

The effects of diurnal temperature rise on tomato fruit quality. Can the management of the greenhouse climate mitigate such effects?

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- 1 The effects of diurnal temperature rise on tomato fruit quality. Can the
- 2 management of the greenhouse climate mitigate such effects?
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13 Abstract

- In the present study, tomatoes were grown under control conditions (CT), a
- moderate (MT, up to 2 °C) or a higher rise in temperature (HT, up to 5 °C), and a
- moderate rise followed by a lower air temperature at the end of the day (ED), in order to
- analyze the effects of increased temperature as well as management system impact on
- 18 fruit quality.
- 19 Rising temperatures up to 5 °C reduced the fruit growth duration by five days
- and greatly reduced the fresh (29.7%) and dry matter (30%) accumulation rate.
- 21 Increasing the temperature during the day and lowering it at the end of the day increased
- 22 citric and malic acid concentrations in the fruit, therefore reducing the sugar/acid ratio.
- 23 The ascorbate concentration and carotenoids increased in the fruits grown under high
- temperature, while, phenolics concentrations did not change. Fruit water conductance
- 25 was strongly reduced under HT (-37%), and so was starch accumulation, in contrast to

hexoses. Dropping the air temperature at the end of the day could help mitigate HT impacts as it slightly increased the pericarp dry matter content. It could also improve seeds quality as it increased their dry matter content. It would be interesting to consider broader changes in ED temperature to confirm its potential role in improving fruits dry matter content.

Keywords: *Solanum lycopersicum* L., fruit growth, temperature, fruit quality, fruit composition, climate change scenario.

1. Introduction

Under glasshouse, different cultural practices and climate management strategies can be used to modulate plant growth conditions in order to obtain the required fruit yield and quality. Among those, temperature control is a useful method for growers to modulate plant growth when other factors such as light, nutrients and water are not limiting (Guichard et al. 2001).

Over the last few decades, several studies have been devoted to identifying the minimal temperatures allowing a high yield of tomatoes while limiting the energy costs of heating. Night temperature has long been known to be the determining factor in tomato fruit setting, with optimum temperatures ranging from 15 °C to 20 °C.. In addition, the variation of daily temperature patterns may affect the plant and fruit growth and metabolism, and ultimately, the fruit quality (Adams et al. 2001; Hurd and Graves, 1985; Truffault et al. 2015; Van der Ploeg and Heuvelink, 2005). In the near future, climate change will cause an increase in temperature, which can affect crop yield, due to a reduced fruit set, and change fruit quality (Garg and Cheema, 2011). Increasing temperature generally increases the fruit growth rate but also accelerates fruit

maturation and ripening so that fruit weight at harvest can be reduced (Adams et al. 2001; Fleisher et al. 2006). Temperatures at the daily scale can affect water and carbon 52 fluxes within the plant. Walker and Ho (1977) reported that increasing fruit temperature favored sugars allocation to the fruits. Similarly, a rapid decrease in air temperature at the end of the day is used by growers to favor assimilate partitioning to the fruits. The tomato fruits dry matter content is not affected by temperature increase from 18 °C to 23 °C (Heuvelink, 1995). However, growing plants at higher temperatures (increasing temperature from 25/14 °C to 30/12 °C, day/night respectively) enhances the tomato fruits dry matter content (Rosales et al. 2011). These changes in dry matter content also reflected changes in the tomato fruits composition (Dorais et al. 2010; Rivero et al. 2001). Rosales et al. (2011) reported that sugars content increased at high temperatures 61 (30-35 °C) while organic acids decreased. Robertson et al. (1995) found a maximum plateau of tomato lycopene concentration between 18 and 26 °C. Similarly, Brandt et al. (2006), Krumbein et al. (2006), and Dannehl et al. (2012) reported that the lycopene 64 content in tomatoes increases with increasing temperature and reaches its maximum 66 concentration at 25 °C. Temperatures below 12 °C strongly inhibit lycopene biosynthesis and temperatures above 32 °C stop the process (Dumas et al. 2003), affecting the activities of enzymes in the metabolic pathway and consequently, the fruit 69 carotenoid composition (Gautier et al. 2008). Likewise, phenolic compounds (e.g. chlorogenic acid, rutin and naringenin) increase with increasing mean temperatures 70 (Dannehl et al. 2012). In contrast, increasing the temperature from 27 to 32 °C reduces 71 ascorbate accumulation (Gautier et al. 2008). Thus, high temperature could also have a significant impact on the quality of fruits due to changes in primary or secondary 74 metabolism (Moretti et al. 2010).

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In this sense, in the present work, we intend to study the consequences of different climates (moderate to larger temperature increase) in order to mimic a possible future scenario under climate change. The data obtained may help in the interpretation of the effects of diurnal temperatures in the development and dry mass allocation as well as in the quality and composition of tomatoes. It is imperative to acquire such knowledge since a moderate or larger temperature increase may have different effects.

2. Materials and methods

2.1. Tomato plant culture and treatments

In June 2013, tomato seeds (*Solanum lycopersicum* L. cv. "Moneymaker") were sown into seed trays containing a peat-moss and vermiculite (50/50 v/v) and were germinated in a glasshouse at 25 °C in Avignon (Southern France, 44° N). Ninety plants with five growing leaves were transplanted per treatment into 15 cm diameter pots containing 5 L of potting soil, in a multispan Venlo-type greenhouse, N-S oriented. Plant nutrition, pests and diseases control were carried out in accordance with commercial practices. Water was supplied to the plants using a drip irrigation system to maintain a 20-30% drainage. The flowers were pollinated by vibration once a day during the anthesis phase, the inflorescences of each truss were pruned to six flowers, and lateral shoots were removed as soon as they appeared.

Plants were grown in four different compartments, and temperature treatments started on September 2013. The air temperature and humidity inside the different compartments of the greenhouse were measured using ventilated shelters equipped with temperature and humidity probes (HD 9817T1, Delta ohm, France). Temperatures were regulated by various techniques: heating, free ventilation by opening the aeration vents, forced ventilation, and evaporative cooling (cool box). These systems were gradually

implemented by the automaton that manages the greenhouse climate. The side walls of the compartments had been bleached to homogenize the radiation and the temperature of each compartment.

Temperature scenarios were applied one week before truss flowering and until fruit harvest, by heating the glasshouse during the warmest hours of the day, from 11:00 to 19:00, thus increasing diurnal temperature (+2 or +5 °C). We also used cooling to rapidly decrease temperature at the end of the day and study the possibility to mitigate the consequences of increased temperatures on fruit growth, physiology and quality.

The control treatment (CT) was similar to the outdoor climate, and the three other treatments had higher diurnal temperature:

- the High temperature Treatment (HT), achieved by heating from 11:00 to 19:00 with an average offset of 2.6 °C up to 5 °C for maximum diurnal temperature (Table 1),
- the Moderate temperature Treatment (MT), achieved by heating from 11:00 to 19:00 with an average offset of 0.5 °C up to 2 °C for maximum diurnal temperature,
- the 'end-of-day' treatment (ED), which was similar to MT except at the end of the day, from 19:00 to 22:00, when the temperature was reduced by 3°C in order to favor sugar allocation to the fruits (Walker and Ho, 1977).

Table 1. Air temperature and relative humidity recorded during the fourth truss development.

Treatments (conditions) and standard deviation					
Temperature (°C)	СТ	MD	ED	HT	
Maximal	25.3 ± 1.5	26.2 ± 1.6	26.1 ± 1.5	29.4 ± 1.5	
Average diurnal	21.7 ± 2.3	22.2 ± 3.1	22.1 ± 2.4	24.3 ± 2.5	
Average nocturnal	17.5 ± 2.9	17.5 ± 3.0	17.0 ± 2.8	18.0 ± 2.8	

Minimal	16.6 ± 1.5	16.6 ± 1.5	16.4 ± 1.4	17.0 ± 1.6
Relative humidity (%)	CT	MD	ED	HT
Maximal	92 ± 3	90 ± 2	92 ± 3	89 ± 3
Average	82 ± 6	80 ± 4	81 ± 5	78 ± 5
Minimal	66 ± 10	61 ± 7	63 ± 7	63 ± 7

CT = Control; MT = moderate increased temperature (from 11:00 to 19:00); ED = moderate increased temperature (from 11:00 to 19:00) followed by a decreased temperature applied at the end of the day (from 19:00 to 22:00); HT = High temperature (from 11:00 to 19:00).

2.2. Fruit sampling for biochemical analysis

The flowers of the fourth cluster of each plant were tagged at anthesis. Then, tagged fruits were harvested from nine days after anthesis (DAA) until ripeness (light red fruits), at nine development stages named A-09, A-16, A-23, A-30, A-37, A-44, A-51, O, R, corresponding to 9, 16, 23, 30, 37, 44 and 51 DDA, orange stage, and red stage, respectively. At each development stage, at least three fruits were harvested and pooled to form a replicate per treatment; four replicates were collected per stage and per treatment. Fruits were rinsed and dried on blotting paper, then each fruit was weighed, and equatorial and distal diameter were measured to determine fruit volume.

Fruits from a replicate were dissected, the jelly was removed, the placenta and seeds were extracted, and two opposite quarters of pericarp were weighed and dried in a stove at 70 °C during 72 hours to determine the percentage of dry weight as well as its partitioning between pericarp and seeds. The two other quarters were pooled and immediately frozen in liquid nitrogen, and stored at -80 °C for biochemical analyses. Then, tomato fruits were ground in liquid nitrogen to obtain a fine powder and stored at -80 °C before analyses.

An aliquot of 10 mg of ground freeze-dried tomato powder was used to determine soluble sugars, starch, and organic acids, following the methodology described by Gomez et al. (2002, 2007), with extraction and separation from a methanol-chloroform-water solution. The soluble sugars and organic assays were estimated using HPLC. Starch was hydrolyzed into glucose molecules, and the determination of NADH formed during the reaction was measured with a micro-plate reader (Multiskan Ascent V1.24) at 340 nm, which corresponds to NADH absorption.

The ascorbic acid (AsA) assay is based on the reduction of ferric ions to ferrous ions and the measurement of absorbance at 550 nm, following the method reported by Stevens et al. (2006), with an additional measurement to subtract other reducing compounds. Three aliquots were assessed per sample to determine the reduced ascorbate and the total ascorbate, after the addition of DTT (DL-Dithiothreitol) to reduce the oxidized form of vitamin C, or after the addition of ascorbate oxidase to estimate other reducing compounds that could bias the assay.

The carotenoid extraction was performed according to Sérino et al. (2009). The assay was performed using HPLC with a DAD UV-visible detector (UV6000LP, Thermo Separation Products, Rivera Beach, FL), with four working wavelengths, 474 nm for lycopene, 454 nm for β -carotene, 286 nm for phytoene, and 448 nm for lutein.

The polyphenols extraction was performed on 20 mg of cold-dried fruit powder and a mix solution of methanol/water (70:30 v/v), the samples were agitated (grinder vibrant–oscillating, Retsch GmbH, Rheenische, StraBe 36, 42781 Haan, Germany, with microtube adaptor) for 1 min at 30 osc/s. Then, the samples were centrifuged for 5 min (4 °C; 13200 rpm), and finally, the supernatant was filtered (GHP 0.2 μ m, 13 mm of diameter). The assay was performed using HPLC (Uptisphere, UP3HDO\$15QK, L = 150 mm, \emptyset = 3 mm, particles = 3 μ m). The spectral data for all the peaks were

accumulated in the range 200-800 nm and the chromatograms were recorded at 330 and 356 nm, with specific retention time for rutin, naringenin chalcone, caffeic acid glucoside, chlorogenic acid, caffeic acid derivative 1, and caffeic acid derivative 2 at 330 nm.

Structural dry matter (sDM) was calculated by subtracting the reducing sugars and organic acids to fruit dry matter. Consequently, fluctuations in water, sugar or acid contents do not influence the estimate of the fruit content of other components when expressed on a sDM basis (Rahmati et al. 2015).

2.3. Fruit sampling for conductance measurement

The skin fruit conductance was assessed for ten fruits per treatment at light red ripe stage. Ten fruits per treatment were harvested at 09:00, and after the pedicles were sealed with paraffin, the fruits were allowed to freely transpire in a controlled environment under forced and large ventilation, i.e. with a negligible boundary layer resistance. Subsequently, measuring the fruit water loss against time allowed the estimation of the fruit surface conductance according to Fishman and Génard (1998).

2.4. Statistical analyses

Statistical differences among assayed variables were determined by analyses of variance, considering the experiment as a complete randomized design, and the development stage, the treatments, and their interactions as explanatory factors. The comparison between means was carried out using Tukey's test ($\alpha = 0.05$ unless specified). The statistical analyses were performed with XLSTAT (Addinsoft Inc. Long Island City, USA).

3. Results

In the present experiment, the mean temperatures were of 21.7 °C (CT) and 24.3 °C (HT) during the day, and of 17.0 °C (ED) and 18.0 °C (HT) during the night (Table 1). The average maximal temperatures were 25.3 °C (CT) and 29.4 °C (HT). On a daily scale, the various climate management systems allowed differentiating the treatments with increased temperatures from CT to HT, MT, and ED, showing intermediate temperature variations (Figure 1A). The impact of the end-of-day treatment (ED) was only visible in air temperature at 20:00. Relative humidity ranged from 61 to 92%, with a night and day cycle and a mean value around 80% (Figure 1B, Table 1).

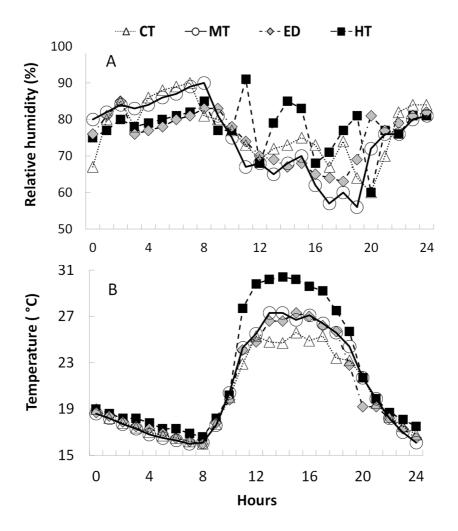


Figure 1. Example of a typical daily evolution of relative humidity (recorded on September 22, 2013) (A) and air temperature (B) within the four greenhouse

compartments depending on the four treatments applied: control treatment (CT); moderate temperature increase (MT); high temperature increase (HT); and temperatures similar to those of the MT treatment during the day and night, with the exception of a rapid drop in temperature at the end of the day (ED).

Fruit under HT started ripening sooner on a daily basis compared to other treatments. The fruit growth duration, defined as the time between anthesis and the fruit light red stage, was reduced by five days from 63.1 to 58.5 days (Table 2) from CT to HT. The fruits grown under the intermediate temperature treatments (ED, MT) showed an intermediate maturation time of around 60 days. We calculated the fruit growth duration (from anthesis to red ripe stage) on a thermal time basis, considering a thermal time basis (TTB) of 5.7 °C according to Adams et al. [4], 10 °C according to Calado and Portas (1987) and Okello et al. (2014), or 0 °C (Table 2). Considering a thermal time basis of 0 °C was the only way to find a constant fruit growth duration whatever the temperature treatment was (average value 1191 °C d).

Table 2. Changes in physical and biochemical traits of ripe tomato fruits (stage R) according to treatment. Data are mean + SE.

Fruit quality traits	CT	MT	ED	HT	P value
Days to reach maturity	63.10 ± 1.12 a	60.16 ± 0.98 ab	60.72 ± 1.18 ab	58.50 ± 1.01 b	0.03
GDDa to reach maturity (TTB 0 °C)	1185 ab	1173 ab	1165 a	1239 b	0.142
GDD to reach maturity (TTB 5.7 °C)	826 a	830 a	819 a	906 b	0.021
GDD to reach maturity (TTB 10 °C)	554 a	571 a	558 a	655b	0.003
Fruit fresh weight (FW g)	$96.0 \pm 7.3 \text{ b}$	$84.2 \pm 6.1 \text{ ab}$	$77.1 \pm 3.2 a$	$78.1 \pm 4.3 a$	0.05
Fruit water conductance (cm/h)	35.34 ± 0.04 a	32.49 ± 0.05 a	31.81 ± 0.04 a	$22.31 \pm 0.06 \text{ b}$	0.028
Pericarp fresh weight (g)	$72.97 \pm 3.74 \text{ b}$	64.03 ± 4.16 ab	60.20 ± 3.85 a	$58.57 \pm 3.50 \text{ a}$	0.09
Pericarp dry weight content (% FW)	4.57 ± 0.13 a	4.60 ± 0.55 a	4.84 ± 0.13 a	4.54 ± 0.13 a	0.39
Seeds fresh weight (g)	20.69 ± 2.53 a	18.97 ± 2.33 a	18.77 ± 1.86 a	$17.40 \pm 1.87 \text{ a}$	0.44
Seeds dry weight content (% FW)	6.77 ± 0.81 b	6.45 ± 0.79 ab	$6.82 \pm 0.65 \text{ b}$	6.22 ± 0.75 a	0.04
Macro compounds (g kg ⁻¹ FW)					
Glucose	$7.54 \pm 0.45 \text{ a}$	7.26 ± 0.33 a	7.80 ± 0.32 a	7.81 ± 0.32 a	0.60
Fructose	8.51 ± 0.49 a	8.00 ± 0.30 a	8.64 ± 0.30 a	8.89 ± 0.33 a	0.28
Sucrose	0.15 ± 0.04 a	0.19 ± 0.05 a	0.26 ± 0.06 a	0.24 ± 0.04 a	0.33
Starch	0.30 ± 0.03 a	0.37 ± 0.08 a	0.36 ± 0.04 a	0.25 ± 0.02 a	0.20
Citric acid	1.93 ± 0.08 a	2.08 ± 0.09 ab	$2.20 \pm 0.07 \text{ b}$	1.95 ± 0.09 a	0.10

Malic acid	1.40 ± 0.04 a	$1.72 \pm 0.12 \text{ b}$	$1.73 \pm 0.09 \text{ b}$	1.45 ± 0.08 a	0.01
Structural dry weight (g g ⁻¹ DW)	0.568 ± 0.012 a	0.573 ± 0.009 a	0.567 ± 0.010 a	0.547 ± 0.009 a	0.22
Micro compounds (mg kg ⁻¹ FW)					
AsA + DHA	89.70 ± 5.46 ab	93.72 ± 6.02 ab	87.62 ± 6.21 b	104.27 ± 5.17 a	0.14
AsA	78.55 ± 5.39 a	81.03 ± 4.06 a	77.52 ± 6.57 a	$103.08 \pm 5.47 \text{ b}$	0.002
Lutein	$0.39 \pm 0.03 \text{ b}$	0.49 ± 0.04 ab	0.41 ± 0.04 ab	0.51 ± 0.04 a	0.06
Lycopene	11.96 ± 2.28 b	14.59 ± 2.55 b	$15.20 \pm 2.94 \text{ b}$	32.66 ± 5.84 a	0.0001
β-Carotene	2.40 ± 0.11 a	2.52 ± 0.11 a	2.46 ± 0.11 a	2.51 ± 0.09 a	0.85
Phytoene	$3.34 \pm 0.71 \text{ b}$	$3.53 \pm 0.72 \text{ b}$	$3.54 \pm 0.64 \text{ b}$	7.96 ± 1.48 a	0.0008
Caffeic acid glucoside	5.28 ± 0.47 a	4.74 ± 0.26 a	5.77 ± 0.50 a	5.12 ± 0.52 a	0.45
Chlorogenic acid	11.34 ± 1.53 a	11.88 ± 1.62 a	9.19 ± 1.25 a	13.31 ± 1.33 a	0,23
Chlorogenic acid derivative 1	1.78 ± 0.16 a	1.50 ± 0.10 a	1.70 ± 0.18 a	1.85 ± 0.10 a	0.21
Chlorogenic acid derivative 2	1.30 ± 0.13 a	1.08 ± 0.09 a	1.34 ± 0.19 a	1.23 ± 0.09 a	0.45
Rutin	7.17 ± 0.69 a	6.19 ± 0.78 a	6.68 ± 0.50 a	$7.84 \pm 0.77 \text{ a}$	0.34
Naringenin chalcone	74.58 ± 12.07 a	63.40 ± 9.84 a	67.12 ± 8.86 a	83.00 ± 9.11 a	0.44

GDD^a: Growing Degree Day; TTB: Thermal time basis; AsA: ascorbic acid; DHA: dehydroascorbic acid; CT= Control conditions; ED = End-of-day temperature conditions (from 19:00 to 22:00); HT = High temperature conditions (from 11:00 to 19:00); LT = Low temperature condition (11:00 to 19:00); FW = Fruit fresh weight (g); DW: Dry weight. Different letters in the same column indicate a significant difference between means according to Tukey's test (α = 0.1), significant differences are highlighted in bold.

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The slight reduction of growth duration, corresponding to a reduction of 7%, had a strong and larger impact on the fruit fresh weight accumulation. The fruit fresh mass decreased by 18.6%, from 96 g to 78 g. Such fruit growth reduction was mostly due to a decreased pericarp fresh weight (minus 20%, Figure 2A), and to a minor extent, to a decrease in the seeds (-16%, Table 2). As usually observed, the pericarp dry weight (DW) decreased during fruit development. However, the pericarp DW remained slightly greater under the ED treatment ($\alpha = 0.1$, Figure 2B) and until maturity (Table 2, $\alpha = 0.09$). Moreover, it decreased with increasing temperature. The fruit surface conductance was significantly modified by temperature treatments (Table 2).

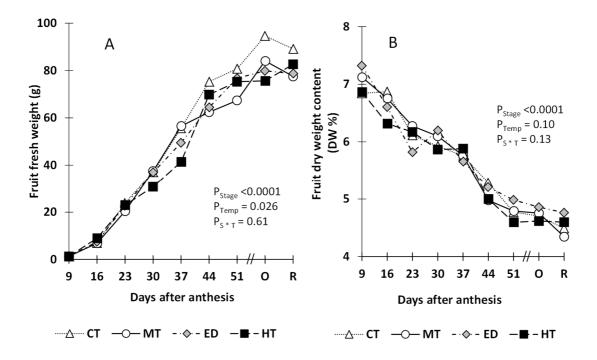


Figure 2. Evolution of fruit fresh weight (A) and fruit dry weight (B) contents as the fruits develop from nine days after anthesis (to light red stage) depending on the four temperature treatments applied (for treatments legends, please refer to Figure 1 caption). The fruits (a total of 290 fruits) were harvested from nine to 51 days after anthesis,

orange (O) and red (R) stages. The data represent average values. p-values were obtained from an ANOVA considering the effect of fruits development stage (Stage), the temperature of each treatment (Temp) and the interaction between fruit Stage and Treatment (S * T).

HT reduced the fruit surface conductance by 37% compared to CT, but there was no significant effect for fruits harvested on plants grown under MT or ED. Expressed on a fresh weight basis (FW), the pericarp sugars composition was not modified at harvest, whatever the treatment. In contrast, organic acid concentrations were affected, with lower concentrations of malic and citric acid under CT and HT, and higher concentrations for fruits grown under the ED treatment (+24 %) compared to the control. Temperature treatments also affected minor compounds accumulation. At harvest, fruit grown under HT had a significant increased amount of ascorbate (+31%) compared to fruits under control conditions. In contrast, moderate increased temperatures (ED, MT) did not affect the fruit ascorbate content. Similarly, carotenoids contents (except β-carotene) were higher under HT; lutein, phytoene and lycopene contents increased by 31%, 138% and 173%, respectively under HT compared to CT, but no significant changes were recorded under moderate temperature increase (ED or MT). Additionally, whatever the temperature treatment, there was no significant effect of moderate or stronger temperature increase on phenolic compounds.

We followed the kinetics of primary and secondary compounds accumulation of within the fruits, expressed per structural dry matter. When expressed on a structural dry matter basis, any change in primary or secondary metabolite content is related to changes in the fruit metabolism and not to a dilution (or concentration) effect triggered by the accumulation (or the shortage) of water or other compounds.

If the starch content evolution during fruit growth showed similar patterns whatever the treatment, the maximal content was affected by the treatments, and especially by the HT treatment, where there was a significant reduction in the starch accumulation (Figure 3A).

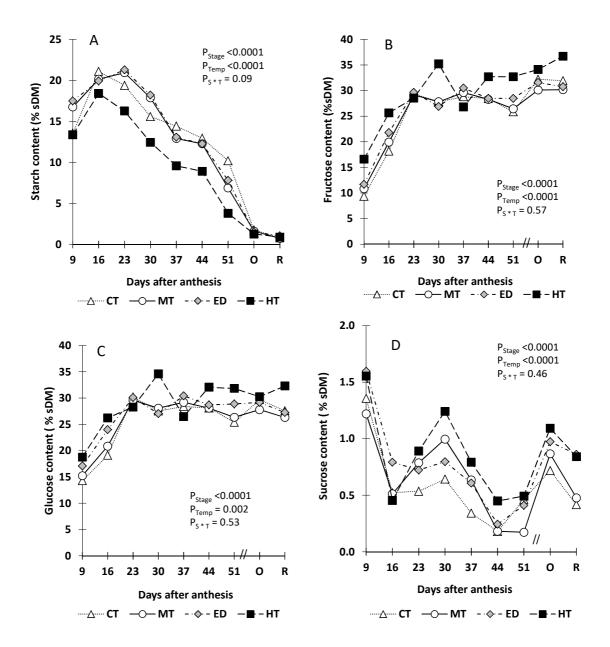
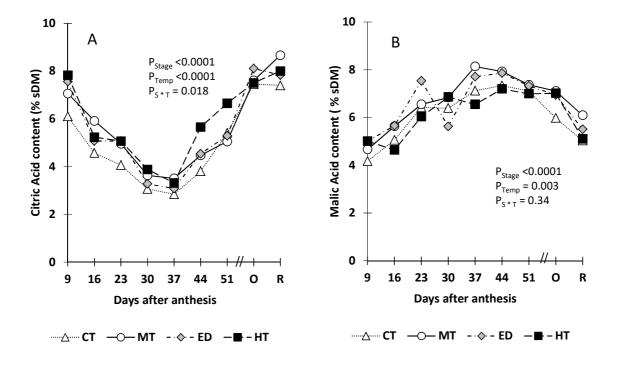


Figure 3. Evolution of the contents of starch (A), fructose (B), glucose (C) and sucrose (D) to the structural dry weight content as the fruits develop from nine days after

anthesis (to light red stage) depending on the four temperature treatments applied (for treatments legends, please refer to Figure 1 caption).

Glucose, fructose and sucrose contents increased in fruit growing under the HT treatment (Figures 3B, C, D). Similarly, the time course of citric acid was affected by HT, with an increased accumulation from 37 DAA. However, there was no difference at maturity (Figure 4A). On the other hand, malic acid accumulation was reduced by the HT treatment, also without significant difference at maturity (Figure 4B). Consequently, the ratio between total soluble sugars and acids was greater under HT when compared to the control (Figure 4C).



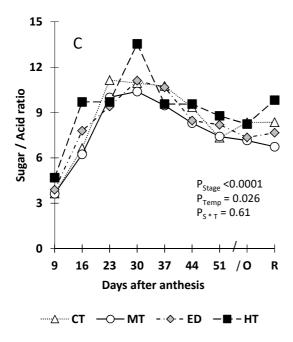


Figure 4. Evolution of the contents of citric acid (A), malic acid (B) and sugar/acid ratio (C) to the structural dry weight content as the fruits develop from nine days after anthesis (to light red stage) depending on the four temperature treatments applied (for treatments legends, please refer to Figure 1 caption).

Total ascorbate concentrations (tAsA) tended to increase under HT treatments (α = 0.14, Table 2). This was due to the accumulation of the reduced form (ascorbic acid content, rAsA) which significantly increased under HT treatments, particularly from 51 DAA until ripening stage (Figure 5). Thus, the proportion of vitamin C in a reduced form (rAsA/tAsA ratio) was significantly greater for fruits harvested under HT (Table 2).

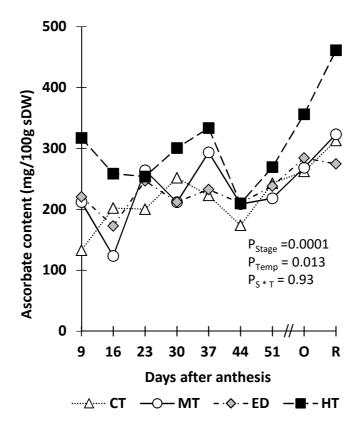


Figure 5. Evolution of the content of reduced ascorbate to the structural dry weight content as the fruits develop from nine days after anthesis (to light red stage) depending on the four temperature treatments applied (for treatments legends, please refer to Figure 1 caption).

4. Discussion

Despite the fact that the average temperature differences were rather small and close to climate projections for the near future in France, we observed that they affected different processes and resulted in reduced fruit growth and changes in fruit quality traits at harvest. In the present experiment, the most visible quality trait impacted by the temperature treatments was the final fruit size at harvest. We observed that the fruit growth duration decreased in a linear way with increased temperatures: the higher the temperature, the lower the duration of fruit development (from anthesis until red ripe stage). Similar findings have been reported with a reduced developmental duration to reach ripe stage for tomato fruits grown under increased temperature, especially during the fruit later stages of maturation (Adams et al. 2001; Fleisher et al. 2006; Hurd and Graves, 1985). In other species like chili, heat stress following anthesis or from 30 DAA until harvest also reduced the growth period of chili fruits by 15 and 10 days, respectively (Pagamas and Nawata, 2008). High temperatures generally accelerate fruit maturation and ripening, and this could be an adaptive mechanism to stress conditions, enabling the plant to disperse its seeds faster when under stress (Mizhari, 1982). Experiments using locally heated fruits showed that heating fruits reduced fruit size, since it modified the cell division orientation (Fanwoua et al. 2012) and reduced cell expansion (Okello et al. 2014). In another study, rising temperatures from 20/20 day/night to 25/25°C had no effect on tomato fruit size (Bertin, 2005). This might be linked to the fact that i) the plant remains at optimal temperature regarding fruit growth, or that ii) different processes are affected by increased temperatures, but they compensate one another. Indeed, Bertin (2005) reported a compensatory effect between a reduced duration of fruit cell division (which reduced the number of fruit cells per fruit) and an increased period of cell expansion, where the cell expansion rate was not affected. When the temperature further increases, some processes will not be at their

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optimal temperature and will not be compensated. In addition, other processes can be affected; so the reduction of fruit size could be related to other factors such as limited assimilate supply, as it was reported for higher fruit load under HT (Gautier et al. 2005). Indeed, the fruit dry weight accumulation was reduced under HT (Figure 2), which could be linked to the increased carbohydrate loss by respiration. Furthermore, the increase in temperature causes a higher vapor pressure deficit (VPD), which could lead to a larger water loss by transpiration. However, this higher climatic demand could be partly, fully, or over-compensated by the observed decrease of the fruit skin conductance associated with the HT treatment. Similar findings have been reported for leaves growing under water stress (Cameron et al. 2006), which could be the consequence of changes in the cuticular wax composition or thickness (Vogg et al. 2004). If the water conductance of young growing fruits is also decreased, it will impair fruits photosynthesis and transpiration (Blanke and Lenz, 1989). Even if their photosynthetic activity remains low (Carrara et al. 2001), it can contribute to 10 or 15% of the carbon skeleton of the fruits (Tanaka et al. 1974). Thus, reduced fruit photosynthesis alters carbon allocation to seeds and may impact the seeds quality (Lytovchenko et al. 2011).

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In addition, reducing fruit transpiration helps maintaining fruit water pressure and limits the entry of water and carbon into the fruits through the phloem. This could partly explain the lower rate of fruit growth observed under HT. Similar data have been reported by Li et al. (2001). in covered peach fruits: a decrease in transpiration, respiration, and photosynthesis rates could contribute to reducing the quantity of carbohydrates entering into the fruits, which in turn would result in reducing soluble solids content. We observed a 37% reduction in fruit water conductance from the

control to the HT treatment (at red ripe stage), which parallels the 33% decrease in dry mass accumulation.

In the present experiment, dry matter and structural dry matter contents were not affected by HT applied during tomato fruits development (Figures 1C and D). However, we observed that the ED treatment tended to increase the pericarp DM content. Under moderate temperatures, Heuvelink (1995) reported no significant difference in the DM content of fruits harvested on plants grown at 18 °C or 24 °C. In contrast, under higher temperatures, Rosales et al. (2011) found increased dry matter contents in tomato fruits produced at 30/12 °C compared to 25/14 °C (day/night respectively), which could also be linked to water limitation in the summer conditions. In the present experiment, the temperature decrease at the end of the day may have been too low, and it would be interesting to consider broader changes in ED temperature to confirm its potential role in improving fruits dry matter content. In addition, we observed that HT tended to reduce both the carbon influx to the fruit and its partitioning to the seeds, and consequently, seed storage and quality could be altered (Lytovchenko et al. 2011).

The present work gives a better understanding of the influence of HT on processes linked to fruit quality traits, considering both visual traits, such as fruit size, and fruit composition. In our study, the time course of starch accumulation and degradation was significantly affected by temperature; the HT treatment reduced starch accumulation particularly from 16 to 58 DAA (Figure 3A). The period during the first 10-30 DAA following the fruit set in tomato fruits is the one when the rate of starch biosynthesis is potentially at its highest levels (Walker and Ho, 1977). Similar data were obtained when increasing temperature in potatoes, which perturbs starch synthesis and related enzymatic activities, increasing leaf respiration and carbon loss (Geigenberger et al. 1998). These changes in starch metabolism were accentuated by the effect of HT on

fruits phenology, which accelerates fruit maturation and consequently starch degradation. This more rapid degradation of starch is in agreement with our observed increased soluble sugars content (glucose, fructose and sucrose content, Figures 3B, C and D).

Rosales et al. (2011) reported that sugar contents increased at HT while organic acids contents diminished. Similarly, Wang and Camp (2000) also reported increased sucrose contents in two strawberry varieties when increasing temperatures from 18/12 to 25/22 °C day/night, respectively. In tomatoes, the effect of HT on organic acids accumulation was different according to each acid: citric acid contents increased by the HT treatment from 37 to 58 DAA, and then decreased at ripe stage (65 DAA) (Figure 4A). Malic acid contents were reduced by the temperature treatments applied during tomato fruits growth (Figure 4B); consequently, the ratio between soluble sugars and organic acids was increased by the HT treatment during tomato fruits growth (Figure 4C).

Ascorbic acid contents (AsA) increased in fruits grown under HT, and particularly during ripening (Figure 5). At this stage, most of the vitamin C was reduced, which is the active form that helps the plant to resist oxidative stress. Rosales et al. (2009) reported similar findings when studying the seasonal change in tomato oxidative compounds: they found the greatest amount of ascorbate during the hottest period of the year.

Lutein contents in tomato fruits were not changed by temperature treatments and decreased during fruit growth and ripening (Table 2). Lycopene, β -carotene and phytoene were increased under the HT treatment (Table 2). Temperature may have contrasting effects on carotenoids contents as previously reported by Dorais et al. (2008), and a decrease in the content of carotenoids has been found under extreme

temperatures (either high above 32 °C or low below 12 °C) (Dumas et al. 2003). In our experiment, temperatures were lower and close to optimal conditions around 22-26 °C. Thus, lycopene and other carotenoids' syntheses were not inhibited.

This study reinforces our knowledge of the effects of temperature on the different processes involved in building tomato quality. Even a moderate increase in temperature can affect the accumulation of water and carbon in the fruit, the synthesis and accumulation of starch, the transpiration of the fruit, and the primary and secondary metabolism, so that the growth and quality of the fruit at maturity are affected. This study also aimed to determine if crop and climate management can prevent the detrimental effects of increased temperatures and help growers cope with consumers' demand for tastier fruits. The ED treatment applied increased the pericarp dry matter content, but the temperature drop at the end of the day was moderate and it did not promote fruit growth duration, nor did it increase the sugar acid ratio. In a context of climate change, crop species will have to undergo an adaption to temperature increases, either through genetics, by investigating the variability of the sensitivity of these processes to temperature, or through cultural practices that mitigate climate changes.

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Conflict of interests

The authors have no conflict of interest to declare.

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