

Kinetic biased signaling: towards a system biology definition of drugs selectivity

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Romain Yvinec

BIOS, INRAE Tours

BIOS group





INRAE Tours, Physiologie de la Reproduction et des Comportements

Biology & Bioinformatics of Signaling Systems : multidisciplinary approaches, linking biology, mathematics and information technology to experimental biology, in order to decipher the intracellular effects induced by reproductive hormones through the activation of their cognate receptors.

Functional selectivity, biased signaling

What is Drugs Selectivity?

• Several reaction pathways are generally associated to a given receptor, and lead to various cell response.



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- Differential activation of those reaction pathways, that differs between (natural or synthetic) ligand



Functional selectivity, biased signaling

What is Drugs Selectivity?

- Several reaction pathways are generally associated to a given receptor, and lead to various cell response.
- Differential activation of those reaction pathways, that differs between (natural or synthetic) ligand
- Drugs Selectivity = Liganddependent selectivity for certain signal transduction pathways at one given receptor



- ◊ Drugs Selectivity (or Biased Signaling) is a key concept to be distinguish from
 - Partial or full agonist.
 - Antagonist, inverse agonist.
 - Affinity (K_d) , potency (EC_{50}) , efficacy (E_{max}) .

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 - Partial or full agonist.
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 - Affinity (K_d), potency (EC_{50}), efficacy (E_{max}).
- A bias might be context-dependent (cell type, physiological state, etc.)
- ♦ Biased agonism is becoming a major tool in drug discovery.
- $\Rightarrow\,$ Candidate screening requires to accurately quantify bias.

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 - Partial or full agonist.
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 - Affinity (K_d) , potency (EC_{50}) , efficacy (E_{max}) .
- A bias might be context-dependent (cell type, physiological state, etc.)
- ◊ Biased agonism is also a powerful tool to challenge our knowledge of signaling systems.
- \Rightarrow "Perturbation" experiments with different biased ligands.

A receptor may adopt several spatial conformations, each of which has different activation pathway profiles.

Conformational selectivity = Ligand-specific modification of the energetic landscape, changing affinities and efficacies of signaling patways.





Kenakin, J Pharmacol Exp Ther (2011)

Theoretical foundation

A receptor may adopt several spatial conformations, each of which has different activation pathway profiles.

Conformational selectivity = Ligand-specific modification of the energetic landscape, changing affinities and efficacies of signaling patways.

Similar concept : modulating bias





Kenakin and Christopoulos, Nat. Rev. Drug Discov. (2013)

To speak about signaling bias, one necessarily needs **two** ligands and **two** responses, in a **same** cellular context.



 \Rightarrow We always compare a ligand with respect to a reference one.

Some examples

Bias quantification - standard method : operational model

Time-dependent bias

Biased quantification using dynamical model

Serotonine receptor $5 - HT_{2C}$

- Quipazine is biaised towards *PI* accumulation with respect to *AA* production, *compared to the reference agonist DOI*.
- LSD is not biased.



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Berg et al., Mol.
Pharmacol. (1998)
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Serotonine receptor $5 - HT_{2C}$

- Quipazine is biaised towards *PI* accumulation with respect to *AA* production, *compared to the reference agonist DOI*.
- LSD is not biased.
- $\Rightarrow \text{ Bias due to an } E_{max}$ difference.





Berg et al., *Mol. Pharmacol.* (1998)

Serotonine receptor $5 - HT_{2A}$



• (R) - 2C - B - CB is biaised towards *PI* accumulation with respect to *AA* production, *compared to the reference agonist DOB*.



Urban et al., J Pharmacol Exp Ther (2007)

Serotonine receptor $5 - HT_{2A}$



- (R) 2C B CB is biaised towards *PI* accumulation with respect to *AA* production, *compared to the reference agonist DOB*.
- \Rightarrow Bias due to an *EC*₅₀ difference.



Urban et al., J Pharmacol Exp Ther (2007)

Many more examples on G Protein Coupled Receptor

Many GPCR's are known to have biased ligands

 G vs β-arrestin dependent signaling pathway



Kenakin, *Chem Rev* (2017)



Biased at more "integrated" response : Steroidogenesis modulated by NAM

Some negative allosteric modulators (NAM) can biased Progesterone production with respect to Testosterone production, under stimulation of LH/CG receptor by hCG.

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Ayoub et al., *Mol. Cell. Endocrinol* (2016)



Biased at more "integrated" response : Steroidogenesis modulated by NAM

Some negative allosteric modulators (NAM) can biased Progesterone production with respect to Testosterone production, under stimulation of LH/CG receptor by hCG.

⇒ Selective (biased) allosteric modulation



Ayoub et al., *Mol. Cell. Endocrinol* (2016)



Some examples

Bias quantification - standard method : operational model

Time-dependent bias

Biased quantification using dynamical model

Dose-response data are fitted with the function

$$y = E_{tot} \frac{\tau^n [L]^n}{([L] + Ka)^n + \tau^n [L]^n} \,.$$

- Response at equilibrium of a Michaelis-Menten type model.
- *Ka* = **Dissociation constant** of the couple Ligand/Receptor
- $\tau = \text{Efficacy coefficient}$ of the transduction pathway



Dose-response data are fitted with the function

$$y = E_{tot} \frac{\tau^n [L]^n}{([L] + Ka)^n + \tau^n [L]^n} \,.$$

For n = 1,

- $EC_{50} = \frac{\kappa_a}{\tau+1}$
- Efficacy $y_{\infty}/E_{tot} = \frac{\tau}{\tau+1}$

J. W. Black and P. Leff



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For n = 1,

- $EC_{50} = \frac{Ka}{\tau+1}$
- Efficacy $y_{\infty}/E_{tot} = \frac{\tau}{\tau+1}$

Then, we define

 \Rightarrow Transduction coefficient :

$$R := \log\left(\frac{\tau}{Ka}\right)$$

J. W. Black and P. Leff



Bias quantification : with the operational model



Two ligands (j = 1, 2) and **two** measured responses (i = 1, 2): Each dose-response data is fitted with the operational model :

$$y_{ij} = E_i \frac{\tau_{ij}^{n_i} [L]^{n_i}}{([L] + Ka_{ij})^{n_i} + \tau_{ij}^{n_i} [L]^{n_i}}.$$

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For a given response *i*, we calculate $\Delta_i \log(\tau/Ka) = \log(\tau_{i2}/Ka_{i2}) - \log(\tau_{i1}/Ka_{i1}).$

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For a given response *i*, we calculate $\Delta_i \log(\tau/Ka) = \log(\tau_{i2}/Ka_{i2}) - \log(\tau_{i1}/Ka_{i1}).$ The **Bias** is then defined by

$$\Delta\Delta\log(au/ extsf{Ka}) = \Delta_2\log(au/ extsf{Ka}) - \Delta_1\log(au/ extsf{Ka})$$

Statistical consideration : parameter confidence interval and (un-)identifiability



Some examples

Bias quantification - standard method : operational model

Time-dependent bias

Biased quantification using dynamical model

Time-dependent bias?

The role of kinetic context in apparent biased agonism at GPCRs

Carmen Klein Herenbrink¹, David A. Sykes², Prashant Donthamsetti^{3,4}, Meritxell Canals¹, Thomas Coudrat¹, Jeremy Shonberg⁵, Peter J. Scammells⁵, Ben Capuano⁵, Patrick M. Sexton¹, Steven J. Charlton², Jonathan A. Javitch^{3,4,6}, Arthur Christopoulos¹ & J Robert Lane¹

- Bias value may change according to the response time after stimulation.
- Kinetic explanation : Ligands with a slow binding kinetics may have changing bias value according to time.

Klein Herenbrink et al., *Nat. Commun* (2016)



Time-dependent bias?

The role of kinetic context in apparent biased agonism at GPCRs

Carmen Klein Herenbrink¹, David A. Sykes², Prashant Donthamsetti^{3,4}, Meritxell Canals¹, Thomas Coudrat¹, Jeremy Shonberg⁵, Peter J. Scammells⁵, Ben Capuano⁵, Patrick M. Sexton¹, Steven J. Charlton², Jonathan A. Javitch^{3,4,6}, Arthur Christopoulos¹ & J Robert Lane¹

- Bias value may change according to the response time after stimulation.
- Kinetic explanation : Ligands with a slow binding kinetics may have changing bias value according to time.
- ⇒ We need to take into account dynamic patterns in bias quantification



Physiological role of a kinetic profile (PTHR)



The mitogen-activated protein kinase ERK activation profile is the sum of two different activation pathways (G vs β -arrestin dependent), linked to two different spatial distribution of activated ERK (nucleus vs cytoplasm).

The balance of activation of the two pathways controls cell fate outcome.



Physiological role of a kinetic profile (PTHR)



The mitogen-activated protein kinase ERK activation profile is the sum of two different activation pathways (G vs β -arrestin dependent), linked to two different spatial distribution of activated ERK (nucleus vs cytoplasm).

The balance of activation of the two pathways is finely controlled by G protein coupled receptors kinases.



Physiological role of a kinetic profile (PTHR)

Short and Long acting Ligands activates the same secondary effector molecules (cAMP) trough different mechanisms, and lead to different physiological responses.



Vilardaga et al., Nat Chem Biol (2014)

Some examples

Bias quantification - standard method : operational model

Time-dependent bias

Biased quantification using dynamical model
• Ligand-specific modification of the energetic landscape controls signaling bias





Kenakin, J Pharmacol Exp Ther (2011)

- Ligand-specific modification of the energetic landscape controls signaling bias
- → However this information is barely accessible.





Kenakin, J Pharmacol Exp Ther (2011)

- Ligand-specific modification of the energetic landscape controls signaling bias
- But we have access to the kinetic profile of receptor downstream signaling





Poupon and Reiter, Cell. Endocrin. in Health and Disease. (2014)

- Ligand-specific modification of the energetic landscape controls signaling bias
- But we have access to the kinetic profile of receptor downstream signaling
- \rightarrow Dynamic modeling.





Poupon and Reiter, Cell. Endocrin. in Health and Disease. (2014)

Dynamic data (on FHSR in HEK cells)

Instead of focusing on dose-response curves, we deal with **kinetic data** performed at several doses (here : induced BRET data)



Dynamic data (on FHSR in HEK cells)

Instead of focusing on dose-response curves, we deal with **kinetic data** performed at several doses (here : induced BRET data)



I) We use chemical reaction network and ODE modeling (based on mass action law) to generate time series SHR SHR SHF Cell

I) We verify the network is able to accurately fit the data (one **separate fit** for each Ligand)



II) We fit **all data at once**, using some **common** parameters (initial concentration of molecules, measurement parameters...) and some **different** ones (kinetic parameters...)



II) We fit **all data at once**, using some **common** parameters (initial concentration of molecules, measurement parameters...) and some **different** ones (kinetic parameters...)



III) We use L^1 -penalization to find ligand specific parameters





Data2Dyanmics : Steiert, Timmer and Kreutz, *Bioinformatics* (2016)

III) We use L^1 -penalization to find **ligand specific parameters**, keeping the fit 'as good as before'



Steiert, Timmer and Kreutz, Bioinformatics (2016)

IV) After re-optimization, the set of distinct (ligand-specific) kinetic parameters gives us an accurate description of ligand specificity.



V) Significant differences between parameters is assessed by statistical methods (Profile Likelihood)



comparison to FSH

Practical problems...



Practical problems...





We obtain a slightly worse fit





And "better" parameter identifiability



C3 is biased towards β -arr, compared to cAMP, in comparison to FSH.



- Notion of signaling bias to quantify differential activation of several pathways by a Ligand at a given receptor.
- Standard quantification has several drawbacks (no time, limited to sigmoid scenario, et).
- We gave a kinetic interpretation of Ligand biased, which rely on kinetic data and dynamic (ODE) modeling, with numerical parameter estimation and L¹ penalization to reduce combinatorial complexity.

- Notion of signaling bias to quantify differential activation of several pathways by a Ligand at a given receptor.
- Standard quantification has several drawbacks (no time, limited to sigmoid scenario, et).
- We gave a kinetic interpretation of Ligand biased, which rely on kinetic data and dynamic (ODE) modeling, with numerical parameter estimation and L¹ penalization to reduce combinatorial complexity.
- \Rightarrow How to deal with "fuzzy/noisy" PLE?
- \Rightarrow How to deal with non uniqueness of optimal parameters?
- ⇒ How to perform a model reduction that would lead to both a satisfactory fit and identifiable parameters?

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M. Ayoub et al., Molecular and Cellular Endocrinology 436 (2016)

L. Riccetti et al., Scientific Reports 7 :940 (2017)

R.Y. et al., Methods in Molecular Biology, in press (2018)



We systematically calculate bias value using standard method (operational model on dose-response curves :)



Bias=2.3 : C3 is biased towards β -arr, compared to cAMP, in comparison to FSH.

We systematically calculate bias value using standard method (operational model on dose-response curves :)



Bias=2.64 : C3 is biased towards β -arr, compared to cAMP, in comparison to FSH.

We systematically calculate bias value using standard method Different times gives (slightly) different bias values



C3 is biased towards β -arr, compared to cAMP, in comparison to FSH

We systematically calculate bias value using standard method Uncertainty can be large according to the time of measurement







Original "raw" data



"Adjusted" data



"Adjusted" data



Dose-dependent bias



Barak and Peterson et al., *Biochem.* (2012)

Extension of the operational model



Kenakin, *Chem. Rev.* (2017) Method based on Intrinsic activities and rank ordering



Onaran et al., *Sci. Rep.* (2017)



Is bias calculation intuitive? (simulated data)



A strong bias is usually 'apparent' on dose-response curves or bias plot

Is bias calculation intuitive? (simulated data)



But there may be counter-intuitive situation...
Is bias calculation intuitive? (simulated data)



But there may be counter-intuitive situation...

Is bias calculation intuitive? (simulated data)



But there may be counter-intuitive situation...

... and those situations occur in real life!

