (100)	0.14 (0.064)	302 (101)
0912D	1000 (100)	0.1 (0.062)	201 (114)
0913D	100 (100)	0.11 (0.07)	210 (103)

1313	Table 2: Details of SEM analysis including samples volume, scanned area of filter and Si
1314	wafer, the number of bioaerosol images collected.
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PBAP type	Typical size (μm) and number conc. (m ⁻³ of air)	General morphological features	References
Fungal spores	Size: 1-50; most frequently 2-10 Concentration: 10 ⁴	(i) shape: globose, elliptical, fusiform, asymmetric, lemon- shaped, barrel-shaped, curved (ii) wall characteristics: smooth, granular, reticulate, spines, warts (iii) attachment scars, attachment pegs (iv) other features: branched hyphae	Després et al. (2012) and references therein, Priyamvada et al. 2017, Li et al. 2020, Huffman et al. 2012, Cybelli et al. 2018, China et al. 2016,
Bacteria	Size: 0.5-4 Concentration: 10 ⁴ (over land), 10-1000 (over ocean)	(i) shape: spherical, rodlike, helical (ii) wall: flagella, pili, capsules (iii) Arrangement: pairs, tetrads, clusters, chains	Després et al. (2012) and references therein, Li et al. 2020
Pollen	Size: 10-50 Concentration: 10-1000	 (i) shape: ellipsoid, fusiform, globose/spheroidal (ii) presence of forrows (iii) outer exine is usually sculptured (iv) exine ornamentation like fossulate or microrugulate 	Després et al. (2012) and references therein, Cybelli et al. 2018 and references therein, Xu et al. 2013
Plant/animal detritus	Size: brochosomes-0.5, leaf litter 0.5-5 Concentration: NA	Various shapes (i) shape: football (brochosomes), spiral (epicuticular wax of plants), elongated (leaf litter)	Després et al. (2012) and references therein, Wittmaack, 2005
Algae	Size: 1-10 Concentration: 100	(i) shape: Unicellular flagellates or amoeboids, colonial and nonmotile forms	Després et al. (2012) and references therein

Table 3: Typical size ranges, concentrations in air and general morphological characteristics
of different PBAPs, based on literature available. Based on this information, PBAPs have been
classified into different groups. Also enlisted are various studies referred to for classification
of PBAPs.

PBAP group	Typical	Mean number
	Size	concentration (m ⁻³ air)
		from SEM analysis
Fungal spores	1-10	7.6×10^4
and associated	μm	$(\pm 2.8 \times 10^4)$
fragments		
Bacteria and	0.5-2	4.2×10^{5}
their fragments	μm	$(\mp 1.98 \times 10^5)$
Pollen and their	6-50	1.07×10^{3}
fragments	μm	(∓673)
Viral,	0.2-5	4.1×10^{5}
plant/animal	μm	$(\pm 1.8 \times 10^5)$
detritus		

1338	Table 4 : General size and mean number concentration of various PBAP groups based on our
1339	SEM analysis.
1340	
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Symbol	Value
$\gamma_{fng}, \gamma_{pln}, \gamma_{bct}, \gamma_{dts}$	0.5 C ⁻¹
ω_{FNG} ,1*	$2.1 \times 10^{-4} \mathrm{m}^2$
$\omega_{PLN,1*}$	0.1 m ²
$\omega_{BCT,1*}$	$2.51 \times 10^{-4} \text{ m}^2$
$\omega_{DTS,1*}$	0.1 m ²
$T_{FNG,1}, T_{fng,2}$	−7 °C, −2 °C
$T_{PLN,1}, T_{pln,2}$	−10 °C, −7 °C
$T_{BCT,1}, T_{bct,2}$	−7 °C, −2 °C
$T_{DTS,1}, T_{dts,2}$	−10 °C, −7 °C

Table 5: Details about most empirical parameters mentioned in section 4.1 and 4.2 together

1353 with their values.

1	2	r	<u>_</u>
	-	n	n
-	-	~	~

Species	Reference	Obs. Mass (kg m ⁻³) or Number	IN conc. (m ⁻³) (in bracket
Fungi	1. O'Sullivan et al. (2015):	Mass:	For 0911N: 0.38
	Nucleation sites g $^{-1}$ at –5, –10, and –15 $^{\rm o}C$ are 2.5 \times 10 $^{5},$ 4.× 10 $^{7},$ 2.× 10 8	1.51×10^{-9}	(-5), 60.63 (-10),
	respectively	(0911N)	303.16 (-15)
		1.52×10^{-9}	For 0912D: 0.37
		(0912D)	(-5), 60.5 (-10),
		1.17×10^{-9}	302.53 (-15)
		(0913D)	For 0913D: 0.29
			(-5), 46.8 (-10),
			234.4 (-15)
	2. Haga et al. (2014): Active IN per fungi at -10 and -15° C are 5.× 10^{-4} , 7.× 10^{-4}	Conc.	For 0911N: 52.80
	respectively.	1.01×10^{5}	(-10), 73.93 (-15)
		(0911N)	For 0912N: 36.9
		0.73×10^{5}	(-10), 51.70 (-15)
		(0912D)	For 0913N: 24.81 (-10), 34.74 (-15)
		0.49×10^{5}	(,
		(0913D)	
Pollen	1. O'Sullivan et al. (2015) (used in construction of scheme):	Mass:	For 0911N:
	Nucleation sites g^{-1} at -10 , $-15^{\circ}C$ are 2×10^3 , 10×10^3 respectively.	5.76×10^{-10}	1.15×10^{-3} (-
		(0911N)	10), 5.76 ×
		5.14×10^{-10}	10 ⁻³ (-15)
		(0912D)	For 0912D:
		1.18×10^{-10}	1.02×10^{-3} (-
		(0)15D)	10), 5.14 ×
			10 ⁻³ (-15)
			For 0913D:
			2.36×10^{-4} (-
			10), 1.18 ×
			10 ⁻³ (-15).
	2. Murray et al. (2012)		Pollen as Potential IN is 2×10^{-3} (-
Bacteria	1. Joly et al. (2014): Bacterial conc. was variable from 1.6 \times 10 ³ to 3.4 \times 10 ⁴ mL ⁻¹		0.32 (-6), 5.8 (-8),
	Mean fraction of ice nucleation active bacteria at -6, -8, and -12 $^{\rm o}{\rm C}$ are 0.01, 0.18, and		32.5 (-10)
	1.02% respectively. Liquid water content was assumed 0.5 g/m ³ .		
Leaf Litter	1. Schnell and Tan-Schnell (1982): Nucleus concentration (g ⁻¹) at -6.5°C and -10°C is	Mass:	For 0911N: 2.3 ×
	60 and 800 respectively (used in construction of scheme).	3.86×10^{-10}	10 ⁻⁵ (-6.5),
		(0911N)	3.08×10^{-4} (-
		2.56×10^{-11}	10)
		(0912D)	

		2.67×10^{-10}	For 0912D: 1.5 ×
		(0913D)	10 ⁻⁶ (-6.5), 2.×
			10 ⁻⁵ (-10)
			For 0913D: 1.6 ×
			10 ⁻⁵ (-6.5),
			2.13×10^{-4} (-
			10)
	2. Schnell and Vali (1976): Nucleus concentration (g ⁻¹) at $-7^{\circ}C$ and $-12.7^{\circ}C$ is 100 and		For 0911N: 3.8 ×
	at is 1000 respectively. (for leaf litter in tropical climate zone)		10^{-5} (-7), 3.8 ×
			10 ⁻⁴ (-12),
			For 0912D: 2.5 ×
			10 ⁻⁵ (-7).
			2.5×10^{-4} (-12).
			For 0913D:
			2.67×10^{-5} (-7),
			2.67×10^{-4} (-
			12),
Total biological	Pouzet et al. 2017 (Figure 1).		0.06 (-5), 1.65 (-
lin	Rainwater content was assumed 0.3 gm/m ³ to convert concentrations to per m ³ of the air.		10), 6 (-12)

Table 6: Origin of estimations of various biological IN concentrations shown in figure 7. The mass of bioaerosol particles was calculated assuming aerosol particles to be spherical with a density of 1000 kg m⁻³. Observation of IN activity of pollen and leaf litter used in the construction of the scheme are mentioned in brackets.

Notation	Description	Value and Units
D	Diameter	m
f _{algae}	Frozen fraction of algal particles from Wilson et al. (2015).	
H_X	Fraction reducing ice nuclei activity at low S_i , warm T	
MAX	Maximum value	
MIN	Minimum value	
n _{IN_BIO,X}	Contribution to total biological IN from group <i>X</i>	kg ⁻¹
n_{s_i}	Ice-active surface site density	m ⁻²
n_m	Ice-active mass site density	kg ⁻¹
n_X	Number mixing ratio of particles in PBAP group X	kg ⁻¹
n _{IN_BIO}	Total number m.r. of active biological IN predicted	kg ⁻¹
$n_{IN_BIO,X}$	Contribution to n_{IN_BIO} from PBAP group X	kg ⁻¹
$n_{X,a}$	Number of aerosols in PBAP group X	kg ⁻¹
$n_{X,cn}$	Number mixing ratio of contact IN for group X	kg ⁻¹
$n_{X,a,liq}$	Number of interstitial IN in group X lost by becoming immersed	kg ⁻¹
$n_{X,\text{imm}}$	Number of IN from group X immersed in cloud liquid	kg ⁻¹
Q_r	Mixing ratio of rain	kg kg ⁻¹
Q_X	Mixing ratio of aerosol in group X	kg kg ⁻¹
RH _{ice}	Relative humidity over ice	%
S _i	Saturation ratio of vapor with respect to ice	
t	Time	S
<i>T</i>	Temperature	°C
$T_{X,1}$	Colder threshold temperature for onset of freezing for group X	°C
$T_{X,2}$	Warmer threshold temperature for onset of freezing for group X	°C
v_t	Fall speed of raindrop in given size bin	m s ⁻¹
W	Vertical velocity of air	m s ⁻¹
X	Label for four PBAP groups (Fungal spores and associated	
	fragments (FNG), Pollen and their fragments (PLN), Bacteria	
	and their fragments (BCT), Viral, Plant/animal detritus (DTS),	
	and Algae (ALG))	0 7
$\xi(T)$	Fraction that is zero for $T > T_{X,2}$ (°C) and 1 for $T < T_{X,1}$ (°C),	$0 \leq \xi \leq$
	being $\delta_1^0(T, T_{X,1}, T_{X,2})$ for $T_{X,1} < T < T_{X,2}$ (°C) for $X = FNG$,	1
	PLN, BCT, DTS	5 13
Ω_X	Total surface area of all bioaerosols > 0.1 μ m in diameter in	
	group X.	m^{2} [air]
0	Interstitial component of 0	Kg [aerosol]
²² X,int	interstitial component of M_X	
		kg ⁻¹
Ωxmain	Total surface area of all aerosols with $D > 0.1$ µm in group X	[aerosol]
,1 utit	inside raindrops	m ² [air]
	1 I	kg ⁻¹

$\delta_a^b(v, v_1, v_2)$	Cubic interpolation function equal to a at $y < y_1$ and to b for $> y_2$		
- u () / J I/ J Z)	$\langle \gamma_2 \rangle$		
	While $\delta_a^b = a_0 + a_1 y + a_2 y^2 + a_3 y^3$ for $y_1 < y < y_2$, where		
	$a_{0}=B, a_1 = Ay_1y_2, a_2 = -\frac{A(y_1+y_2)}{2}$, and $a_3 = \frac{A}{3}$ with $A =$		
	$\frac{6(a-b)}{(y_2-y_1)^3}$ and $B = a + \frac{Ay_1^3}{6} - \frac{Ay_1^2y_2}{2}$ (adopted from Phillips et al.		
	2008)		
μ_X	Average number of ice embryos per aerosol particle		
γ_X	Slope value of the fit for $X =$	°C ⁻¹ .	
	FNG, PLN, BCT, and DTS each of four PBAP group.		
α_{x}	Fraction of $n_{IN,1,*}$ from IN activity group X= {DM, BC, O,	{2/3, 1/3-	
	solO}	0.06,	
	,	0.06}	
$\omega_{X,1*}$	Baseline coefficient of PBAP for <i>X</i> = FNG, PLN, BCT, and	m ²	
,_	DTS		
Δt	Model time step	S	
ΔT_{CIN}	Diff. in freezing temperature between contact and bulk water	4.5°C	
CIIV	modes.		

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Table A1: Symbols used in this paper, along with their descriptions and units.



FIG. 1. Location of the ATTO site and Manaus in the central Amazon rainforest (source Google
maps) (a). The sampling location at the ATTO site include a triangular mast on which a Total
Suspended Particles (TSP) inlet was installed at 60 m above ground (b). The HYSPLIT sevendays (168 h) ensemble backward trajectories (c, d, and e) for three typical sampling days (Sept.
11-13, 2018) are shown for an endpoint altitude of 60 m.



FIG. 2. Representative scanning electron microscopy images of brochosomes and leaf litter
(group: plant and animal detritus) (a), fungal spore (b), bacteria (c) and pollen (d) from 0911N,

- 1406 0912D and 0913D. Scale is varying for each image and is shown separately.



FIG. 3. The size distribution of PBAPs based on SEM analysis of samples 0911N, 0912D, and 0913D (a, b, and c). The size distribution of PBAPs shown here is from the combination of 5 and 1.2 μ m filters. The total PBAP size distribution from each day is compared with the literature data (d). The total PBAP size distribution from our observations include unidentified PBAPs in addition to the other PBAP groups considered for classification. The combined aerosol size distribution from SMPS and APS on 0914 (thin black line) is also shown for reference.

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FIG. 4. Comparison of total bioaerosol particle number concentration ($D > 0.3 \mu m$) measured by Scanning Electron Microscopy (SEM; left bars) with Flow Cytometry (FCM; right bars). Error bars for number concentrations measured by FCM were estimated based on independent runs of FCM for the same sample.







FIG. 5. The concentration of IN activated by deposition and condensation freezing mode in the FRIDGE-STD chamber at -20° C (a), -25° C (b), and -30° C (c). The dotted vertical lines in a, b, and c indicate saturation w.r.t. water. Cumulative IN concentrations activated by immersion freezing mode from separate experiments in FRIDGE-IMM are shown(d). The cumulative immersion freezing IN concentrations for 0911N, 0912D, 0913D from analysis in LINDA are also shown.

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FIG. 6. Cumulative concentrations of biological (heat sensitive, bold lines) and non-biological
(heat-resistance, shaded area) IN. The sum of two categories corresponds to the concentrations
of total IN. The lower bound was considered for values below the detection limit. The IN
concentrations shown here are from 5 and 1.2 µm filters processed in LINDA.



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1460 FIG. 7. Prediction of biological ice nuclei concentrations by new empirical parameterization 1461 based on size distribution of PBAPs, shown as a function of temperature for various bioaerosol 1462 species. The total biological ice nuclei concentrations from the empirical parametrization (thick black line) are compared with the observed values (thick red) from the drop freezing 1463 experiments with LINDA. The prediction is based on the total size distribution $(5 + 1.2 \,\mu\text{m})$ for 1464 1465 each sampling day shown here. In addition, predicted values of biological IN are compared to previous observations listed in Table 5. Biological ice nuclei concentration of fungi based on 1466 1467 active fractions measured by Haga et al. (2014) (green crosses) and O'Sullivan et al. (2015) (green triangles) are shown for comparison. The number concentrations of pollen as biological 1468 ice nuclei are shown based Murray et al. (2012) (blue diamonds) and O'Sullivan et al. (2015) 1469 (blue squares). The bacterial ice nuclei concentrations based on Joly et al. 2014 (magenta 1470 circles). The leaf litter as ice nuclei are also shown based on Schnell Tan-Schnell (1982), 1471
1473	lower	bounds	in	predicting	ice	nuclei	concentrations	due	to	uncertainties	in	collection
1474	efficie	ncy.										
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Schnell and Vali (1976). Black dotted lines in the plot of $1.2 \,\mu m$ filter represent the upper and



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FIG. 8. Ice-active surface site densities (n_s) (a) and ice-active mass site density (b) for various PBAP groups from the proposed empirical parameterization (dotted lines) are shown as a function of temperature. The values for these two parameters from previous studies are also shown for a few biological species of PBAPs.

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FIG. 9. The number concentration of biological ice nuclei predicted by empirical parameterization as a function of supercooling in a water-saturated air parcel. The number concentrations of dust, soot, and biological ice nuclei predicted by the empirical ice nucleation scheme by Phillips et al. (2013) (mentioned as PH13 in the plot) are also shown for comparison. The error bars of observed IN concentrations indicates the standard deviations estimated based on the uncertainties in the measurements. An error bar in bold red on the Biological (model) line indicated uncertainties in the estimated biological IN at temperature colder than -15° C.



FIG. 10. The pie chart shows the relative contributions to the total IN concentration of the different PBAP groups as well as dust and soot particles at $T = -15^{\circ}C$ (a) and $T = -25^{\circ}C$ for the parcel model simulation shown in Figure 9.





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Figure C1: Prediction of biological ice nuclei concentrations by new empirical parameterization based on size distribution of PBAPs in 0911N, 0912D and 0913D shown as a function of temperature for various bioaerosol species. The total biological ice nuclei concentrations from the empirical parametrization (thick black line) are compared with the

1547 observed values (thick red) from the drop freezing experiments with LINDA. For each case, 1548 the prediction is based on PBAP size distribution from 5 μ m, and 1.2 μ m and is shown 1549 separately. Black dotted lines in the plot of 1.2 μ m filter represent the upper and lower bounds 1550 in predicting ice nuclei concentrations due to uncertainties in collection efficiency.