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2,5-diketopiperazines mitigate the amount of advanced glycation end-products accumulated with age in human dermal fibroblasts

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ABSTRACT

OBJECTIVE: Glycation is a common non-enzymatic reaction between proteins and sugars, resulting in the formation of advanced glycation end products (AGEs) in the human body. As can be seen in diabetic patients, the accumulation of AGEs in the skin has aesthetic consequences (wrinkles, brown spots, yellowish complexion). Therefore, the objective of this work was to find compounds isolated from natural sources that could eliminate the final AGEs accumulated in the skin with aging.

METHODS & RESULTS: A preliminary screening performed on a bank of microbial extracts and pure compounds showed that 2,5-Diketopiperazines (DKPs), as well as the extract of *Sphingobacterium* sp (SNB-CN13), reduced the presence of AGEs in fibroblasts by -28% and -23% respectively. In this article, we present the dereplication approach used to reveal the presence of 26 different DKPs in the crude extract of *Sphingobacterium* sp. Bioguided fractionation has led to the isolation of 12 of them, whose identity has been confirmed by HRMS and NMR. A green synthesis approach has been developed to synthesize 3 symmetrical DKPs. The biological activity of all DKPs was evaluated by the development of an in vitro test using immunocytochemistry to reveal the presence of AGE carboxymethyl-lysine in human dermal fibroblasts.

CONCLUSION: Our work shows for the first time that DKPs decrease the amount of carboxymethyl-lysine AGE in elderly human dermal fibroblasts grown in vitro. Therefore, diketopiperazines can be considered as compounds of interest for dermatological and cosmetic applications with an anti-ageing aim.

Cell culture ; chemical analysis ; skin physiology ; 2,5-diketopiperazines ; molecular networks ; AGEs/CML clearance.

INTRODUCTION

Louis-Camille Maillard described the glycation reaction for the first time in 1912 (1). This naturally occurring non-enzymatic reaction between reducing sugars and amino-acid residues of proteins leads to the formation of a complex and heterogenous group of compounds named advanced glycation end-products (AGEs).

AGEs accumulate in the whole body during the lifetime, both in the extra- and intracellular environments (2). Long lifespan proteins of the body, like collagen, elastin and hemoglobin are well-known targets of irreversible modifications caused by the glycation reaction (3).

At the extracellular level, proteins crosslinking induces inhibition of cell growth, impaired cell adhesion, and tissue dysfunction (4). At the intracellular level, AGEs cause cell damage directly by altering protein functions, and indirectly by producing reactive oxygen species (ROS) (5). Excessive levels of ROS can then alter cell detoxification systems such as the proteasome, as well as the enzymes involved in cell repair (6). AGEs can also bind to the Receptor of Advanced Glycation End products (RAGE), triggering the release of inflammatory mediators by the NF- κ B pathway (7). Finally the inflammation and oxidation reactions triggered by AGEs will result in the formation of more AGEs in cells. These phenomena also reduce the antioxidant and repair potential of cells, which will eventually lead to dysfunction of tissues and organs (8). Consequently, AGEs are considered to be the main agents responsible for many age-related diseases (3) such as diabetic vascular complications (9), atherosclerosis (10), or Alzheimer's disease (11).

The glycation reaction does not spare the skin. AGEs have been shown to accumulate in both the dermis and epidermis (12,13). Pentosidine, a fluorescent AGE, and carboxymethyl-lysine, an AGE that forms protein adducts, are the two main final glycation products found in the skin (12). The accumulation of AGEs in the intra- and

extracellular skin environment causes, among other things, a stiffening of collagen, elastin or fibronectin fibers, as well as an increase in melanogenesis (14,15). The skin is thus less elastic and more sensitive to the various mechanical stimuli it endures. Also, the complexion is more yellow and less homogeneous, which inevitably affects its youthful appearance overtime (15–17), as seen in people with diabetes (19,20). Most compounds on the cosmetic market act by preventing or reversing the first step of the glycation reaction, *i.e.*, the binding between proteins and sugars leading to the formation early glycation products, also called Amadori products. Looking at their mode of action, none of these ingredients really affect the reduction in the amount of final AGEs present in elderly cells. Therefore, the dermatological, cosmetic and therapeutic interest in discovering compounds that reduce the amount of AGEs accumulated in skin is important.

In this context, we screened a bank of pure compounds and microbial extracts to find suitable candidates to reduce both the total amount of AGEs, and the amount of pentosidine in human dermal fibroblasts (21). During screening, molecules belonging to the family of 2,5-diketopiperazines, and an ethyl acetate extract of *Sphingobacterium* sp. (SNB-CN13) showed a reduction in AGE content of fibroblasts by more than 20%. In this article, we present for the first time the activity of naturally occurring 2,5-DKP on the elimination of carboxymethyl lysine AGE (CML) in human dermal fibroblasts and evaluate their potential as active ingredients in dermatology and cosmetics.

MATERIAL AND METHODS

General experimental procedures. Optical rotations were measured on an Anton Paar MCP 300 polarimeter in a 100 mm-long 350 μ L cell. UV spectra were recorded using a Perkin-Elmer Lambda 5 spectrophotometer. CD spectra were measured at 25°C on a

JASCO J-810 spectropolarimeter. Nuclear Magnetic Resonance (NMR) spectra were recorded on a Bruker 500 MHz spectrometer or a Bruker 600 MHz spectrometer equipped with a 1 mm inverse detection probe. Chemical shifts (δ) are reported in ppm based on TMS signal, and coupling constants (J) are reported in Hertz. HRESIMS measurements were performed using a Waters Acquity UHPLC system with column bypass coupled to a Waters Micromass LCT premier Time-of-Flight mass spectrometer (Milford, Ma, USA) equipped with an electrospray interface (ESI). X-ray diffraction data for compound **11** were obtained at room temperature on a Rigaku Rapid II diffractometer equipped with a rotating anode mm007 HF generator and Osmic mirrors (Cu $K\alpha$ radiation, $\lambda = 1.54187 \text{ \AA}$) using ω -scans. Data were indexed, integrated and scaled using CrystalClear(22). They were also corrected for polarization, Lorentz and absorption effects (FS_ABSCOR). X-ray diffraction data for compound **20** were obtained at room temperature on a Rigaku XtaLabPro diffractometer equipped with a microfocus source (MicroMax003_Mo) and multilayer confocal mirrors (Mo $K\alpha$ radiation, $\lambda = 0.71073 \text{ \AA}$). Data were indexed, integrated and scaled using CrysAlisPro. They were also corrected for polarization, Lorentz and absorption effects (SCALE3 ABSPACK). For each compound, the structure was solved with the ShelXT (23) structure solution program using Direct Methods and refined with the ShelXL (24) refinement package using Least Squares minimization. All non-hydrogen atoms were refined with anisotropic displacement parameters and H atoms have been added geometrically and treated as riding on their parent atoms. Molecular graphics were computed with Ortep 3 (25). Flash chromatography was performed on a Grace Reveleris system with dual UV and ELSD detection equipped with a 120 g C18 column. For UV-based experiments, effluents were monitored at 254 and 280 nm. Analytical and preparative HPLCs were conducted on a Gilson system equipped with a 322 pumping device, a GX-271 fraction collector, a

171 diode array detector and a prepELSII detector electrospray nebulizer. Columns used for these experiments included: a Phenomenex Luna C₁₈ or PFP (2) 5 μm 4.6 x 250 mm analytical column, and Phenomenex Luna C₁₈ or PFP (2) 5 μm 21.2 x 250 mm preparative column. The flow rate was set to 1 or 21 mL/min respectively, using a linear gradient of water mixed with an increasing proportion of acetonitrile. All solvents were HPL grade and supplemented with 0.1% of formic acid. The Mueller-Hinton (MH) was purchased from Conda. TritonX-100 was from Euromedex (Euromedex France, Souffelweyersheim, France). DMSO was obtained from Carlo Erba (Carlo Erba Reagents S.A.S, Val de Reuil, France). Tween-20 was obtained from Acros Organics (Acros Organics, Geel, Belgium). Bovine serum albumin (BSA) was obtained from Dutscher (Dominique Dutscher SAS, Brumath, France). Paraformaldehyde was obtained from Electron Microscopy Science (EMS, Hartfield, PA, U.S.A). Mouse anti-CarboxyMethyl-Lysine (CML) antibody (KH011) was obtained from Cosmo Bio Co (Cosmo Bio Co LTD, Tokyo, Japan). Goat anti-mouse secondary antibody Alexa-488 conjugate (A11017) was obtained from Lifetechnologies (ThermoFisher Scientific, Waltham, MA, U.S.A). Mounting medium with DAPI was obtained from Vector (Vector Laboratories Inc, Burlingame, CA, U.S.A).

Isolation and identification of *Sphingobacterium spp.* SNB-CN13. Pieces of termite nests were collected and cultured as described in our previous work (26). The procedure adopted for identification of the strain was also describe in the same article (26), and the strain was saved at -80 °C at the Métabolites de micro-organismes symbiotiques – Bioinspiration laboratory, ICSN/CNRS, France. The bacteria rDNA sequence obtained was aligned with sequences from GenBank, NCBI (<http://www.ncbi.nlm.nih.gov/genbank/> consulted on December 15, 2019), indicating

that SNB-CN13 was closely related to *Sphigobacterium* species. The sequence was deposited on the GenBank with the accession number FJ959369.1.

Culture of bacterium, extraction and isolation of compounds. *Sphigobacterium* sp. was cultivated on solid Mueller-Hinton medium at 28°C for one week on 101 Petri dishes that were 14 cm in diameter (total 1.4 m²). The contents of the Petri dishes were cut into small pieces, transferred into a large container and macerated with ethyl acetate for 24 hours. The organic solvent was collected via filtration, washed with water in a separating funnel, and evaporated to dryness under reduce pressure to yield 3.24 g (2.3 g/m²).

A portion of the ethyl acetate extract (2.6 g) was fractionated using flash chromatography using a gradient of water mixed with increased proportions of acetonitrile (H₂O:CH₃CN 95:5 – 80:20 – 50:50 – 20:80 – 0:100) at 80 mL/min using a C18 120 g column. Five fractions were then collected based on the solvent steps described above. During fractionation, a precipitate was formed in a tube of fraction F2, corresponding to compound **8** (2 mg). Fractions F1 (18.7 mg) and F2 (191 mg) retained biological activity and were subjected to further fractionation. Fraction F1 was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 95:5 over 35 minutes, flow rate 0.6 mL/min) to yield compound **18** (1.5 mg, RT = 8.2 min) and **15** (2.6 mg, RT = 26.5 min). Fraction F2 was purified by preparative HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃CN + 0.1% FA, isocratic elution 88:12 over 30 minutes, flow rate 21 mL/min) to afford compound **6** (2.3 mg, RT = 14.8 min), **11** (4.2 mg, RT = 9.0 min) and sub-fractions F2-4, F2-5, F2-6, F2-7, F2-8, F2-9, F2-11, and F2-13 that were subjected to further purification. Sub-fraction F2-4 (4.3 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O

+ 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 85:15 over 40 minutes, flow rate 1 mL/min) to afford compounds **9** (0.4 mg, RT = 31.7 min), and **10** (1.2 mg, RT = 34.0 min). Sub-fraction F2-5 (1.5 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 77:23 over 20 minutes, flow rate 1 mL/min) to afford compound **4** (0.4 mg, RT = 17.8 min). Sub-fraction F2-6 (10 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 80:20 over 30 minutes, flow rate 1 mL/min) to afford compounds **17** (1.5 mg, RT = 15.5 min), and **5** (0.4 mg, RT = 25.1 min). Sub-fraction F2-7 (1.5 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 80:20 over 35 minutes, flow rate 1 mL/min) to yield compound **13** (0.2 mg, RT = 17.6 min). Sub-fraction F2-8 (6 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 80:20 over 25 minutes, flow rate 1 mL/min) to afford compounds **14** (2.1 mg, RT = 13.7 min), **7** (1.3 mg, RT = 17.0 min), and **2** (0.7 mg, RT = 19.3 min). Sub-fraction F2-9 (3.1 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 80:20 over 35 minutes, flow rate 1 mL/min) to yield compound **12** (1.4 mg, RT = 26.9 min). Sub-fraction F2-11 (2.4 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 80:20 over 30 minutes, flow rate 1 mL/min) to afford compounds **3** (0.4 mg, RT = 21.7 min), and **1** (0.5 mg, RT = 23.0 min). Finally, sub-fraction F2-13 (4.9 mg) was purified by analytical HPLC (Luna PFP (2), mobile phase H₂O + 0.1% FA / CH₃OH + 0.1% FA, isocratic elution 60:40 over 12 minutes, flow rate 1 mL/min) to afford compound **16** (0.7 mg, RT = 8.0 min).

All chemical descriptions of the isolated compounds (NMR, HREIMS) are available in the additional material section of this publication.

Cell preparation method. The skin explant of a 53-year-old donor comes from abdominal surgery and is prepared (hypodermis eliminated) and provided by the BioEC Laboratory (Longjumeaux, France).

The explant (epidermis + dermis) is then cut into discs of about 1 cm in diameter. The discs are placed in a T75 culture flange, the dermis in contact with the plastic of the flange. After a few minutes of rest the discs adhere to the culture flange. Culture medium is added (10 mL of DMEM, 20% FBS, 1% antibiotic/antifungal medium) so that the skin discs are not completely covered.

The flask is then placed in the oven at 37°C, 5% CO₂, and the culture medium is changed every 3 days. About a week later, a heterogenous culture of fibroblasts and keratinocytes is observed in the mother box. When the cells have proliferated over the entire surface of the flask, the cells are detached with a trypsin solution and stored in liquid nitrogen. During the first culture starting again from the cryotubes, only the fibroblasts colonize the flask, in DMEM medium with 10% FBS.

Before and during each test, cells are stained with trypan blue and counted using an automatic cell counter TC-20 (Bio-Rad, Marnes-la-Coquette, France). The cell viability of the counted sample is automatically analysed by the device according to the trypan blue exclusion principle. The percentage of viability of treated cells has always been above 95%.

Biological activity – Immunofluorescence of CML. Dermal fibroblasts were plated at 5000 cells/cm² on glass coverslips set into a 24-wells-plate and cultured in DMEM supplemented with 10 % FBS for 24 hours. On day 2, treatment with the molecules was done for 24 hours, at the concentration of 1 µg/mL. On day 3, medium containing the

treatment was removed, cells were washed with PBS, and fresh DMEM + 10 % FBS medium was added in wells. On day 4, the fibroblasts were fixed with 2 % paraformaldehyde solution for 12 minutes at room temperature, permeabilized with 0,2% TritonX-100 solution for 10 minutes and finally blocked with 5 % BSA, 0,2 % Tween-20 solution for further 10 minutes. Cells were then incubated with the primary antibody anti-CML (mouse) at a concentration of 7 $\mu\text{g}/\text{mL}$ at 4°C overnight, followed by incubation with Alexa-488 anti-mouse secondary antibody diluted at 1:200 for 2 hours at room temperature. After washing steps, coverslips were mounted onto microscope slides using a DAPI containing mounting medium and observed under a confocal photonic microscope (Spinning-Disk microscope, Roper/Nikon). Five images were recorded by coverslips (ten pictures per treatment condition). All pictures were taken with the same exposure time. Fluorescence quantification of images recorded was done using ImageJ software.

UPLC-MS/MS analysis of SNB-CN13 extract

Extract was diluted at 2 mg/mL in MeOH. Metabolome profiling was conducted with a Thermo UHPLC-HRMS system. Analyses of extract (1.0 μL injected, 2 μg on column) was performed in electrospray positive ionisation mode in the 133.4–2000 Da range in centroid mode. The mass detector was an Orbitrap MS/MS FT Q-Exactive focus mass spectrometer. The analysis was conducted in FullMS data dependent MS² mode. In FullMS, resolution was set to 70,000 and AGC target was $3 \cdot 10^6$. In MS², resolution was 17,500, AGC target 10^5 , isolation window 0.4 Da, normalized collision energy 30, with 20 s dynamic exclusion. UHPLC column was a Phenomenex Luna Omega polar C₁₈ 150×2.1 mm, 1.6 μm . The column temperature was set to 42 °C, and the flow rate was 0.5 mL/min. The solvent system was a mixture of water (A) with increasing proportions of

acetonitrile (B), both solvents modified with 0.1% formic acid. The gradient was as follows: 2 % B 3 min before injection, then from 1 min to 13 min, a linear increase of B up to 100 % followed by 100 % B for 5 min. The flow was discarded (not injected into the mass spectrometer) before injection and up to 1 min after injection.

MZmine 2 Data preprocessing Parameters

Raw files were directly processed using MZmine 2.32. The mass detection was realized keeping the noise at 20. The chromatogram building was achieved using a minimum time span of 0.01 min, minimum height of 1.0E7 and m/z tolerance of 0.0015 or 5 ppm. The local minimum search deconvolution algorithm was used with the following settings: chromatographic threshold = 1 %, minimum retention time range = 0.05 min, minimum relative height = 30 %, minimum absolute height = 5.0E5, minimum ratio of peak top/edge = 1, and peak duration range = 0.01 – 5.0 min. Chromatograms were deisotoped using the isotopic peaks grouper algorithm with an m/z tolerance of 0.0015 or 5 ppm, and an RT tolerance of 0.1 min. Peak alignment was performed using the RANSAC aligner method with the following parameters: m/z tolerance = 0.0015 or 5 ppm, absolute RT tolerance = 0.1 min, absolute RT tolerance after correction = 0.5 min, threshold value = 1). The peak list was gap filled with the peak finder module (intensity tolerance at 50%, m/z tolerance at 0.0015 or 5 ppm and absolute RT tolerance of 0.1 min).

Molecular Network analysis

A molecular network was created using the online workflow at Global Natural Products Social Networking (GNPS) (<http://gnps.ucsd.edu>) (27). The data was filtered by removing all MS/MS peaks within +/- 17 Da of the precursor m/z . MS/MS spectra were

window filtered by choosing only the top 6 peaks in the +/- 50 Da window throughout the spectrum. Precursor ion mass tolerance and fragment ion mass tolerance were set at 0.01 Da. Further, consensus spectra that contained less than 1 spectrum were discarded. A network was then created where edges were filtered to have a cosine score above 0.7 and more than 4 matched peaks. Further edges between two nodes were kept in the network if and only if each of the nodes appeared in each other's respective top 10 most similar nodes. The spectra in the network were then searched against GNPS' spectral libraries. The library spectra were filtered in the same manner as the input data. All matches kept between network spectra and library spectra were required to have a score above 0.7 and at least 4 matched peaks. Finally, molecular network was visualized using Cytoscape 3.4.0.

General procedure for the synthesis of symmetric 2,5-diketopiperazines. Thionyl chloride (SOCl₂, 1.67 mL, 22.9 mmol) was added dropwise to a solution of amino acid (15.19 mmol) in methanol (MeOH, 25 mL) at 0 °C under argon. The mixture was stirred for 19 h at room temperature. The solution was evaporated under reduced pressure. The residue was cooled at 0 °C and mixed with an aqueous solution of K₂CO₃ (50 %, 2.5 mL) for 15 minutes. The solution was then extracted with *tert*-butyl methyl ether (TBME), and the organic phase was dried with Na₂SO₄ before being evaporated under reduced pressure. The oily residue was then heated for 17 h between 80 and 105 °C depending on the amino acid used (Leu: 80 °C; Met: 80 °C; Pro: 105 °C). Then TBME (2 x 3 mL) was added to the reaction mixture and the precipitate was collected by filtration after cooling to afford the corresponding symmetric 2,5-diketopiperazine. The NRM, HRMS and crystallographic descriptions of the synthesized molecules can be found in the supplementary material.

RESULTS AND DISCUSSION

A preliminary examination of a bank of pure compounds and microbial extracts revealed that 2,5-diketopiperazines and an ethyl acetate extract of *Sphingobacterium* sp. SNB-CN13 reduced the AGE content of dermal fibroblasts by 28 and 23 %, respectively (21). The working hypothesis of this work was that the activity of the *Sphingobacterium* sp. extract could be attributed to the presence of DKPs.

First, the SNB-CN13 extract was analyzed by LC-MS². The collected data were used to build a molecular network to visualize the chemical diversity of the *Sphingobacterium* sp. extract (Fig. 1). Based on comparison with spectral databases and spectral similarities, we have identified two groups of related nodes containing DKPs in the network. By examining in detail the clustered MS² spectra in these nodes, we have identified successive characteristic losses of CO (-27.9 Da), NH₃ (-45.0 Da), and CO again (-73.0 Da) from the parent ion, as well as characteristic fragment ions of amino acids residues formed after the cleavage of the amide bonds. A typical fragmentation scheme is provided in the supporting information (Scheme S1). Overall, this information confirmed the identification of the parent ions as pseudomolecular diketopiperazines ions (28). Overall, 26 presumed DKPs were found in the crude extract of *Sphingobacterium* sp. (Table S1).

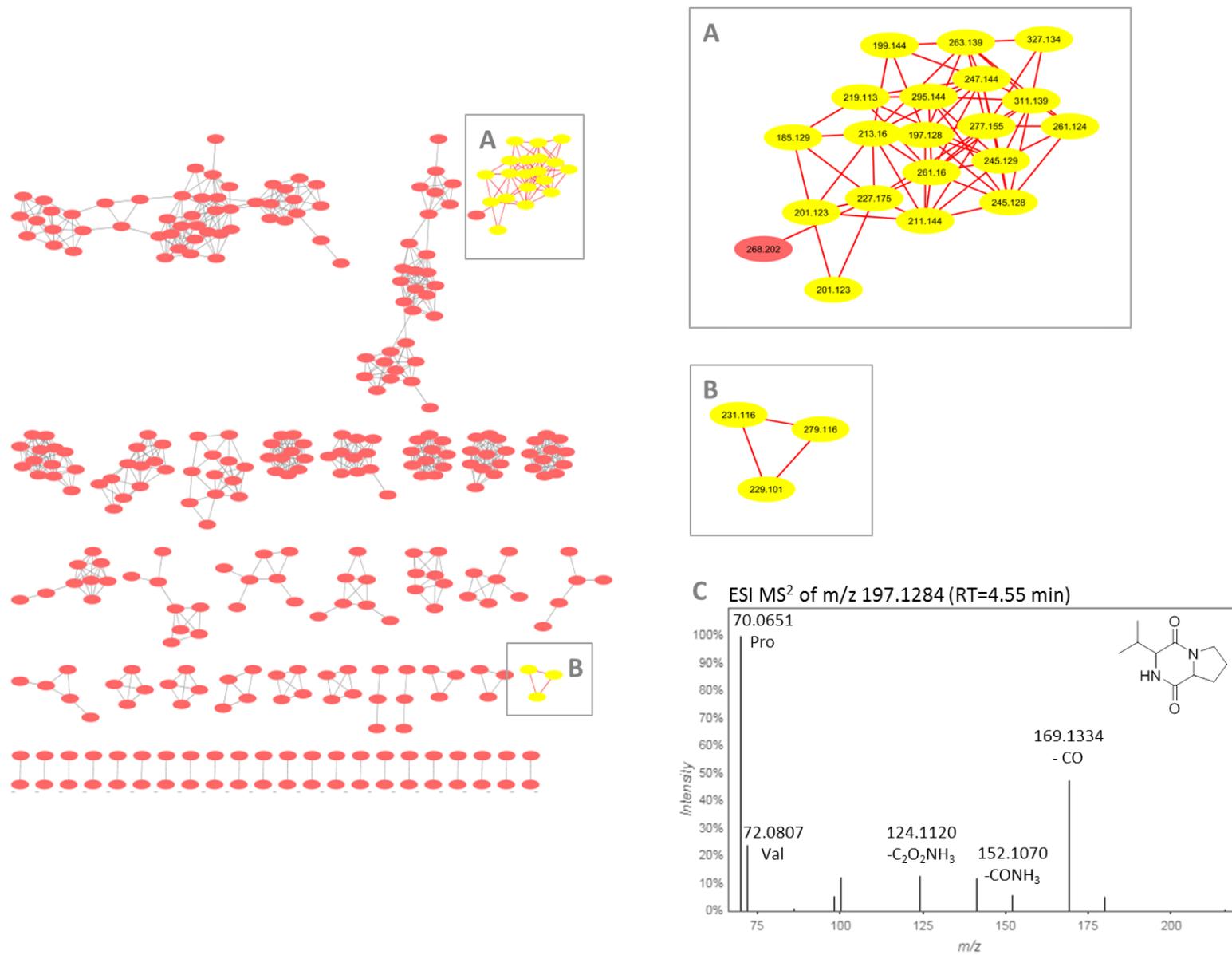


Figure 1. Molecular network of the crude extract of *Spingobacterium* sp.. Clusters A and B contain DKP structures (yellow nodes). C. Example of MS² spectrum (m/z 197.1284; RT 4.55 min) showing ion fragments characteristic of amino acids and losses characteristic of the DKP skeleton.

Bioguided fractionation of the crude extract was then carried out by flash chromatography and preparative HPLC, and twelve DKPs were isolated: cyclo(dehydroalanine-L-leucine) (**1**) (29), cyclo(dehydroalanine-L-isoleucine) (**2**) (29), cyclo(L-phenylalanine-L-alanine) (**3**) (30), cyclo(L-leucine-L-glutaric acid) (**4**) (31), cyclo(L-leucine-L-alanine) (**5**) (32), cyclo(L-tyrosine-L-valine) (**6**) (33), cyclo(L-tyrosine-L-tyrosine) (**7**) (34), cyclo(L-tyrosine-L-leucine) (**8**) (35), cyclo(L-proline-L-methionine) (**9**) (36), cyclo(L-tyrosine-L-proline) (**10**) (36), cyclo(L-proline-L-valine) (**11**) (37), and cyclo(L-proline-L-isoleucine) (**12**) (38) (Fig. 2).

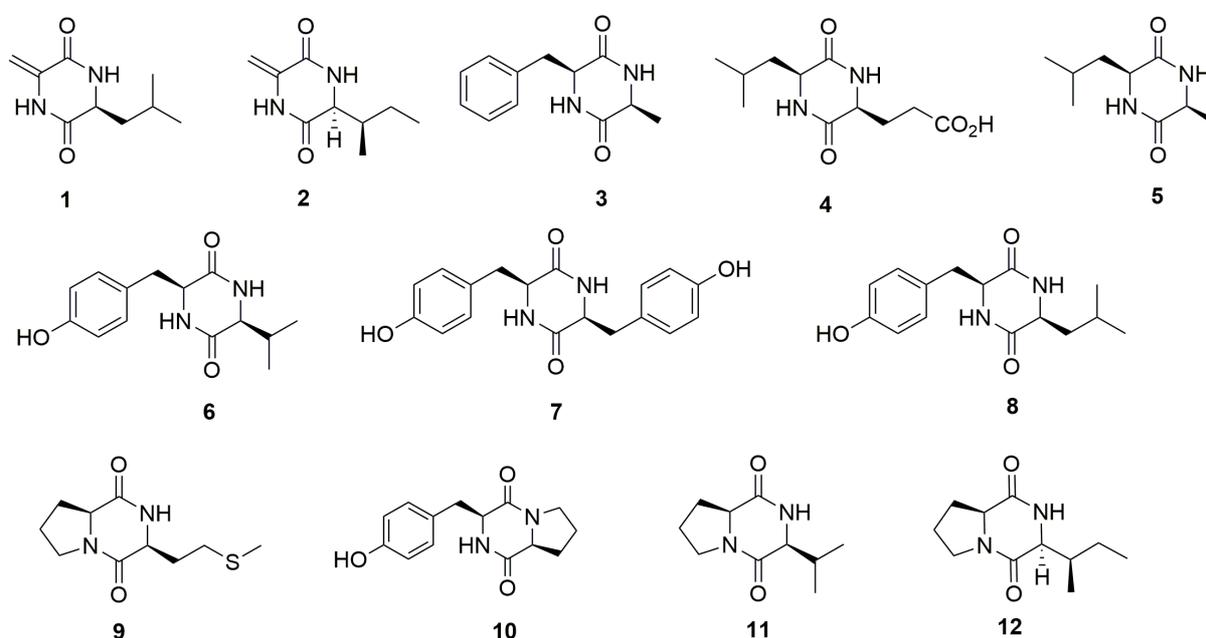


Figure 2. 2,5-Diketopiperazines **1-12** isolated from the crude extract of SNB-CN13 by bioguided fractionation.

Six other molecules were also isolated during the fractionation: benzamide (**13**) (39), 2-pyrrolicarboxylic acid (**14**) (40), glutaric acid (**15**) (41), 4-hydroxyphenylacetic acid (**16**) (42), 3-methylsulfanylpropionic acid (**17**) (43), and 3-methylsulfinylpropanoic acid (**18**) (43).

All compounds were identified by comparison of their spectroscopic data and physicochemical properties with the reported in the literatures, and determined to be identical.

Following these results, the synthesis of diketopiperazines was studied. In order to develop a cosmetic ingredient for the organic market, the development strategy turned to a synthesis protocol based on the principles of green chemistry. In this process, amino acids were esterified with MeOH/HCl, and then condensed by heating after deprotonation of the amino group (Fig. 3) (44).

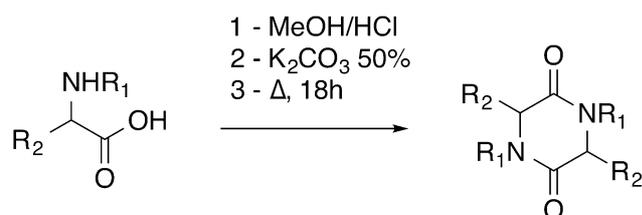


Figure 3. Synthesis process of symmetrical 2,5-DKP.

Using this protocol, the three symmetrical DKPs cyclo(L-leucine-L-leucine) (**19**), cyclo(L-methionine-L-methionine) (**20**), cyclo(L-proline-L-proline) (**21**) were obtained without epimerization (Fig. 4).

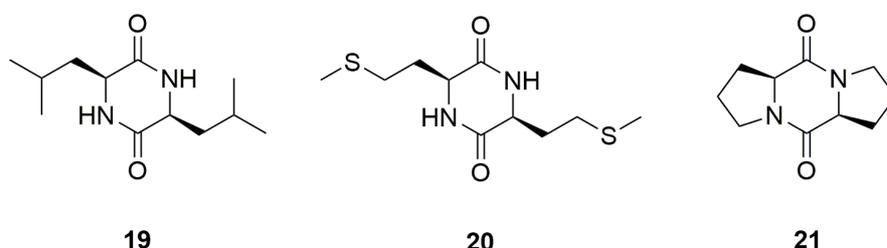


Figure 4. Symmetrical 2,5-DKPs synthesized.

The activity of the synthetic and isolated DKPs on the elimination of AGEs was then evaluated in human dermal fibroblasts. The *in vitro* test developed makes it possible to detect and quantify intracellular carboxymethyl lysine (CML) naturally formed and accumulated in human cells during aging, without using any glycation inducer. The decrease of the amount of CML in cells was evaluated by immunofluorescence. Compounds were tested at a concentration of 1 $\mu\text{g}/\text{mL}$ for 24 hours.

The intracellular level of CML in untreated dermal fibroblasts from a 53-year-old donor was quite high, resulting in significant green fluorescence of the cells during photonic confocal microscopy observation (Fig. 5a – Fibroblasts treated with DMSO-control). Isolated (compounds **1-12**) and synthesized (compounds **19-21**) 2,5-diketopiperazines had a different impact on the green fluorescence of CML in fibroblasts. As shown in Fig. 5b, cyclo(L-Pro-L-Met) (**9**) have little or no effect on the intensity of green fluorescence of CML in treated fibroblasts. Fig. 5c (fibroblasts treated with cyclo(L-Leu-L-Leu) (**19**)), Fig. 5d (fibroblasts treated with cyclo(L-Met-L-Met) (**20**)) and Fig. 5e (fibroblasts treated with cyclo(L-Pro-L-Pro) (**21**)) show a reduction in green fluorescence of CML, indicating that these compounds have an effect on the elimination of CML accumulated with aging in dermal fibroblasts.

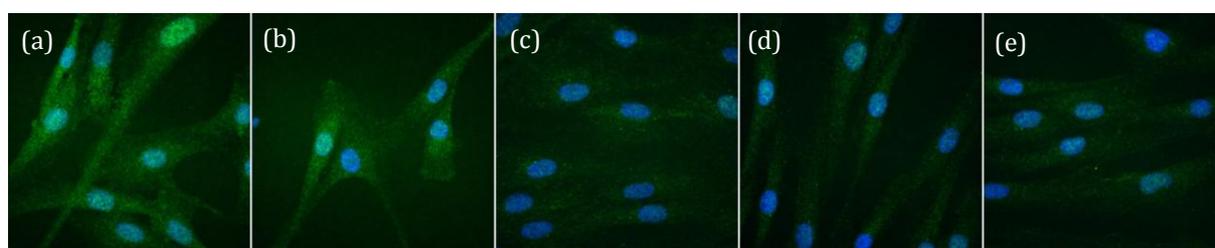


Figure 5. Confocal microscopy images (x 40) of human dermal fibroblasts treated for 24 h with (a) DMSO as control, (b) compound **9**, (c) compound **19**, (d) compound **20**, and (e) compound **21** at 1 $\mu\text{g}/\text{mL}$. The nucleus was labelled by DAPI (blue); The immunological labeling of CML was performed with Alexa 488 (green).

The fluorescence of the images recorded under the confocal photonic microscope was quantified. The expression of CML in fibroblasts treated with molecules **1-12** and **19-21** was compared to the control. The results were expressed as a percentage of the decrease in CML compared to the control group for each treatment and are presented in Table I.

Table I. Percentage of decrease in CML in fibroblasts treated with molecules **1-21**.

Molecules	Decrease of CML content (% compared to control)	p-value^a
SNB-CN13	23.0 ± 4.0	***
1 cyclo(Dha-L-Leu)	19.0 ± 1.6	***
2 cyclo(Dha-L-Ile)	5.5 ± 1.9	ns
3 cyclo(L-Phe-L-Ala)	16.1 ± 1.7	***
4 cyclo(L-Leu-L-Glu)	6.8 ± 2.5	ns
5 cyclo(L-Leu-L-Ala)	23.7 ± 2.1	***
6 cyclo(L-Tyr-L-Val)	22.6 ± 1.4	***
7 cyclo(L-Tyr-L-Tyr)	8.9 ± 1.4	*
8 cyclo(L-Tyr-L-Leu)	23.4 ± 1.1	***
9 cyclo(L-Pro-L-Met)	0.0 ± 4.2	ns
10 cyclo(L-Tyr-L-Pro)	17.0 ± 1.5	***
11 cyclo(L-Pro-L-Val)	23.3 ± 1.6	***
12 cyclo(L-Pro-L-Ile)	9.4 ± 1.7	*
19 cyclo(L-Leu-L-Leu)	28.0 ± 1.0	***
20 cyclo(L-Met-L-Met)	28.0 ± 1.0	***
21 cyclo(L-Pro-L-Pro)	27.0 ± 1.0	***

^aSignificance levels when compared to negative control were calculated by one-way Anova followed by Dunett's multiple comparison test. *** p < 0.001, * p < 0.05, ns = not significant

Among molecules **1-12** and **19-21**, twelve of them significantly decreased the intracellular amount of CML in fibroblasts, with values ranging from 8.9 % for cyclo(L-Tyr-L-Tyr) (**7**) to 28 % for cyclo(L-Leu-L-Leu) (**19**). Cyclo(L-Met-L-Met) (**20**). Cyclo(Dha-L-Ile) (**2**), cyclo(L-Leu-L-Glu) (**4**) and cyclo(L-Pro-L-Met) (**9**) were inactive. It

should also be noted that the symmetrical synthetic DKPs cyclo(L-Leu-L-Leu) (**19**), cyclo(L-Met-L-Met) (**20**) and cyclo(L-Pro-L-Pro) (**21**) showed the most important activity by significantly decreasing the intracellular amount of CML by 27.0–28.0 %.

Cell viability was assessed during cell counting using the trypan blue exclusion principle and revealed no decrease in fibroblast viability following treatment with diketopiperazines (viability >95%).

CML is a chemically very stable molecule. It is unlikely that the biological activity of DKP described in this paper results from a direct chemical action on CML. At the same time, it has been described that DKPs, due to their small steric size, are capable of binding to many cellular receptors (45). One of the hypotheses, which remains to be demonstrated, to explain the activity of diketopiperazines would be related to the AGER receptor. The AGER receptor has been described in many cells including fibroblasts (6). Its subunit 1 (AGER-1, OST 48), present on both the cell surface and the surface of the endoplasmic reticulum, specifically fixes terminal glycation products, without binding early glycation products such as Amadori products (46–48). The binding of intra- and extracellular AGEs to this receptor triggers their degradation, and has a protective effect against oxidative stress (6,46). With age or in the case of diabetes, the level of AGEs in the body is high. In both cases, there is also a decrease in the expression of the AGER-1 receptor (6,46).

This hypothesis that the activity of diketopiperazines on the elimination of AGEs in dermal fibroblasts is supported by an increase of a cellular detoxification process will have to be demonstrated in order to provide more details on the mechanism of action of diketopiperazines.

CONCLUSION

In this article we describe for the first time that some molecules belonging to the 2,5-diketopiperazine family act as detoxifying agents, capable of eliminating CML, a final AGE accumulated with age in dermal fibroblasts. 2,5-DKPs are natural cyclic dipeptides, produced by 90% of Gram-negative bacteria, but also by Gram-positive bacteria, fungi, plants and mammals. These molecules are also found in the human body (in the brain and human fluids) (49,50), and in industrially processed food (beer, bread, meat, chocolate...) (33,51).

We demonstrated that a de-replication strategy combining molecular network technique and MS2 data analysis was an effective tool to detect the presence of 26 potential DKPs in the crude extract of *Sphingobacterium* sp. SNB-CN13 and to predict their amino acid composition. The chemical study led to the isolation of 12 of the 26 diketopiperazines identified by the molecular network technique. Evaluation of the biological activity of the isolated DKPs showed that these molecules can remove advanced glycation end products present in the skin with age. It is interesting to note that symmetrical DKPs synthesized following the principles of green chemistry are also active on the same target.

Although the mode of action of the diketopiperazines remains to be confirmed, and given the elimination activity of advanced glycation end-products and the relative ease of synthesis of symmetrical compounds, diketopiperazines can be considered as compounds of interest for dermatological and cosmetic applications with an anti-ageing aim.

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