

A temperature- and photoperiod-driven model reveals complex temporal population dynamics of the invasive box tree moth in Europe

Christelle Suppo, Audrey Bras, Christelle Robinet

▶ To cite this version:

Christelle Suppo, Audrey Bras, Christelle Robinet. A temperature- and photoperiod-driven model reveals complex temporal population dynamics of the invasive box tree moth in Europe. Ecological Modelling, 2020, 432, pp.109229. 10.1016/j.ecolmodel.2020.109229. hal-03123286

HAL Id: hal-03123286 https://hal.science/hal-03123286

Submitted on 27 Jan 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	1	A temperature- and photoperiod-driven model reveals complex temporal
2 3 4	2	population dynamics of the invasive box tree moth in Europe
5 6 7	3	
9 10 11	4	Christelle Suppo ^a , Audrey Bras ^{b,c} and Christelle Robinet ^b
12 13 14 15	5 6 7	^a Institut de Recherche sur la Biologie de l'Insecte, UMR 7261, CNRS –Université de Tours, F-37200 Tours, France ^b INRAE, URZF, F-45075 Orléans, France
16 17	8	^c Unit of Chemical Ecology. Department of Plant Protection Biology. Swedish University of Agricultural
18	9	Sciences Alnarn Sweden
19 20	10	
21 22	10	
23	11	
24 25	12	
26 27 28	13	* Corresponding author: christelle.suppo@univ-tours.fr
28 29		
30 31	14	
32	15	
33 34	16	
35 26	17	
36 37	17	
38 39	18	
40	19	
41 42	20	
43	24	
44 45	21	
46	22	
48	23	
49 50	24	Number of tables: 5
52	25	<u>Number of figures</u> : 6
53 54 55	26	
56	27	Journal: Ecological Modelling, species issue "ISEM 2019"
57 58 59	28	<u>Type of paper</u> : Original research paper
60		
61 62		1
63		L
64 65		

Abstract

The box tree moth, Cydalima perspectalis, is an invasive insect that has rapidly colonised Europe, damaging to natural and ornamental box trees. In its native habitat in China, the number of generations per year is variable, but the number of generations observed in European climates remained unclear. This is a key issue for understanding the rapid increase in population size and range, and for optimising control. We developed a temperature- and photoperiod-driven model to simulate the life cycle of this insect and development rates for each life stage. The model was calibrated on published data and validated with observations obtained in France and Switzerland. Model stability analysis showed that minimal temperature for larval development to be the most important parameter to estimate. Diapause parameters had little effect. We then explored the effects of temperature increases of 1 and 2°C. The number of generations ranged from two to four at the various study sites. Climate warming will accelerate the insect life cycle, making it possible for the occurrence of one more generation per year. The key finding of this study was the complexity of population dynamics for this species. Some generations overlapped, making it difficult to identify the adult flight period clearly for each generation. Furthermore, various stages were potentially able to overwinter, not just diapausing larvae. Climate warming may also enhance this phenomenon in the future. Further explorations of the complex dynamics of this species are required, notably it remains unclear how successfully the various life stages survive winter temperatures. Further model refinements are also required to obtain more accurate estimates of box tree moth phenology. However, this is the first phenology model for box tree moth to be published, and our findings provide useful information for improving control of this pest.

Keywords: climate change; Cydalima perspectalis; phenology; diapause; modelling

53 1. Introduction

The number of biological invasions worldwide has steadily increased since 1800, but has not yet reached saturation point (Seebens et al. 2017). The number of exotic arthropod invasions, in particular, has increased spectacularly since the 16th century, with a clear acceleration in the second half of the 20th century (Roques 2010). This increase in the number of invasions is partly attributable to the introduction of phytophagous species with the intensification of the international ornamental plant trade (Dehnen-Schmutz et al. 2010; Roques 2010).

Increases in the number of entries and the spread of exotic species within a new territory account for the surge in biological invasions. Following their arrival in new territories, exotic insect species spread at various rates, depending on the time since their introduction and their feeding guild. For example, species discovered since the 1990s tend to spread three to four times more rapidly than those arriving before this time point (Roques et al. 2016). Exceptionally high dispersal capabilities generally drive this rapid range expansion. Some species can naturally disperse over very long distances (e.g., the invasive yellow-legged hornet; Robinet et al. 2017), can rapidly evolve the capacity for fast dispersal (e.g., the bean beetle; Ochocki & Miller 2017), or may be transported over long distances by human activities, thanks to the free circulation of goods within the European Union, for example (Roques et al. 2016). However, high dispersal capabilities alone cannot account for the rapid spread observed. Population growth is another key factor to be taken into account, as it might explain how possible Allee effects are overcome at low population density (Taylor & Hastings 2005). Together with dispersal capacity and population carrying capacity, population growth could drive the rate at which the invasion wave spreads (Shigesada & Kawasaki 1997; Haond et al. 2018). Thus, insects that produce several generations per year are likely to spread more rapidly (Fahrner & Aukema 2018).

Population phenology modelling can be used to estimate the number of generations and the time window associated with each life stage. The phenology model classically used to describe the development of plants and poikilothermic organisms, such as insects, is the degree-day model. This

model has proved accurate for many species over the years (Bonhomme 2000, Damos & Savopoulou-Soultani 2012, Rebaudo & Rabhi 2018). It is based on the principle that a number of thermal units above a given temperature threshold must be accumulated to complete the development of a given life stage, leading to definition of the heat accumulation required for development. In this model, we assume a linear relationship between temperature and development rate. Non-linear models have been developed to refine this modelling framework and to provide more subtle performance curves than linear relationships (Rebaudo & Rabhi 2018). However, these models require data for the fitting of these non-linear functions (e.g., experimental data for development rates as a function of temperature), which is feasible for only a set of well-documented species. These models, based on temperature-driven processes, can also be improved by considering: 1) photoperiod thresholds, particularly for organisms that have a dormancy or diapause period (Pollard et al. 2020) and 2) population growth (fecundity and mortality factors) (e.g., the agent-based model developed for the invasive brown marmorated stink bug, Halyomorpha halys, model, Nielsen et al. 2016).

A good knowledge of the biology of the species concerned is required for the parameterisation of such models and for the development of effective control measures limiting the spread of invasive species. However, the biology of invasive species is generally poorly understood, as most of these species cause no significant damage in their native area and some are even unknown to science (Seebens et al. 2017). In addition, the build-up of the population in the invaded area is subject to founder effects and genetic bottlenecks (Dlugosch & Parker 2008), and may not, therefore, be entirely representative of the characteristics of individuals in their native area. Careful study of the invasive population is, therefore, required, to assess its potential growth and spread.

99 The box tree moth, *Cydalima perspectalis* (Walker, 1859) (Lepidoptera: Crambidae), is emblematic of 100 a rapid invasion (Roques et al. 2016). It is native to Asia and it develops on box trees. It was first 101 detected in Europe in Germany in 2007, and has since invaded a large area extending from France to 102 Iran (Bras et al. 2019) and causing significant damage to European *Buxus* trees. It has a rate of spread

in Europe of up to 155 km/year. This rapid spread probably reflects multiple introductions from China on imported box trees (Bras et al. 2019), in addition to other factors, such as human-mediated dispersal through the box tree trade within the European Union (Bras 2018), and its own potential for population growth. The box tree moth has three to five generations per year in China (Wan et al. 2014) but only two generations per year have been recorded in Switzerland (Nacambo et al. 2014). It remains unclear how many generations this moth could potentially produce in the various climates found in Europe. This species overwinters at the larval stage: from the 2nd to the 5th larval instar in China, depending on the precise geographic area (Wan et al. 2014), and mostly at the 3rd-4th larval instar stages in Europe (Nacambo et al. 2014; Bras 2018). The conditions required for diapause induction are relatively well known (day length of 13:30 hours in Europe; Nacambo et al. 2014 and day length of between 12:50 and 13:36 hours in China; Xiao et al. 2011), but those for the end of diapause remain unclear, making it difficult to predict when insects will resume their development in the spring.

In this study, we aimed to develop a temperature- and photoperiod-driven model (based mostly on a phenology model) to explore box tree moth phenology in detail and to determine how many generations this species can produce in Europe. The results of this study will ultimately improve our understanding of the spatiotemporal dynamics of this invasive species, and will also facilitate the temporal optimisation of pest management methods targeting particular stages in the life cycle.

2. Materials and Methods

50 123

2.1 Biology of the study species in its invaded range

The box tree moth is a multivoltine species (Fig. 1).





Fig. 1. Box tree moth life cycle and timeline for three generations per year, as is thought to occur in some parts of Europe. The life cycle is illustrated for Generation 3. The period of the year during which the moth is active is shown in green and the overwintering period is shown in blue. The first generation (Generation 1) is the overwintering generation.

Throughout most of the invaded range, this insect is active from late March to late October. Two to 132 three generations per year have been recorded in several parts of Europe (Nacambo et al. 2014; Matošević et al. 2017), but it is thought that up to four generations per year may occur in Georgia 133 (Gninenko et al. 2014). The moth overwinters at the larval stage, in cocoons stuck to the leaves of box 135 trees. The overwintering cocoons can be observed from September (when day length falls below about 13 h) until the spring. The factors involved in diapause termination remain unclear, but 136 137 exposure to cold temperatures for at least one and a half months may be required for the insect to resume its development (Nacambo et al. 2014). The overwintering larvae usually complete their 138 development in spring, when temperatures start to increase. The first adults are observed in June 140 and the second and third generations are produced during the summer.

2.2. Description of the phenology model



For each life stage, from eggs to pupae, we consider a degree-day model (Fig. 2).

Fig. 2. Diagram of the *Cydalima perspectalis* life stage model. Grey rectangles indicate life stages for which a degree-day model is applied.

We considered the accumulation of temperature above a given lower developmental threshold (TL) and below a given upper developmental threshold (TU), and assumed that the development of the stage was complete when the number of degree-days required for the stage development (DD) was reached. We used the double-sine wave method to calculate the number of degree-days accumulated (Zalom et al. 1983). In this method, temperatures are interpolated from the daily minimal and maximal temperatures, by half-days, with a sinusoidal curve and the area under the curve between TL and TU is calculated. For larvae, we did not consider each of the individual larval stages (from L1 to L7), because too few data were available to parameterise the model. Instead, we considered three groups of larval stages: G1 corresponding to L1-L2, G2 corresponding to L3-L4, and G3 corresponding to L4-L7, because only G2 is likely to enter diapause for winter and, therefore, to experience an interruption of larval development.

The dates of diapause induction and termination were calculated from the photoperiod and temperature datasets (daily minimal and maximal temperatures). Diapause can begin when photoperiod falls below a day length of DI. For any day after this date and before November (after November, we considered the temperature to be too low), cold was accumulated and we determined the date by which sufficient exposure to cold had been accumulated (DLcold days at which the minimal temperature was below TLcold). After this date, heat began to accumulate, and we calculated the date at which sufficient heat accumulation had been achieved (DUwarm consecutive days during which the maximal temperature was above TUwarm). Once sufficient heat has been accumulated, overwintering ends and the larvae can continue their development. We also calculated a mortality rate for the diapause period, based on cold accumulation (see model parameterisation).

The model was initiated with a flight curve providing the number of individuals flying each day, to represent the number of adults emerging each day. We assumed that the males and females emerged at the same time (Göttig 2017) and that the sex ratio was 0.5 (Tabone et al. 2015, Göttig 2017). We then needed to know the number of fertilised females to calculate the number of eggs. Assuming that males and females are randomly distributed in space, the probability of mating success follows a Poisson probability distribution (Boukal & Berec 2002). The number of fertilised females is therefore: $N_f \times (1 - \exp(-N_m))$, where N_f and N_m are the number of adult females emerging ona given day and the number of adult males present on that day, respectively (Robinet et al. 2007). This quantity rapidly tends toward N_f as N_m increases, and is therefore weakly constrained by the number of males. We simulated egg-laying according to this flight curve, by distributing the number of eggs laid by each female (FEC) over a period of time after emergence (DayEqq). We then applied the degree-day model for eggs (with TL=TLeggs and DD=DDeggs), for larva groups G1, G2 and G3 (with TL=TLlarvae1, TLlarvae2, TLlarvae3, and DD=DDlarvae1, DDlarvae2, DDlarvae3, respectively), and for pupae (with TL=TLpupae and DD=DDpupae). For each stage, we applied a mortality rate (meggs, mlarvae1, mlarvae2, mlarvae3, mpupae, madult) before simulating the

development of the next stage. Once pupae complete their development, the adults emerge and are ² 186 assumed to live for a certain period (longevity). The females then lay eggs and the algorithm moves on to the next loop (Fig. 2).

In addition, for G2 (L3-L4), we also tested whether the date (considered incrementally, day after day) could be considered a date on which they could enter diapause, for each day of their development. Larvae entering diapause were considered to stop developing, with development beginning again after the end of diapause.

In total, 24 input parameters must be provided to simulate the phenology of the box tree moth (Table 1). The model was coded in R language (R Core Team 2019; Robinet & Suppo 2020).

Parameter	Default value	Description
NF	1000	Number of female adults at time t=0
DI	13 h	Photoperiod triggering winter diapause
TLcold	2°C	Temperature threshold below which cold is
		accumulated for diapause
DLcold 45 days		Number of days during which minimal
		temperature should be below <i>TLcold</i> to
		accumulate cold for diapause
TUwarm	vector(5,10,15,20)	Maximal temperature thresholds (°C) for
		determining the end of diapause
DUwarm	vector(28,9,7,3)	Number of consecutive days during which
		maximal temperature should be above
		TUwarm to achieve diapause. As soon as a
		condition is fulfilled (e.g., the second
		condition, daily maximal temperature is
		>10°C during 9 consecutive days), we
		considered the heat required to complete
		diapause to have accumulated.
F	2	Number of eggs laid by each female
Deggs	Day 4 to 14 after adult	Days after adult emergence on which eggs
	emergence (spread over 11 days)	are laid by females
L	14 days	Adult longevity
TLeggs	10.91°C	Temperature threshold above which
		degree-days are accumulated for eggs
DDeggs	48.54°C days	Sum of degrees above <i>TLeggs</i> necessary for
		egg development
TLlarvae	8.38°C	Temperature threshold above which
		degree-days are accumulated for larvae
DDlarvae1	44.5°C days	Sum of degrees above TLlarvae necessary
		for the development of larvae (L1-2)
DDlarvae2	66.7°C days	Sum of degrees above <i>TLlarvae</i> necessary

		for the development of larvae (L3-4)
DDlarvae3	211.3°C days	Sum of degrees above TLlarvae necessary
		for the development of larvae (L5-7)
TLpupae	11.5°C	Temperature threshold above which
		degree-days are accumulated for pupae
DDpupae	133.33°C days	Sum of degrees above <i>TLpupae</i> necessary
		for pupae development
meggs	0.1	Mortality rate of eggs
mlarvae1	0.2	Mortality rate of larvae L1-2 (G1)
mlarvae2	0.05	Mortality rate of larvae L3-4 (G2)
mlarvae3	0.05	Mortality rate of larvae L5-7 (G3)
трирае	0.02	Mortality rate of pupae
madults	0.05	Mortality rate of adults
fmort	function	Mortality (as a %) for diapausing larvae
		(derived from Nacambo et al. 2014)

Table 1. List of model parameters.

2.3. Model parameterisation

Development thresholds. The estimates of TL and DD for each stage were mostly obtained from the work of Nacambo et al. (2014), who studied an invasive population in Switzerland (Europe). We thus 35 200 considered TLeggs = 10.91°C and DDeggs = 48.54 degree-days, and TLpupae = 11.5°C and DDpupae = 133.33 degree-days. For larval stage, Nacambo et al. (2014) showed that the lower temperature threshold was 8.38°C and that 322.58 degree-days were required for development. Since we considered three groups of larval stages (L1-2, L3-4 and L5-7), we assumed that TLlarvae1 = TLlarvae2 = TLlarvae3 = 8.38°C. In a study conducted in Asia, Maruyama & Shinkaji (1991) found that stages L1 to L5 each completed development within three days, and that stage L6 completed development within eight days at 25°C. Since we have observed that L1 individuals originating from the invasive population develop much more rapidly than L2-L5 larvae, we considered only one day of development to be required for L1. We also assumed that L7 larvae developed at the same rate as L6 larvae (i.e. within 8 days). Thus, at 25°C, the total duration of larval development from L1 to L7 would be 29 days. According to Tabone et al. (2015), the duration of the larval stage is between 29 and 33

days for a population sampled within the invaded range in Europe. The two estimates are, thus, consistent. We then derived the number of degree-days above 8.38°C required from the number of days of development at 25°C, and we found that: DDlarvae1 = 44.5 degree-days, DDlarvae2 = 66.7 degree-days, and DDlarvae3 = 211.3 degree-days. As no information was available about the upper developmental threshold, we assumed that *TUeqqs* = *TUlarvae1* = *TUlarvae2* = *TUlarvae3* = *TUpupae* = + infinity. This is a conservative assumption for the sites considered here, since individuals can probably develop at temperatures of up to 35°C (Nacambo et al. 2014), and the model was applied to regions in which maximal temperatures in summer are generally around this threshold. More rigorous testing of the effects of heatwaves, during which temperatures may reach 40°C, would require improvements in the assessment of this parameter. In this study, we focus on average temperatures, ignoring extreme climate events.

Diapause conditions. Photoperiod is an important factor for diapause induction. Taking into account the work of Nacambo et al. (2014) and Xiao et al. (2010), we assumed that diapause was induced as soon as the photoperiod fell below 13 h of daylight (DI = 13 h). The conditions for the end of diapause still remained uncertain. Diapausing larvae must first accumulate a certain amount of cold. In accordance with the findings of Nacambo et al. (2014), we considered that minimal temperatures had to be below *TLcold* = 2°C for a period *DLcold* = 45 days. After accumulating cold, the larvae have to accumulate heat to end diapause. Based on the results of laboratory experiments on L4 under a photoperiod of 10-12 hours (Bras 2018; Poitou 2018), we assumed that a maximal temperature of at least 5, 10, 15, or 20°C (TUwarm) for 28, 9, 7 or 3 consecutive days (DUwarm), respectively, was required. The larvae can continue their development if one of these conditions is fulfilled.

Mortality rates. Except for eggs, no data were available for the accurate estimation of mortality at each stage. Based on observations and extrapolations from field data, we considered: *meggs* = 10% (Lepilleur et al. 2017), *mlarvae1* = 20%, *mlarvae2* = 5%, *mlarvae3* = 5%, *mpupae* = 2%, *madult* = 5% (Elisabeth Tabone, Pers. Com.). These estimates are subject to a high degree of uncertainty, but this

uncertainty affects the population level rather than the time window for each stage and generation.
In addition to these mortality rates, we considered mortality during the winter diapause. Nacambo et
al. (2014) showed that mortality rate during diapause depends on the duration for which larvae were
exposed to a temperature of 2°C or lower during the overwintering period. We used the following
function to estimate this mortality rate (in %), which ranged from 90% when cold accumulation was
very short (<1 month) to 10% when it lasted 3.5 months:

$$fmort(x) = \begin{cases} 90 & ifx < 30days \\ 100 - (2.67(x - 45) + 50) & if30 \le x < 45days \\ 100 - (0.67(x - 60) + 60) & if45 \le x < 60days \\ 100 - (0.67(x - 75) + 70) & if60 \le x < 75days \\ 100 - (0.67(x - 90) + 80) & if75 \le x < 90days \\ 100 - (0.67(x - 105) + 90) & if90 \le x < 105days \\ 100 - (-0.67(x - 120) + 80)if105 \le x < 120days \\ 100 + 1.33(x - 180) & ifx \ge 120days \end{cases}$$

with x the number of days at which the daily minimal temperature was below 2°C during the diapause period.

Other biological parameters. According to Tabone et al. (2015), female adults can live for 12 days and male adults can live for 15 days under laboratory conditions (25°C). We therefore assumed that adults of both sexes could live for L = 14 days in the field. Each female lays a mean of 800 eggs (± 300 eggs) (Tabone et al. 2015). As we aimed to study the phenology of the insects rather than their abundance, we used a fecundity F = 2 in subsequent simulations, to avoid a surge in the size of the population and problems visualising the number of individuals. We also needed to enter the number of females initially present, *NF*, to initiate the model. We chose an arbitrary value for this, *NF* = 1000.

2.4. Model validation

The model was validated in four ways: a preliminary check and three validations on independent datasets: 1) for four sites at which adult flights were recorded with pheromone traps in two

consecutive years, 2) for one site at which adult flights were recorded in four consecutive years and
3) for one site at which all life stages were followed for a whole year (Pineau et al. 2020).

257 <u>Preliminary check</u>. We compared the predicted phenology (duration of each life stage) at a constant
 258 temperature of 25°C with the laboratory observations reported by Tabone et al. (2015).

Validation 1. We compared the predicted time-window of adult occurrence, according to simulations, with trap captures at several locations. We considered trap captures in 2017 and 2018 at Orléans (47.83°N, 1.91°E, 105 m asl), Contrevoz (45.80°N, 5.63°E, 370 m asl), and Caen (49.18°N, -0.37°W, 10 m asl) in France. We also considered the data of Nacambo et al. (2014) for Basel and Riehen (Switzerland) in 2011 and 2012. Basel and Riehen are only 5 km apart. We therefore considered them to constitute a single site, referred to hereafter simply as "Basel". We initiated the model on the first flight of the first year and validation was performed in the second year. We calculated the correctness rate of presences and absences, between observations and simulation results. Temperature data were retrieved from the nearest weather station, at Orléans, Belley (about 6 km from Contrevoz), Caen, and Basel. The temperature data for French sites were provided by Météo-France and those for the Swiss site were retrieved from <u>http://www.ecad.eu</u> (Klein Tank et al. 2002). Photoperiod data were retrieved from <u>http://www.soda-pro.com (</u>SoDa team 2019).

<u>Validation 2.</u> We explored the cumulative error of the model over the years, using the data from
Nacambo et al. (2014) for Basel in 2009 (first flight) to initiate the model, and simulating insect
phenology until 2012. We calculated the correctness rate of presences and absences, between
observations and simulation results for 2010, 2011 and 2012.

⁴⁷ 275 <u>Validation 3.</u> Finally, we validated the model with a more comprehensive dataset (from eggs to adults) collected in Orléans from 2018 to 2019 (Pineau et al. 2020). Trap captures were checked over
⁵¹ a longer period (i.e., from April 28th to November 6th 2018, and from April 24th 2019 to November
⁵³ 278 29th 2019), to ensure that both the earliest and latest flights were recorded. Two pheromone traps
⁵⁶ 279 were frequently checked (two to seven times per week, following the flight season). At the INRAE
⁵⁸ 280 nursery in Orléans, 30 box trees (1.50 m high) were checked for three minutes each. The numbers of

eggs, small larvae (G1: L1-L2), medium-sized larvae (G2: L3-L4), large larvae (G3: L5-L7), pupae and diapausing larvae were counted. Life cycle stages were observed weekly from June 21st 2018 to October 31st 2018, monthly during the cold season (from November 2018 to May 2019) and then weekly again until June 21st 2019. The larvae found in the dense silk cocoons on the box tree leaves were assumed to be diapausing larvae. To our knowledge, this dataset, providing phenological data for all stages of the life cycle of this moth in the field, is the only one of its kind to date. The model was initiated with the first adult flight in 2018 (before Julian day 200), and phenology was simulated until June 2019. For each life stage and for all observation dates, we calculated the correctness rate of presences and absences between observations and simulations.

2.5. Model stability

Given the uncertainty on the values of the parameters, we performed a model stability analysis. We focused exclusively on parameters involved in the phenology of this species, disregarding the others (e.g., fecundity or mortality rates). We considered either a range of likely values according to our knowledge (uncertainty analysis) or a standard range (e.g., degree days ± 20% and temperature threshold ± 2°C) (sensitivity analysis) when it was not possible to determine the likely range of values. The stability analysis was performed on the temperature data for Orléans in 2016 and 2017. The output variables for this analysis were the date at which diapause ended and the peak date of adult flights for each generation. When analysing the effects of each parameter, we calculated the elasticity coefficient $\left(\frac{(y_1-y_0)/y_0}{(x_1-x_0)/x_0}\right)$ with x_0 as the default value of the parameter, x_1 as the new value of the parameter, y_0 as the output variable corresponding to x_0 , and y_1 as the output variable corresponding to x_1 . We then took the absolute value of the elasticity coefficient and calculated the maximal absolute value over the range of values of the input parameter and the output variables. It was not possible to calculate this value for all parameters tested (Deggs and DUwarm), so we also calculated the rate of variation $(y_1 - y_0)/y_0$, and the maximum of these absolute values.

2.6. Model simulations in a context of climate warming

We explored the possible effects of climate warming, by considering temperature increases of +1°C and +2°C relative to the daily minimal and maximal temperatures recorded at Orléans, Caen, Contrevoz and Basel. We compared the number of generations and calculated the percentage of days on which adult flights occurred, simulated under a climate warming scenario, for the periods of adult flight simulated for the current climate (hereafter referred to a "% similarity"). For all generations in the second year, we also compared the dates of the simulated peak numbers of adults in the current climate and in conditions of climate warming. We calculated the advance, in days, of the peaks for temperature increases of +1°C and +2°C relative to current climate conditions. Then, we calculated the mean and the maximum advances of the generations for the two climate warming scenarios.

3. Results

3.1 Model validation

Preliminary check. Each generation (from laying of the first egg to death of the last adult) took 48 days in simulations of phenology at 25°C (excluding the diapause submodel), and 51-63 days in the laboratory (according to Tabone et al. 2015). The life cycle (from laying of the first egg of one generation to laying of the first egg of the following generation) was completed within 38 days in the simulation, assuming that females began to lay eggs four days after emergence (against mean of 45 days and range of 40-50 days in the laboratory). Egg development took four days in the simulation (against 1-5 days in the laboratory). Development of the larval of groups G1, G2, and G3 took 3, 4 and 13 days, respectively, corresponding to a total of 20 days for complete larval development (against 29-33 days in the laboratory). Pupal development took 10 day in the simulation (against 9-10 days in the laboratory). The simulated life cycle at 25°C was completed slightly faster in the simulation than

under laboratory conditions (Tabone et al. 2015). This was mostly due to the shorter time required ² 332 for larval development in the simulations. Following activation of the diapause submodel in the simulation, the population was unable to survive because it never met diapause conditions (the conditions for diapause induction and cold accumulation).

Validation 1. Correctness rates exceeded 60% at Orléans and Contrevoz, and 80% at Caen and Basel (validation in the following year, 2010) (Table 2 and Fig. 3).

Site	Year of initialisation	Year of validation	% correctness
Orléans	2017	2018	66.3
Contrevoz	2017	2018	62.47
Caen	2017	2018	80
Basel	2009	2010	81.64
Basel	2009	2011	51.23
Basel	2009	2012	85.48

Table 2. Model validation based on the presence/absence of adult stages at Orléans, Contrevoz, Caen and Basel (initialisation on first flight in 2017 and validation in 2018 for Orléans, Contrevoz and Caen, and initialisation on the first flight in 2009 and validation in subsequent years for Basel). This table provides the correctness rate of presences/absences predictions compared to observations.

However, the model simulated a third flight for Orléans at the end of 2018 that was not actually observed (Fig. 3).





Fig. 3. Model validation at four sites. The model was initiated by the first flight of the first year and was validated on adult flights observed in subsequent years. The black curves and the red dotted curves represent the adult density (normalised by year) according to model simulations (black) and observations (red dotted). To improve clarity, the horizontal red dashed lines at the top indicate the days on which adults were observed, and the horizontal black dashed lines indicate the days on which adults were predicted to be present in model simulations for the years considered for validation purposes.

<u>Validation 2.</u> The model did not accumulate much error over the years. The fitting success of the model for Basel exceeded 80% (in 2010 and 2012) (Table 2). However, the simulated time window for the presence of adults was relatively large in 2011 and the model predicted earlier and later flights than were actually observed for this particular year (Fig. 3).

359 <u>Validation 3.</u> A comparison of the periods corresponding to each life stage at Orléans in 2018-2019
360 yielded the following results (Fig. 4; Table 3).



Fig. 4. Model validation on life stage data collected at Orléans in 2018-2019. Black curves represent the abundance of the life stage based on simulations. Horizontal red dashed lines indicate the days on which a particular life stage was observed and horizontal black dashed lines indicate the days on which the corresponding life stage was predicted to be present in model simulations. For larva 2 (G2), asterisks indicate the days on which diapausing larvae were observed. Three generations (numbered 1, 2 and 3) were followed. The adults of generation 1 (numbered 1) provided the eggs numbered 1, which provided the larvae 1 numbered 1, and so on. Larva 1, 2 and 3 correspond to larval groups G1, G2 and G3, respectively.

(1) The second adult flight was correctly estimated, but the simulations predicted a third adult flight at the end of 2018 (at around day 300 - end of October 2018), which was not observed in the field despite the presence of two pheromone traps until early November (Fig. 4). (2) Egg occurrence was

not correctly estimated (Table 3). (3) The occurrence of larvae of G1 was correctly estimated. The model even predicted the first time of occurrence in early 2019. (4) The occurrence of G2 larvae was also correctly estimated. (5) The occurrence of G3 larvae was correctly estimated, and (6) the occurrence of pupae was relatively well estimated. Most correctness rates exceeded 90% (Table 3).

Life stage	Number of weeks of occurrence	Number of weeks of observation	% correctness
Adult (j>200)	8	34	94.44
Egg	5	39	61.11
Larva G1	20	39	97.09
Larva G2	23	39	95.35
Larva G3	25	39	95.35
Pupa	28	39	95.35

Table 3. Validation of the model of subsequent life stages at Orléans in 2018-2019. This tableprovides, for all observation dates, the correctness rate of presences/absences predictionscompared to observations. For adults, this calculation was performed only for observations afterday 200, because the data for observations before this time point were used for modelinitialisation.

According to these simulations (Fig. 4), the second adult flight provides individuals that overwinter as G2 larvae, whereas the third adult flight provides individuals that could overwinter as eggs, producing larvae in early 2019. This implies that the generations overlap, which was confirmed by representations of the time windows corresponding to the development of each generation (Fig. 5).



day 304 (October 31st 2018). In the field, diapausing larvae were observed from day 228 (August 16th 2018) onwards, and more than half the population (>50%) was already in diapause by day 278 (October 5th 2018). According to the model, diapause was predicted to end on day 417 (February 21st 2019), regardless of when individuals entered diapause. Observations showed that the number of diapausing larvae decreased to 0 between day 453 (March 29th 2019) and day 488 (May 5th 2019). The simulations therefore appear to predict too early an end to diapause (given that the larvae in cocoons observed in the field are effectively still in diapause).

 43 401

3.2 Model stability

The parameter with the greatest effect was clearly TLlarvae, the minimal threshold above which the heat accumulation occurs in the description of larval development (Table 4, SM1). This parameter 408 should, therefore, be carefully assessed to ensure a good estimate of the species phenology.

² 409 Parameters relating to diapause (*DI*, *TLcold*, *DLcold* and *DUwarm*) had surprisingly little effect.

DI 12 and 14h TLcold 0 and 4°C DLcold 35 and 55 days DUwarm -2 and + 2 days (structure) Deggs Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (structure)	simultaneously for each $aas = day 4 to 10$ to	0.024 0.002 0.008	0.002 0.002 0.002 0.024 0.012
TLcold 0 and 4°C DLcold 35 and 55 days DUwarm -2 and + 2 days (response) value) value) Deggs Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with Detter 18 days) TLeggs 9 and 13°C (±2°C) DDeggs DDeggs 38.8 and 58.2°C (response)	simultaneously for each	0.002 0.008	0.002 0.002 0.024 0.012
DLcold 35 and 55 days DUwarm -2 and + 2 days (response) value) Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with Deresson 18 days) TLeggs 9 and 13°C (±2°C) DDeggs	simultaneously for each $aas = day 4 to 10$ to	0.008	0.002 0.024 0.012
DUwarm -2 and + 2 days (x value) value) Deggs Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with De 18 days 13°C (±2°C) 38.8 and 58.2°C (x)	simultaneously for each		0.024
value) Deggs Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (10)	aas = day 4 to 10 to		0.012
Deggs Day 4 to 7 Day 8 to 11 Day 11 to 14 L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (±2°C)	aas = day 4 to 10) to		0.012
Day 8 to 11 Day 11 to 14 L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (±2°C)	aas = day 4 to 10 to		
Day 11 to 14 L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (±2°C)	aas = day 4 to 10) to		
L 10 days (with De 18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C (±2°C)	aas = day 4 to 10 to		
18 days TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C	$ggs = uuy + io \pm oj io$	0.054	0.015
TLeggs 9 and 13°C (±2°C) DDeggs 38.8 and 58.2°C	18 days		
(±2°C) DDeggs 38.8 and 58.2°C (9 and 13°C		0.023
DDeggs 38.8 and 58.2°C	(±2°C)		
(+ 200()	days	0.060	0.012
(± 20%)	(± 20%)		
<i>TLlarvae</i> 6.5 and 10.5 °C	6.5 and 10.5 °C		0.043
(±2°C)	(±2°C)		
DDlarvae1 35.6 and 53.4°C	days (± 20%)	0.051	0.010
DDlarvae2 53.4 and 80.1°C	days	0.044	0.009
DDlarvae3 169.0 and 253.5°	°C days	0.156	0.031
<i>TLpupae</i> 9.5 and 13.5°C		0.154	0.027
DDpupae 106.7 and 160.0°	°C days (± 20%)	0.126	0.025
Table 4. Stability analysis. Elast	icity represents the maxim	num of absolute	values for elas

numbers in bold are the highest values.

3.3 Model simulations under climate warming

Insect phenology changed considerably in simulations of climate warming, but the predicted
changes differed between sites. Most adult flights would be observed within the same time window
as under current climatic conditions at Orléans and Contrevoz (Table 5).

Site	Scenario	% similarity	Mean advance in peak of adult occurrence (days)	Maximum advance in peak of adult occurrence (days)
		21		

Orléans	+1°C	90.9	15	29
Orléans	+2°C	91.8	28	46
Contrevoz	+1°C	88.0	8	15
Contrevoz	+2°C	88.5	16	24
Caen	+1°C	62.5	3	5
Caen	+2°C	71.9	33	33
Basel	+1°C	79.2	12	12
Basel	+2°C	75.1	24	25

or +2°C) relative to simulations of adult flights based on temperatures in 2017-2018 for Orléans, Contrevoz and Caen, and 2011-2012 for Basel. The mean and maximum numbers of days of advance in the peak of adults' occurrence in conditions of climate warming are compared with the adult flights in current conditions, in 2018.

Table 5. Changes in adult flight patterns in the context of different climate warming scenarios (+1°C

The predicted changes were largest for Caen and Basel, at which flight similarity was lower than at other sites (Table 5). On average, at all four sites, the peak of adult flights would generally occur earlier for an increase of +1°C (mean of 9.5 days) and still more for an increase of +2°C (mean of 25 days). The number of generations could reach three at Orléans, Caen and Basel, and even four at Contrevoz for an increase in temperature of 2°C (Fig. 6).



adults present under observed current climatic conditions (2017-2018 for Orléans, Contrevoz and Caen; and 2011-2012 for Basel), orange curves represent adult density in a scenario involving a +1°C increase in temperature, and red curves represent adult density in a scenario including a +2°C increase in temperature. The adult density was normalised by scenario and year. Generations are numbered 1, 2, 3 and 4 for the scenario with a temperature increase of +2°C. For the sake of clarity, 40 438 horizontal black, orange and red dashed lines indicate the days on which adults are predicted to be present in model simulations for the three scenarios (baseline conditions, +1°C and +2°C, respectively).

4.1 What do the model simulations and discrepancies tell us?

 4.

Discussion

This study provides the first version of a phenology model for the invasive box tree moth. The model provided rough estimates of the time window for each life stage and did not accumulate much error

446 over the years. It predicted two or three generations per year, which is consistent with the available 447 observations. This model revealed a complex dynamics of invasive populations of box tree moth: 1) 448 several generations overlap, making it difficult to distinguish between flight periods, and 2) the insect 449 may potentially overwinter not only as a larva, but also at other life stages, and 3) climate warming 450 may enhance this phenomenon in the future. However, these model simulation outputs require 451 confirmation in the field.

In cases of discrepancy between simulations and observations, it was not possible to provide a clear explanation for the differences, which may result from an error in the model simulations or from a bias in the observations. Two main discrepancies require further investigation: 1) the presence of eggs and young larvae in the winter, and 2) diapause duration.

1) The simulations suggested that eggs might be present in the winter, whereas they are not observed in the field during this period. Nevertheless, it is very difficult to detect eggs, which are tiny and transparent, and generally located on the underside of box tree leaves. The model suggested that eggs could overwinter and provide first instar larvae (L1) in the spring, which was an unexpected result. It will therefore be important to address the following question: are eggs (and young larvae) able to survive the cold temperatures in winter or to survive to a very low development during this cold period? The collection of more field data will be required to answer this question. Meanwhile, for exploration of the effects of cold temperatures, we modified the model to include an additional mortality parameter related to low temperatures during the winter (data not shown). However, the modified model was unable to simulate the occurrence of young larvae in early spring, as observed in the field. The presence of young larvae in early spring is consistent with the presence and survival of eggs during the winter, which could then develop into L1 during the spring. Furthermore, our simulations suggest that, with climate warming, we will probably observe more eggs and young larvae in winter (data not shown). This issue of the possible survival of both eggs and young larvae in 470 winter is, therefore, extremely important in the context of climate warming. Laboratory experiments471 could be performed to address this question.

2) The time window for diapause duration was underestimated by the model in validation 3. However, the larvae observed in cocoons in the field may not necessarily actually be in diapause (i.e., interrupted development). If they build their cocoon before entering diapause and remain within the cocoon for a short period after the end of diapause, this would bias the observation dataset relating to diapause in the field. Accurate validation of the diapause period on the basis of field data is complex. Another issue concerning diapause emerged from the preliminary check (at a constant temperature of 25°C). In simulations without the diapause submodel, the development rates obtained for each life stage were very similar to those observed in laboratory conditions. Indeed, no diapause is observed in the constant conditions of the laboratory (temperature 25°C, photoperiod 16L:8D). The photoperiod is constant and longer than the threshold period of 13 h, preventing the induction of diapause. When we simulated phenology at 25°C with the diapause submodel, the simulations showed a population crash in the winter (data not shown). The model should be refined further, to determine whether larvae fail to enter diapause under some temperature conditions.

According to our findings, the box tree moth can have more than two generations in Europe, and may even produce four generations in conditions of climate warming with an increase in temperature of 2°C, at the sites studied. Climate warming may therefore accelerate the already very rapid spread of this invasive species. In addition, we set female fecundity to a very low value (F = 2) to avoid a population explosion. Population growth is a key factor in the rapid spread of this pest and high population abundance will clearly have a major effect on box trees, some of which may be heavily, or even completely defoliated, leading to their death (Kenis et al. 2013, Mitchell et al. 2018). At this point, low resource availability might lead to a decrease in the abundance of box tree moths.

4.2 Limitations of the model

The current model is not spatially explicit. However, it could be used to explore the spatial impact of different temperature conditions (and climate, in the long term) on temporal dynamics. It could be applied to sites in different countries, provided that the box tree moth populations at these sites originate from the same source population as the population studied in Switzerland (development thresholds from Nacambo et al. 2014), and thus present similar thermal development thresholds. In Europe, the invasion pathway followed by this moth appears to be complex, probably involving several genetic processes (Bras et al. 2019). Nevertheless, an eastern Chinese origin has been proposed for all invasive populations across Europe and Asia Minor studied to date (Bras et al. 2019), suggesting that all these populations may have similar development thresholds, making it possible to use our model at a larger scale. In addition, we did not consider an upper temperature threshold for development. If the model is applied to a site in an area warmer than those studied here (e.g. southern Europe), where temperatures frequently exceed 35°C, an upper threshold should be considered. Without such a threshold, the model may be subject to limitations at high temperatures, resulting in a possible overestimation of development rates.

In this study, we considered the maximum and minimum daily temperatures recorded by weather stations. We assumed these temperatures to be representative of the temperatures actually experienced by the box tree moth individuals. However, microclimate conditions are known to display considerable spatial variation and to have a large impact on insects (Kearney and Porter 2009, Pincebourde et al. 2016). Eggs and larvae are generally found on box tree leaves. Only diapausing larvae and pupae are likely to be protected, within their cocoon, wrapped in a leaf. The outputs of simulations could, therefore, be refined by considering temperature at a local scale rather than the temperatures recorded by weather stations.

4.3 Usefulness of the model in terms of pest management

517 Box trees do not have a particularly high economic value, but the decline of ornamental box trees 518 and natural box tree stands would nevertheless have a major ecological, cultural and social impact

(Mitchell et al. 2018). The development of effective control measures against box tree moth has therefore received considerable attention. Our model provides an appropriate framework for improving the management of this multivoltine species in the near future. Indeed, several environmentally friendly control measures could be applied, targeting particular life stages of the moth.

Hereafter, we shortly review three of them. 1) Pheromone traps capture adult males, and can therefore be installed during adult flight periods, which may be quite long. If the traps are in place unnecessarily for several months, the pheromone baits should be regularly replaced, to ensure that the traps remain effective. Precise assessments of the time window for adult flights with the model presented here would be useful for optimising this method, ensuring that traps were installed only when necessary (either for population monitoring or for a tentative of mass trapping). The catching of adult females would also be of great interest, and a female-targeting lure should become available in the near future (Molnár et al. 2019). 2) Bacillus thuringiensis sprays can be used against young larvae (Göttig 2017). If applied at the wrong time (i.e., too early, on eggs, or too late, on older larval instars) this method may not efficiently protect ornamental box trees against feeding larvae. For this control method, our model can estimate the timing of larval emergence and provide an indication of the time window during which first larval instars are present, making it possible to optimise treatment efficacy. 3) Classical biological control involves releasing a specific natural enemy targeting the egg stage, to control the population before the larvae have an opportunity to feed on and defoliate box trees (e.g. using Trichogramma sp. as biological agents; Göttig 2017, Enriquez et al. 2015). The model can predict the time window for egg occurrence, corresponding to the period in which the parasitoids should be released.

Identifying the appropriate time window for each control method would increase control efficacy.
Furthermore, in newly invaded regions, such as North America, where this insect was first detected
in August 2018 (Frank 2019), our model is of major potential interest to stakeholders to facilitate the

implementation of efficient management strategies at early stages of box tree moth invasion. This
model could also feed into platforms, such as the Degree-Days, Risk, and Phenological event mapping
(DDRP) platform (Barker et al. 2020).

4.4 Key message

The model developed here goes far beyond a simple degree-day model, as it accounts for photoperiod, diapause processes, and population growth parameters. Models of this type appear to be particularly useful for identifying knowledge gaps and disentangling overlaps between the life stages of invasive species (Nielsen et al. 2016). This model is also a potentially useful tool for assessing potential population growth (including the number of generations) in invaded areas and, thus, for fine-tuning estimates of the risk of rapid invasions due to the potential growth rate of the pest. In addition, in the context of climate warming, population phenology may become more erratic, and models of this type can provide an important cornerstone in efforts to improve control methods targeting specific life stages.

- 557 References
- Barker, B.S., Coop, L., Wepprich, T., Grevstad, F., Cook, G., 2020. DDRP: real-time phenology and
 climatic suitability modeling of invasive insects. BioRxiv, doi:10.1101/2020.05.18.102681

560 Bonhomme, R., 2000. Bases and limits to using 'degree.day' units. Eur. J. Agron., 13, 1–10.

Boukal, D. S., and Berec, L., 2002. Single-species models of the Allee effect: extinction boundaries, sex ratios and mate encounters. J. Theor. Biol., 218(3), 375-394.

https://doi.org/10.1006/jtbi.2002.3084

Bras, A., (2018). Identification des facteurs sous-tendant l'invasion fulgurante d'un insecte asiatique en Europe, la pyrale du buis: approche génétique et biologique. PhD thesis, Université d'Orléans (France), pp. 243.

Bras, A., Avtzis, D.N., Kenis, M., Li, H., Vétek, G., Bernard, A., Courtin, C., Rousselet, J., Roques, A.,
Auger-Rozenberg, M-A., 2019. A complex invasion story underlines the fast spread of the invasive
box tree moth (*Cydalima perspectalis*) across Europe. J. Pest. Sci., 92, 1187-1202.

Damos, P., Savopoulou-Soultani, M., 2012. Temperature-driven models for insect development and vital thermal requirements. Psyche, Article ID 123405, doi:10.1155/2012/123405

572 Dehnen-Schmutz, K., Holdenrieder, O., Jeger, M.J., Pautasso, M., 2010. Structural change in the
573 international horticultural industry: some implications for plant health. Sci. Hortic-Amsterdam,
574 125, 1-15.

Dlugosch, K.M., Parker, I.M., 2008. Founding events in species invasions: genetic variation, adaptive evolution, and the role of multiple introductions. Mol. Ecol., 17, 431–449.

Enriquez, T., Giorgi, C., Venard, M., Colombel, E., Gaglio, F., Buradino, M., Martin, J-C., Tabone, E., 2015. Des souches de trichogramme contre la pyrale du buis. Phytoma, 685, 21-24.

Fahrner, S., Aukema, B.H., 2018. Correlates of spread rates for introduced insects. Global. Ecol.
Biogeogr., 27, 734-743.

Frank, S., 2019. Watch for potential new boxwood pest. The Purdue Landscape Report, 19(2),1–4.

Gninenko, Yu.I., Shiryaeva, N.V., Shurov, V.I., 2014. The box tree moth - a new invasive pest in the Caucasian Forests. Plant Health - Research and Practice, 1, 32–39.

584 Göttig, S.G., 2017. Development of eco-friendly methods for monitoring and regulating the box tree 585 pyralid, *Cydalima perspectalis* (Lepidoptera: Crambidae), an invasive pest in ornamentals. PhD

 to predict species' ranges. Ecol. Lett., 12, 334-350.
Kenis, M., Nacambo, S., Leuthardt, F.L.G., Di Dominico, F., Haye, T., 2013. The box tree moth: *Cydalima perspectalis*, in Europe: horticultural pest or environmental disaster? Aliens, 33, 38-41.
Klein Tank, A.M.G. and Coauthors, 2002. Daily dataset of 20th-century surface air temperature and precipitation series for the European Climate Assessment. Int. J. of Climatol., 22, 1441-1453.
Lepilleur, T., Brancaccio, L., Ben Soussan, T., Baby, P., Aveline, S., Duval, C., 2017. Efficacité des trichogrammes contre la pyrale du buis. Phytoma, 705, 20-25 http://www.phytomaldv.com/revue-1802-PHYTOMA-705
Maruyama, T., Shinkaji, N., 1991. The life-cycle of the box-tree pyralid, *Glyphodes perspectalis* (Walker) (Lepidoptera: Pyralidae). II. Developmental characteristic of larvae. Jpn J Appl Entomol Zool, 35, 221-230.
Matošević, D., Lukić, I., Bras, A., Lacković, N., Pernek, M., 2017. Spatial distribution, genetic diversity and food choice of box tree moth (*Cydalima perspectalis*) in Croatia. SEEFOR, 8, 41–46.

thesis, Technische Universität Darmstadt (Germany), pp. 130.

ouvertes.fr/hal-01877823/document

http://tuprints.ulb.tu-darmstadt.de/6855/1/Dissertation%20Stefanie%20G%C3%B6ttig.pdf

Haond, M., Morel-Journel, T., Lombaert, E., Vercken, E., Mailleret, L., Roques, L., 2018. When higher

Kearney, M., Porter, W., 2009. Mechanistic niche modelling: combining physiological and spatial data

carrying capacities lead to faster propagation. bioRxiv, 307322; https://hal.archives-

https://doi.org/10.15177/seefor.17-06

Mitchell, R., Chitanava, S., Dbar, R., Kramarets, V., Lehtijärvi, A., Matchutadze, I., Mamadashvili, G.,
Matsiakh, I., Nacambo, S., Papazova-Anakieva, I.P., Sathyapala, S., Tuniyev, B., Vétek, G.,
Zukhbaia, M., Kenis, M. 2018. Identifying the ecological and social consequences of a decline in *Buxus* forests in Europe and the Caucasus. Biol. Invasions, 20, 3605-3620.

Molnár, B.P., Kárpáti, Z., Nagy, A., Szarukán, I., Csabai, J., Koczor, S., Tóth, M., 2019. Development of
 a female-targeted lure for the box tree moth *Cydalima perspectalis* (Lepidoptera: Crambidae): a
 preliminary report. J. Chem. Ecol., 45, 657-666.

Nacambo, S., Leuthardt, F.L.G., Wan, H., Li, H., Haye, T., Baur, B., Weiss, R.M., Kenis, M., 2014.
Development characteristics of the box-tree moth *Cydalima perspectalis* and its potential
distribution in Europe. J. Appl. Entomol., 138, 14-26.

- Nielsen, A.L., Chen, S., Fleischer, S.J., 2016. Coupling developmental physiology, photoperiod, and
 temperature to model phenology and dynamics of invasive heteropteran *Halyomorpha halys*.
 Front. Physiol., 7, 165. <u>https://doi.org/10.3389/fphys.2016.00165</u>
- Ochocki, B.M., Miller, T.E.X., 2017. Rapid evolution of dispersal ability makes biological invasions
 faster and more variable. Nat. Commun., 8, 14315. <u>https://doi.org/10.1038/ncomms14315</u>
 - Pincebourde, S., Murdock, C. C., Vickers, M., Sears, M. W., 2016. Fine-scale microclimatic variation can shape the responses of organisms to global change in both natural and urban environments. Integr. Comp. Biol., 56, 45-61. https://doi:10.1093/icb/icw016
 - Pineau, P., Lorme, P., Bernard, A., Bras, A., Poitou, L., Rousselet, J., Robinet, C., 2020. Records of the box tree moth, *Cydalima perspectalis*, phenology in Orleans and Contrevoz (France). https://doi.org/10.5281/zenodo.3719293

Poitou, L., 2018. Rôles de la température et de la photopériode sur la levée de la diapause hivernales
chez une espèce dont l'invasion est fulgurante en Europe, la pyrale du buis, *Cydalima perspectalis*(Lepidoptera, Crambidae). Master thesis, Université de Bordeaux (France), pp. 23.

- Pollard, C.P., Griffin, C.T., de Andrade Moral, R., Duffy, C., Chuche, J., Gaffney, M.T., Fealy, R.M.,
 Fealy, R., 2020. phenModel: A temperature-dependent phenology/voltinism model for a
 herbivorous insect incorporating facultative diapause and budburst. Ecol. Model., 416.
 https://doi.org/10.1016/j.ecolmodel.2019.108910
- Rebaudo, F., Rabhi, V.-B., 2018. Modeling temperature-dependent development rate and phenology
 in insects: review of major developments, challenges, and future directions. Entomol. Exp. Appl.,
 166, 607–617.
 - Robinet, C., Liebhold, A., Gray, D., 2007. Variation in developmental time affects mating success and
 Allee effects. Oikos, 116: 12271.

Robinet, C., Suppo, C., Darrouzet, E., 2017. Rapid spread of the invasive yellow-legged hornet in
France: the role of human-mediated dispersal and the effects of control measures. J. Appl. Ecol.,
54, 205-215.

- Robinet, C., Suppo, C., 2020. R script to simulate the phenology of the box tree moth, *Cydalima perspectalis*. <u>https://doi.org/10.5281/zenodo.3719273</u>
- R Core Team (2019) R: A language and environment for statistical computing. R Foundation for
 Statistical Computing, Vienna, Austria. URL <u>https://www.R-project.org/</u>.

Roques, A., 2010. Taxonomy, time and geographic patterns, in: Roques, A. et al. (Eds), Alien
terrestrial arthropods of Europe. BioRisk, 4, 11-26.

Roques, A., Auger-Rozenberg, M-A., Blackburn, T.M., Garnas, J., Pyšek, P., Rabitsch, W., Richardson,
D.M., Wingfield, M.J., Liebold, A.M., Ducan, R.P., 2016. Temporal and interspecific variation in
rates of spread for insect species invading Europe during the last 200 years. Biol. Invasions, 18,
907-920.

Seebens, H., Blackburn, T.M., Dyer, E.E., Genovesi, P., Hulme, P.E., Jeschke, J.M., Pagad, S., Pyšek, P.,
Winter, M., Arianoutsou, M. et al., 2017. No saturation in the accumulation of alien species
worldwide. Nat. Commun., 8, 14435.

Shigesada, N., Kawasaki, K., 1997. Biological Invasions: theory and practice. Oxford University Press,
 pp. 205.

SoDa team (2019) Solar Energy Services for Professionals. Available at: <u>http://www.soda-pro.com</u>. Accessed 14 January 2019.

Tabone, E., Enriquez, T., Giorgi, C., Venard, M., Colombel, E., Gaglio, F., Buradino, M., Martin, J-C.,
2015. Mieux connaître la pyrale du buis *Cydalima perspectalis*. Phytoma, 685, 18-20.

Taylor, C.M., Hastings, A., 2005. Allee effects in biological invasions. Ecol. Lett., 8, 895-908.

Wan, H., Haye, T., Kenis, M., Nacambo, S., Xu, H., Zhang, F., Li, H., 2014. Biology and natural enemies of *Cydalima perspectalis* in Asia: is there biological control potential in Europe? J. Appl. Entomol., 138, 715-722. doi:10.1111/jen.12132

Xiao, H-J., Xin, H-Q., Zhu, X-F., Xue, F-S., 2011. Photoperiod and temperature of diapause induction in
 Diaphania perspectalis (Lepidoptera: Pyralidae). Chinese J. Appl. Entomol., 48(1), 116–120.

Zalom, F.G., Goodell, P.B., Wilson, L.T., Barnett, W.W., Bentley, W.J., 1983. Degree-days: the
calculation and use of heat units in pest management. University of California Division of
Agriculture and Natural Resources, leaflet 21373, pp. 10.

670 Acknowledgements

We thank Laura Poitou and Mathieu Laparie (INRAE URZF) for their work on the box tree moth diapause and for providing data to feed the model, Jérôme Rousselet (INRAE URZF) for supervising field observations, Patrick Pineau and Philippe Lorme (INRAE URZF) for collecting phenology data in Orléans, Alexis Bernard (INRAE URZF) for installing and retrieving pheromone traps and solving technical field problems, Elisabeth Tabone, Jean-Claude Martin and Maurane Buradino (INRAE UEFM) for providing useful information about the biology of this species, Eric Hell (CRPF) and Olivier Baubet (DSF) for helping us to find an appropriate site at Contrevoz and the townhall of Contrevoz for hosting a pheromone trap, Benoit Coiffiers (FREDON) for providing phenology data in Caen, and Marc Kenis (CABI) for providing data in Switzerland. We sincerely thank Marie-Anne Auger-Rozenberg for the coordination of the INCA project in which this study was conducted. We thank INRAE Agroclim and Météo-France for providing daily minimum and maximum temperatures for the French sites.

83 Funding

We acknowledge support for this work from the INCA project (Invasion fulgurante de la pyrale du
buis *Cydalima perspectalis* en Région Centre Val de Loire) funded by French region *Région Centre Val de Loire*.