

Real-time sonoporation through HeLa cells

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- 1 Real-time sonoporation through HeLa cells
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- 3

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10 Abstract. The purpose of this study was to investigate the physical mechanisms of

- sonoporation, to understand and ameliorate ultrasound-assisted drug and gene delivery.
- 12 Sonoporation is the tran-sient permeabilisation of a cell membrane with help of ultrasound
- 13 and/or an ultrasound contrast agent, allowing for the trans-membrane delivery and cellular
- 14 uptake of macromolecules between 10 kDa and 3 MDa. We studied the behaviour of
- 15 ultrasound contrast agent microbubbles near can- cer cells at low acoustic amplitudes. After
- administering an ultrasound contrast agent, HeLa cells were subjected to 6.6-MHz
- 17 ultrasound with a mechanical index of 0.2 and observed with a high- speed camera.
- 18 Microbubbles were seen to enter cells and rapidly dissolve. The quick dissolution after
- 19 entering suggests that the microbubbles lose (part of) their shell whilst entering. We have
- 20 demonstrated that lipid-shelled microbubbles can be forced to enter cells at a low
- 21 mechanical index. Hence, if a therapeutic load is added to the bubble, ultrasound-guided
- 22 delivery could be facilitated at diagnostic settings. However, these results may have
- 23 implications for the safety regulations on the use of ultrasound contrast agents for
- 24 diagnostic imaging.
- 25 Keywords: Sonoporation, Ultrasound, Microbubbles, HeLa Cancer cells, Drug delivery, Low
- 26 Mechanical Index
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- 28
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35 INTRODUCTION

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37 In medical-diagnostics, guidelines state an MI<0.3 can be considered safe for pregnant 38 women and neonatals, but yet diagnostic imaging machines allow the use of MI up to 1.9. 39 Sonoporation is the transient permeabilisation and resealing of a cell membrane with the 40 help of ultrasound and/or an ultrasound contrast agent, allowing for the trans- membrane 41 delivery and cellular uptake of macromolecules between 10 kDa and 3 MDa [1]. Previous 42 studies on non-invasive, ultrasound-induced therapeutics used acoustic amplitudes 43 corresponding to mechanical indices between 0.2 and 7.0 [2, 3]. Many studies have 44 demonstrated increased drug and gene uptake of sites under sonication [4, 5, 6]. These 45 studies presumed that a physical membrane disruption mechanism, i.e. sonoporation, 46 caused the increased uptake, as opposed to naturally occurring active uptake processes, 47 such as endocytosis, which are controlled by the system biology. Here we examin the effect of low MI ultrasound (MI=0.2) in combination with lipid shelled, gas filled microbubbles on 48 49 HeLa cancer cells. Figure 1 shows six possible mechanisms of sonoporation.

52 METHODS

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54 An overview of the experimental setup is shown in Figure 2. We have described our 55 experimental setup extensively in Delalande et al. [7]. In short, a signal consisting of 40 56 cycles with a centre frequency of 6.6 MHz and a pulse repetition frequency of 10 kHz, was 57 generated by an AFG3102, dual channel arbitrary function generator (Tektronix, Inc., 58 Beaverton, OR), amplified by a 2100L, 50-dB RF amplifier (Electronics & Innovation Ltd., 59 Rochester, NY) and fed to a custom-built 6.6-MHz ultrasound transducer. The peak-negative 60 acoustic pressure corresponds to an MI of 0.2. The transducer was placed in a custom-built, 61 260×160×150 (mm)3 Perspex sonication chamber, in which an OptiCell cell culture chamber 62 (Nunc GmbH & Co. KG, Langenselbold, Germany) was placed. One side of the cell culture chamber contained a monolayer of HeLa cells. A customised BXFM-F microscope unit with 63 64 an LCAch N 20×/0.40 NA PhC (OlympusDeutschland GmbH, Hamburg, Germany) and a LUMPlanFL 60×/0.90 NA water- immersion objective (Olympus) was placed on top of the 65 66 sonication chamber with the objective lens immersed in the water. The colour charge coupled device (CCD) of a PHOTRON FastCam MC-2.1 high-speed camera (VKT Video 67 68 Kommunikation GmbH, Pfullingen, Germany) was connected to the microscope.

71 RESULTS AND DISSCUSSION

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73 Lipid-shelledmicrobubbles were forced into cells using pulsed ultrasound at MI=0.2 at 74 transmit frequencies of 1.0 MHz and 6.6 MHz. This phenomenon typically takes 2 s from the 75 moment a bubble contacts the cell membrane, to complete dissolution of the gas inside the 76 cell. Mostbubble---cell penetration occurred within 8s from the start of sonication. These 77 results were easily reproducible, independent of the setup geometry. We are the first to 78 observe the translation of entire microbubbles into cells. Figure 3 shows a 3D reconstruction 79 of the results seen. Targeted drug delivery down to the cellular level, with the use of 80 encapsulated bubbles will allow the use of high-toxicity drugs to be injected into the body, 81 but only delivered to a specific area. Thus, leaving healthy tissue unaffected. Our 82 sonoporation observations could be attributed to the long pulse lengths used. Although cells 83 themselves are acoustically active, this acoustic activity is probably negligible to that of microbubbles in high concentrations. Therefore, we expect bubble-cell interaction to be 84 85 more likely in very low bubble concentrations.

88 CONCLUSION

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- 90 It has been demonstrated that lipid-shelled microbubbles can be forced to enter cells at a
- 91 low MI. Hence, if a therapeutic load is added to the bubble, ultrasound guided delivery
- 92 could be facilitated at diagnostic settings. In addition, these results may have implications
- 93 for the safety regulations on the use of ultrasound contrast agents for diagnostic imaging.

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- 120



- 123 Figure 1. Possbile mechanisms of sonoporation.



- 127 Figure 2. Confocal fluorescence configuration used to simultaniously observe and sonicate
- 128 HeLa cells in the pressance of fluorescence coated microbubbles.



- 133 Figure 3. 3D reconstruction of microbubble translation through a cell membrane. Once the
- 134 microbubble has translated through the membrane, the pore re-seals and the microbubble
- is seen to disolve.