

Modeling Ovarian Folliculogenesis: Morphogenesis and Population Dynamics

Celine Bonnet, Keltoum Chahour, Frederique Clement, Marie Postel, Frédérique Robin, Romain Yvinec

▶ To cite this version:

Celine Bonnet, Keltoum Chahour, Frederique Clement, Marie Postel, Frédérique Robin, et al.. Modeling Ovarian Folliculogenesis: Morphogenesis and Population Dynamics. REPROSCIENCES 2019, Apr 2019, Toulouse, France. hal-03115058

HAL Id: hal-03115058

https://hal.science/hal-03115058

Submitted on 19 Jan 2021

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



MODELING OVARIAN FOLLICULOGENESIS: MORPHOGENESIS AND POPULATION DYNAMICS



C. Bonnet ¹, K. Chahour ², F. Clément ³, M. Postel ⁴, F. Robin ³ and <u>R. Yvinec</u> ⁵ Ecole Polytechnique ² Université Côte d'Azur ³ Inria Paris-Saclay ⁴ Sorbonne Université ⁵ INRA Tours

INTRODUCTION

We present stochastic population dynamic models applied to several situations to understand ovarian folliculogenesis

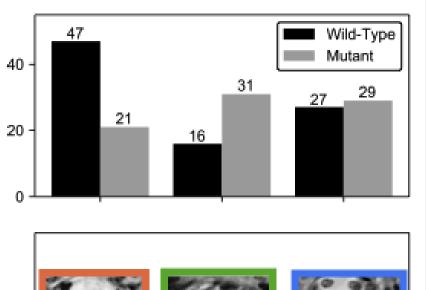
- Somatic cell population dynamics in a single follicle (either in the activating or basal growing phase)
- Whole follicle population dynamics during the reproductive lifespan

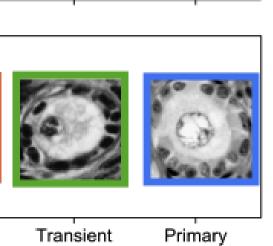
CELL DYNAMICS IN AN ACTIVATING FOLLICLE

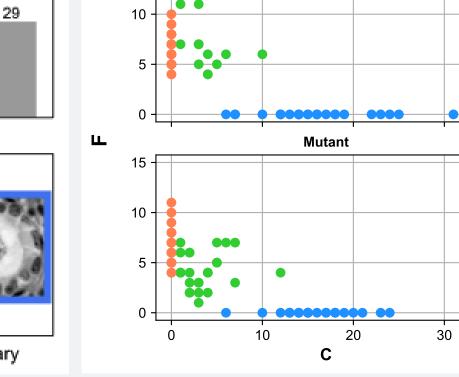
Somatic cell transition and proliferation during follicle activation

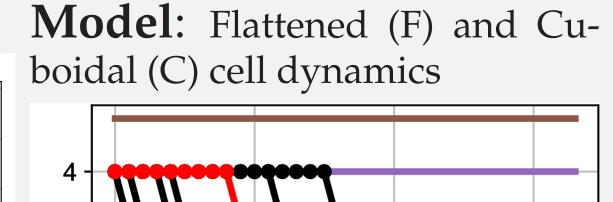
Key questions: What are the kinetics of follicle activation? Are transition and proliferation concomitant?

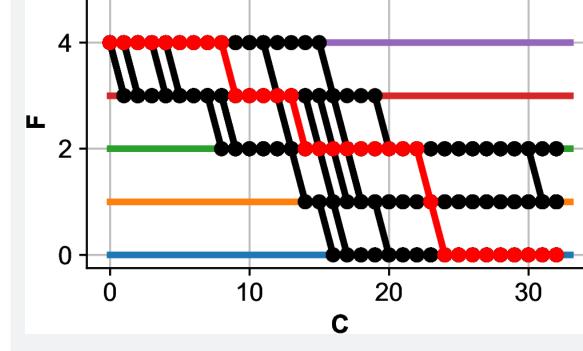
Data: Cell counts (Lundy et al., J. Reprod. Fertil., 115 (1999); Wilson et al. Biol Reprod 64 (2011))



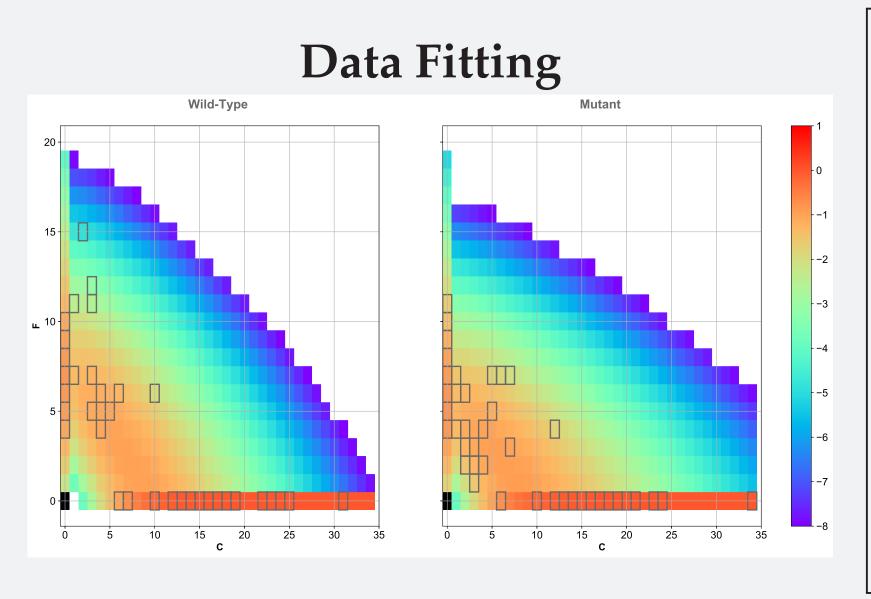








Cell events	Rate
$(F,C) \to (F-1,C+1)$	αF
$(F,C) \to (F-1,C+1)$	$\beta \frac{FC}{F+C}$
$(F,C) \to (F,C+1)$	γC
$(F,C) \to (F,C+1)$	$ \delta F $



Results: see [3]

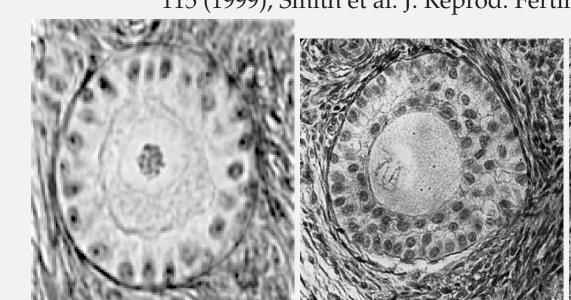
- Clear separation between transition and proliferation in Wildtype.
- Auto-amplification is not mandatory, especially for the Mutant dataset

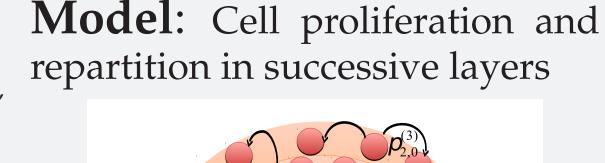
CELL DYNAMICS IN A GROWING FOLLICLE

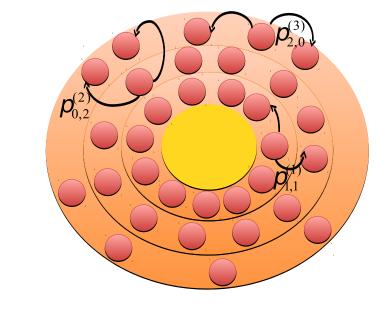
Somatic cell proliferation during basal follicle growth

Key questions: What is the rate of growth? Is proliferation oocyte-dependent?

Data: Total cell numbers, Oocyte and Follicle diameters at three time points (Lundy et al., J. Reprod. Fertil., 115 (1999); Smith et al. J. Reprod. Fertil., 100 (1994))

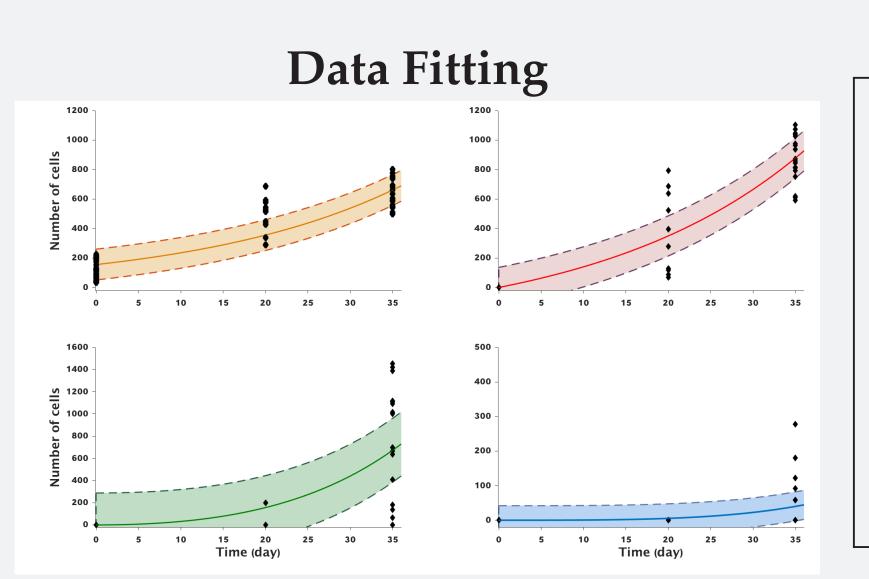






Stochastic Linear Model	
Both daughter cells stay on mother's layer	
One daughter cell into next mother's layer	
Both daughter cells into next mother's layer	

Cell events	Rate
$N^j o 2N^j$	$p_{2,0}^j b_j N^j$
$N^j \rightarrow N^j + N^{j+1}$	$p_{1,1}^{j^{\prime}}b_{j}N^{j}$
$N^j o 2N^{j+1}$	$p_{0,2}^{j}b_{j}N^{j}$



Results: see [2] Data are compatible with

- exponential an growth phase.
 - layer-dependent growth rate is decreasing with oocyte distance

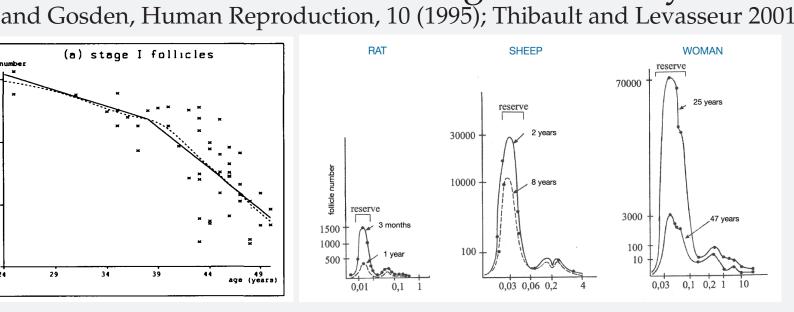
WHOLE FOLLICLE POPULATION DYNAMICS

Nonlinear interactions between follicular populations

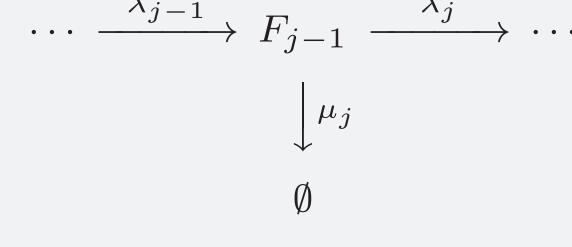
Key questions: How a quasi-stable maturity repartition is achieved? What is the role of nonlinear interactions?

Data: Follicle counts according to maturity class
Faddy and Gosden, Human Reproduction, 10 (1995); Thibault and Levasseur 2001

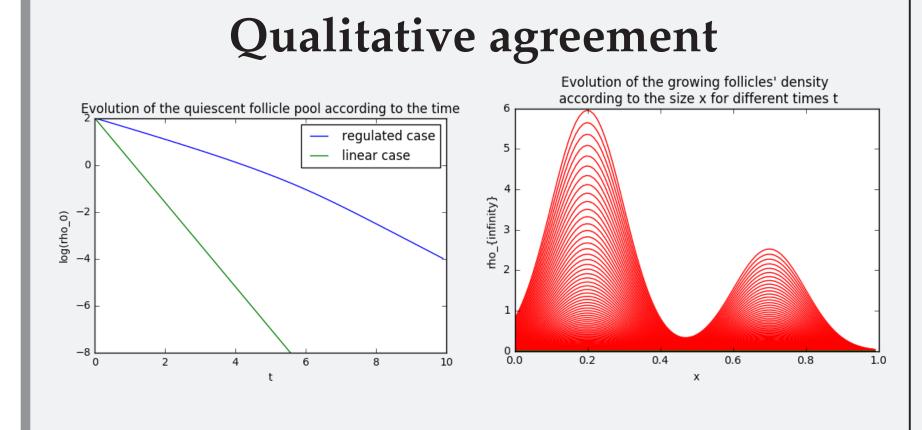
Model: Population-dependent follicle maturity and atrosia



follicle maturity and atresia



Stochastic Nonlinear Model	Follicle events	Rate
Follicle activation	$F_0 \to F_1$	$\varepsilon \frac{\lambda_0}{1+K\sum_{j=1}^d a_j F_j} F_0$
Follicle maturation Follicle atresia	$F_j \to F_{j+1}$ $F^j \to \emptyset$	$\lambda_j F_j$ $\mu_j F_j$
Tomcie attesia		$\mu_{\mathcal{I}}$



Results: see [1]

- Time-scale separation explains quasi-stable maturity repartition.
- Removal of actiinhibition vation explains acceleration of reserve exhaustion

MATHEMATICAL TOOLBOX

- (Structured) Population dynamics model: deterministic and stochastic models.
- Long-time and asymptotic analysis, time-scale separation.
- Transient analysis: analytical and numerical solutions, first passage time.
- Statistical methods: parameter estimation and identifiability analysis.

CONCLUSION AND PERSPECTIVE

Mathematical modeling helps:

- to test different follicle growth scenario and make prediction
- ♦ to challenge biological knowledge on follicle dynamics.

Current study and future projects will focus on different scales:

- * Intra-cellular scale: FSHR signaling network and its role in follicle selection (for a review of gonadotropin signaling models, see [5]).
- * Follicle scale: coupling cell dynamics with biomechanics model to model antrum formation (extending and revisiting previous model [4]).
- * Ovarian scale: Complement follicle population dynamics with (i) ovarian reserve formation and (ii) ovarian cycle dynamics. Comparative physiology approaches will also be taken into account.

REFERENCES

- [1] C. Bonnet, K. Chahour, F. Clément, M. Postel, and R. Yvinec. Multiscale population dynamics in reproductive biology: Singular perturbation reduction in deterministic and stochastic models. Arxiv preprint: 1903.08555v1, Mar. 2019.
- [2] F. Clément, F. Robin, and R. Yvinec. Analysis and Calibration of a Linear Model for Structured Cell Populations with Unidirectional Motion: Application to the Morphogenesis of Ovarian Follicles. SIAM Journal on Applied Mathematics, 79(1):207–229, Jan. 2019.
- [3] F. Clément, F. Robin, and R. Yvinec. Stochastic nonlinear model for somatic cell population dynamics during ovarian follicle activation. Arxiv preprint: 1903.01316v1, Mar. 2019.
- [4] F. Clément, P. Michel, D. Monniaux, and T. Stiehl. Coupled somatic cell kinetics and germ cell growth: multiscale model-based insight on ovarian follicular development. Multiscale Modeling & Simulation, 11(3):719–746, 2013.
- [5] R. Yvinec, P. Crépieux, E. Reiter, A. Poupon, and F. Clément. Advances in computational modeling approaches of pituitary gonadotropin signaling. Expert Opinion on Drug Discovery, 13(9):799–813, Sept. 2018.

Acknowledgement: We thank Ken McNatty for providing experimental datasets and Danielle Monniaux for helpful discussion (and histological images!).