

Deciphering ascending thoracic aortic aneurysm hemodynamics in relation to biomechanical properties

Giuseppe de Nisco, Paola Tasso, Karol Calò, Valentina Mazzi, Diego Gallo, Francesca Condemi, Solmaz Farzaneh, Stéphane Avril, Umberto Morbiducci

▶ To cite this version:

Giuseppe de Nisco, Paola Tasso, Karol Calò, Valentina Mazzi, Diego Gallo, et al.. Deciphering ascending thoracic aortic aneurysm hemodynamics in relation to biomechanical properties. Medical Engineering & Physics, 2020. hal-03139655

HAL Id: hal-03139655 https://hal.science/hal-03139655v1

Submitted on 12 Feb 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Deciphering ascending thoracic aortic aneurysm hemodynamics in relation to biomechanical properties Giuseppe De Nisco¹, Paola Tasso¹, Karol Calò¹, Valentina Mazzi¹, Diego Gallo¹, Francesca Condemi², Solmaz Farzaneh², Stéphane Avril², Umberto Morbiducci^{1†} ¹ Polito^{BIO}Med Lab, Department of Mechanical and Aerospace Engineering, Politecnico di Torino, Turin, Italy ² Mines Saint-Etienne, Université de Lyon, INSERM, U 1059 SAINBIOSE, F - 42023 Saint-Etienne France [†]Address for correspondence: Umberto Morbiducci, Ph.D. Department of Mechanical and Aerospace Engineering, Politecnico di Torino Corso Duca degli Abruzzi, 24 - 10129 Turin, Italy Tel.: +39 011 0906882 Fax: +39 011 5646999 E-mail: umberto.morbiducci@polito.it

Abstract

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

The degeneration of the arterial wall at the basis the ascending thoracic aortic aneurysm (ATAA) is a complex multifactorial process, which may lead to clinical complications and, ultimately, death. Individual genetic, biological or hemodynamic factors are inadequate to explain the heterogeneity of ATAA development/progression mechanisms, thus stimulating the analysis of their complex interplay. Here the disruption of the hemodynamic environment in the ATAA is investigated integrating patient-specific computational hemodynamics, CT-based in vivo estimation of local aortic stiffness and advanced fluid mechanics methods of analysis. The final aims are (1) deciphering the ATAA spatiotemporal hemodynamic complexity and its link to near-wall topological features, and (2) identifying the existing links between arterial wall degeneration and hemodynamic insult. Technically, two methodologies are applied to computational hemodynamics data, the wall shear stress (WSS) topological skeleton analysis, and the Complex Networks theory. The same analysis was extended to the healthy aorta. As main findings of the study, we report that: (1) different spatiotemporal heterogeneity characterizes the ATAA and healthy hemodynamics, that markedly reflect on their WSS topological skeleton features; (2) a clear link (stronger than canonical WSS-based descriptors) emerges between the variation of contraction/expansion action exerted by WSS on the endothelium along the cardiac cycle, and ATAA wall stiffness. The findings of the study suggest the use of advanced methods for a deeper understanding of the hemodynamics disruption in ATAA, and candidate WSS topological skeleton features as promising indicators of local wall degeneration.

Highlights

52

- Different levels of spatiotemporal heterogeneity characterize the aneurysmal and healthy ascending aorta hemodynamics, reflecting on wall shear stress topological skeleton.
- Peculiar wall shear stress topological skeleton features are linked to local ascending thoracic
 aortic aneurysms stiffness.
- The topological shear variation index, a measure of wall shear stress luminal
 contraction/expansion action variation along the cardiac cycle, is an indicator of local aortic
 wall degradation, performing better than canonical wall shear stress-based descriptors of flow
 disturbances.
- Wall shear stress topological skeleton analysis, combined with Complex Networks theory,
 contributes to better determine whether arterial wall degeneration, in combination with
 hemodynamic insult, leads to aneurysmal progression/rupture.

Introduction

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

Ascending thoracic aortic aneurysm (ATAA) is an unphysiological localized ballooning of the aorta at the ascending segment proximal to the brachiocephalic trunk, which can grow in size and eventually rupture, leading to serious complications and death. The common clinical treatment of ATAA consists in surgical repair by replacing the diseased aortic segment with a synthetic graft [1,2]. Due to significant mortality rates associated to prophylactic surgery (3-5 %) [1], indications for surgical treatment depend on the estimated maximum aortic diameter, with a suggested critical threshold value of 5.5 cm (except for patients with Marfan syndrome) [2–4]. However, the diameter-based criterion for surgical intervention has been widely recognized as inadequate [5,6]. To find complementary or alternative markers of ATAA risk of rupture, a plethora of studies have focused on, e.g., genetic, biological, structural and biomechanical factors involved in the ATAA wall dilatation/degeneration [7–13]. However, current evidence is still insufficient to rely on biomechanical factors alone [10,12], and rather suggests a possible relation between ATAA hemodynamics and wall mechanical properties, which might better explain the complexity of ATAA development and progression. In particular, the analysis of aortic hemodynamics, with the assessment of distinguishable intravascular flow features and wall shear stress (WSS) distribution on the luminal surface as obtained from in vivo [14–17] and patient-specific computational studies [18–22], have been suggested to derive markers of rupture risk, giving the supposed link between hemodynamic features and ATAA wall mechanical properties. For instance, previous follow-up studies reported a significant association between the luminal exposition to relatively low WSS values and ascending aorta wall weakening and dilatation in ATAA patients [21,23,24]. In this context, we propose a framework combining patient-specific computational fluid dynamics (CFD), advanced fluid mechanics analysis and personalized in vivo estimates of the local aortic

stiffness. The aims are to interpret the complex role played by local hemodynamics in ATAA and to decipher the existing links between near-wall features, intravascular flow structures and ATAA wall stiffness. More specifically, changes in hemodynamics associated with pathologic aortic dilatation were analyzed by applying two recently proposed methodologies on two subjectspecific (1 healthy - 1 diseased) CFD models. The first approach relies on the analysis of WSS vector field topological skeleton, essentially consisting of a collection of critical points (i.e., locations at the luminal surface of the vessel where the WSS vanishes), and special lines which separate the luminal surface into areas of different WSS behavior, ultimately identifying where an expansion/contraction action is exerted by WSS on the endothelium (which is expected to have biological relevance) [25][26]. The second one exploits Complex Networks (CNs) theory to describe the spatiotemporal heterogeneity in near-wall and intravascular flow patterns, revealing information embedded in hemodynamic quantities waveforms (along the cardiac cycle) and useful to describe nonlinear phenomena involving them [27]. Both approaches were combined here to relate near-wall hemodynamics and intravascular flow patterns, which are expected to markedly affect WSS features [28,29]. Finally, ATAA local stiffness was obtained through a recently proposed inverse method based on gated CT scans [30]. Spatial correlations with WSS features were extracted from the topological skeleton analysis to ultimately associate ATAA complex hemodynamic features and wall mechanical properties.

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

Methods

One 36 years old healthy subject with no history of cardiovascular diseases, and one 59 years old subject with bicuspid aortic valve (BAV), moderate aortic valve insufficiency and a 6.03 cm x 5.95 cm ATAA were enrolled in the study after informed consent. An overview of the methods applied in this study is provided in Figure 1 and detailed onwards.

Medical imaging and geometry reconstruction

The two subjects were imaged by a 3T MR scanner (Siemens Magnetom Prisma) without contrast agent, using the same 4D flow MRI protocol exhaustively detailed elsewhere [22]. Based on 4D flow MRI images, the 3D geometries of each subject were reconstructed using CRIMSON (CardiovasculaR Integrated Modelling and SimulatiON) [31]. The resulting geometries included the ascending thoracic aorta, the aortic arch, the descending thoracic aorta and the supra-aortic vessels (brachiocephalic artery - BCA, left common carotid artery - LCCA, and left subclavian artery - LSA, as shown in Figure 1).

Additionally, the diseased subject underwent ECG gated CT imaging in order to identify ATAA wall local stiffness with a recently proposed inverse method [30] detailed below.

In vivo aortic wall stiffness estimation

A recently proposed non-invasive inverse approach was applied here to identify aortic wall stiffness for the diseased subject. Briefly, ATAA geometries at ten phases along the cardiac cycle, including diastolic and systolic phases, were (1) reconstructed from gated CT scans using MIMICS (v.10.01, Materialise NV), and (2) meshed ensuring a set of nodes attached to the same material points at all the recorded cardiac phases. Based on the Fourier series analysis of temporal changes in position of each node, the local strain distribution was reconstructed and the wall stiffness at

every location was derived by satisfying the local equilibrium equation under the acting tensions. The strategy applied to estimate local wall stiffness in vivo is exhaustively detailed elsewhere [30]. Computational hemodynamics The finite element-based open-source code SimVascular (http://simvascular.github.io/) was adopted to solve the governing equations of fluid motion in the two models. In detail, the Navier-Stokes equations, in their discretized form and under unsteady flow conditions, were solved using a stabilized finite element method supporting the use of linear tetrahedral elements (P1-P1) in the SimVascular flow solver for velocity and pressure [32]. Newtonian blood rheology (with prescribed constant density and dynamic viscosity respectively equal to 1050 kg m⁻³ and 0.0035 Pa s) and rigid wall with no-slip condition were assumed. To ensure a grid-independent solution, based on a mesh sensitivity analysis, an average tetrahedral element size of 1.05·10⁻³ m with a near-wall refinement consisting of 12 tetrahedral boundary layers with a decreasing ratio of 0.90 was adopted. The resulting computational grids consisted of 4.13 and 7.97 million elements for healthy and diseased model, respectively. Boundary conditions were prescribed using patient-specific 4D flow MRI measurements. In detail, measured fully 3D phase velocity data were extracted along the cardiac cycle and imposed in terms of Dirichlet inflow boundary conditions (BCs) at the ascending aorta (AAo) inlet section (as detailed in [33]). As regards outflow BCs, 4D flow MRI measured flow rates were prescribed at the supra-aortic vessels [34] in terms of fully developed velocity profiles. A three element Windkessel model was prescribed as BC at the descending aorta (DAo) outlet section, by applying a 3D-0D coupling scheme [22,31]. The three element Windkessel model components (i.e., the impedance -Z_C, the distal resistance - R, and the capacitor - C) were tuned as described in [22], obtaining patient specific values for the healthy ($Z_C = 1.36 \cdot 10^7 \text{ kg m}^{-4} \text{ s}^{-1}$; $R = 2.28 \cdot 10^8 \text{ kg m}^{-4} \text{ s}^{-1}$; $C = 1.50 \cdot 10^{-8}$

130

131

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

kg⁻¹ m⁴ s²) and the pathologic subject ($Z_C = 5.32 \cdot 10^6$ kg m⁻⁴ s⁻¹; $R = 9.02 \cdot 10^7$ kg m⁻⁴ s⁻¹; $C = 3.18 \cdot 10^{-8}$

154 $kg^{-1} m^4 s^2$).

On the simulated flow fields in the healthy and diseased models, advanced post-processing tools

were applied to better decipher the aortic hemodynamics complexity.

- 157 Deciphering aortic hemodynamics WSS topological skeleton analysis
- 158 Based upon dynamical systems theory, the topological skeleton of a vector field is made of fixed
- points and manifolds, where: (1) a fixed point is a point where the vector field vanishes; (2) vector
- 160 field manifolds, connecting fixed points, identify contraction/expansion regions (respectively,
- 161 stable/unstable manifolds).
- Here, a very recently proposed Eulerian method was considered to analyze the topological
- skeleton of the WSS vector field across the aortic luminal surface [25]. Based on the Volume
- 164 Contraction Theory, it has been demonstrated [25] that specific features of the WSS vector field
- topological skeleton, i.e., WSS manifolds, can be easily captured using the divergence of the
- 166 normalized WSS vector:

DIV =
$$\nabla \cdot \mathbf{\tau}_u = \nabla \cdot \left(\frac{\mathbf{\tau}}{|\mathbf{\tau}|}\right)$$
, (1)

- where τ_u is the unit vector of WSS vector τ . Eq. (1), neglecting the WSS vector magnitude
- variation, identifies the WSS spatial contraction/expansion configuration patterns [25]: negative
- values of DIV identify contraction regions, approximating attracting manifolds; positive values of
- DIV identify expansion regions, approximating repelling manifolds (Figure 2a).
- 172 According to the scheme proposed elsewhere for the complete WSS topological skeleton
- extraction [25], the identification of WSS fixed points location at the luminal surface was
- performed by computing a mesh-independent topological index, named Poincaré index. Then, the
- identified fixed points were classified using the eigenvalues of the Jacobian matrix, which provide
- information about fixed points nature (Figure 2). Technically, the adopted eigenvalue-based

criterion allowed to distinguish between fixed point node or focus configurations. The

combinations of the Poincaré index values and of eigenvalues λ_i (i = 1, 2, 3) of the Jacobian matrix

corresponding to the possible WSS fixed points configurations are summarized in Figure 2b.

As a first step, here the WSS topological skeleton of the cycle-average WSS vector field $\bar{\tau}$ at the

luminal surface of both the healthy and diseased model was analyzed. The unsteady nature of the

WSS vector field fixed points along the cardiac cycle was investigated using the WSS fixed points

weighted residence time along the cardiac cycle [25]:

184
$$RT\nabla_{x_{fp}}(e) = \frac{\bar{A}}{A_o} \frac{1}{T} \int_0^T \mathbb{I}_e(\mathbf{x}_{fp}, t) |(\nabla \cdot \mathbf{\tau})_e| dt, \qquad (2)$$

where $x_{fp}(t)$ is the WSS fixed point position at time $t \in [0, T]$, e is the generic triangular element 185 of the superficial mesh of area A_e , \bar{A} the average surface area of all triangular elements of the 186 superficial mesh, $\mathbb I$ is the indicator function and $(\nabla \cdot \mathbf \tau)_e$ is the instantaneous WSS divergence. 187 188 Eq. (2) quantifies the fraction of cardiac cycle spent by a fixed point inside a generic mesh surface element e on the aortic luminal surface emphasizing the contribution of fixed points surrounded 189 by a marked local contraction/expansion action. In fact, from a physical point of view, the term 190 $|(
abla \cdot oldsymbol{ au})_e|$ in eq. (2) allows to measure the strength of the contraction/expansion action of shear on 191 the wall. 192 193 Additionally, here a quantity based on WSS divergence, named Topological Shear Variation Index (TSVI), is introduced to measure the amount of variation in WSS contraction and expansion action 194 exerted at the luminal surface of the vessel. To do that, the root mean square of the normalized 195 WSS divergence is computed as follows: 196

197
$$TSVI = \left\{ \frac{1}{T} \int_0^T \left[\nabla \cdot (\mathbf{\tau}_u) - \overline{\nabla \cdot (\mathbf{\tau}_u)} \right]^2 dt \right\}^{1/2}, \tag{3}$$

198 Roughly speaking, eq. (3) allows to localize regions on the luminal surface of the vessel exposed to
199 large variations in the shear contraction/expansion action exerted by the flowing blood along the

cardiac cycle.

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

information about node connectivity as follows:

200

Deciphering aortic hemodynamics - Complex Networks analysis The spatiotemporal heterogeneity of intravascular and near-wall flow patterns was here investigated applying a recently introduced approach based on the application of the CNs theory to computational hemodynamics data [27,35], where the nodes of the network are represented by the waveforms, along the cardiac cycle, of specific hemodynamic quantities at the nodes of the aortic computational grid. Here, the CNs-based analysis was applied to the WSS vector magnitude ($|\tau|$) waveforms along the cardiac cycle, whereas intravascular hemodynamics was described in terms of axial velocity (V_{ax}) and kinetic helicity density (H_k). In detail, V_{ax} is the component of blood velocity vector along the local axial direction, identified as the direction of the tangent to the vessel's centerline [36]. By definition, V_{ax} is representative of the main flow direction. H_k is defined as the internal product between local velocity and vorticity vectors and its sign is an indicator of the (right- or left-handed) direction of rotation of helical blood flow patterns. The latter descriptor was involved in the analysis because of the established physiological significance of helical blood flow in main arteries [29,37,46,38–45]. For both investigated aortic models, three different CNs were built, starting from V_{ax} , H_k and $|\tau|$ time-histories, respectively. In the here adopted approach [27], two generic nodes i and j of the CN are connected by a topological link $\{i, j\}$ if the Pearson correlation coefficient R_{ij} between the waveforms of each considered hemodynamic quantity at the two nodes is larger than a given threshold \hat{R} . In this sense, each CN can be represented by its *adjacency matrix* A_{ij} containing the

223
$$A_{ij} = \begin{cases} 0, & \text{if } \{i, j\} \notin E \text{ or } i = j, \\ 1, & \text{if } \{i, j\} \in E. \end{cases}$$
 (4)

- Here, for each aortic model, and for each of the three CNs based on the hemodynamic quantities
- V_{ax} , H_k and $|\tau|$, respectively, the median values of the single R_{ij} distributions were set as threshold
- values \hat{R} to build up *adjacency matrices*, as proposed elsewhere [27,35].
- 227 Three metrics, based on A_{ij}, were used to characterize the structure of each CN. In detail, here we
- 228 considered the degree centrality DC_i of node i, defined as the percentage of nodes of the network
- connected to node *i*, i.e., the so-called *neighbors* of *i*:

230
$$DC_i = 100 \left(\frac{1}{N-1} \sum_{j=1}^{N} A_{ij} \right) = 100 \left(\frac{1}{N-1} dc_i \right).$$
 (5)

- In eq. (5) dc_i is the non-normalized degree centrality of node i. To allow comparisons between the
- healthy and the diseased model networks, characterized by different number of nodes (N), dc_i was
- 233 normalized to the total number (N-1) of possible neighbors of i. In summary, for a specific
- 234 hemodynamic descriptor, DC_i measures the fraction of time-histories correlated above threshold
- 235 with the time-history in node i, quantifying the spatiotemporal homogeneity/heterogeneity of the
- considered hemodynamic quantity waveform in each node, with respect to the whole network
- (i.e., the fluid volume for V_{ax} and H_k , or the luminal surface for $|\tau|$).
- The second CNs metric used in this study was applied to the V_{ax} and H_k -based networks to provide
- a quantitative measure of the anatomical length of persistence of the correlation of axial velocity
- and helical flow patterns inside the aorta. To do that, the so-called normalized average Euclidean
- 241 distance (AED_i) [27,47] of each node i of the network from all its neighbors n(i) was evaluated,
- 242 defined as:

$$243 AED_i = \frac{1}{D_{max}} \frac{\sum_{j \in n(i)} l_{ij}}{dc_i}, (6)$$

- where D_{max} is the maximum AAo diameter in the region upstream of the proximal supra-aortic
- trunk, and l_{ij} is the Euclidean distance between neighbor nodes i and j. In eq. (6), AED_i was

normalized with respect to D_{max} , in order to account for AAo geometric variability between the healthy and the diseased model. To investigate the effect that the presence of the ATAA has on the anatomical extent of the spatiotemporal correlations between V_{ax} and H_k structures, in both models AED_i was computed only in the aortic region from the inlet section to the first supra-aortic vessel bifurcation point.

The length of persistence of the correlation of WSS spatiotemporal patterns at the aortic luminal surface was evaluated in topological terms in the $|\tau|$ -based network of the two aortic models. This was done computing the *average shortest path length* $(\overline{SPL_i})$ of node i, defined as:

$$\overline{SPL}_i = \frac{1}{N-1} \sum_{j \in V, i \neq j} d_{ij} , \qquad (7)$$

where V is the set of nodes of the network, and $d_{ij} \in Z$ is the shortest topological distance between nodes i and j, i.e., the minimum number of links connecting node i with node j [48] (Figure 1). In synthesis, \overline{SPL}_i is a measure of the topological length of the correlation persistence of the $|\tau|$ waveform at each luminal surface node (i.e., the smaller the \overline{SPL}_i , the larger the topological persistence length of the correlation between i and the rest of the network). In order to be consistent with the computation of the anatomical persistence length of correlation (i.e., the AED_i metric), also \overline{SPL}_i was computed only in the aortic region from the inlet section to the first supraaortic trunk bifurcation point.

Deciphering aortic hemodynamics – Canonical WSS-based hemodynamic descriptors

In addition to the advanced methods for analyzing aortic hemodynamics introduced above, flow disturbances were evaluated here also in terms of canonical WSS-based descriptors (Table 1), namely the time-averaged wall shear stress (TAWSS), oscillatory shear index (OSI) [49], and relative residence time (RRT) [50]. Moreover, a descriptor of WSS multidirectionality was considered (Table 1), i.e., the transversal WSS (transWSS) [51], defined as the average WSS component acting orthogonal to the time-averaged WSS vector direction.

270 Aortic hemodynamics vs. wall stiffness

To investigate the existence of possible relations between ATAA hemodynamics and the mechanical properties of the aortic wall, an analysis of co-localization between exposure to "disturbed" hemodynamics vs. wall stiffness was carried out according to schemes proposed elsewhere [28,29,52]. More specifically, objective thresholds for WSS-based hemodynamic descriptors and for wall stiffness were identified as the 2nd tertile of the respective luminal distribution (1st tertile for TAWSS). The co-localization of luminal surface areas (SAs) characterized by stiffness and each one of OSI, RRT, transWSS, or *TSVI* values higher (lower for TAWSS) than the respective thresholds was quantitatively assessed by the similarity index (SI) [52]:

SI =
$$\frac{2(SA_{high \, stiffness} \cap SA_j)}{SA_{high \, stiffness} + SA_j},$$
 (8)

with *j* indicating a generic WSS-based hemodynamic descriptor. SI ranges between 0 (the SAs have no spatial overlap) and 1 (the SAs are equivalent and perfectly spatially overlapped).

Results

282

305

ATAA wall stiffness in vivo estimation 283 The in vivo estimated local wall stiffness distribution on the AAo of the diseased subject is 284 presented in Figure 3. Consistent with previous observations [53,54], local variations in the wall 285 286 mechanical properties are evident in the investigated model, where the highest values of the 287 stiffness are observed at the outer AAo wall. For this reason, in order to investigate the possible role played by hemodynamic features in the development and progression of aneurysm disease, 288 289 the analysis was here focused at the proximal AAo outer wall of both healthy and diseased 290 models, where the biomechanical wall alteration mainly occurred in the ATAA subject. WSS topological skeleton analysis 291 As suggested elsewhere [12], here the analysis of the WSS vector field topological skeleton at the 292 293 aortic luminal surface was applied in a two-step strategy. 294 As a first step, the aortic cycle-average WSS vector field topological skeleton was identified and 295 analyzed (Figure 4a) aiming at identifying (on average) differences between the healthy and ATAA 296 model. It emerged that on both models a line of marked WSS expansion was located at the 297 proximal AAo outer wall, as identified by positive DIV values. However, the healthy aorta only presented there an unstable node. Moreover, marked contraction lines characterized the WSS 298 299 topological skeleton on the ATAA outer wall, but they were not present on the luminal surface of 300 the healthy aorta. In the second step, the impact of the WSS vector field dynamics (along the cardiac cycle) on 301 topological skeleton features was analyzed. The analysis of the luminal surface distribution of fixed 302 points weighted residence time ($RTV_{x_{fn}}$, Figure 4b) highlighted the presence of a wide region at 303 304 the AAo outer-wall of the diseased model where instantaneous WSS fixed points appear and

reside for a longer fraction of cardiac cycle. Such region is surrounded by luminal regions with a

high local WSS contraction/expansion strength. On the other hand, a completely different distribution was observed on the outer wall of the healthy model (Figure 4b), mainly characterized by low $RT\nabla_{x_{fp}}$ values.

The analysis of the *TSVI* luminal distributions (Figure 4c) highlighted that: (1) the highest local variations in the contraction/expansion action exerted by the WSS on the endothelium along the cardiac cycle are located at two distinct regions of the AAo outer wall luminal surface, in both healthy and diseased models; (2) in the diseased model, high *TSVI* and high $RT\nabla_{x_{fp}}$ regions are markedly co-localized on the outer wall luminal surface.

CNs analysis

In the attempt to better describe differences between healthy and ATAA models accounting for possible links between the biomechanical alteration of the wall and hemodynamics, CNs were applied to the aortic flow and WSS fields.

The DC distributions of the V_{ax^-} , H_{k^-} and $|\tau|$ -based CNs are displayed in Figure 5 for both models. For visualization purposes, in each CN the DC values were classified in "high" and "low" based on the median value of the DC distribution of the healthy subject. Regarding the axial flow in the AAo of the diseased model (Figure 5a), a predominance of V_{ax} waveforms with a neighborhood which is representative of a portion of the whole aortic axial flow smaller than in the healthy model emerged, as highlighted by the low value DC distribution in that region.

Regarding H_k , the DC distribution in the AAo (Figure 5b): (1) confirms the presence of counterrotating helical flow patterns both in the healthy and in the ATAA model [22,36,38,44], as suggested elsewhere [27]; (2) highlights that helical flow (characterized considering H_k waveforms along the cardiac cycle) presents a spatiotemporal heterogeneity that in the ATAA model is markedly different from the healthy one, and which could be the consequence of the combination of the AAo dilatation and of the different shape of the (measured and imposed) inflow velocity

profile (reminding that (1) the investigated ATAA subject is affected by BAV, and (2) that the presence of valvulopathy could markedly affect the AAo hemodynamics [55]. Figure 5 confirms that the differences in the spatiotemporal intravascular complexity characterizing the ATAA and healthy models reflect WSS magnitude waveforms heterogeneity. In detail, the distal segment of the AAo of the ATAA model is characterized by an extended region of the luminal surface of the dilated aorta exposed to $|\tau|$ waveforms presenting a neighborhood which is smaller than in the healthy model (low DC values, Figure 5c). Therefore, in the diseased subject the pathologic expansion of the aortic wall where stiffness is higher (Figure 3) translates into WSS magnitude waveforms with a shape markedly different from the rest of the vessel. The impact of ATAA on the spatiotemporal hemodynamic heterogeneity in the AAo was evaluated also in terms of anatomical persistence length of the correlation between neighbor waveforms. AED maps (Figure 6a-b) highlight how the aortic dilation disrupts the persistence of the correlation of the V_{ax} and H_k waveforms, breaking the topological connections among waveforms in the AAo and shortening the length of correlation, compared to the healthy model. Finally, the heterogeneity of WSS magnitude spatiotemporal features induced by AAo dilatation, already emerged from the DC maps, was confirmed by \overline{SPL} maps. More specifically, it was observed that the luminal surface of the distal dilated AAo presents larger areas where the functional length of correlation of WSS magnitude waveforms is smaller than the healthy AAo (Figure 6c).

Aortic hemodynamics vs. wall stiffness

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

Here we assess the co-localization of the *in vivo* estimated wall stiffness at AAo outer wall of the ATAA model with the WSS topological skeleton-based quantities, and with canonical WSS-based descriptors of disturbed shear. A visual inspection of Figure 7 suggests that an association might exist between instantaneous WSS fixed points dynamics at the luminal surface and the aortic stiffness. Interestingly, instantaneous WSS fixed points undergo longer combinations of residence

time and marked contraction/expansion strength of the WSS vector field (as expressed by the quantity $RTV_{x_{fp}}$) on those ATAA luminal surface areas where stiffness is high (Figure 7). To explore more in depth the possible connections between peculiar WSS features and the altered biomechanical properties of the diseased aortic wall, the extent of the co-localization of high aortic wall stiffness regions with SAs exposed to presumably disturbed shear was quantified. By visual inspection of Figure 8a, it emerged that high stiffness co-localizes with high TSVI better than with canonical WSS-based descriptors. Of note, it was observed that SAs exposed to low TAWSS poorly co-localize with high stiffness regions. These observations were quantitatively confirmed by the SI values (Figure 8b), remarking the stronger capability of the WSS vector field divergence-based quantity TSVI as indicator of altered arterial wall mechanical properties, than the canonical WSS-based descriptors. Indeed, TSVI co-localizes with ATAA wall stiffness 206% more than TAWSS, 15% more than OSI and transWSS, and 45% more than RRT.

Discussion

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

Wall dilatation/degeneration of the ascending aorta is a complex multifactorial process promoted by a unique bio-chemo-mechanical environment, which may ultimately lead to aortic wall dissection/rupture [7–13]. Despite a large body of literature focused on the role played by individual biological, chemical, or biomechanical factors in the ATAA development [12], the heterogeneity characterizing ATAA disease cannot be exhaustively explained by individual features and the analysis of their complex interplay is still poorly investigated [10,12]. In this context, here we aim to highlight the possible links between ATAA hemodynamics, obtained by CFD simulations and explored with advanced methods, and in vivo estimated wall mechanical properties, inversely derived from gated CT scans [30]. The findings of this study confirm, on the basis of an in-depth quantitative exploration, that the near-wall and intravascular hemodynamic environment characterizing the ATAA model markedly differs from the healthy aorta. More in detail, the analysis of the WSS topological skeleton highlighted peculiar features on the ATAA outer wall, which are not or are poorly present on the luminal surface of the healthy ascending aorta (Figure 4), i.e.: (1) the presence of localized marked WSS contraction regions; (2) the appearance and persistence of instantaneous WSS fixed points along the cardiac cycle (as quantified by $RTV_{x_{fn}}$, which additionally weights the strength of the local contraction/expansion action exerted by the WSS); (3) a large luminal surface area undergoing marked variation in the WSS contraction/expansion action on the endothelium over the cardiac cycle (as indicated by TSVI). From the CNs-based analysis, marked differences between ATAA and healthy aorta emerged in the organization of the intravascular hemodynamic patterns. In detail, here for the first time the disruption of spatiotemporal homogeneity of intravascular hemodynamic features induced by the

AAo dilatation was quantified in terms of the Euclidian length of persistence of the correlation between flow quantities. As indicated by AED, the presence of aortic dilation seems to reduce the anatomical length of correlation persistence both for the axial velocity and kinetic helicity density waveforms, compared to the healthy case (Figure 6). This induced-by-aortic-dilation increase in AAo intravascular hemodynamics spatiotemporal heterogeneity reflects on a reduced functional length of correlation in WSS magnitude waveforms , localized on the distal outer wall, as indicated by \overline{SPL} (Figure 6). Such differences between ATAA and healthy models are further confirmed by the observed WSS magnitude DC luminal distribution, which highlights that in the dilated aorta the topologically isolated region is more extended than in the healthy one (Figure 5c). The latter result can be explained by the demonstrated causative role of helical flow in determining WSS patterns at the aortic luminal surface [28,29]. The evidences emerged from the WSS topological skeleton analysis and from the CNs-based analysis support the hypothesis that a complex interplay exists between hemodynamic features, aortic wall dilatation and its degradation in terms of wall mechanical properties. This statement is supported by the main finding of the study, i.e., that in the distal AAo outer wall of the ATAA model, extended high stiffness regions (indicating a degradation of the mechanical properties of the wall), co-localize with luminal surface areas of: (1) high instantaneous WSS fixed points residence time (weighted by WSS divergence value) (Figure 7); (2) large variations in the WSS contraction/expansion action exerted by the flowing blood along the cardiac cycle, here quantified by TSVI (Figure 8). It is worth noting that the cross-sectional nature of this study does not allow to establish whether the observed hemodynamic features are responsible for wall dilatation and degradation, or whether they are consequence of the dilated geometry. However, previous studies suggested that

391

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

altered aortic flow predisposes the ascending thoracic aorta to wall dilatation/degradation [21,23] as a consequence of the purported role of endothelial shear in the pathogenesis of ATAA [56]. In this sense, the findings of this study suggest that future studies addressing the role of hemodynamics in the ascending thoracic aorta to wall dilatation/degradation should include descriptors based on the WSS topological skeleton, such as *TSVI* (Figure 8). In fact, such descriptors show a higher co-localization with regions at the aortic wall characterized by high stiffness than canonical WSS-based descriptors. The hemodynamic quantities presented here may improve the prediction of ATAA development and progression and contribute to a deeper understanding of the underlying hemodynamics-driven processes.

Limitations

Several limitations could weaken the findings of this study. The main limitation is that the present analysis is based on only one ATAA model and one healthy aorta model. For this reason, we acknowledge that the here reported association between the WSS topological skeleton-based descriptor *TSVI* and ATAA wall weakening indicator should be confirmed on a larger dataset. The inverse approach here adopted to estimate local wall stiffness *in vivo* is based on the assumption that the aortic wall behaves as a membrane with no through-thickness shear [30,57]. The regions near the supra-aortic branches may not satisfy such assumption and were therefore excluded from the wall stiffness analysis. Moreover, in the perspective of extending the analysis of aortic stiffness at different stages of ATAA growth, an extensive use of the inverse approach may be limited by the use of X-ray during the gated CT scans procedure. In this regard, other techniques, such as ultrasounds [58] or MRI [59], may be adopted as a source of dynamic images of the aortic wall during the cardiac cycle.

Finally, the main limitation regarding the numerical settings of the CFD study is represented by the rigid wall assumption. However, recent observations have demonstrated that aortic wall motion has a minor impact on WSS-based descriptors of disturbed shear [60], and on intravascular flow topology. Concerning the latter, for example, similar helical fluid structures have been observed *in vivo* [38,44,61] and *in vitro* both in rigid and distensible aortic phantoms [62,63]. Therefore, we believe that the assumption on aortic wall distensibility does not entail the generality of the present results.

Conclusions

In this study, the integration of computational hemodynamics, *in vivo* stiffness estimation and advanced fluid mechanics methods allowed to explore more in depth the existence of links between the altered hemodynamics in the ATAA and the wall biomechanical degradation.

Moreover, combining WSS topological skeleton analysis and CNs-based analysis has highlighted that WSS topological skeleton features across the luminal surface of the ascending aorta are markedly affected by the different level of spatiotemporal heterogeneity characterizing the ATAA and healthy subjects' hemodynamics.

In conclusion, despite the exploratory nature of the study, advanced methods of analysis of the aortic blood flow have shown their very strong potential to decipher the links between local hemodynamics and the degradation of the mechanical properties of the aortic wall. In this sense, WSS topological skeleton analysis [25,26] and CNs-based analysis promise to be very effective tools [25,26].

References

- Isselbacher EM. Thoracic and abdominal aortic aneurysms. Circulation 2005;111:816–28. https://doi.org/10.1161/01.CIR.0000154569.08857.7A.
- Erbel R, Aboyans V, Boileau C, Bossone E, Bartolomeo R Di, Eggebrecht H, et al. 2014 ESC
 Guidelines on the diagnosis and treatment of aortic diseases: Document covering acute and
 chronic aortic diseases of the thoracic and abdominal aorta of the adult. The Task Force for
 the Diagnosis and Treatment of Aortic Diseases of the European . Eur Heart J 2014;35:2873–
 926. https://doi.org/10.1093/eurheartj/ehu281.
- Coady MA, Rizzo JA, Hammond GL, Kopf GS, Elefteriades JA. Surgical intervention criteria for thoracic aortic aneurysms: a study of growth rates and complications. Ann Thorac Surg 1999;67:1922–8. https://doi.org/10.1016/s0003-4975(99)00431-2.
- Johansson G, Markstrom U, Swedenborg J. Ruptured thoracic aortic aneurysms: a study of incidence and mortality rates. J Vasc Surg 1995;21:985–8. https://doi.org/10.1016/s0741-5214(95)70227-x.
- Pape LA, Tsai TT, Isselbacher EM, Oh JK, O'gara PT, Evangelista A, et al. Aortic diameter >or = 5.5 cm is not a good predictor of type A aortic dissection: observations from the International Registry of Acute Aortic Dissection (IRAD). Circulation 2007;116:1120–7. https://doi.org/10.1161/CIRCULATIONAHA.107.702720.
- Chau KH, Elefteriades JA. Natural history of thoracic aortic aneurysms: size matters, plus moving beyond size. Prog Cardiovasc Dis 2013;56:74–80. https://doi.org/10.1016/j.pcad.2013.05.007.
- 480 [7] Elefteriades JA, Farkas EA. Thoracic aortic aneurysm clinically pertinent controversies and uncertainties. J Am Coll Cardiol 2010;55:841–57.
 482 https://doi.org/10.1016/j.jacc.2009.08.084.
- Poullis MP, Warwick R, Oo A, Poole RJ. Ascending aortic curvature as an independent risk factor for type A dissection, and ascending aortic aneurysm formation: a mathematical model. Eur J Cardiothorac Surg 2008;33:995–1001. https://doi.org/10.1016/j.ejcts.2008.02.029.
- 487 [9] Gallo A, Agnese V, Coronnello C, Raffa GM, Bellavia D, Conaldi PG, et al. On the prospect of 488 serum exosomal miRNA profiling and protein biomarkers for the diagnosis of ascending 489 aortic dilatation in patients with bicuspid and tricuspid aortic valve. Int J Cardiol 490 2018;273:230–6. https://doi.org/https://doi.org/10.1016/j.ijcard.2018.10.005.
- 491 [10] Agnese V, Pasta S, Michelena HI, Minà C, Romano GM, Carerj S, et al. Patterns of ascending 492 aortic dilatation and predictors of surgical replacement of the aorta: A comparison of 493 bicuspid and tricuspid aortic valve patients over eight years of follow-up. J Mol Cell Cardiol 494 2019;135:31–9. https://doi.org/https://doi.org/10.1016/j.yjmcc.2019.07.010.
- 495 [11] Pasta S, Agnese V, Di Giuseppe M, Gentile G, Raffa GM, Bellavia D, et al. In Vivo Strain 496 Analysis of Dilated Ascending Thoracic Aorta by ECG-Gated CT Angiographic Imaging. Ann 497 Biomed Eng 2017;45:2911–20. https://doi.org/10.1007/s10439-017-1915-4.

498 [12] Wagenseil JE. Bio-chemo-mechanics of thoracic aortic aneurysms. Curr Opin Biomed Eng 2018;5:50-7. https://doi.org/10.1016/J.COBME.2018.01.002. 499 Petit C, Mousavi SJ, Avril S. Review of the Essential Roles of SMCs in ATAA Biomechanics. 500 [13] Adv Biomech Tissue Regen 2019:95-114. https://doi.org/10.1016/B978-0-12-816390-501 502 0.00006-6. Bollache E, Guzzardi DG, Sattari S, Olsen KE, Di Martino ES, Malaisrie SC, et al. Aortic valve-503 [14] mediated wall shear stress is heterogeneous and predicts regional aortic elastic fiber 504 505 thinning in bicuspid aortic valve-associated aortopathy. J Thorac Cardiovasc Surg 2018;156:2112-2120.e2. https://doi.org/https://doi.org/10.1016/j.jtcvs.2018.05.095. 506 [15] Suwa K, Rahman OA, Bollache E, Rose MJ, Rahsepar AA, Carr JC, et al. Effect of Aortic Valve 507 Disease on 3D Hemodynamics in Patients With Aortic Dilation and Trileaflet Aortic Valve 508 509 Morphology. J Magn Reson Imaging 2020;51:481–91. https://doi.org/10.1002/jmri.26804. [16] Dux-Santoy L, Guala A, Teixidó-Turà G, Ruiz-Muñoz A, Maldonado G, Villalva N, et al. 510 511 Increased rotational flow in the proximal aortic arch is associated with its dilation in 512 bicuspid aortic valve disease. Eur Hear J - Cardiovasc Imaging 2019;20:1407-17. https://doi.org/10.1093/ehjci/jez046. 513 Rodríguez-Palomares JF, Dux-Santoy L, Guala A, Kale R, Maldonado G, Teixidó-Turà G, et al. 514 [17] 515 Aortic flow patterns and wall shear stress maps by 4D-flow cardiovascular magnetic resonance in the assessment of aortic dilatation in bicuspid aortic valve disease. J 516 517 Cardiovasc Magn Reson 2018;20:28. https://doi.org/10.1186/s12968-018-0451-1. [18] Youssefi P, Gomez A, Arthurs C, Sharma R, Jahangiri M, Alberto Figueroa C. Impact of 518 519 Patient-Specific Inflow Velocity Profile on Hemodynamics of the Thoracic Aorta. J Biomech 520 Eng 2017;140. https://doi.org/10.1115/1.4037857. 521 [19] Pasta S, Gentile G, Raffa GM, Bellavia D, Chiarello G, Liotta R, et al. In Silico Shear and Intramural Stresses are Linked to Aortic Valve Morphology in Dilated Ascending Aorta. Eur J 522 Vasc Endovasc Surg 2017;54:254-63. 523 524 https://doi.org/https://doi.org/10.1016/j.ejvs.2017.05.016. [20] Mendez V, Di Giuseppe M, Pasta S. Comparison of hemodynamic and structural indices of 525 ascending thoracic aortic aneurysm as predicted by 2-way FSI, CFD rigid wall simulation and 526 527 patient-specific displacement-based FEA. Comput Biol Med 2018;100:221–9. 528 https://doi.org/https://doi.org/10.1016/j.compbiomed.2018.07.013. [21] Condemi F, Campisi S, Viallon M, Croisille P, Fuzelier J-F, Avril S. Ascending thoracic aorta 529 530 aneurysm repair induces positive hemodynamic outcomes in a patient with unchanged 531 bicuspid aortic valve. J Biomech 2018;81:145-8. https://doi.org/10.1016/j.jbiomech.2018.09.022. 532 [22] Condemi F, Campisi S, Viallon M, Troalen T, Xuexin G, Barker AJ, et al. Fluid- and 533 534 Biomechanical Analysis of Ascending Thoracic Aorta Aneurysm with Concomitant Aortic 535 Insufficiency. Ann Biomed Eng 2017;45:2921–32. https://doi.org/10.1007/s10439-017-1913-6. 536

Burk J, Blanke P, Stankovic Z, Barker A, Russe M, Geiger J, et al. Evaluation of 3D blood flow

[23]

538 539		patterns and wall shear stress in the normal and dilated thoracic aorta using flow-sensitive 4D CMR. J Cardiovasc Magn Reson 2012;14:84. https://doi.org/10.1186/1532-429X-14-84.
540 541 542 543	[24]	Guzzardi DG, Barker AJ, van Ooij P, Malaisrie SC, Puthumana JJ, Belke DD, et al. Valve-Related Hemodynamics Mediate Human Bicuspid Aortopathy: Insights From Wall Shear Stress Mapping. J Am Coll Cardiol 2015;66:892–900. https://doi.org/10.1016/j.jacc.2015.06.1310.
544 545 546	[25]	Mazzi V, Gallo D, Calo K, Najafi M, Khan MO, De Nisco G, et al. A Eulerian method to analyze wall shear stress fixed points and manifolds in cardiovascular flows. Biomech Model Mechanobiol 2019. https://doi.org/10.1007/s10237-019-01278-3.
547 548	[26]	Arzani A, Shadden SC. Wall shear stress fixed points in cardiovascular fluid mechanics. J Biomech 2018;73:145–52. https://doi.org/10.1016/j.jbiomech.2018.03.034.
549 550 551 552	[27]	Calo K, Gallo D, Steinman DA, Mazzi V, Scarsoglio S, Ridolfi L, et al. Spatiotemporal Hemodynamic Complexity in Carotid Arteries: an Integrated Computational Hemodynamics & Complex Networks-Based Approach. IEEE Trans Biomed Eng 2019. https://doi.org/10.1109/TBME.2019.2949148.
553 554 555	[28]	De Nisco G, Kok AM, Chiastra C, Gallo D, Hoogendoorn A, Migliavacca F, et al. The Atheroprotective Nature of Helical Flow in Coronary Arteries. Ann Biomed Eng 2019;47:425–38. https://doi.org/10.1007/s10439-018-02169-x.
556 557 558	[29]	De Nisco G, Hoogendoorn A, Chiastra C, Gallo D, Kok AM, Morbiducci U, et al. The impact of helical flow on coronary atherosclerotic plaque development. Atherosclerosis 2020. https://doi.org/10.1016/J.ATHEROSCLEROSIS.2020.01.027.
559 560 561	[30]	Farzaneh S, Trabelsi O, Avril S. Inverse identification of local stiffness across ascending thoracic aortic aneurysms. Biomech Model Mechanobiol 2019;18:137–53. https://doi.org/10.1007/s10237-018-1073-0.
562 563 564	[31]	Dillon-Murphy D, Noorani A, Nordsletten D, Figueroa CA. Multi-modality image-based computational analysis of haemodynamics in aortic dissection. Biomech Model Mechanobiol 2016;15:857–76. https://doi.org/10.1007/s10237-015-0729-2.
565 566 567	[32]	Updegrove A, Wilson NM, Merkow J, Lan H, Marsden AL, Shadden SC. SimVascular: An Open Source Pipeline for Cardiovascular Simulation. Ann Biomed Eng 2017;45:525–41. https://doi.org/10.1007/s10439-016-1762-8.
568 569 570 571	[33]	Morbiducci U, Ponzini R, Gallo D, Bignardi C, Rizzo G. Inflow boundary conditions for image-based computational hemodynamics: impact of idealized versus measured velocity profiles in the human aorta. J Biomech 2013;46:102–9. https://doi.org/10.1016/j.jbiomech.2012.10.012.
572 573 574 575	[34]	Gallo D, De Santis G, Negri F, Tresoldi D, Ponzini R, Massai D, et al. On the use of in vivo measured flow rates as boundary conditions for image-based hemodynamic models of the human aorta: implications for indicators of abnormal flow. Ann Biomed Eng 2012;40:729–41. https://doi.org/10.1007/s10439-011-0431-1.
576 577	[35]	Calò K, De Nisco G, Gallo D, Chiastra C, Hoogendoorn A, Steinman DA, et al. Exploring wall shear stress spatiotemporal heterogeneity in coronary arteries combining correlation-based

578 579		analysis and complex networks with computational hemodynamics. Proc Inst Mech Eng H 2020:(under review).
580 581 582 583 584	[36]	Morbiducci U, Gallo D, Cristofanelli S, Ponzini R, Deriu MA, Rizzo G, et al. A rational approach to defining principal axes of multidirectional wall shear stress in realistic vascular geometries, with application to the study of the influence of helical flow on wall shear stress directionality in aorta. J Biomech 2015;48:899–906. https://doi.org/10.1016/j.jbiomech.2015.02.027.
585 586 587 588	[37]	Gallo D, B. BP, Morbiducci U, Ye Q, (Joyce) XY, Maryam E, et al. Segment-specific associations between local haemodynamic and imaging markers of early atherosclerosis at the carotid artery: an in vivo human study. J R Soc Interface 2018;15:20180352. https://doi.org/10.1098/rsif.2018.0352.
589 590 591 592	[38]	Morbiducci U, Ponzini R, Rizzo G, Cadioli M, Esposito A, De Cobelli F, et al. In vivo quantification of helical blood flow in human aorta by time-resolved three-dimensional cine phase contrast magnetic resonance imaging. Ann Biomed Eng 2009;37:516–31. https://doi.org/10.1007/s10439-008-9609-6.
593 594 595	[39]	Liu X, Sun A, Fan Y, Deng X. Physiological significance of helical flow in the arterial system and its potential clinical applications. Ann Biomed Eng 2015;43:3–15. https://doi.org/10.1007/s10439-014-1097-2.
596 597 598	[40]	Stonebridge PA, Suttie SA, Ross R, Dick J. Spiral Laminar Flow: a Survey of a Three-Dimensional Arterial Flow Pattern in a Group of Volunteers. Eur J Vasc Endovasc Surg 2016;52:674–80. https://doi.org/10.1016/j.ejvs.2016.07.018.
599 600 601	[41]	Gallo D, Steinman DA, Bijari PB, Morbiducci U. Helical flow in carotid bifurcation as surrogate marker of exposure to disturbed shear. J Biomech 2012;45:2398–404. https://doi.org/10.1016/j.jbiomech.2012.07.007.
602 603 604	[42]	Gallo D, Steinman DA, Morbiducci U. An insight into the mechanistic role of the common carotid artery on the hemodynamics at the carotid bifurcation. Ann Biomed Eng 2015;43:68–81. https://doi.org/10.1007/s10439-014-1119-0.
605 606 607	[43]	Morbiducci U, Ponzini R, Grigioni M, Redaelli A. Helical flow as fluid dynamic signature for atherogenesis risk in aortocoronary bypass. A numeric study. J Biomech 2007;40:519–34. https://doi.org/10.1016/j.jbiomech.2006.02.017.
608 609 610 611	[44]	Morbiducci U, Ponzini R, Rizzo G, Cadioli M, Esposito A, Montevecchi FM, et al. Mechanistic insight into the physiological relevance of helical blood flow in the human aorta: an in vivo study. Biomech Model Mechanobiol 2011;10:339–55. https://doi.org/10.1007/s10237-010-0238-2.
612 613 614	[45]	De Nisco G, Kok AM, Chiastra C, Gallo D, Hoogendoorn A, Migliavacca F, et al. The Atheroprotective Nature of Helical Flow in Coronary Arteries. Ann Biomed Eng 2019;47. https://doi.org/10.1007/s10439-018-02169-x.
615 616 617	[46]	Kilner PJ, Yang GZ, Mohiaddin RH, Firmin DN, Longmore DB. Helical and retrograde secondary flow patterns in the aortic arch studied by three-directional magnetic resonance velocity mapping. Circulation 1993;88:2235–47. https://doi.org/10.1161/01.cir.88.5.2235.

618 619	[47]	Scarsoglio S, Iacobello G, Ridolfi L. Complex networks unveiling spatial patterns in turbulence. Int J Bifurc Chaos 2016;26:1–12. https://doi.org/10.1142/S0218127416502230.
620 621	[48]	Boccaletti S, Latora V, Moreno Y, Chavez M, Hwang DU. Complex networks: Structure and dynamics. Phys Rep 2006;424:175–308. https://doi.org/10.1016/j.physrep.2005.10.009.
622 623 624	[49]	Ku DN, Giddens DP, Zarins CK, Glagov S. Pulsatile flow and atherosclerosis in the human carotid bifurcation. Positive correlation between plaque location and low oscillating shear stress. Arteriosclerosis 1985;5:293–302. https://doi.org/10.1161/01.atv.5.3.293.
625 626 627 628	[50]	Himburg HA, Grzybowski DM, Hazel AL, LaMack JA, Li X-M, Friedman MH. Spatial comparison between wall shear stress measures and porcine arterial endothelial permeability. Am J Physiol Heart Circ Physiol 2004;286:H1916-22. https://doi.org/10.1152/ajpheart.00897.2003.
629 630 631	[51]	Peiffer V, Sherwin SJ, Weinberg PD. Computation in the rabbit aorta of a new metric - the transverse wall shear stress - to quantify the multidirectional character of disturbed blood flow. J Biomech 2013;46:2651–8. https://doi.org/10.1016/j.jbiomech.2013.08.003.
632 633 634	[52]	Gallo D, Steinman DA, Morbiducci U. Insights into the co-localization of magnitude-based versus direction-based indicators of disturbed shear at the carotid bifurcation. J Biomech 2016;49:2413–9. https://doi.org/10.1016/j.jbiomech.2016.02.010.
635 636 637	[53]	Choudhury N, Bouchot O, Rouleau L, Tremblay D, Cartier R, Butany J, et al. Local mechanical and structural properties of healthy and diseased human ascending aorta tissue. Cardiovasc Pathol 2009;18:83–91. https://doi.org/10.1016/j.carpath.2008.01.001.
638 639 640 641	[54]	Khanafer K, Duprey A, Zainal M, Schlicht M, Williams D, Berguer R. Determination of the elastic modulus of ascending thoracic aortic aneurysm at different ranges of pressure using uniaxial tensile testing. J Thorac Cardiovasc Surg 2011;142:682–6. https://doi.org/10.1016/j.jtcvs.2010.09.068.
642 643 644 645	[55]	Pirola S, Jarral OA, O'Regan DP, Asimakopoulos G, Anderson JR, Pepper JR, et al. Computational study of aortic hemodynamics for patients with an abnormal aortic valve: The importance of secondary flow at the ascending aorta inlet. APL Bioeng 2018;2:26101. https://doi.org/10.1063/1.5011960.
646 647 648	[56]	Michel JB, Jondeau G, Milewicz DiM. From genetics to response to injury: Vascular smooth muscle cells in aneurysms and dissections of the ascending aorta. Cardiovasc Res 2018;114:578–89. https://doi.org/10.1093/cvr/cvy006.
649 650 651	[57]	Farzaneh S, Trabelsi O, Chavent B, Avril S. Identifying Local Arterial Stiffness to Assess the Risk of Rupture of Ascending Thoracic Aortic Aneurysms. Ann Biomed Eng 2019;47:1038–50. https://doi.org/10.1007/s10439-019-02204-5.
652 653 654 655	[58]	Wittek A, Karatolios K, Fritzen C-P, Bereiter-Hahn J, Schieffer B, Moosdorf R, et al. Cyclic three-dimensional wall motion of the human ascending and abdominal aorta characterized by time-resolved three-dimensional ultrasound speckle tracking. Biomech Model Mechanobiol 2016;15:1375–88. https://doi.org/10.1007/s10237-016-0769-2.
656 657	[59]	Arani A, Arunachalam SP, Chang ICY, Baffour F, Rossman PJ, Glaser KJ, et al. Cardiac MR

658		amyloidosis. J Magn Reson Imaging 2017;46:1361–7. https://doi.org/10.1002/jmri.25678.
659 660 661	[60]	Lantz J, Renner J, Karlsson M. Wall shear stress in a subject specific human aorta - Influence of fluid-structure interaction. Int J Appl Mech 2011;03:759–78. https://doi.org/10.1142/S1758825111001226.
662 663 664 665	[61]	Frydrychowicz A, Berger A, Munoz Del Rio A, Russe MF, Bock J, Harloff A, et al. Interdependencies of aortic arch secondary flow patterns, geometry, and age analysed by 4-dimensional phase contrast magnetic resonance imaging at 3 Tesla. Eur Radiol 2012;22:1122–30. https://doi.org/10.1007/s00330-011-2353-6.
666 667 668 669	[62]	Gallo D, Gülan U, Di Stefano A, Ponzini R, Lüthi B, Holzner M, et al. Analysis of thoracic aorta hemodynamics using 3D particle tracking velocimetry and computational fluid dynamics. J Biomech 2014;47:3149–55. https://doi.org/https://doi.org/10.1016/j.jbiomech.2014.06.017.
670 671 672 673	[63]	Gulan U, Luthi B, Holzner M, Liberzon A, Tsinober A, Kinzelbach W. Experimental Investigation of the Influence of the Aortic Stiffness on Hemodynamics in the Ascending Aorta. IEEE J Biomed Heal Informatics 2014;18:1775–80. https://doi.org/10.1109/JBHI.2014.2322934.
674		

675 **Tables**

676

677

678 679 Table 1: Definition of WSS-based hemodynamic descriptors of "disturbed flow".

WSS-based hemodynamic descriptors

Time Average WSS (TAWSS) $TAWSS = \frac{1}{T} \int_0^T |\mathbf{\tau}| \, dt$ $\left[\left| \int_0^T \mathbf{\tau} \, dt \right| \right|$

Oscillatory Shear Index (OSI)
$$\text{OSI} = 0.5 \left[1 - \left(\frac{\left| \int_0^T \mathbf{\tau} \, dt \right|}{\int_0^T |\mathbf{\tau}| \, dt} \right) \right]$$

Relative Residence Time (RRT)
$$RRT = \frac{1}{TAWSS \cdot (1 - 2 \cdot OSI)} = \frac{1}{\frac{1}{T} \left| \int_0^T \mathbf{\tau} \, dt \right|}$$

Transversal WSS (transWSS)
$$\operatorname{transWSS} = \frac{1}{T} \int_0^T \left| \mathbf{\tau} \cdot \left(\mathbf{n} \times \frac{\int_0^T \mathbf{\tau} \, dt}{\left| \int_0^T \mathbf{\tau} \, dt \right|} \right) \right| dt$$

 τ is the WSS vector; T is the period of the cardiac cycle; \mathbf{n} is the unit vector normal to the arterial surface at each element.

680 Figures captions

- 681 **Figure 1. Schematics of the study.** The diagram shows how imaging data contribute to define vessel
- geometry, hemodynamic variables and wall mechanical properties. BCs: boundary conditions; CFD:
- computational fluid dynamics; ATAA: ascending thoracic aorta aneurysm; CNs: complex networks.
- 684 Figure 2. WSS topological skeleton features. (a) Configuration of each fixed point-type and
- contraction and expansion regions, colored by blue and red respectively. (b) Identification and
- classification of fixed points based on Poincaré index and eigenvalues of the Jacobian matrix.
- Figure 3. Estimated wall stiffness distribution at the luminal surface of ATAA model.
- 688 Figure 4. Luminal distributions of WSS topological skeleton-based metrics. Aortic luminal
- distributions of (a) cycle-average WSS topological skeleton, (b) WSS fixed points weighted residence
- time $(RT\nabla_{x_{fn}})$, and (c) TSVI. In panel (a), contraction/expansion regions are colored by blue/red,
- 691 respectively, and Line Integral Convolution technique is adopted to visualize WSS vector. The ATAA
- and healthy aortic models are displayed in the top and bottom panel, respectively.
- 693 **Figure 5.** *DC* distributions of hemodynamic descriptors-based CNs. The *DC* distributions of the (a)
- 694 $V_{\alpha x}$, (b) H_k , and (c) $|\tau|$ CNs are displayed for the diseased (top) and healthy (bottom) subjects.
- 695 Figure 6. Anatomical and topological length of correlation persistence of hemodynamic
- spatiotemporal patterns. The AED distributions of the (a) V_{ax} , and (b) H_k CNs, and (c) \overline{SPL}
- distribution of the $|\tau|$ CNs are displayed for the diseased (top) and healthy (bottom) subjects. AED
- and \overline{SPL} were computed at the ascending aorta segment of each model.
- 699 Figure 7. Wall stiffness vs. weighted residence time analysis. Contour maps of estimated wall
- stiffness (left) and WSS fixed points weighted residence time ($RT\nabla_{x_{fp}}$ right) at the luminal surface
- of ATAA model. The black contour line represents the 2nd tertile of stiffness luminal distribution at
- 702 the ATAA outer wall.

- 703 Figure 8. Wall stiffness vs. deranged aortic hemodynamics analysis. (a) co-localization maps of high
- stiffness and disturbed shear; (b) similarity indices quantifying the co-localization of luminal SAs
- 705 characterized by high stiffness and each descriptor of deranged hemodynamics (i.e., low TAWSS;
- 706 high OSI, RRT, transWSS, TSVI).

Figures

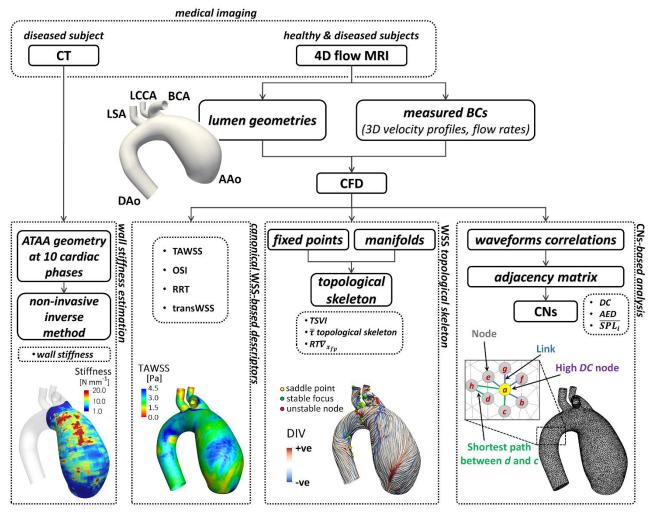


Figure 1. Schematics of the study. The diagram shows how imaging data contribute to define vessel geometry, hemodynamic variables and wall mechanical properties. BCs: boundary conditions; CFD: computational fluid dynamics; ATAA: ascending thoracic aorta aneurysm; CNs: complex networks.

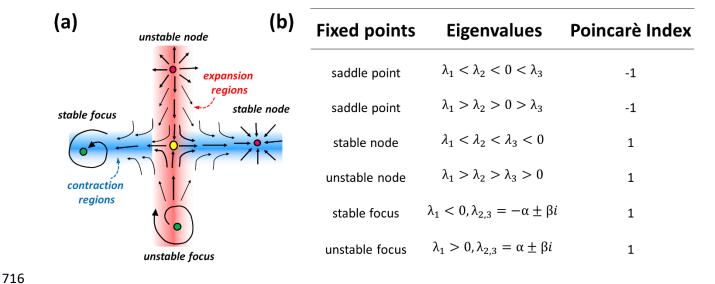


Figure 2. WSS topological skeleton features. (a) Configuration of each fixed point-type and contraction and expansion regions, colored by blue and red respectively. (b) Identification and classification of fixed points based on Poincaré index and eigenvalues of the Jacobian matrix.

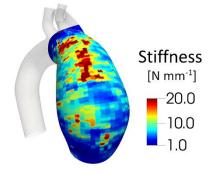


Figure 3. Estimated wall stiffness distribution at the luminal surface of ATAA model.

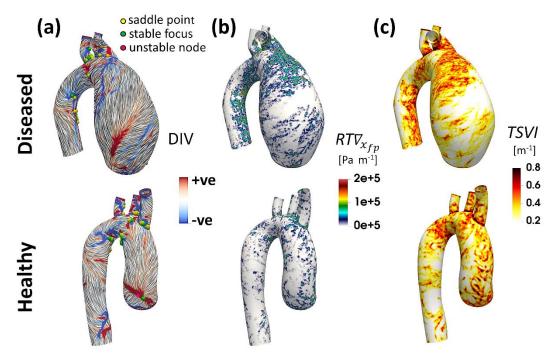


Figure 4. Luminal distributions of WSS topological skeleton-based metrics. A ortic luminal distributions of (a) cycle-average WSS topological skeleton, (b) WSS fixed points weighted residence time ($RTV_{x_{fp}}$), and (c) TSVI. In panel (a), contraction/expansion regions are colored by blue/red, respectively, and Line Integral Convolution technique is adopted to visualize WSS vector. The ATAA and healthy a ortic models are displayed in the top and bottom panel, respectively.

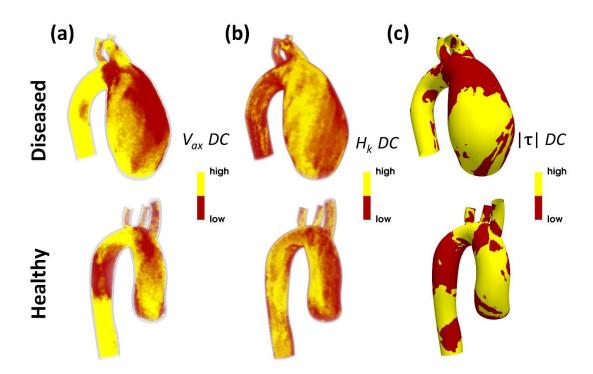


Figure 5. *DC* distributions of hemodynamic descriptors-based CNs. The *DC* distributions of the (a) $V_{\alpha x}$, (b) H_k , and (c) $|\tau|$ CNs are displayed for the diseased (top) and healthy (bottom) subjects.

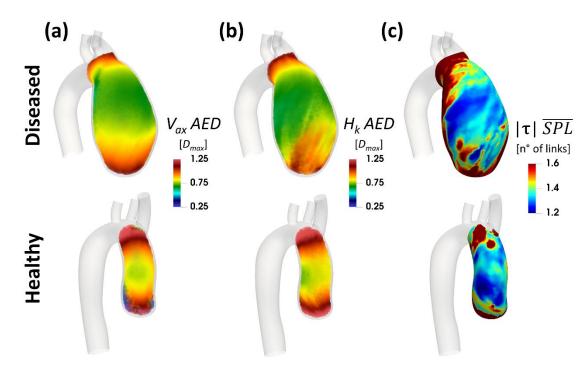


Figure 6. Anatomical and topological length of correlation persistence of hemodynamic spatiotemporal patterns. The *AED* distributions of the (a) V_{ax} , and (b) H_k CNs, and (c) \overline{SPL} distribution of the $|\tau|$ CNs are displayed for the diseased (top) and healthy (bottom) subjects. *AED* and \overline{SPL} were computed at the ascending aorta segment of each model.

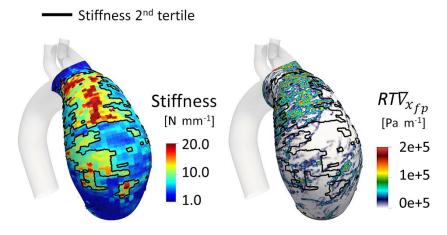


Figure 7. Wall stiffness vs. weighted residence time analysis. Contour maps of estimated wall stiffness (left) and WSS fixed points weighted residence time ($RTV_{x_{fp}}$ - right) at the luminal surface of ATAA model. The black contour line represents the 2nd tertile of stiffness luminal distribution at the ATAA outer wall.

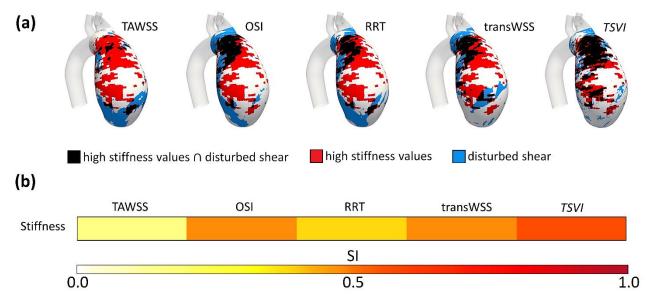


Figure 8. Wall stiffness vs. deranged aortic hemodynamics analysis. (a) co-localization maps of high stiffness and disturbed shear; (b) similarity indices quantifying the co-localization of luminal SAs characterized by high stiffness and each descriptor of deranged hemodynamics (i.e., low TAWSS; high OSI, RRT, transWSS, *TSVI*).