

# Dynamical modeling of reaction networksMathematics of system biology

Romain Yvinec

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## Dynamical modeling of reaction networks Mathematics of system biology

Romain Yvinec

Systems Biology and Networks

Motivations and Objectives

Chemical reaction network formalism

Parameter estimation

**Applications** 

### Outline

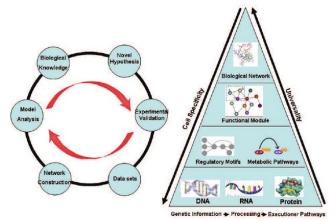
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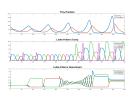
Oltvai and Barabasi, Science 25:763-764, 2002.

## Small networks in Population dynamics

(Interactions between populations, Epidemiology)

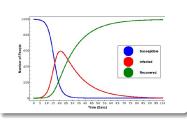
#### Lotka-Volterra model

$$\begin{array}{ccc}
\varnothing & \xrightarrow{k_1} & A \\
A+B & \xrightarrow{k_2} & 2B \\
B & \xrightarrow{k_3} & \varnothing
\end{array}$$



#### S.I.R model

$$\begin{array}{ccc} S + I & \xrightarrow{k_1} & I + I \\ I & \xrightarrow{k_2} & R \end{array}$$

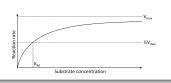


## Small networks in molecular biology

('Toy' molecular models with isolated components)

#### **Enzymatic kinetics**

$$E + S \xrightarrow[k_1^-]{k_1^+} ES \xrightarrow[k_2^-]{k_2^+} E + P$$



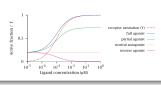
#### Pharmacology model

$$R_{i} \rightleftharpoons R_{a}$$

$$A + R_{i} \rightleftharpoons AR_{i}$$

$$A + R_{a} \rightleftharpoons AR_{a}$$

$$AR_{a} \rightleftharpoons AR_{i}$$



## (Single) Gene Expression

$$G \xrightarrow{\lambda_{1}} G + M$$

$$M \xrightarrow{\lambda_{2}} M + P$$

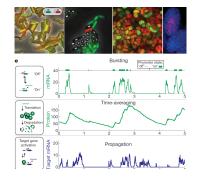
$$M \xrightarrow{\gamma_{1}} \varnothing$$

$$P \xrightarrow{\gamma_{2}} \varnothing$$

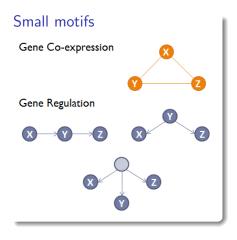
$$G \xrightarrow{k_{off}} G_{off}$$

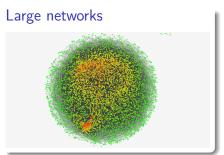


Eldar and Elowitz (Nature 2010)

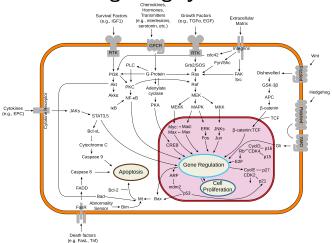


## Co-expression genes network

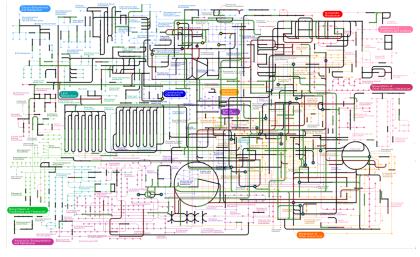




## Signaling system



### **Metabolomic Network**



### Outline

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## Possible applications of mathematical modelling

- Understand non-trivial behavior of a biological system (by reproducing this behavior with an understandable model, starting from 'first principles')
- ▶ Help to identify key regulatory process in signaling cascades
- Quantify some non-observables quantities, in particular : molecules concentrations, reaction rates.

## Possible applications of mathematical modelling

**Today :** Understand the mathematical formalism of dynamical reactions network

## Possible applications of mathematical modelling

**Today :** Understand the mathematical formalism of dynamical reactions network

- Build a model from a network of interactions [Cell Designer]
- Parameter calibration with kinetic data [GraphPad Prism / Copasi]

## Warning!



#### **#Ihatemathematics**

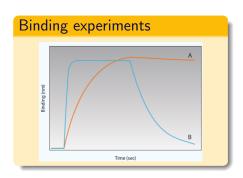
The following couples of slides contain some abstract notions... Why that?

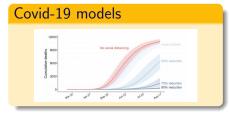
## Maths is about structures...and is generic!



## Reactions networks applications range from

- Chemistry
- molecular biology
- epidemiology
- ▶ and beyond!





### Outline

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#### Definition

A chemical reaction network is given by thee sets (S, C, R):

- Species,  $S := \{S_1, \dots, S_d\}$ : molecules that undergo a serie of chemical reactions.
- Reactant / Product,  $C := \{y^1, \dots y^n\}$ : Linear combination of species, that represent either 'what is consumed', or 'what is produced', in any reaction.
- Reaction,  $\mathcal{R} := \{y^k \to y^{k'}, y^k, y^{k'} \in \mathcal{C}\}$ : ensemble of reactions between species or combination of species (directed graph between Reactant / Product).

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- Mass-action law,  $\kappa$ : a list of positive parameter (kinetic rate) for each reaction in  $\mathcal R$

#### Example

$$A \stackrel{k^+}{\rightleftharpoons} B$$

Species 
$$\mathcal{E} := \{A, B\}$$
  
R / P  $\mathcal{C} := \{A, B\}$   
Reaction  $\mathcal{R} := \{A \to B, B \to A\}$   
Rate  $\{k^+, k^-\}$ 

#### Example (minimal cAMP production model)

$$L + R \xrightarrow{k_{on}} LR$$

$$ATP + LR \xrightarrow{k^{+}} cAMP + LR$$

$$cAMP \xrightarrow{k^{-}} AMP$$

$$Species \mathcal{E} := \{L, R, LR, ATP, cAMP, AMP\}$$

$$R / P \mathcal{C} := \{L + R, LR, ATP + LR, cAMP + LR, cAMP, AMP\}$$

$$Reaction \mathcal{R} := \{L + R \rightarrow LR, LR \rightarrow L + R, ATP + LR \rightarrow cAMP + LR, cAMP \rightarrow AMP\}$$

$$Rate \{k_{on}, k_{off}, k^{+}, k^{-}\}$$

## Chemical Reaction Network, FSHR-induced cAMP signals

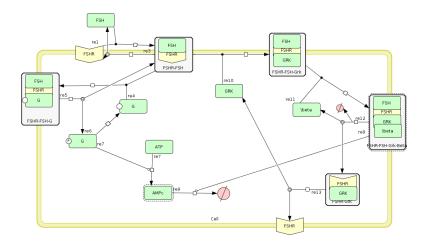
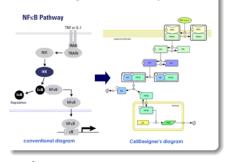


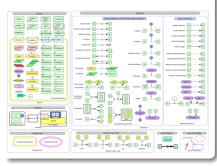
Figure – FSHR (G $\alpha$ s-coupled GPCR) models build on CellDesigner

## Cell Designer

#### About diagrams and layout...



#### ...with well-defined conventions



Funahashi, A., Tanimura, N., Morohashi, M., and Kitano, H., CellDesigner: a process diagram editor for gene-regulatory and biochemical networks, BIOSILICO, 1:159-162, 2003

## Chemical Reaction Network, "real" example

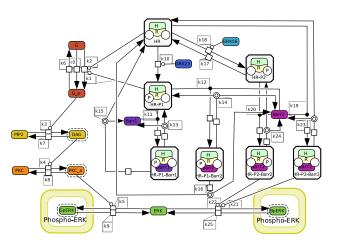


Figure – ERK Phosphorylation pathways build on CellDesigner, Heitzler et al. MSB 2012

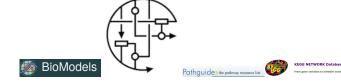
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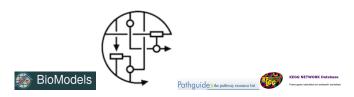
NB: there exists public databases of reaction network models.



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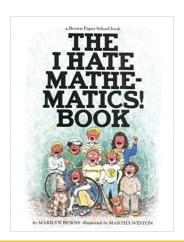


NB (bis) : A reaction network is a network... but a network is **NOT** a reaction network!

#### We have seen

- Many examples of dynamical system biology models
- ▶ The formalism of *reaction network* models
- ▶ How to build a reaction network within Cell Designer
- What about dynamics?

## Warning!



#### **#Ihatemathematics**

The following couples of slides contain some abstract notions...let's try to catch the meaning!

#### We build a model that

- Keep track of concentration of species along time.
- Satisfy Law of Mass action: The velocity of a reaction is proportional to the concentrations of its reactants.
- Is a system of Ordinary Differential Equations, in which reactions are "added" on top of each other, e.g. they happens continuously and simultaneously.

#### Example

$$A \stackrel{2}{\underset{0.1}{\rightleftharpoons}} B$$

$$\begin{array}{lcl} \frac{dx_A}{dt}(t) & = & -0.1x_A(t) + 2x_B(t) \,, & x_A(t=0) = A_{tot} \\ \frac{dx_B}{dt} & = & +0.1x_A(t) - 2x_B(t) \,, & x_B(t=0) = 0 \end{array}$$

 $x_A(t)$  = time dependent concentration of species A

#### Example

$$L + R \xrightarrow{k_{on}} LR$$

$$\begin{array}{lcl} \frac{dx_L}{dt}(t) & = & -k_{on}x_L(t)x_R(t) + k_{off}x_{LR}(t) \,, & x_L(0) = Dose \\ \frac{dx_R}{dt}(t) & = & -k_{on}x_L(t)x_R(t) + k_{off}x_{LR}(t) \,, & x_R(0) = R_{tot} \\ \frac{dx_{LR}}{dt}(t) & = & k_{on}x_L(t)x_R(t) - k_{off}x_{LR}(t) \,, & x_{LR}(0) = 0. \end{array}$$

#### Example (minimal cAMP production model)

$$L + R \xrightarrow{k_{on}} LR$$

$$ATP + LR \xrightarrow{k^{+}} cAMP + LR$$

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$$\frac{dx_{L}}{dt} = -k_{on}x_{L}x_{R} + k_{off}x_{LR}, \quad x_{L}(0) = Dose$$

$$\frac{dx_{R}}{dt} = -k_{on}x_{L}x_{R} + k_{off}x_{LR}, \quad x_{R}(0) = R_{tot}$$

$$\frac{dx_{LR}}{dt} = k_{on}x_{L}x_{R} - k_{off}x_{LR}, \quad x_{LR}(0) = 0$$

$$\frac{dx_{LR}}{dt} = k^{+}x_{ATP}x_{LR} - k^{-}x_{CAMP}, \quad x_{CAMP}(0) = 0$$

$$\frac{dx_{CAMP}}{dt} = -k^{+}x_{ATP}x_{LR} \quad x_{ATP}(0) = ATP_{tot}.$$

## But what is an "Ordinary Differential Equation"? A math theory in one slide!

The equation

$$\frac{dx}{dt} = v(x),$$

is numerically solved by successive time-step iteration, of small length  $\Delta t \ll 1$  :

1) Start at a given initial condition  $x_0$  at time  $t_0 = 0$ 

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- 2) To calculate the value of x at the first time step, remember that (assuming constant speed)

Final Position = Initial Position + velocity \* Time,

which becomes, in mathematical notations,

$$x(\Delta t) = x_0 + v(x_0) * \Delta t,$$

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Final Position = Initial Position + velocity \* Time,

which becomes, in mathematical notations,

$$x(\Delta t) = x_0 + v(x_0) * \Delta t,$$

Iterate: To calculate the value of x at the next time step, use

$$x((i+1)*\Delta t) = x(i*\Delta t) + v(x(i*\Delta t))*\Delta t,$$

# But what is an "Ordinary Differential Equation"? A math theory in one slide...and a figure!

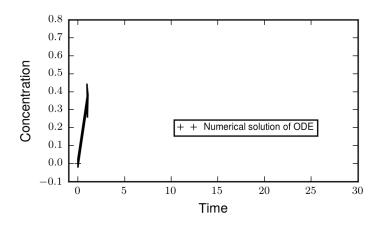


Figure – Solving an ODE

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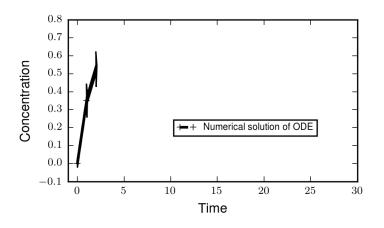


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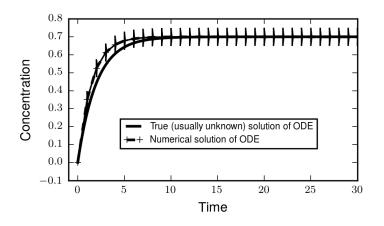
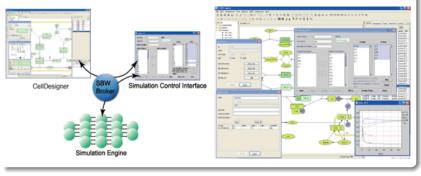


Figure – Solving an ODE

### Solving an ODE in practice : no need to code!

### ODE solver within Cell Designer



Funahashi, A., Tanimura, N., Morohashi, M., and Kitano, H., CellDesigner: a process diagram editor for gene-regulatory and biochemical networks, BIOSILICO, 1:159-162, 2003

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NB : You don't need to code, but you need to specify kinetic rate and initial condition values to simulate a reaction network.

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NB: You don't need to code, but you need to specify kinetic rate and initial condition values to simulate a reaction network.

NB (bis): You can play with this tools to "explore" the behavior of a model. But that can be time consuming and inefficient...

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What about inferring those values from data?

### Outline

Systems Biology and Networks

Motivations and Objectives

Chemical reaction network formalism

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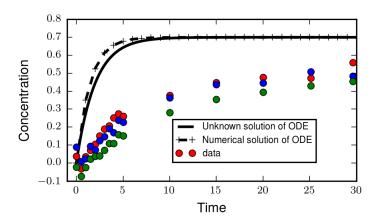
**Applications** 

## Parameter and network inference in Chemical Reaction Network

Goal: Given some time series data, find the minimal (biologically plausible) reaction network with its parameter (=reaction rates and initial conditions) that fits consistently the data.

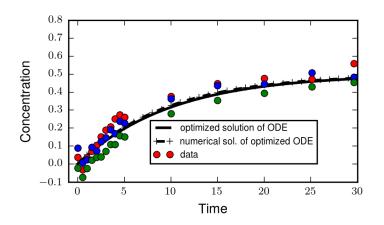
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## Regression analysis and Parameter estimation with time series : What is difficult?

- ▶ In linear models, there exists a *unique optimal* solution
- Yet in practice, (generalized) linear models do not perform well on biochemical data due to Heteroscedasticity and highly dependent time point data.

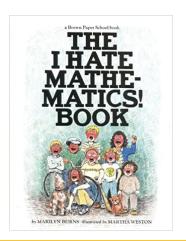
## Regression analysis and Parameter estimation with time series : What is difficult?

- ► For most of (nonlinear) reaction network, there is no guarantee to find a unique optimal solution.
- Reaction network models allows to perform multifactorial analysis ("Anova-like")

## Regression analysis and Parameter estimation with time series : What is difficult?

- ► For most of (nonlinear) reaction network, there is no guarantee to find a unique optimal solution.
- Reaction network models allows to perform multifactorial analysis ("Anova-like")
- Many other tools exists from the statistical field of time series analysis.

## Warning!



#### **#Ihatemathematics**

The following couples of slides contain some abstract notions...but that the last ones!

Goal: Given some time series data, find the minimal (biologically plausible) reaction network with its parameter (=reaction rates and initial conditions) that fits consistently the data.

**Strategy** 1) From a given network  $(\mathcal{S}, \mathcal{C}, \mathcal{R})$ , with given parameter values, solve the ODEs,

$$\frac{dx}{dt} = v(x,k), \quad x(0) = x_0,$$

and compute a distance between the solution and the data.

**Goal :** Given some time series data, find the minimal (biologically plausible) reaction network with its parameter (=reaction rates and initial conditions) that fits consistently the data.

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**Strategy** 2) Using **optimization** algorithms, find the best parameter values  $k, x_0$ , to minimize the distance

**Strategy** 3) If needed, change the reaction network (add or delete species/reactions)

Goal: Given some time series data, find the minimal (biologically plausible) reaction network with its parameter (=reaction rates and initial conditions) that fits consistently the data.

Statistics There exists a well developed statistical theory to assess the quality of a fit and to give confidence interval on parameter values (-> See Likelihood maximization or Bayesian statistics).

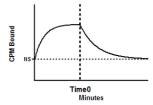
#### Parameter estimation in



## Predefined or user-defined time-dependent equations

#### Model

Radioligand=HotNM\*1e-9
Kob=[Radioligand]\*Kon+Koff
Kd=Koff/Kon
Eq=Bmax\*radioligand/(radioligand + Kd)
Association=Eq\*(1-exp(-1\*Kob\*X))
YatTime0 = Eq\*(1-exp(-1\*Kob\*Time0))
Dissociation= YatTime0\*exp(-1\*Koff\*(X-Time0))
Y=IF(X<Time0. Association. Dissociation) + NS



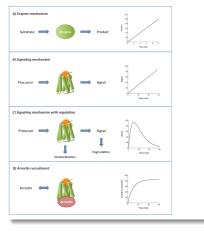
#### Parameter estimation in



- Limited to solvable models
- Adapted to analyze one single output at a time, assuming excess of Ligand.

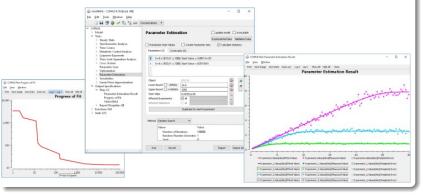


## Predefined or user-defined time-dependent equations



Hoare et al., Analyzing kinetic signaling data for G-protein-coupled receptors, Scientific Reports 10(1):12263 2020

### Parameter estimation in Copasi

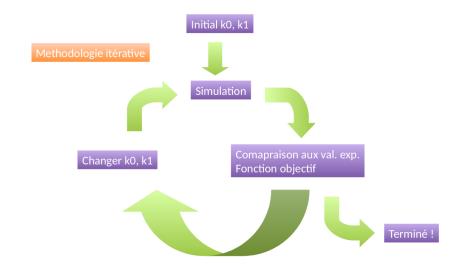


- Models can be imported from Cell Designer.
- Supports both graphical interface and command line.

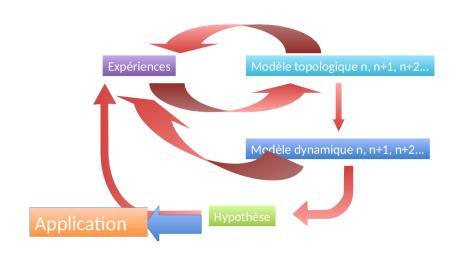
Bergman et al. COPASI and its applications in biotechnology, Journal of Biotechnology 261:215-220, 2017.

Hoops et al. COPASI: a COmplex PAthway SImulator. Bioinformatics 22:3067-74, 2006.

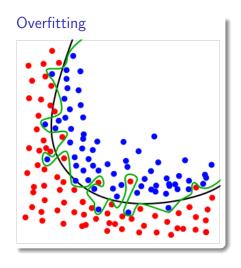
### Remember! it's an iterative and interdisciplinary workflow!



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### Is the monkey who typed Hamlet actually a good writer?



- There is a trade-off between toy minimal models and detailed biochemistry pathways.
- Overfitting leads to unreliable prediction and meaningless model / parameter value.
- (Advanced) statistical tools exist to sort this out: model selection (especially for hierarchical models) and parameter identifiability.

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- Many examples of dynamical system biology models
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- How to build and simulate a reaction network model within Cell Designer.
- How to calibrate parameters of a dynamical reaction network model with GraphPad Prism and/or Copasi.

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 $\ensuremath{\mathsf{NB}}$  : The full workflow can be long and require collaboration with statistician / applied mathematician.

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NB bis : **What about applications?** Go to session 8 : Computational approaches!

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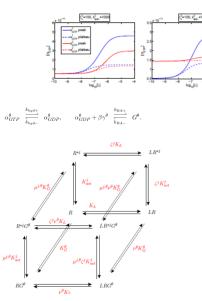
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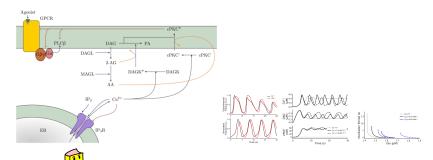
**Applications** 

# Some applications : Understanding G protein activation cycle



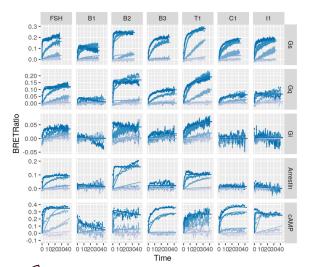
Bridge, Meads, Frattini, Winfield, Ladds, Modelling and simulation of biased agonism dynamics at a G protein-coupled receptor, J. Theoret. Biol. 442:44–65. 2018

# Some applications : Shedding light on GPCR-induced Calcium oscillations in Astrocytes



De Pittà, Ben-Jacob, Berry, G protein-coupled receptor-mediated calcium signaling in astrocytes, in Computational Glioscience, Springer 2019.

# Some applications : Revisiting signaling bias using dynamical model





De Pascali, ..., R.Y,..., in preparation.

### Conclusions

- Dynamical reaction network framework has many different applications.
- Its a powerful framework to reveal comprehensive spatio-temporal patterns behind GPCR signaling complexities.
- Its a powerful framework to analyze quantitatively time series data in GPCR signaling.
- Adequate tools foster necessary interdisciplinary collaborations by providing a common language.